RESEARCH LETTER

Superior effect of MP-AzeFlu compared to monotherapy with fluticasone propionate or azelastine on GILZ, MKP-1 and TTP anti-inflammatory gene expression in healthy and inflamed upper airway mucosa

To the Editor,

Allergic rhinitis (AR) is a Th2 IgE-mediated disease with elevated levels of pro-inflammatory mediators, eosinophil infiltration of the nasal mucosa, and mucus hypersecretion. Antihistamines and intranasal corticosteroids, including MP-AzeFlu (intranasal fluticasone and azelastine), are recommended as the first-line therapy for AR.¹ Intranasal corticosteroids are also the first-line treatment for chronic rhinosinusitis with nasal polyps (CRSwNP) or without (CRSsNP), while antihistamines are only indicated in chronic rhinosinusitis (CRS) with concomitant AR.²

MP-AzeFlu has demonstrated efficacy in AR, and a superior effect compared to these drugs administered individually, on nasal and ocular symptoms and quality of life; moreover, MP-AzeFlu has shown earlier and faster AR control in moderate-to-severe AR.³ While MP-AzeFlu is guideline-recommended and its clinical efficacy has been widely studied, the mechanisms by which this therapy exerts its effect in AR are largely unknown.

Our laboratory has used an *in vitro* model of eosinophilic inflammation based on the interaction between cultured primary isolated nasal mucosa epithelial cells and isolated peripheral blood eosinophils to study the effect and potency of anti-inflammatory drugs.^{4,5} Using this model, we have previously reported greater anti-inflammatory effects of the combination of an intranasal corticosteroid and an antihistamine, including MP-AzeFlu, in both nasal mucosa and nasal polyp epithelial cells, compared to the effect of these drugs administered alone.⁵ We have also demonstrated that glucocorticoids induce the transcription of anti-inflammatory genes such as glucocorticoid-induced leucine zipper (GILZ), mitogenactivated protein kinase (MAPK) phosphatase 1 (MKP-1) and tristetraprolin (TTP) in nasal mucosa fibroblasts.⁶

While detailed insights are lacking regarding the mechanisms of pathogenesis in CRSwNP, it is now clear that IgE-mediated Th2 inflammatory pathways play a critical role in this disease.⁷ H_1 receptor antagonists decrease eosinophil survival⁴ and inhibit the release of pro-inflammatory mediators by mast cells⁸ and the production of pro-inflammatory cytokines by nasal epithelial cells.^{4,9} In addition, azelastine (AZE) has been shown to enhance the anti-inflammatory effect

of budesonide and improve nasal symptoms in AR patients through the induction of MKP-1 expression.⁹ As such, it is reasonable to hypothesize that an increased induction of anti-inflammatory genes could partially explain the superior anti-inflammatory effect of MP-AzeFlu when compared to fluticasone propionate (FP) or AZE alone.

In the present study, we propose to analyse the anti-inflammatory effect of MP-AzeFlu, compared to monotherapy with FP or AZE, on GILZ, MKP-1 and TTP gene expression in healthy and inflamed upper airway mucosa.

1 | MATERIALS AND METHODS

1.1 | Study population

Nasal mucosa (NM) tissues were obtained from patients undergoing nasal-corrective surgery for nasal deviation and/or turbinate hypertrophy. Patients with current upper respiratory tract infections were excluded. Nasal polyps (NP) were obtained from patients undergoing nasal polypectomy and/or endoscopic sinus surgery for CRSwNP. Patients with asthma, nonsteroidal anti-inflammatory drug-exacerbated respiratory disease (N-ERD) or other nasal and nasosinusal diseases (i.e. vasculitis, granulomatosis, benign or malignant tumours) were excluded.

1.2 | Experimental design

Fibroblasts were isolated using a specific and selective growth culture medium. The purity of fibroblast cultures was confirmed by positive immunostaining to vimentin (fibroblast marker) and negative to cytokeratin 1 (epithelial cell marker).⁶ Cells were incubated with serum-free medium and treated for 2–24 h with MP-AzeFlu (dilution $1:10^2-1:10^4$) or equivalent dilutions of FP and AZE. mRNA and protein expression of GILZ, MKP-1 and TTP (at 2, 6 and/or 24 h) were assessed by real-time polymerase chain reaction (RT-PCR) and western blot, respectively.

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1.3 | Ethics issues

Prior to study initiation, the protocol was approved by the HCB Ethics Committee. Written informed consent was obtained from all patients prior to their participation.

1.4 | Statistical methods

All results are expressed as mean \pm standard error of the mean (SEM). The Wilcoxon tests were used for the non-parametric paired comparisons. Analyses were performed using GraphPad 8.4.0 with an alpha level set at p < .05.

