

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active. Contents lists available at ScienceDirect



Review

Veterinary Parasitology: Regional Studies and Reports

journal homepage: www.elsevier.com/locate/vprsr



Neospora caninum, A potential cause of reproductive failure in dairy cows from Northern Greece



M. Lefkaditis*, R. Mpairamoglou, A. Sossidou, K. Spanoudis, M. Tsakiroglou

Veterinary Faculty, School of Health Sciences, University of Thessaly, Greece

ARTICLE INFO	A B S T R A C T	
<i>Keywords:</i> Neospora caninum Abortion Infertility Cow	Neospora caninum infection has been reported in a large number of intermediate hosts, such as ruminants, rabbits, mice, etc. but neosporosis has emerged as a serious disease in cattle and dogs worldwide. Abortions and other infertility issues have been reported in the infected cows, leading to great economic losses in farmers. The aim of our study was to assess <i>N. caninum</i> seroprevalence in dairy cattle from Northern Greece (region of Xanthi) by using the indirect fluorescent antibody technique. Blood samples were collected from 875 Holstein – Friesian dairy cows and tested for <i>Neospora caninum</i> antibodies. Among the cows that were studied, 184 (21.03%) were positive for <i>N. caninum</i> antibodies and concurrently their farms had a known previous history of infertility problems, such as abortions, increased number of artificial inseminations needed for conception, increased rate of returning to estrus and retention of fetal membranes.	

1. Introduction

Neospora caninum (Apicomplexa: Coccidia) is an obligate intracellular parasite that is the etiologic agent of the polysystemic disease neosporosis (Donahoe et al., 2015; Dubey et al., 2013). This apicomplexan parasite was initially recognized in 1984 in dogs in Norway.

(Bjerkas et al., 1984) and described as a new genus and species in 1988 (Dubey et al., 1988). In the *N. caninum* life cycle, dogs and other related canids are the only definitive hosts that shed through their feces the unsporulated oocysts into the environment, beside their role of intermediate host (Dubey and Schares, 2011; King et al., 2010; Gondim et al., 2004; Dubey et al., 2002; Basso et al., 2001; Lindsay et al., 2001; Lindsay et al., 2002; Basso et al., 2001; Lindsay et al., 2001; Lindsay et al., 2002; Basso et al., 2001; Lindsay et al., 2001; Lindsay et al., 2002; Dijkstra et al., 2001; Schares et al., 2001; Lindsay et al., 2001; Schares et al., 2001; Lindsay et al., 1999a; Lindsay et al., 1999b; McAllister et al., 1998). Thus, dogs play an important role in the horizontal transmission and maintenance of *N. caninum* infection in dairy cattle (Dubey and Schares, 2011; King et al., 2010; Gondim et al., 2004; McAllister et al., 1998).

N. caninum has been reported in a large number of intermediate hosts, such as ruminants, rabbits, mice, etc. (Dubey et al., 2007), but neosporosis has emerged as a serious disease in cattle and dogs worldwide (Dubey and Schares, 2011; Dubey et al., 2007). While this

disease has a considerable impact on reproduction in cattle, in adult and older dogs appears to be asymptomatic (Silva and Machado, 2016; Kul et al., 2015; Lindsay et al., 1999a). It has been shown that 12–42% of the aborted bovine fetuses worldwide are infected with *N. caninum* (Piagentini et al., 2012; Xu et al., 2012; Dubey et al., 2007; Hall et al., 2005; Jenkins et al., 2002).

N. caninum causes abortions in both dairy and beef cattle. The abortions can occur starting with month three of gestation until delivery (Dubey et al., 2013; Reiterová et al., 2009; Dubey et al., 2007) in an epidemic or endemic manner (Wouda et al., 1999). *N. caninum* can also cause fetal viability disorders or neurological birth defects in newborn calves (Lassen et al., 2012; Malaguti et al., 2012) and those younger than 2 months of age (Dubey, 2003). The *N. caninum*-infected young calves may present neurologic signs, low birth weight (Dubey and Schares, 2011), difficulties to rise and stand, flexed or hyper-extended hind and/or forelimbs, and in some cases exophthalmia or asymmetrical appearance of the eyes. However, most of the calves born congenitally-infected remain clinically healthy animals (Dubey, 2003; Bielanski et al., 2002).

