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## Paediatric Respiratory Reviews

Clinical usefulness

# Re-opening the pediatric pulmonary function laboratory during the ongoing COVID-19 pandemic

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#### **Educational** Aims

The reader will come to appreciate that:

- risk will vary with local community viral activity and individual patient.
- local testing environment must be assessed for aerosol exposure risk.
- different pulmonary function tests require different levels of protection.

#### ARTICLE INFO

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#### ABSTRACT

The COVID-19 pandemic continues with new waves of intensification. This review provides an update based on international recommendations concerning the conduct of pulmonary function testing in a manner to limit risk to both patient and tester.

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# BACKGROUND

In the November 2020 issue of Paediatric Respiratory Reviews [1], an international perspective on pediatric pulmonary function testing during the COVID-19 pandemic was published. Since that time, several national and international societies have published guidelines around testing [2–4]. The pandemic has waxed and waned and resurged around the world, reflecting local public health recommendations on mask wearing, social distancing, vaccination, travel, access to effective vaccination, and viral mutations. As such, the SARS-CoV-2 virus is becoming endemic. This article will address current recommendations during persistence of COVID-19 within our communities as we emerge from lockdowns.

During the lockdowns that many of us experienced, the marked decrease in non-SARS-CoV-2 respiratory viral infections, such as influenza and, for a while, respiratory syncytial virus (RSV) high-lighted how successful mask wearing, handwashing, and social distancing could be [5,6]. We all saw a dramatic decrease in exacerbations of chronic lung conditions, such as asthma, Cystic Fibrosis and Primary Ciliary Dyskinesia [7,8], underscoring the role of respiratory viruses in pulmonary exacerbations. With loosening

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of restrictions, there has been a rebound in viral infections, particularly in young children not previously exposed to viruses. We are now much more cognizant of the consequences of respiratory virus exposure and that viruses can travel and be contagious through both droplet and aerosol spread [9,10].

Depending on local COVID-19 activity, pulmonary function laboratories may be closed, partially open, or fully open. A Stringency Index to assess local viral activity has been developed by the WHO [11].

#### PREPARING FOR PULMONARY FUNCTION TESTING

#### Who should undergo pulmonary function testing?

Many people, particularly children, infected with SARS-CoV-2 are asymptomatic, so that all individuals coming for testing should be considered as potentially contagious. Even more than during the pre-pandemic era, it is important to decide on whether a test is really needed. On the practical side in considering the number of tests that can be conducted on a given day, contamination precautions result in the need for more time between individuals to be tested, so fewer tests can be performed.







#### Hierarchy of tests

For the purpose of assigning risk and therefore level of protection required, pulmonary function tests have often been segmented into those with a higher risk of aerosol generation and those with a lower risk. More recently, it has been recognized that regular talking can generate aerosols and that the exhortations of the tester can certainly generate aerosols, often more than generated by the test itself [12–16]. Cough is a significant aerosolgenerating activity, and 50% of people will cough during or after standard pulmonary function testing [17]. Therefore, the majority of standard pulmonary function tests should be considered as atrisk procedures for exposure of the personnel. Bronchoprovocation testing, particularly with nebulized methacholine, and exercise testing require extra precautions. Personal protective equipment (PPE) requirements are discussed below.

#### Screening of patients

The goal of screening is risk assessment. Most laboratories question people in the 72 hours prior to them coming to the appointment and the day of their appointment. People are often not forthcoming about the presence of symptoms, so that questioning has limited reliability when negative. Some laboratories require negative COVID-19 testing prior to coming to the laboratory, or at least testing those suspected of having COVID-19. Routine testing of asymptomatic or low risk patients (e.g. vaccinated, low community viral prevalence) consumes a lot of health care resources and inconveniences the patient and family. Pulse oximetry and temperature screening may be warranted in some settings [18]. If the patient is suspected to have COVID-19, in the absence of testing, then testing should be delayed until there is resolution of symptoms and a minimum of three weeks have passed since the onset of symptoms [19,20]. Patients with chronic respiratory disease who are acutely ill can have symptoms mimicking COVID-19 and should not be tested until back to their usual status. Patients above the age of 5 years and caregivers should be masked. If possible, only the patient should be in the testing room.

#### Waiting room

The waiting room should be configured ideally for physical spacing. If possible, a minimum of 1- meter, and preferably 2-meter distance should be kept [21]. Hand sanitizer should be readily available. Wait times should be minimized.

#### Personal protective equipment (PPE)

PPE is an important component of the overall approach to preventing and controlling infection. The recommended facemask is the fit-tested N95, in Europe known as the FFP2, which filters out 95% of airborne particles. In the context where both patient and tester have been vaccinated and community prevalence is low, then a gown, gloves, face shield and surgical mask is sufficient to protect both the patient and tester [22]. A fit-tested mask (at a minimum N95, with some recommending N99) should be worn for those activities with high risk of aerosol generation (wet nebulization, bronchoprovocation challenge, exercise testing).

