

Telomerase as a Cancer Target. Development of New Molecules



D.L. Mengual Gomez^a, R.G. Armando^a, C.S. Cerrudo^b, P.D. Ghiringhelli^b and D.E. Gomez^a,*

^{*a}</sup>Laboratory of Molecular Oncology.Department of Science and Technology. Quilmes National University. Bernal, Buenos Aires. Argentina.; ^{<i>b*}Laboratory of Genetic Engineering and Cellular and Molecular Biology, Quilmes National University, Bernal, Buenos Aires, Argentina</sup>



D.E. Gomez

Abstract: Telomeres are the terminal part of the chromosome containing a long repetitive and noncodifying sequence that has as function protecting the chromosomes. In normal cells, telomeres lost part of such repetitive sequence in each mitosis, until telomeres reach a critical point, triggering at that time senescence and cell death. However, in most of tumor cells in each cell division a part of the telomere is lost, however the appearance of an enzyme called telomerase synthetize the segment that just has been lost, therefore conferring to tumor cells the immortality hallmark. Telomerase is significantly

overexpressed in 80–95% of all malignant tumors, being present at low levels in few normal cells, mostly stem cells. Due to these characteristics, telomerase has become an attractive target for new and more effective anticancer agents. The capability of inhibiting telomerase in tumor cells should lead to telomere shortening, senescence and apoptosis. In this work, we analyze the different strategies for telomerase inhibition, either in development, preclinical or clinical stages taking into account their strong points and their caveats. We covered strategies such as nucleosides analogs, oligonucleotides, small molecule inhibitors, G-quadruplex stabilizers, immunotherapy, gene therapy, molecules that affect the telomere/telomerase associated proteins, agents from microbial sources, among others, providing a balanced evaluation of the status of the inhibitors of this powerful target together with an analysis of the challenges ahead.

Keywords: Telomere, Telomerase, Inhibitor, Cancer, Preclinical, Clinical trials.

Received: August 31, 2015

Revised: September 15, 2015

Accepted: October 25, 2015

1. INTRODUCTION

Since the discovery of telomerase [1], an enzyme able to elongates telomeres. and the following discovery that such enzyme was mostly active in tumors [2], telomerase become a prominent target and different ways of inhibitions towards it were attempted. [3]. Telomerase has different components, a catalytic subunit, hTERT, a RNA component, hTR and associated proteins. But telomeres also comprise special structures, proteins that are associated with them and a variety of conformations, all of them allowing or not the activity of telomerase It is necessary then, to decode the structure of the complex telomere/telomerase in order to understand how each possible inhibitor functions and gain perspective about the chances of new developments.

1.1. Telomere and Telomerase Structure

The telomere is a nucleoprotein complex found in the extremes of the chromosomes, where their structure is different from the rest of the chromatin [4] consisting in short and repetitive sequences of d[TTAGGG] [5, 6]. The G-rich strand of telomeric DNA is always oriented 5'-3' towards the terminal portion of the chromosome and had a protruding extreme of ~200 nucleotides [7] as consequence of the problem of terminal replication. The 3' protruding G-rich strand can form complex structures of telomeres [8,9]. We also can observe different telomeric proteins that bind to mammal telomeres. In humans, telomeres are bound by a six-protein complex called shelterin, [10,11] comprised of TRF1 and TRF2 [12, 13, 14] which in turn recruits RAP1, TIN2, TPP1 [15]. and POT1 which interacts with DNA.

Another important structural parameter governing telomere function is that they also contain RNA, called TERRAs [telomere repeat containing RNAs] [16], implicated in the negative regulation of telomerase [17].

There are also TRF1 and TRF2 associated factors. The main factor associated to TRF1 is tankyrase, a positive regulator of telomere length [18]. PINX1 is a TRF1-associated telomerase inhibitor, which associates with TRF1 [19]. PINX1 a negative regulator of telomere length is able to simultaneously interact with the telomerase catalytic subunit providing the enzyme a physical link with TRF1 [20].

There is also a physical link between the human shelterin complex and telomerase providing new insight into the mechanism of processive telomere synthesis. [21].