Natural *N. caninum* infections can occur via horizontal (lateral) or transplacental (vertical, congenital) transmission (Dubey et al., 2007). In cattle and other domesticated bovine species, the transplacental transmission is the most frequent route of infection, being observed in up to 93.7% of cases (Dubey et al., 2007; Schares et al., 1998). In the definitive canid hosts, the horizontal transmission through ingestion of

https://doi.org/10.1016/j.vprsr.2019.100365

Received 9 June 2019; Received in revised form 11 November 2019; Accepted 6 December 2019 Available online 09 December 2019

2405-9390/ © 2019 Published by Elsevier B.V.

^{*} Corresponding author. *E-mail addresses:* mlefkaditis@vet.uth.gr (M. Lefkaditis), kyspanou@vet.uth.gr (K. Spanoudis).

tissues infected with tachyzoites, tissue cysts or food and water contaminated with sporulated oocyst is the predominant infection route (Donahoe et al., 2015; Dubey et al., 2007). The lactogenic transmission of *N. caninum* has been demonstrated experimentally in newborn calves fed with colostrum infected with tachyzoites, but there is an ongoing debate regarding whether or not this occurs naturally (Davison et al., 2001). It has been shown that dogs fed with milk infected with *N. caninum* tachyzoites do not shed oocysts (Dijkstra et al., 2001).

Neosporosis is recognized as one of the most important cause of reproductive issues and abortion in cattle worldwide (Reichel et al., 2013; Dubey et al., 2007; Haddad et al., 2005). The abortions and neonatal mortality can cause severe financial loss, especially when the disease is endemic or epidemic. The economic impact is directly related with the costs associated with abortion and indirectly with the cost of veterinary services, rebreeding, loss of milk yield and replacement if cows that aborted are culled (Ansari-Lari et al., 2017).

Knowledge of the infected and non-infected cows in a region would increase our understanding of the economic impact due to *N. caninum* infection and would help us eradicate the disease.

The aim of this study was to assess *N. caninum* seroprevalence in dairy cattle from Northern Greece (region of Xanthi) by using the indirect fluorescent antibody technique (IFAT).

2. Materials and methods

2.1. Cattle and herd management

This was a prospective study conducted between March 2016 and May 2018 in 5 Holstein–Friesian dairy farms located in the prefecture of Xanthi (Northern Greece). All farms reported low fertility rates and high rates of miscarriage and provided us with the reproductive history of their cows. A number of 875 Holstein–Friesian dairy cows (mean age 4.28 years) were included in the study. The herds were kept in free-stall housing and were divided according to the stage of reproduction cycle and milk production. All cows received a balanced feed ratio and were vaccinated against infectious bovine rhino tracheitis (IBR), bovine viral diarrhea (BVD), Coronavirus, Rotavirus and *Escherichia coli*, according to the national vaccination program. There was no seasonal breeding management in the herd, but artificial insemination was performed throughout the year.

2.2. Indirect fluorescent antibody technique

Blood samples were collected from the jugular vein of each cow at the time of early pregnancy detection through rectal examination (around 28 days after insemination in heifers and around 34 days in multiparous). Within 30 min after collection, the samples were centrifugated at 2000 rpm for 10 min and the serum was separated and stored in labelled testing tubes at -4 °C, pending examination for the presence of *N. caninum* antibodies.

Many serological tests are available for the diagnosis of bovine *N.caninum* infection (Wapenaar et al., 2007; Dubey and Schares, 2006).

We used IFAT, based on the protocol described by Dubey et al., 1988. This technique is commonly used for diagnosis of protozoan infections and is based on the labeling of antibodies with a fluorescent stain (fluorescein) that gives a green-yellow color in ultraviolet light. An antibody titer of 1:200 or higher was considered as positive, and a titer < 1:200 was considered as negative (Silva et al., 2007; Dubey, 2003; Reichel and Drake, 1996).

2.3. Herd follow-up during the study period

The number of artificial inseminations, birth interval and pregnancy outcomes were assessed. In case of abortion, the aborted fetuses were submitted to necropsy. Calves born of *N. caninum* positive dams were clinically examined.

2.4. Statistical analysis

Statistical analyses were carried out with SPSS software (IBM DPDD Statistics[®], Version 19.0). Data were analysed by applying the Chi – Square test. The difference between groups (*N. caninum* seropositive dams vs *N. caninum* seronegative dams) was considered statistically significant if $p \leq .05$.

3. Results

3.1. Serological status of the herds

Among the 875 cows that were included in the study, 184 (21.03%) were seropositive for the detection of *N. caninum* antibodies by IFAT and 691 were seronegative (78.97%).