#### The testing room

Ventilation is key. The ideal setup is a negative pressure room with a high number (at least 12) of air exchanges per hour. The air exchange rate is the number of room volumes provided by the ventilation system every hour, not the number of times per hour that all the air in the room is changed. A minimum number of 6 air exchanges per hour is recommended for the standard hospital room. With such ventilation, 95% of contaminants are removed in 30 min [23]. Note that with 12 air exchanges per hour. it takes 15 min to remove 95% of the contaminants. Central ventilation systems should be configured in order to avoid returning some of the filtered air back into the room. All ventilation systems should be verified on a recurring basis. If an enhanced number of air exchanges cannot be provided, the use of high efficiency particulate air (HEPA) room filtration should be considered, whether built in, or as portable systems. Ultraviolet systems are an alternative to HEPA filtration. When there is a central system or HEPA system, windows should be sealed. When there is inadequate ventilation, opening windows will contribute to air exchange but raises the risk of contamination from the outside. Further such inconsistent aeration will cause temperature and humidity fluctuations that can interfere with the performance and accuracy of the testing equipment. Many institutions have placed plexi-glass (perspex) dividers between the patient and tester. This will prevent droplet spread, but not aerosol spread.

#### The equipment

Disposable bacterial/viral filters between the mouthpiece and the equipment should be employed unless they adversely affect test results and changed between patients [24]. The equipment and surfaces should be wiped down between patients according to local practice.

#### CONDUCTING PULMONARY FUNCTION TESTING

#### Spirometry pre- and post-bronchodilators

Spirometry will generate both aerosols and frequently droplets. The device needs a high efficiency in-line filter. The equipment exposed to the patient may be covered with disposable protective covers or wiped down after each patient. A plexi-glass barrier can be added to reduce droplet exposure of the tester. The room should be highly ventilated, and the tester wearing gloves, face-shield/ goggles, and gown. The preferred mask is an N95, although if risk

#### Table 1

Pulmonary Function Test Filter and PPE Requirements.

Test	Filter	Mask	Gown + Faceshield/ Goggles
Spirometry	Yes	N95	Yes
Bronchodilator (MDI/Dry powder)	No	N95	Yes
Bronchodilator	Yes	N95+	Yes
(Wet nebulized)			
Filter on expiratory port			
Bronchoprovocation Testing	Yes	N95+	Yes
Filter on expiratory port for wet			
nebulized			
Negative pressure room			
recommended			
Lung volumes	Yes	N95	Yes
Carbon Monoxide Diffusion	Yes	N95	Yes
Oscillometry	Yes	N95	Yes
Multiple Breath Washout	Yes	N95	Yes
Exhaled Nitric Oxide	Yes	N95	Yes
Use in-circuit filter			
Nasal Nitric Oxide	Yes	N95	No
Use in-circuit filter			
Respiratory muscle strength	Yes	N95	Yes
Mouth			
Respiratory muscle strength	No	N95	Yes
Sniff	NI-	NOF	V
Cardiopulmonary Exercise Testing	No	N95+	Yes

Legend: Derived from McGowan et al. (3). N95+: some recommend N99.

is considered low, a surgical mask can be used. The patient should remain on the mouthpiece for 2–3 tidal breaths following the manoeuvre. When the patient comes off the mouthpiece they should replace their facemask. If delivering a bronchodilator, then it is preferred to use a metered dose inhaler with a valved spacing device. A dry powder device can also be used. Note that a single deep breath hold followed by exhalation increases the amount of exhaled aerosol particles from the distal airway be a factor of 4 [25]. If wet nebulization is employed, it must be recognized that nebulization can result in environmental aerosol exposure, and SARS-CoV-2 is viable for at least 3 hours following wet nebulization [19]. For wet nebulization a N99 (FFP3) rather than an N95 (or if low risk, surgical facemask) (plus gloves, gown, and faceshield/goggles) is recommended for the tester Table 1.

#### Bronchoprovocation testing

Bronchoprovocation testing can provoke coughing and vigilant patient screening is recommended. Note that cough is particularly frequent in the case of using dry powder mannitol [26]. Using wet nebulization of methacholine can result in infected reservoirs in the case of wet nebulizers. Using a breath-activated nebulizer will reduce this is risk. A vibrating mesh nebulizer will avoid the reservoir coming into contact with infected particles. Even with the use of a breath-activated or vibrating mesh nebulizer, a high-quality filter should be placed on the expiratory limb. Bronchoprovocation testing is a high-risk procedure for generating aerosols. The room should have high air exchanges, or a ventilation system that does not recirculate air, a HEPA or ultraviolet cleaning capacity. For such testing it is recommended that the tester wear an N99 mask (plus gloves, gown, and face-shield/goggles). If possible, such testing should occur in a negative pressure setting, or otherwise adequate time be given for air exchanges as would be done for induced sputum collection for tuberculosis.