In most mammals, the maintenance of telomeric length is carried out mainly by telomerase. The human holoenzyme telomerase is a ribonucleoprotein composed by a catalytic subunit, hTERT and an RNA component [hTR] which acts as a template for the addition of a short repetitive sequence [dTTAGGG]n in the 3' end of the telomeric DNA and species-specific accessory proteins. These accessory proteins regulate telomerase biogenesis, subcellular localization and

^{*}Address correspondence to this author at the Laboratory of Molecular Oncology, Department of Science and Technology. Quilmes National University, Bernal, Buenos Aires, Argentina. R. Saenz Peña 352, (1876) Buenos Aires, Argentina; Tel: 54 11 43657100 ext 5644; E-mail: degomez@unq.edu.ar

function *in vivo*. For instance, analysis of affinity-purified telomerase from HeLa cells has identified integral protein components of human telomerase: dyskerin, NHP2; NOP10, pontin/reptin, Gar1 and TCAB1 [22] Fig. (1). As described the complex telomere/telomerase is integrated by numerous molecules with different functions elegantly reviewed by Rubtsova *et al* [23].



Fig. (1). Schematic representation of telomerase and its associated proteins.

1.2. Telomerase Inhibiting Strategies

As we just observed the complexity of the telomere/telomerase complex, we can understand that there is a wide variety of strategies to inhibit telomerase. This complexity allowed the development of several inhibitors and paves the way to the development of new ones. Although the numerous strategies and molecules can be classified in different ways, we choose to do so, based in the general approach to inhibition and then analyzing each molecule belonging to that group, but also understanding that one molecule can belong to more than one category

1.2.1. Nucleosides

3-Azido-2,3 -dideoxythymidine [azidothymidine [AZT] or zidovudine] was the first reported telomerase inhibitor Fig. (2A) The similarity between HIV retrotranscriptase and telomerase led to the discovery that AZT was preferentially integrated into the telomeric region of CHO DNA [24]. Similar results, but by quantitative methods were found by us also [25]. Later, different groups demonstrated that AZT inhibited telomerase and/or reduce telomerase length [26, 27]. Moreover, we demonstrated that telomere shortening by AZT was an irreversible process, [28]. Similar results were founded by other researchers. [29, 30]. Similarly, synergistic interactions between paclitaxel and AZT [31] and between AZT and 5-fluorouracil [32] were described. In 2001, we found that chronic in vitro AZT exposure on F3II mouse mammary carcinoma cells with 800 µM AZT for at least 30 passages completely inhibited telomerase activity on F3II mammary carcinoma cells, leading to senescence and apoptosis [33], also corroborated by other authors [34]. Azidothymidine is used to treat several virus-associated human cancers [35]. In non-viral tumors, AZT has been used in phase I and II clinical trials alone or in combination for different solid tumors showing some rate of regression [36]. More clinical trials using AZT are needed to understand the full potential of this agent in a clinical setting.