3.2. Number of artificial inseminations

The total number of artificial inseminations used in the group of seropositive cows was 721 dosages, while in the group of seronegative cows was 1651 dosages. The mean number of artificial inseminations performed until conception was 2.39 in seronegative cows, whereas in seropositive cows increased to 3.92 (p < .05) (Table 1). In the group of seropositive cows 170 of 184 (92%) returned to estrus, while in the group of seronegative cows 206 of 691 (30%) returned to estrus.

3.3. Birth interval and gestation outcomes

The birth interval was 199 days in the *N. caninum* seropositive cows, whereas in the seronegative cows was significantly lower 120 days (p < .05). The number of abortions was significantly higher in seropositive cows compared to the seronegative cows: 52 versus 18, respectively (p < .05) (Table 1). Following abortion, retention of fetal membranes was recorded in 11 seropositive dams. Necropsy of the aborted fetuses revealed inflammatory lesions in the internal organs, mostly in the brain and skeletal muscles. Autolysis was also observed in the fetal tissues harvested from the aborted fetuses. Six neonatal calves

Table 1

Artifical insemination, birth interval and gestation outcomes.

Category	Study groups		Statistical outcome <i>p</i> -value
	<i>N. caninum</i> seropositive dams $N = 184$	<i>N. caninum</i> seronegative dams $N = 691$	_
Number of artificial inseminations, mean ± SD	3.92 ± 1.56	2.39 ± 1.1	< 0.05
Birth interval (number of days), n	199	120	< 0.05
Number (percentage) of abortions, n (%)	52 (61.9%)	18 (2.6%)	< 0.05
Number (percentage) of calves born with clinical signs of neosporosis, n (%)	6 (3.3%)	0 (0.0%)	< 0.05

N, number of dams; SD, standard deviation.

born of seropositive dams presented clinical signs of neosporosis, including exophthalmia, ataxia, incoordination and weakness, hydrocephalus and hyperextension of the hind limbs.

4. Discussion

This is the first report in Greece when *N. caninum* seroprevalence was assessed in dairy cows by IFAT. To date, there was only a previous report in which *N. caninum* infection was evaluated in milk samples by the enzyme-linked immunosorbent assay (ELISA) (Sotiraki et al., 2008). IFAT is the first serological test used for diagnosis of neosporosis (Dubey et al., 1988) and is the accepted technique in *N. caninum*-antibody research, being extensively used for the diagnosis of *N. caninum* infection in epidemiological studies conducted in other domestic and wild animals (Björkman and Uggla, 1999).

In the present study, the percentage of abortions in N. caninum seropositive dams was 10 times higher than in the seronegative ones (61.9% versus 2.6%, respectively). These findings are in line with those from a study in Iran, which also found a percentage of abortions 3 times higher in seropositive dams compared to the seronegative ones (21.6% versus 7.3%, respectively) (Mazuz et al., 2014). Spilovska et al. showed a correlation between the presence of N. caninum specific antibodies and occurrence of abortions (Spilovska et al., 2015). It has also been shown that < 5% of cows may have repeated abortions due to neosporosis (Dubey et al., 2007; Pabon et al., 2007). In addition, a cohort study showed that the N. caninum infected dams with abortion history had a 5.6-fold-higher abortion risk than the congenitally infected cows that had not experienced an abortion before (Thurmond and Hietala, 1997). In our study, the proportion of seropositive cows with a history of abortion (28.26%) was significantly higher than the proportion of cows with the same clinical signs but seronegative (2.60%). This finding provides an indirect evidence that N. caninum may be involved in abortions of cows from the studied region.

N. caninum seroprevalence we found by IFAT in dairy cattle was 21.03%, being higher than the seroprevalence reported in Hungary (3.4%) (Hornok et al., 2006), Germany (4.1%) (Conraths et al., 1996), Ireland (12.6%) (McNamee et al., 1996), Spain (11.2%) (Caetano-da-Silva et al., 2004) and in the Czech Republic (3.2%) (Václavek et al., 2003), and lower than in Belgium (29%) (De Meerschman et al., 2002), Germany (27%) (Schares et al., 1998), Italy (24.4%) (Magnino et al., 1999), Slovakia (24.1) (Spilovska et al., 2015) and Denmark (22%) (Jensen et al., 1999). In some European countries (Germany, The Netherlands, Spain and Sweden) the prevalence of N. caninum ranges between 16% and 76% (Bartels et al., 2006). Bartels et al. performed a study in The Netherlands and the average prevalence in random herds was 10.8% (range: 1.0% to 48.7%), while in epidemic-abortion herds, the average was 31.2% (range: 11.3% to 61.6%) (Bartels et al., 2006). In the western Romania, neosporosis prevalence range was shown to be 10% to 52.2% in different regions (Imre et al., 2012).