2 | RESULTS

2.1 | GILZ

MP-AzeFlu and FP up-regulated GILZ mRNA expression at all time-points (from 2 h to 24 h) and dilutions in both NM and NP. AZE induced a mild up-regulation of GILZ mRNA expression at dilution $1:10^2$ at some time-points. MP-AzeFlu at dilution $1:10^2$ showed a superior effect in the up-regulation of GILZ mRNA expression at 2 and 6 h in NM and at 6 and 24 h in NP compared to monotherapy with FP or AZE (Figure 1). Neither MP-AzeFlu

Key Messages

- MP-AzeFlu has demonstrated efficacy in allergic rhinitis, and a superior effect compared with FP and AZE administered in monotherapy.
- We analysed the anti-inflammatory effect of MP-AzeFlu, compared with FP or AZE, on GILZ, MKP-1 and TTP gene expression in healthy and inflamed upper airway mucosa.
- The superior clinical effect of MP-AzeFlu compared to FP and AZE monotherapy may be related to greater upregulation of anti-inflammatory GILZ gene expression and, to a lesser extent, MKP-1 and TTP.

nor FP significantly up-regulated GILZ protein expression at any time-point or dilution.

2.2 | MKP-1

MP-AzeFlu and FP up-regulated MKP-1 mRNA expression at all time-points and dilutions in both NM and NP. AZE at dilution $1:10^2$ up-regulated MKP-1 mRNA expression at 2 h in both NM and NP.



FIGURE 1 Superior effect of MP-AzeFlu compared to monotherapy with fluticasone propionate (FP) or azelastine (AZE) on GILZ mRNA expression in nasal mucosa (NM) and nasal polyps (NP) in a time course (2, 6 and 24 h). *p < .05; **p < .01; (*)p = .06 compared to negative control

		MKP-1 mRNA		TTP mRNA	
		NM	NP	NM	NP
2 h	MP-AzeFlu	$5.9 \pm 1.1^{\dagger}$	$5.6 \pm 1.4^{*}$	$1.7\pm0.1^{*,\dagger}$	1.9 <u>+</u> 0.3
	FP	4.5 ± 0.8	4.1 ± 1.0	1.3 ± 0.1	1.6 ± 0.4
	AZE	2.2 ± 0.4	4.7 ± 1.6	1.3 ± 0.2	1.4 <u>+</u> 0.3
6 h	MP-AzeFlu	$6.9 \pm 2.1^{\ddagger}$	7.7 <u>+</u> 3.2	$1.8 \pm 0.1^{\ddagger}$	$2.9 \pm 0.2^{\dagger}$
	FP	5.3 <u>±</u> 0.8	6.4 ± 2.6	1.6 ± 0.1	2.1 ± 0.3
	AZE	1.6 ± 0.5	3.0 ± 1.2	1.0 ± 0.1	1.0 ± 0.1

TABLE 1 Superior effect of MP-AzeFlu compared with fluticasone propionate and azelastine monotherapy on MKP-1 and TTP mRNA expression (2 and 6 h)

Note: Data expressed as fold change (mean \pm SEM) from untreated cells (control = 1).

Abbreviations: AZE, azelastine; FP, fluticasone propionate; NM, nasal mucosa; NP, nasal polyps.

*p < .05 compared to FP; [†]p < .05 compared to AZE; [‡]p < .01 compared to AZE.

MP-AzeFlu at dilution 1:10² showed a partial superior effect in the up-regulation of MKP-1 mRNA expression compared to monotherapy with FP in NP (at 2 h) and AZE in NM (at 2 h and 6 h; Table 1). MKP-1 protein expression was not detected when cells were incubated with MP-AzeFlu and FP.

2.3 | TTP

MP-AzeFlu and FP at dilution $1:10^2$ increased TTP mRNA expression in NM (at 2 h and 6 h) and NP (at 6 h).

AZE had no effect on TTP mRNA expression. MP-AzeFlu at dilution $1:10^2$ showed a superior effect in the up-regulation of TTP mRNA expression compared to monotherapy with FP and AZE in NM (at 2 h) and with AZE in NM and NP (at 6 h; Table 1). The effect of drugs on TTP protein expression was not studied due to very low gene expression.

3 | DISCUSSION

In this study, MP-AzeFlu, FP and AZE up-regulated GILZ mRNA expression, with MP-AzeFlu showing a superior effect compared with that of monotherapy. MP-AzeFlu, FP and AZE also up-regulated MKP-1 mRNA expression; however, no significant differences were observed between MP-AzeFlu and monotherapy. MP-AzeFlu and FP up-regulated TTP mRNA expression, while AZE did not.