Other nucleosides have been studied as potential inhibitors of telomerase. It has been demonstrated that carbovir, induced senescence-like processes in cultures of immortal mouse fibroblasts [37]. Also, it was reported that both Azdd GTP and C.OXT-GTP, the triphosphate derivatives of 3azido-2,3-dideoxyguanosine [AZddG] and carbocyclic oxetanocion G [C.OXT-g] showed potent telomerase-inhibitory activity and induce telomere shortening in human HL60 cells [38]. Later on, the same group found that AZddAA caused telomere shortening in the same model [39]. Tendian et al studied the interaction of five doxyguanosine nucleotide analogs, 6-thio-2-deoxyguanosine, 5-triphosphate [T-dGTP], 5triphosphate of carbovir [CBV-TP], ddGTP, D-carbocyclic-2-deoxyguanosine 5-triphosphate [D-CdG-TP] and Lcarbocyclic 2.deoxyguanosine 5-triphosphate [L-CdG-TP]. T-dGTP is the active metabolite of both 6-mercaptopurine and 6-thioguanine, which are two drugs used in the treatment of acute leukemia. CBV-TP is the active metabolite of Abacavir, an agent approved for the treatment of AIDS and D-CdG-TP is the active metabolite of D-CdG, an agent with activity against herpes simplex virus, cytomegalovirus, and hepatitis-B virus, founding that all of them inhibited telomerase activity by 50% [40]. Numerous acyclic nucleoside phosphonates [ANPs] possess excellent antiviral activities against a broad spectrum of DNA viruses and retroviruses as well as significant antiproliferative potency. In cells, ANPs are phosphorylated to their diphosphates active antimetabolites, which inhibit viral and/or cellular replicases and terminate nascent DNA chain. The group of Hajec analyzed the antitelomerase activity of 15 of these diphosphates of ANPs and found that the most effective compound studied was the guanine derivative PMEGpp [41]. It has been patented that acyclic nucleoside analogs such us acyclovir, ganciclovir, penciclovir and the corresponding pro-drugs, i.e., valacyclovir, valganciclovir and famciclovir, respectively have been identified as inhibitors or antagonists of telomerase [42]. The telomerase inhibitory effects of purine nucleosides bearing a 3'-down azido group were also investigated. It was found that AZddGTP is a selective inhibitor of telomerase, producing a reproducible telomere shortening [43]. In 2001, a potent telomerase inhibiting nucleoside was developed: 6-thio-7-deaza-2'-deoxyguanosine 5'-triphosphate [TDG-TP] with a low and high specificity [44]. Previously, the same group found human telomerase inhibition by 7-deaza-2'deoxypurine nucleoside triphosphates using a cell-free biochemical telomerase assay [45].

1.2.2. Oligonucleotides

Feng et al reported that antisense oligonucleotides complementary to sequences within or near the human telomeric template RNA resulted in suppression of telomerase activity while antisense oligonucleotides against targets that were more distant from the telomeric template failed to inhibit the action of the ribonucleoprotein [46]. The advances in antisense technology have led to improvements in the introduction of the molecules into cells, stability, lengthening of halflife, and specificity of target binding. Modifications of traditional antisense oligonucleotides used in telomerase inhibition include 2'-5'-oligoadenylate, 2'-O-methyl-RNA, phosphorothioate-modified oligodeoxynucleotides [PS-ODN], peptide nucleic acids [PNA] (Fig. **2B**), and locked nucleic acids [LNA] [47]. PU27 is a sequence specific DNA oligoclenucleotide currently in preclinical stage for a wide variety of tumor type including leukemia, prostate cancer, renal cancer and breast cancer. PU27 has shown growth inhibitory effect because of its ability to bind and inhibit the enzymatic activity of a-enolase. It also demonstrated altered oncogene expression and inhibition of telomerase activity thus selectively inhibit cancer cell metabolism and cell growth. [48]

1.2.3. Small Molecule Inhibitors

Among many small molecules tried with different success aiming to inhibit telomerase activity BIBR1532 [2-[E]-3-naphthalene-2-yl-but-2-enoylylamino]- benzoic acid] is the best known Fig. (**2C**). It is a non-competitive inhibitor of TERT and hTR that *in vitro* reduces telomere length, inhibits cell proliferation, producing finally cell senescence [49]. Although good results have been observed in preclinical studies on breast, prostate and fibrosarcoma cancer cell lines no further progress or entrance in clinical trials has been shown. In the last years, BIBR1532 have been used as a tool to inhibit telomerase and demonstrating in that way that decreases alpha-fetoprotein expression [50] and was also demonstrated that glucose restriction increase the activity of this inhibitor [51].

1.2.4. Stabilization of G Cuadruplexes

As explained in the telomere structure section, one indirect path to inhibit telomerase activity could be the stabilization of the G cuadruplexes preventing hTR of recognizing the unfolded single-stranded telomere overhang. Most of these molecules contain a polycyclic heteroaromatic structure. In this group stands telomestatin (Fig. 2D) [52], RHPS4 and BRACO19 [53]. Although effective they were poorly soluble or inefficient to cross biological barriers [54], therefore reducing their clinical significance. An extensive research has been carried out in the modification of telomestatin to increase its potency. More recently a series of macrocyclic molecules [telomestatin analogues] have been developed, with improved features over telomestatin parental structure [55]. Macrocyclic hexaoxazole L2H2-6M(2)OTD is one of the derivatives of telomestatin that interact with Gquadruplex by p-stacking and electrostatic interactions [56]. Telomestatin is currently under clinical trials [57].