There is some evidence that cattle breed can affect the seroprevalence of *N. caninum* in a herd (Munhoz et al., 2009; Duong et al., 2008; Armengol et al., 2007). A study conducted in Belgium found a higher abortion rate (28.6%) in *N. caninum* seropositive dairy cows compared to seropositive beef cows (14.0%) (De Meerschman et al., 2002).

However, a study conducted Spain found similar rates of abortion between dairy and beef cows (Eiras et al., 2011). Further investigations are needed to determine whether improved breeds of dairy cattle are more susceptible to *N. caninum* infection than the zebu cattle or crossbreeds (Munhoz et al., 2009).

Regarding the number of inseminations applied before each conception, we observed that *N. caninum* seropositive cows have required 1.64 times more inseminations compared to the seronegative ones. The seropositive cows also returned more often to estrus and had a longer period of days open, which is typically associated with reduced profitability. These findings are in line with previous studies (Kamga-

Waladjo et al., 2010; Hall et al., 2005; Tiwari et al., 2005).

The association between returning to estrus after pregnancy and occurrence of *N. caninum* in a herd also remains to be further assessed. A Canadian case-control study suggested that there is a positive association between the occurrence of *N. caninum*-related abortions in a herd and the annual rate of cattle returning to estrus after pregnancy (Hobson et al., 2005). In addition, a high rate of early pregnancy losses could increase the chance of definitive hosts to have access to the infectious material and thereby it could also increase the rate of oocystmediated horizontal transmission. This assumption is supported by the results of four other studies (Waldner, 2005; Muñoz-Zanzi et al., 2004; Waldner et al., 2001; Waldner et al., 1998). However, other epidemiological studies showed no indication that *N. caninum* is able to cause early pregnancy losses (López-Gatius et al., 2004; López-Gatius et al., 2005; Romero et al., 2005; Jensen et al., 1999).

Retention of fetal membranes and subsequent metritis occur rarely among cows infected with *N. caninum* (Dubey, 2003). In our study, the retention of fetal membranes after abortion was 5.98% in the seropositive cows.

Six neonatal calves born of seropositive dams in our study presented clinical signs of neosporosis, such as exophthalmia, ataxia, incoordination, hydrocephalus and hyperextension of the hind limbs. Similar clinical signs, as well as the inability to rise and low birth weight were previously described by Dubey and Schares (2011) in calves born to *N. caninum* seropositive dams). In addition, different studies suggest that calves born of dams with a previous history of *N. caninum* infections and/or neurological dysfunctions at birth, including mild neurologic limb disorders and mild nonsuppurative encephalomyelitis (Barr et al., 1993; Bryan et al., 1994).

From our experience, the preventive measures for *N. caninum* infection in cattle should be focused on limited consumption of raw meat by the canines rather than chemotherapy. The latter appears to be uneconomical because is mostly used only as a preventive measure and its longterm use leads to unacceptable milk or meat residues, followed by withdrawal periods for their consumption and major economic losses.

5. Conclusion

To the authors' knowledge, this is the first report of a sero-epidemiological approach for assessment of *N. caninum* infection in Greece by using IFAT. Our findings suggest that neosporosis has a high seroprevalence in Northern Greece with a much greater geographical distribution than previously believed. We also noted a singificantly increased number of insemination needed for conception in *N. caninum* seropositive dams compared to seronegative dams, as well as a substantial increase in birth interval and abortions in seropositive dams. All these may lead to major financial losses, especially if preventive measures for *N. caninum* spread are not be applied.

We recommend the following preventive measures that could avoid financial losses caused by chemotherapy: quarantine and testing of replacement and other new cows (farmers may decide to remove infected cows or their progeny from the herd), limited contamination of pastures, food and feeding stuff with feces from dogs and other potential definitive hosts, regular control of water supply, regular rodent control to eliminate a potential reservoir of *N. caninum* in livestock and reproductive management control (e.g. the control of embryos transfer from infected dams into uninfected recipients can prevent endogenous transplacental transmission of the parasite, artificial insemination of seropositive dams with semen from beef bulls).

Ethical statement for solid state ionics

None.

Declaration of Competing Interest

None.