Anti-inflammatory genes are activated when glucocorticoids (GCs) diffuse across the cell membrane to bind to GRα. The ligandbound receptor translocates to the nucleus and binds GC-responsive elements (GREs) on the promoter region of target genes.¹⁰ The antiinflammatory effect of MP-AzeFlu has been confirmed by the upregulation of anti-inflammatory gene expression of GILZ, MKP-1 and TTP in our *in vitro* model. These findings suggest some of the molecular mechanisms of action of MP-AzeFlu in the upper airway inflammation (both AR and CRSwNP). However, in vitro models have limitations. In this artificial setting, interactions between cells are lost. Furthermore, the concentration of drugs used do not correlate with clinical doses used in the patient care setting. Thus, these findings may not translate to clinical outcomes and further studies are needed to confirm the findings.

In conclusion, the superior clinical effect of MP-AzeFlu on both NP and NM compared with monotherapy may be partly related to the greater up-regulation of the anti-inflammatory gene expression of GILZ and, to a lesser extent, MKP-1 and TTP expression.

KEYWORDS

azelastine, fluticasone, gene expression, GILZ, MKP-1, MP-AzeFlu, TTP

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CONFLICT OF INTEREST

Joaquim Mullol is or has been a member of national and international scientific advisory boards (consulting), received fees for lectures, and grants for research projects from Allakos, AstraZeneca, Genentech-Roche, Glenmark, GSK, Menarini, MSD, Mitsubishi-Tanabe, MYLAN-MEDA Pharma (Viatris), Novartis, Procter & Gamble, Sanofi-Genzyme & Regeneron, UCB and Uriach Group. SVA, JRF, VT, MF, IA and AV do not have conflicts of interest for the present manuscript.

AUTHOR CONTRIBUTIONS

All authors made substantial contributions to the conception or design of the manuscript, or the acquisition, analysis or interpretation of data for the manuscript, and all authors were involved in drafting the manuscript or revising it critically for important intellectual content. The authors were fully responsible for all content and editorial decisions and received no financial support or other forms of compensation related to the development of this manuscript. All authors had final approval of the manuscript and are accountable for all aspects of the work in ensuring the accuracy and integrity of this manuscript.

ETHICAL APPROVAL

The Dymecos 2 study was approved by the Ethics Committee (CEIm) from Hospital Clinic Barcelona (Catalonia, Spain) on 3 February 2016 with the Registration No. HCB/2016/0007.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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REFERENCES

- Eifan A, Durham S. Pathogenesis of rhinitis. Clin Exp Allergy. 2016;46(9):1139-1151.
- Orlandi RR, Kingdom TT, Hwang PH, et al. International consensus statement on allergy and rhinology: rhinosinusitis. *Int Forum Allergy Rhinol.* 2016;Suppl 1:S22-S209.
- Klimek L, Berger WE, Bousquet J, et al. MP-AzeFlu in moderate-tosevere allergic rhinitis: a literature review. *Int Arch Allergy Immunol*. 2021;182(11):1026-1035.
- Mullol J, de Borja CF, Martínez-Antón MA, et al. Mometasone and desloratadine additive effect on eosinophil survival and cytokine secretion from epithelial cells. *Respir Res.* 2011;12(1):23.
- Roca-Ferrer J, Pujols L, Pérez-González M, et al. Superior effect of MP-AzeFlu than azelastine or fluticasone propionate alone on reducing inflammatory markers. *Allergy Asthma Clin Immunol.* 2018;14:86.
- Fernández-Bertolín L, Mullol J, Fuentes-Prado M, et al. Deficient glucocorticoid induction of anti-inflammatory genes in nasal polyp fibroblasts of asthmatic patients with and without aspirin intolerance. J Allergy Clin Immunol. 2013;132(5):1243-1246.e12.
- Kariyawasam HH, James LK. Chronic rhinosinusitis with nasal polyps: targeting IgE with anti-IgE omalizumab therapy. *Drug Des Devel Ther.* 2020;14:5483-5494.
- 8. Kempuraj D, Huang M, Kandere-Grzybowska K, et al. Azelastine inhibits secretion of IL-6, TNF-alpha and IL-8 as well as NF-kappaB activation and intracellular calcium ion levels in normal human mast cells. *Int Arch Allergy Immunol.* 2003;132(3):231-239.
- Luo X, Ma R, Wu X, et al. Azelastine enhances the clinical efficacy of glucocorticoid by modulating MKP-1 expression in allergic rhinitis. *Eur Arch Otorhinolaryngol.* 2015;272(5):1165-1173.
- Hox V, Lourijsen E, Jordens A, et al. Benefits and harm of systemic steroids for short- and long-term use in rhinitis and rhinosinusitis: an EAACI position paper. *Clin Transl Allergy*. 2020;10(1):1.

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