Daunomycin is basically an anthracyclin isolated from *Streptomyces peucetius* and it is well known for its DNA intercalation and G-quadruplex stabilization.

Distamycin A was isolated from *Streptomyces distallicus*. Distamycin-A stacks on the terminal G-quadrets and interacts with the flanking bases [58]. Distamycin inhibits protein interactions with G-quadruplex DNA. The first report of telomerase inhibitory activity of distamycin derivatives was by Zaffaroni *et al.* They tested the antagonistic activity of MEN 10,716, a derivative of distamycin in JR8 melanoma cell extracts [59]. Chemical modification of this compound has been carried out extensively to increase the potency of inhi-

bition. A study shows that introduction of more number of pyrrole groups allows binding with mixed groove/G-quartet in a stacking mode [60]. Some other compounds more water-soluble have been developed such as quarfloxin, quarflox-in/CX3543 [61] and RHPS4 [62]. Quarfloxin has reached phase II clinical trials, although results are not available.

Ascididemin and Meridine are two marine compounds with pyridoacridine skeletons known to stabilize Gquadruplexes and inhibit telomerase *in vitro*. [63]

The interaction of berberine and 9 different berberine derivatives with human telomeric DNA indicated that these compounds could induce and stabilize the formation of antiparallel G-quadruplex of telomeric DNA. Compared with berberine, the derivatives exhibit stronger binding affinity with G-quadruplex and higher inhibitory activity for telomerase [64]

A cryptolepine derivative containing indole and quinoline structures, SYUIQ-5 has been reported to induce and stabilize G-quadruplex, inhibiting c-myc promoter and telomerase activity [65]

In addition, cationic porphyrins are being studied as possible telomerase inhibitors due to their ability to bind and stabilize G-quadruplexes. The best studied molecule of this group is the cationic porphyrin TMPyP4. [66]

1.2.5. Immunotherapy

Many immunotherapeutic approaches are under development, either at preclinical or clinical levels [67]. Basically, antitelomerase immunotherapy sensitizes immune cells to tumor cells expressing hTERT peptides as surface antigens via the human leukocyte antigen [HLA] class I pathway. Some 26 different hTERT peptides have been utilized to generate an antitelomerase immune response, many of them showing good preclinical and clinical results [68]. In clinical assays, different peptides produce a good immunological response with low toxicity and some promising results were published. For instance, Vx001 and I540 produced in responsive patients, a longer survival time than in those that were non-responsive [69]. However, biomarkers or indicators to point out which patients are going to be responsive remain to be developed.

Many clinical trials are currently ongoing with immunological peptides either alone or in combination. In phase I and phase II clinical trials: GRNVAC1, TERT and Survivin peptide loaded dendritic cells and dendritic cells transfected with TERT, surviving and p53 mRNA [70]. A promising vaccine is GV1001 [Tertomotide] Fig. (2E). This peptide vaccine consists of 15 amino acid epitope of hTERT. It generates telomerase-specific T-helper cells, activates antigenpresenting cells and cytotoxic T cells, generating a good immune response and has successfully already completed several phase I and II clinical trials either alone or in combination with the alkylating agent Temozolomide. Currently has reached phase III clinical trials [Telovac] for non-small cell lung cancer and one NDA were filed for pancreatic cancer. Unfortunately, there was no significant difference in overall survival between the groups that received the vaccine and the control group receiving chemotherapy [71]. Another vaccine currently going through phase I clinical trial for hormone refractory prostate cancer is TeloB-VAX. It is

Telomerase Inhibitors

composed of the patients' own circulating B-lymphocytes harboring a unique patented engineered plasmid DNA belonging to Adamis Pharmaceutical Corporation [72]. Another vaccine being studied by VAXON Biotech is Vx-001, composed of two separate peptides: the native cryptic peptide ARG-Vx001 [TERT572] and its optimized variant TYR-Vx001 [TERT572Y]. The study included *in vivo* experiments in mice, *in vitro* experiments on human lymphocytes, and a phase I/II clinical trial. Vx-001 vaccination of humanized mice protects them against tumor growth *in vivo* [73]. Furthermore, Vx-001 induces anti-tumor immune responses by human lymphocytes *in vitro*. Vx-001 has completed a phase I/II trial with 33 patients with NSCLC [74] demonstrating its safeness and tolerance and a strongly immunogenic response in 70% of patients.