References

- Ansari-Lari, M., Rowshan-Ghasrodashti, A., Jesmani, H., Masoudian, M., Badkoobeh, M., 2017. Association of Neospora caninum with reproductive performance in dairy cows: a prospective study from Iran. Vet. Res. Forum 8, 109–114.
- Armengol, R., Pabon, M., Santolaria, P., Cabezon, O., Adelantado, C., Yaniz, J., Lopez-Gatius, F., Almeria, S., 2007. Low seroprevalence of Neospora caninum infection associated with the Limousin breed in cow-calf herds in Andorra, Europe. J. Parasitol. 93, 1029–1032.
- Barr, B.C., Conrad, P.A., Breitmeyer, R., Sverlow, K., Anderson, M.L., Reynolds, J., Chauvet, A.E., Dubey, J.P., Ardans, A.A., 1993. Congenital Neospora infection in calves born from cows that had previously aborted Neospora-infected fetuses: four cases (1990-1992). J. Am. Vet. Med. Assoc. 202, 113–117.
- Bartels, C.J.M., van Schaik, G., Veldhuisen, J.P., van den Borne, B.H.P., Wouda, W., Dijkstra, T., 2006. Effect of Neospora caninum-serostatus on culling, reproductive performance and milk production in Dutch dairy herds with and without a history of Neospora caninum-associated abortion epidemics. Prev. Vet. Med. 77, 186–198.
- Basso, W., Venturini, L., Venturini, M., Hill, D., Kwok, O., Shen, S., Dubey, J., 2001. First isolation of Neospora caninum from the feces of a naturally infected dog. J. Parasitol. 87, 612–618.
- Bielanski, A., Phipps-Todd, B., Robinson, J., 2002. Effect of < em > Neospora caninum < /em > on in vitro development of preimplantation stage bovine embryos and adherence to the zona pellucida. Vet. Rec. 150, 316–318.
- Bjerkas, I., Mohn, S.F., Presthus, J., 1984. Unidentified cyst-forming sporozoon causing encephalomyelitis and myositis in dogs. Z Parasitenkd. 70, 271–274.
- Björkman, C., Uggla, A., 1999. Serological diagnosis of Neospora caninum infection. Int. J. Parasitol. 29, 1497–1507.
- Bryan, L.A., Gajadhar, A.A., Dubey, J.P., Haines, D.M., 1994. Bovine neonatal encephalomyelitis associated with a Neospora sp. protozoan. Can. Vet. J. 35, 111–113.
- Caetano-da-Silva, A., Ferre, I., Aduriz, G., Álvarez-García, G., Del-Pozo, I., Atxaerandio, R., Regidor-Cerrillo, J., Ugarte-Garagalza, C., Ortega-Mora, L., 2004. Neospora caninum infection in breeder bulls: seroprevalence and comparison of serological methods used for diagnosis. Vet. Parasitol. 124, 19–24.
- Conraths, F., Bauer, C., Becker, W., 1996. Detection of antibodies against Neospora caninum in cows on hessian farms with abortion and fertility problems. Dtsch. Tierarztl. Wochenschr. 103, 221–224.
- Davison, H.C., Guy, C.S., McGarry, J.W., Guy, F., Williams, D.J., Kelly, D.F., Trees, A.J., 2001. Experimental studies on the transmission of Neospora caninum between cattle. Res. Vet. Sci. 70, 163–168.
- De Meerschman, F., Speybroeck, N., Berkvens, D., Rettigner, C., Focant, C., Leclipteux, T., Cassart, D., Losson, B., 2002. Fetal infection with Neospora caninum in dairy and beef cattle in Belgium. Theriogenology. 58, 933–945.
- Dijkstra, T., Eysker, M., Schares, G., Conraths, F., Wouda, W., Barkema, H., 2001. Dogs shed Neosporacaninum oocysts after ingestion of naturally infected bovine placenta but not after ingestion of colostrum spiked with Neosporacaninum tachyzoites. Int. J. Parasitol. 31, 747–752.
- Donahoe, S.L., Lindsay, S.A., Krockenberger, M., Phalen, D., Šlapeta, J., 2015. A review of neosporosis and pathologic findings of Neospora caninum infection in wildlife. Int J Parasitol Parasites Wildl. 4, 216–238.
- Dubey, J.P., 2003. Review of Neospora caninum and neosporosis in animals. Korean J Parasitol. 41 (1), 16.
- Dubey, J.P., Schares, G., 2006. Diagnosis of bovine neosporosis. Vet. Parasitol. 140, 1–34. Dubey, J., Schares, G., 2011. Neosporosis in animals—the last five years. Vet. Parasitol. 180, 90–108.
- Dubey, J.P., Hattel, A.L., Lindsay, D.S., Topper, M.J., 1988. Neonatal Neospora caninum infection in dogs: isolation of the causative agent and experimental transmission. J. Am. Vet. Med. Assoc. 193, 1259–1263.
- Dubey, J., Barr, B., Barta, J., Bjerkås, I., Björkman, C., Blagburn, B., Bowman, D., Buxton, D., Ellis, J., Gottstein, B., 2002. Redescription of Neospora caninum and its differentiation from related coccidia. Int. J. Parasitol. 32, 929–946.
- Dubey, J.P., Schares, G., Ortega-Mora, L.M., 2007. Epidemiology and control of Neosporosis and < em > Neospora caninum < /em > Clin. Microbiol. Rev. 20, 323–367.
- Dubey, J.P., Jenkins, M.C., Kwok, O.C.H., Ferreira, L.R., Choudhary, S., Verma, S.K., Villena, I., Butler, E., Carstensen, M., 2013. Congenital transmission of Neospora caninum in whitetailed deer (Odocoileus virginianus). Vet. Parasitol. 196, 519–522.
- Duong, M.C., Alenius, S., Huong, L.T., Bjorkman, C., 2008. Prevalence of Neospora caninum and bovine viral diarrhoea virus in dairy cows in southern Vietnam. Vet. J. 175, 390–394.
- Eiras, C., Arnaiz, I., Alvarez-Garcia, G., Ortega-Mora, L.M., Sanjuanl, M.L., Yus, E., Dieguez, F.J., 2011. Neospora caninum seroprevalence in dairy and beef cattle from the northwest region of Spain, Galicia. Prev. Vet. Med. 98, 128–132.
- Gondim, L.F.P., Gao, L., McAllister, M., 2002. Improved production of Neospora caninum oocysts, cyclical oral transmission between dogs and cattle, and in vitro isolation from oocysts. J. Parasitol. 88, 1159–1163.
- Gondim, L.F.P., McAllister, M.M., Pitt, W.C., Zemlicka, D.E., 2004. Coyotes (Canis latrans) are definitive hosts of Neospora caninum. Int. J. Parasitol. 34, 159–161.
- Haddad, J.P.A., Dohoo, I.R., VanLeewen, J.A., 2005. A review of Neospora caninum in dairy and beef cattle–a Canadian perspective. Can. Vet. J. 46, 230–243.