#### Lung volume measurement

If a gas washout/dilution technique is used a bacterial/viral filter needs to be in place and the patient should take 2–3 tidal breaths after a vital capacity measurement before coming off the mouthpiece. If using a whole body plethysmograph, the interior needs to be properly cleaned after each patient, with the cleaner wearing PPE. It is recommended that the tester wear an N95 mask (plus gloves, gown, and face-shield/goggles).

#### Diffusing capacity

An inline filter is required and the patient should take 2–3 tidal breaths after a vital capacity measurement before coming off the mouthpiece. It is recommended that the tester wear an N95 mask (plus gloves, gown, and face-shield/goggles).

#### Oscillometry

This test is done with the patient quietly breathing. A high efficiency inline filter is recommended.

#### Multiple breath washout

This test is done with the patient quietly breathing. A high efficiency inline filter is recommended.

#### Exhaled nitric oxide

An inline filter should be used. It is recommended that the tester wear an N95 mask (plus gloves, gown, and face-shield/goggles).

#### Nasal nitric oxide

An in-circuit filter is recommended. If this is not possible, then the circuit should be discarded after each test. For the tester, if a filter is not possible, physical distancing and minimizing exposure time is recommended.

#### Respiratory muscle strength testing

An inline filter is recommended for measurements made through a mouthpiece, but not for sniff testing. Coughing may be provoked. It is recommended that the tester wear an N95 mask (plus gloves, gown, and face-shield/goggles).

#### Cardiopulmonary exercise testing

Exercise testing requires the patient to breathe deeply, forcibly and frequently with significant risk of aerosol and droplet production and prolonged viral shedding. Patient screening is particularly important in this situation. Filters cause resistance particularly as accumulate water vapour and become saturated. This may impact breathing patterns and exercise performance [27] and so are not recommended. Single-use masks, sensors, turbines and gas lines are recommended. A ventilation system without recirculation is recommended. If this is not possible, then it should have a HEPA or ultraviolet cleaning capacity.

#### DIRECTIONS FOR FUTURE RESEARCH

- Pulmonary function testing in the office setting.
- Further clarification is needed of the risk/benefit and cost differential of portable air filtration systems [e.g. Hepafilters] vs implementing the recommended 12 or more cycles of air recirculation per hour in pulmonary function laboratories.

#### **CONFLICTS OF INTEREST**

Nil to declare.

#### References

- Beydon N, Gochicoa L, Jones MJ, Lands LC, Lombardi E, Rosenfeld M, et al. Pediatric lung function testing during a pandemic: An international perspective. Paediatr Respir Rev 2020;36:106–8.
- [2] Fouzas S, Gidaris D, Karantaglis N, Opsimos H, Alexopoulos EI, Douros K, et al. Pediatric pulmonary function testing in COVID-19 pandemic and beyond. A position statement from the Hellenic Pediatric Respiratory Society. Front Pediatr. 2021;9:673322.
- [3] McGowan A, Laveneziana P, Bayat S, Beydon N, Boros PW, Burgos F, et al. International consensus on lung function testing during COVID-19 pandemic and beyond. ERJ Open Res. 2021:00602-2021.
- [4] Canadian Thoracic Society CSoRT. Resumption of Pulmonary Function Testing during the Covid-19 Pandemic version 3.0 [website]. 2021 [Available from: https://cts-sct.ca/wp-content/uploads/2021/11/CTS\_CSRT\_PFT\_Covid-19\_ update3\_IN-PRESS.pdf.
- [5] Redlberger-Fritz M, Kundi M, Aberle SW, Puchhammer-Stöckl E. Significant impact of nationwide SARS-CoV-2 lockdown measures on the circulation of other respiratory virus infections in Austria. J Clin Virol 2021;137:104795.
- [6] Ippolito G, La Vecchia A, Umbrello G, Di Pietro G, Bono P, Scalia S, et al. Disappearance of seasonal respiratory viruses in children under two years old during COVID-19 pandemic: A monocentric retrospective study in Milan. Italy Front Pediatr 2021;9:721005.
- [7] Patel S, Thompson MD, Slaven JE, Sanders DB, Ren CL. Reduction of pulmonary exacerbations in young children with cystic fibrosis during the COVID-19 pandemic. Pediatr Pulmonol 2021;56(5):1271–3.
- [8] Golan-Tripto I, Arwas N, Maimon MS, Bari R, Aviram M, Gatt D, et al. The effect of the COVID-19 lockdown on children with asthma-related symptoms: A tertiary care center experience. Pediatr Pulmonol 2021;56(9):2825–32.
- [9] Wang CC, Prather KA, Sznitman J, Jimenez JL, Lakdawala SS, Tufekci Z, et al. Airborne transmission of respiratory viruses. Science 2021;373(6558).
- [10] Greenhalgh T, Jimenez JL, Prather KA, Tufekci Z, Fisman D, Schooley R. Ten scientific reasons in support of airborne transmission of SARS-CoV-2. Lancet 2021;397(10285):1603–5.