Vx-001 entered a randomized phase IIb clinical trial in HLA-A*0201 positive patients with TERT expressing NSCLC [stage IV and distant recurrent stage I-III] who controlled disease after first line chemotherapy. Results are expected at the end of 2016. [75]

Peptide540-548, peptide611-626, peptide672-686 and peptide766-780, which are derived from human telomerase, constitute the immunogenic component of the GX301 cancer vaccine which is being tested in phase II clinical trial for prostate cancer [76]

1.2.6. Gene Therapy

Labs and companies have been working for a very long time to bring gene therapy to the clinic, yet very few patients have received any effective gene-therapy treatment. However, gene therapy is also a strategy used in the quest for targeting telomerase. Probably the best-known molecule is the antisense oligonucleotide Imetelstat or GRN163L [Geron Corporation] (Fig. 2F), a lipid-conjugated 13-mer oligonucleotide sequence that is complementary to hTR that showed good in vivo and vitro results [77]. The molecule demonstrates high resistance to cellular nucleases, which confers stability in plasma and tissues. Such results led to a number of phases I and II clinical trials either with Imetelstat alone or in combination for multiple oncology and hematologic myeloid malignancies indications. [78]. Interestingly, it has been demonstrated that Imetelstat could cross the bloodbrain barrier. Trials showed good results with the exception of a phase II clinical trial using Imetelstat plus paclitaxel in advanced breast cancer. This trial was stopped in September 2012 due to the results of an interim analysis showing a worse survival time in patients receiving Imetelstat. On November 13, 2014, Geron entered into an exclusive collaboration and license agreement with Janssen Biotech. Since then development of Imetelstat will proceed under a mutually agreed clinical development plan, which includes two phase II studies to be pursued initially, one in myelofibrosis, and one in myelodysplastic syndrome expected to be initiated during 2015.

Other approach involves "suicide gene therapy", viral vectors that are genetically modified to encode a prodrug activating enzyme [i.e. cytosine deaminase or carboxypeptidase G2] which in turn will replicate only in TERT-overexpressing cells, activating the effect of cytotoxic prodrugs like 5-flucytosine or ZD2767P [79].

Furthermore, other strategy has already reached the clinical phases. Telomelysin is an attenuated adenovirus-5 vector whereas TERT promoter element drives expression of E1A and B genes linked with and internal ribosome entry site. In this way, it induces virus-mediated lysis of cancer cells after viral propagation in the TERT-overexpressing cells. The drug is in phase I/II development stage for hepatocellular carcinoma and esophageal cancer [80]

1.2.7. Targeting Telomere and Telomerase-Associated Proteins

One interesting strategy is targeting the associated proteins rather than the main molecules. One interesting case is targeting tankyrases with PARP inhibitors. Also is interesting the approach on inhibition of the chaperone HSP90. Studies show that HSP90-p23 co-chaperone complex is required for maturation and activation of telomerase [78]. With that idea on mind, Geldanamycin [GA] (Fig. **2G**) was used. However since HSP90-P23 has low solubility and high hepatotoxicity, the analogs 17-AAG [Tanespimycin] and 17-DAG were developed and are being tested in clinical trials at the moment [81]. Small interfering small RNAs having as a target TRF1, TRF2 and TIN2 have been studied. Some molecules against POT1 also have been analyzed [82].

1.2.8. Telomerase Inhibitors from Microbial Sources

Telomerase inhibitors were isolated from various fungal, bacterial and actinomycetes sources (for review see [83]. Some of them are chemically modified in order to increase their potency and some were synthesized as such in the laboratory [84].