Hall, C.A., Reichel, M.P., Ellis, J.T., 2005. Neospora abortions in dairy cattle: diagnosis,

Veterinary Parasitology: Regional Studies and Reports 19 (2020) 100365

mode of transmission and control. Vet. Parasitol. 128, 231-241.

- Hobson, J.C., Duffield, T.F., Kelton, D., Lissemore, K., Hietala, S.K., Leslie, K.E., McEwen, B., Peregrine, A.S., 2005. Risk factors associated with Neospora caninum abortion in Ontario Holstein dairy herds. Vet. Parasitol. 127, 177–188.
- Hornok, S., Edelhofer, R., Hajtós, I., 2006. Seroprevalence of neosporosis in beef and dairy cattle breeds in Northeast Hungary. Acta Vet. Hung. 54, 485–491.
- Imre, K., Morariu, S., Ilie, M.S., Imre, M., Ferrari, N., Genchi, C., Darabus, G., 2012. Serological survey of Neospora caninum infection in cattle herds from Western Romania. J. Parasitol. 98, 683–685.
- Jenkins, M., Baszler, T., Bjorkman, C., Schares, G., Williams, D., 2002. Diagnosis and seroepidemiology of Neospora caninum-associated bovine abortion. Int. J. Parasitol. 32, 631–636.
- Jensen, A.M., Bjorkman, C., Kjeldsen, A.M., Wedderkopp, A., Willadsen, C., Uggla, A., Lind, P., 1999. Associations of Neospora caninum seropositivity with gestation number and pregnancy outcome in Danish dairy herds. Prev. Vet. Med. 40, 151–163.
- Kamga-Waladjo, A.R., Gbati, O.B., Kone, P., Lapo, R.A., Chatagnon, G., Bakou, S.N., Pangui, L.J., Diop Pel, H., Akakpo, J.A., Tainturier, D., 2010. Seroprevalence of Neospora caninum antibodies and its consequences for reproductive parameters in dairy cows from Dakar-Senegal, West Africa. Trop. Anim. Health Prod. 42, 953–959.
- King, J.S., Šlapeta, J., Jenkins, D.J., Al-Qassab, S.E., Ellis, J.T., Windsor, P.A., 2010. Australian dingoes are definitive hosts of Neospora caninum. Int. J. Parasitol. 40, 945–950.
- Kul, O., Atmaca, H.T., Anteplioglu, T., Ocal, N., Canpolat, S., 2015. Neospora caninum: the first demonstration of the Enteroepithelial stages in the intestines of a naturally infected dog. J. Comp. Pathol. 153, 9–13.
- Lassen, B., Orro, T., Aleksejev, A., Raaperi, K., Jarvis, T., Viltrop, A., 2012. Neospora caninum in Estonian dairy herds in relation to herd size, reproduction parameters, bovine virus diarrhoea virus, and bovine herpes virus 1. Vet. Parasitol. 190, 43–50.
- Lindsay, D.S., Dubey, J.P., Duncan, R.B., 1999a. Confirmation that the dog is a definitive host for Neospora caninum. Vet. Parasitol. 82, 327–333.
- Lindsay, D.S., Upton, S.J., Dubey, J., 1999b. A structural study of the Neospora caninum oocyst. Int. J. Parasitol. 29, 1521–1523.
- Lindsay, D.S., Ritter, D.M., Brake, D., 2001. Oocyst excretion in dogs fed mouse brains containing tissue cysts of a cloned line of Neospora caninum. J. Parasitol. 87, 909911.
- López-Gatius, F., Pabón, M., Almería, S., 2004. Neospora caninum infection does not affect early pregnancy in dairy cattle. Theriogenology. 62, 606–613.
- López-Gatius, F., Santolaria, P., Almería, S., 2005. Neospora caninum infection does not affect the fertility of dairy cows in herds with high incidence of Neospora-associated abortions. J. Veterinary Med. Ser. B 52, 51–53.
- Magnino, S., Vigo, P.G., Fabbi, M., Colombo, M., Bandi, C., Genchi, C., 1999. Isolation of a bovine Neospora from a newborn calf in Italy. Vet. Rec. 144, 456.