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- [11] World Health Organization. Criteria for releasing COVID-19 patients from isolation [website]. 2020 [Available from: https://www.who.int/newsroom/commentaries/detail/criteria-for-releasing-covid-19-patients-fromisolation.
- [12] Helgeson SA, Lim KG, Lee AS, Niven AS, Patel NM. Aerosol generation during spirometry. Ann Am Thorac Soc 2020;17(12):1637–9.
- [13] Wilson NM, Marks GB, Eckhardt A, Clarke AM, Young FP, Garden FL, et al. The effect of respiratory activity, non-invasive respiratory support and facemasks on aerosol generation and its relevance to COVID-19. Anaesthesia. 2021;76 (11):1465-74.
- [14] Wu JKY, Ryan CM, Hiebert RJ, Han Z, Liu A, Jeong C-H, et al. Aerosol generation during pulmonary function testing: Monitoring during different testing modalities. Can J Respir Crit Care Sleep Med. 2021:1-8.
- [15] Sheikh S, Hamilton FW, Nava GW, Gregson FKA, Arnold DT, Riley C, et al. Are aerosols generated during lung function testing in patients and healthy volunteers? Results from the AERATOR study. Thorax. 2021.
- [16] Greening NJ, Larsson P, Ljungström E, Siddiqui S, Olin A-C. Small droplet emission in exhaled breath during different breathing manoeuvres: Implications for clinical lung function testing during COVID-19. Allergy 2021;76(3):915–7.
- [17] Kimberley L, Swan J, Perera M, Swaffield B, Cranstone K, Wilson N, et al. P36 Cough provoked by lung function testing – should lung function testing be treated as an aerosol generating procedure post COVID-19? Thorax 2021;76 (Suppl 1):A104–5.
- [18] Jouffroy R, Jost D, Prunet B. Prehospital pulse oximetry: a red flag for early detection of silent hypoxemia in COVID-19 patients. Crit Care 2020;24(1):313.
- [19] van Doremalen N, Bushmaker T, Morris DH, Holbrook MG, Gamble A, Williamson BN, et al. Aerosol and surface stability of SARS-CoV-2 as compared with SARS-CoV-1. N Engl J Med 2020;382(16):1564–7.

- [20] Bourouiba L. Turbulent gas clouds and respiratory pathogen emissions: potential implications for reducing transmission of COVID-19. JAMA 2020;323(18):1837–8.
- [21] Crimi C, Impellizzeri P, Campisi R, Spicuzza L, Vancheri C, Crimi N. Resumption of respiratory outpatient services in the COVID-19 era: Experience from Southern Italy. Am J Infect Control 2020;48(9):1087–9.
- [22] Sterr CM, Nickel I-L, Stranzinger C, Nonnenmacher-Winter CI, Günther F, Mukherjee A. Medical face masks offer self-protection against aerosols: An evaluation using a practical in vitro approach on a dummy head. PLoS One 2021;16(3):e0248099.
- [23] Centers for Disease Control and Prevention. Guidelines for environmental; infection control in health-care facilities [Website]. 2003 [Available from: https://www.cdc.gov/infectioncontrol/guidelines/environmental/appendix/ air.html#b1.
- [24] Stanojevic S, Beaucage F, Comondore V, Faughnan M, Kovesi T, McCoy C, et al. Resumption of pulmonary function testing during the post-peak phase of the COVID-19 pandemic. Can J Respir Crit Care Sleep Med 2020;4(3):156–9.
- [25] Larsson P, Bake B, Wallin A, Hammar O, Almstrand A-C, Lärstad M, et al. The effect of exhalation flow on endogenous particle emission and phospholipid composition. Respir Physiol Neurobiol 2017;243:39–46.
- [26] Brannan JD, Anderson SD, Perry CP, Freed-Martens R, Lassig AR, Charlton B. The safety and efficacy of inhaled dry powder mannitol as a bronchial provocation test for airway hyperresponsiveness: a phase 3 comparison study with hypertonic (4.5%) saline. Respir Res 2005;6(1):144.
- [27] Marques MS, Fonseca A, Lima R, Ladeira I, Gomes J, Guimarães M. Effect of a viral filter on cardiopulmonary exercise testing. Pulmonology 2021.