Actinomycetes spp. is the most widely explored microorganism for telomerase inhibitors since possesses benzofuran and benzodipyran rings, which have been found to be potential inhibitors of telomerase. Rubromycins (Fig. 2H) isolated from Streptomyces collinus are extensively studied for their ability to induce apoptosis in cancer cells; however, their telomerase inhibitory activity was explored recently. They are primarily aromatic naphthoquinone and isocoumarin ring systems that competitively interact with the hTERT and/or hTR subunits of telomerase enzyme. Studies proved that the spiroketal moiety of rubromycin is the key pharmacophore for telomerase inhibitory action [85]. Griseorhodins are another group of compounds that possess quinine moieties and inhibit telomerase in vitro. Fungi also have become sources of telomerase inhibitors among them Thelavin A and B, which are isolated from Thielavia terricola [86] and diazaphilonic acid, isolated from Talaromyces flavus [87].

1.3. Other Molecules

Oleic acid is a fatty acid that occurs naturally in various animal and vegetable fats and oils Fig. (21). In chemical terms, oleic acid is classified as a monounsaturated omega-9 fatty acid and was found to be inhibitor of human telomerase [88]. Helenalin, a natural sesquiterpene lactone, is a potent and selective inhibitor for human telomerase [89]. Five new alkaloids, dictyodendrins A-E were isolated from the marine sponge *Dictyodendrilla verongiformis* as telomerase inhibitors. Dictyodendrins are tyramine-based pyrrolocarbazole derivatives containing three or four p-hydroxybenzene



Fig. (2). a) Structure of the most important inhibitory molecules belonging to each group. A) Nucleosides. B) Oligonucleotides. C) Small molecule inhibitors. D) Stabilizators of G quadruplex. E) Immunotherapeutic molecules. F) Gene therapy constructs. G) Molecules that target telomere and telomerase associated proteins. H) Inhibitors from microbial sources. I) Other inhibitors.

b). Mechanism of action of the most important inhibitory molecules belonging to each group. A) AZT: Integrates into the telomeric DNA. B) PNA: This modified antisense oligonucleotide is complementary to sequences within or near the human telomeric template. C) BIBR1532: Competiting inhibitor of TERT and hTR. D) Telomestatin: stabilizes G cuadruplexes preventing hTR of recognizing the unfolded single stranded telomere overhang. E) Tertomide Generates telomerase specific T helper cells, activates antigen presenting cells and cytotoxic T cells, generating a good immune response. F) Imetelstat: A lipid=conjugated 13=mer oligonucleotide sequence that is complementary to hTR. G) Gedanamycin: targets the HSP90.P23 co.chaperone complex, required for maturation and activation of telomerase. H) Rubromycin: competitive interact with the hTERT and or hTR subunits of telomerase enzyme. I) Oleic acid. The three-dimensional structure of the active site of telomerase (i.e., the binding site of the primer and dNTP substrate) might have a "pocket" that could "join" these compounds.

groups. They inhibited telomerase completely at a concentration of 50 microg/mL [90].

CONCLUSION

As has been analyzed many approaches and agents have been directed against telomerase, then it is time to analyze the challenges and perspectives we have ahead.

Although at the moment no therapy has been used as oncology treatment, telomerase inhibition is still one of the best targets to point out in oncology. Let us no forget that telomerase was discovered only 30 years ago and that its relation with cancer was established 20 years ago. Also, as with the years have passed and increasing body of evidence has demonstrated the complex relationship between telomere/telomerase at the different levels of regulation of telomerase activity and with their relationship with associated proteins. Interestingly, those associated proteins could be an excellent target in our fight against cancer.

We should take also into account some possible shortcomings. For instance, in the case of AZT, as the shortening of telomeres is a slow process, the dynamics of the disease could put at risk the life of the patient before the action of the drug is effective. Therefore, concerns must be expressed when attempting to treat advanced tumors with AZT [91]. However, AZT treatment could constitute a good adjuvant therapy in cases where conventional treatments reduce the bulk of the tumor giving time for AZT to act in the remnant surviving tumor cells.

Firstly, the number of AZT-treated passages could be insufficient for a senescence program to be triggered, and secondly telomeres shortening to a critical length could induce a compensatory mechanism of preservation so preventing further losses, known as alternative lengthening of telomeres or ALT [92]. Thirdly, an AZT-resistant phenotype could have developed because of selection following treatment.