- Malaguti, J.M., Cabral, A.D., Abdalla, R.P., Salgueiro, Y.O., Galleti, N.T., Okuda, L.H., Cunha, E.M., Pituco, E.M., del Fava, C., 2012. Neospora caninum as causative agent of bovine encephalitis in Brazil. Rev. Bras. Parasitol. Vet. 21, 48–54.
- Mazuz, M.L., Fish, L., Reznikov, D., Wolkomirsky, R., Leibovitz, B., Savitzky, I., Golenser, J., Shkap, V., 2014. Neosporosis in naturally infected pregnant dairy cattle. Vet. Parasitol. 205, 8591.
- McAllister, M.M., Jolley, W.R., Wills, R.A., Lindsay, D.S., McGuire, A.M., Tranas, J.D., 1998. Oral inoculation of cats with tissue cysts of Neospora caninum. Am. J. Vet. Res. 59, 441444.
- McNamee, P.T., Trees, A.J., Guy, F., Moffett, D., Kilpatrick, D., 1996. Diagnosis and prevalence of neosporosis in cattle in Northern Ireland. Vet. Rec. 138, 419–420.
- Munhoz, A.D., Pereira, M.J.S., Flausino, W., Lopes, C.W.G., 2009. Neospora caninum seropositivity in cattle breeds in the south Fluminense Paraíba Valley, state of Rio de Janeiro. Pesqui. Vet. Bras. 29, 29–32.
- Muñoz-Zanzi, C.A., Thurmond, M.C., Hietala, S.K., 2004. Effect of bovine viral diarrhea virus infection on fertility of dairy heifers. Theriogenology. 61, 1085–1099.
- Pabon, M., Lopez-Gatius, F., Garcia-Ispierto, I., Bech-Sabat, G., Nogareda, C., Almeria, S., 2007. Chronic Neospora caninum infection and repeat abortion in dairy cows: a 3year study. Vet. Parasitol. 147, 40–46.
- Piagentini, M., Moya-Araujo, C.F., Prestes, N.C., Sartor, I.F., 2012. Neospora caninum infection dynamics in dairy cattle. Parasitol. Res. 111, 717–721.
- Reichel, M.P., Drake, J.M., 1996. The diagnosis of Neospora abortions in cattle. N. Z. Vet. J. 44, 151–154.
- Reichel, M.P., Alejandra Ayanegui-Alcérreca, M., Gondim, L.F.P., Ellis, J.T., 2013. What is the global economic impact of Neospora caninum in cattle – the billion dollar question. Int. J. Parasitol. 43, 133–142.
- Reiterová, K., Špilovská, S., Antolová, D., Dubinský, P., 2009. Neospora caninum, potential cause of abortions in dairy cows: the current serological follow-up in Slovakia. Vet. Parasitol. 159, 1–6.
- Romero, J.J., Breda, S.V., Vargas, B., Dolz, G., Frankena, K., 2005. Effect of neosporosis on productive and reproductive performance of dairy cattle in Costa Rica. Theriogenology. 64, 1928–1939.
- Schares, G., Peters, M., Wurm, R., Bärwald, A., J. Conraths, F., 1998. The efficiency of vertical transmission of Neospora caninum in dairy cattle analysed by serological techniques. Vet. Parasitol. 80, 87–98.
- Schares, G., Heydorn, A., Cüppers, A., Conraths, F., Mehlhorn, H., 2001. Hammondia heydorni-like oocysts shed by a naturally infected dog and Neospora caninum NC-1 cannot be distinguished. Parasitol. Res. 87, 808–816.
- Silva, R.C., Machado, G.P., 2016. Canine neosporosis: perspectives on pathogenesis and management. Vet. Med. (Auckland, N.Z.) 7, 59–70.
- Silva, D.A.O., Lobato, J., Mineo, T.W.P., Mineo, J.R., 2007. Evaluation of serological tests for the diagnosis of Neospora caninum infection in dogs: optimization of cut off titers and inhibition studies of cross-reactivity with toxoplasma gondii. Vet. Parasitol. 143, 234244.
- Sotiraki, S., Brozos, C., Samartzi, F., Schares, G., Kiossis, E., Conraths, F.J., 2008.

Neospora caninum infection in Greek dairy cattle herds detected by two antibody assays in individual milk samples. Vet. Parasitol. 152, 79–84.

- Spilovska, S., Reiterova, K., Antolova, D., 2015. Neospora caninum associated abortions in Slovak dairy farm. Iran. J. Parasitol. 10, 96–101.
- Thurmond, M.C., Hietala, S.K., 1997. Effect of congenitally acquired Neospora caninum infection on risk of abortion and subsequent abortions in dairy cattle. Am. J. Vet. Res. 58 13811385.
- Tiwari, A., VanLeeuwen, J.A., Dohoo, I.R., Stryhn, H., Keefe, G.P., Haddad, J.P., 2005. Effects of seropositivity for bovine leukemia virus, bovine viral diarrhoea virus, Mycobacterium avium subspecies paratuberculosis, and Neospora caninum on culling in dairy cattle in four Canadian provinces. Vet. Microbiol. 109, 147–158.
- Václavek, P., Koudela, B., Modrý, D., Sedlák, K., 2003. Seroprevalence of Neospora caninum in aborting dairy cattle in the Czech Republic. Vet. Parasitol. 115, 239–245.
- Waldner, C.L., 2005. Serological status for N. caninum, bovine viral diarrhea virus, and infectious bovine rhinotracheitis virus at pregnancy testing and reproductive performance in beef herds. Anim. Reprod. Sci. 90, 219–242.

- Waldner, C.L., Janzen, E.D., Ribble, C.S., 1998. Determination of the association between Neospora caninum infection and reproductive performance in beef herds. J. Am. Vet. Med. Assoc. 213, 685–690.
- Waldner, C.L., Henderson, J., Wu, J.T., Breker, K., Chow, E.Y., 2001. Reproductive performance of a cow-calf herd following a Neospora caninum-associated abortion epidemic. Can. Vet. J. 42, 355–360.
- Wapenaar, W., Barkema, H.W., VanLeeuwen, J.A., McClure, J.T., O'Handley, R.M., Kwok, O.C.H., Thulliez, P., Dubey, J.P., Jenkins, M.C., 2007. Comparison of serological methods for the diagnosis of Neospora caninum infection in cattle. Vet. Parasitol. 143, 166–173.
- Wouda, W., Bartels, C.J., Moen, A.R., 1999. Characteristics of Neospora caninum-associated abortion storms in diary herds in the Netherlands (1995 to 1997). Theriogenology. 52, 233–245.
- Xu, M.J., Liu, Q.Y., Fu, J.H., Nisbet, A.J., Shi, D.S., He, X.H., Pan, Y., Zhou, D.H., Song, H.Q., Zhu, X.Q., 2012. Seroprevalence of toxoplasma gondii and Neospora caninum infection in dairy cows in subtropical southern China. Parasitology. 139, 1425–1428.