However, one of the advantages of telomerase targeting therapies is that rapidly proliferating cancer cells have shorter telomeres [5kb] compared to normal somatic cells and stem cells [10-20 kb] that have not yet reached critical lengths [78]. Some authors consider that in some cases, functional p53 may be required to induce the response to telomerase inhibitors in cells with critically shortened telomeres. [93]. Other aspect to consider is that clinical results many times are not successful due to poor pharmacokinetics: limited solubility, difficulties to pass through biological barriers, etc [94] which leads to the search of other solubilizers or carriers.

In addition, we should consider that some of these inhibitors start their tumor deleterious effect after a variable amount of time. After telomerase inhibition, the telomere will start to become shorter, but tumor senescence and death will only start when reaching a critical length. Most often patients who join phase I clinical trials have advanced [metastatic] cancer leaving this kind of inhibitor without the chance of demonstrating its effectivity in less advanced tumors. Some authors have mentioned the importance of founding a "window of opportunity" for these inhibitors. Clearly, that will be the case of smaller tumors that requires a bigger number of mitosis, allowing the inhibitors to exert its action. Some other authors are suggesting changes in clinical trials policies to allow this kind of molecules to have the chance to demonstrate its effectiveness without compromising the safety of the patient or the seriousness of the trial. Although clinical trials are, the basis where daily clinical practice should be based on, such evidence is scarce at the end-of life of cancer patients. Research in this patient's population is hampered by the lack of clear definition of the study population, the study design, the definition of meaningful endpoints and ethical considerations [95] In the meantime, some authors have advanced the path of therapy combination with established oncological treatments. This approach is promising since it is tested as maintenance/consolidation treatments to prolong remission in patients with advanced cancers. Some examples are combinations with radiotherapy [96], trastuzumab [97], paclitaxel [98], doxorubicin [99], docetaxel [100] and etoposide [101]. Since many telomerase targeting molecules have a long lag time to produce critically shorter telomeres a combination therapy of telomerase inhibitors and standard of care may be the best approach to target effectively cancer cells.

With their advantages and pitfalls, telomerase inhibition remains as one of the hottest targets in the quest for new antitumor drugs. More research in the subject will guarantee the answers to our questions, and eventually the finding of a blockbuster molecule.

CONFLICT OF INTEREST

None of the author of this paper has a financial or personal relationship with other people that could inappropriately influence or bias the content of the paper. Dr. Mengual Gomez, Dr.Armando and Dr.Gomez were founded for the study by the Ministry of Science and Technology and Quilmes National University. Dr. Cerrudo and Dr. Ghiringhelli were founded by Quilmes National University.

ACKNOWLEDGEMENTS

Dr. Mengual Gomez and Dr. Armando contributed to the selection and analysis of the literatures as well as with the design. Dr. Cerrudo and Dr. Ghiringhelli provided careful analysis of the molecules enumerated as inhibitors. Dr. Gomez was in charge of the design and general direction of the paper. The authors want to express their gratitude to DelveInsight for helping us to obtain important and significant information for this work.

REFERENCES

- Greider, C.W.; Blackburn, E.H. Identification of a specific telomere terminal transferase activity in Tetrahymena extracts. *Cell*, 1985, 43(2 Pt 1), 405-413.
- [2] Kim, N.W.; Piatyszek, M.A.; Prowse, K.R.; Harley, C.B.; West, M.D.; Ho, P.L.; Coviello, G.M.; Wright, W.E.; Weinrich, S.L.; Shay, J.W. Specific association of human telomerase activity with immortal cells and cancer. *Science*, **1994**, *266*(5193), 2011-2015.
- [3] Mocellin, S.; Pooley, K.A.; Nitti, D. Telomerase and the search for the end of cancer. *Trends Mol. Med.*, **2013**, *19*(2), 125-133.
- [4] Giraud-Panis, M-J.; Pisano, S.; Poulet, A.; Le Du, M-H.; Gilson, E. Structural identity of telomeric complexes. *FEBS Lett.*, 2010, 584(17), 3785-3799.
- [5] Nersisyan, L.; Arakelyan, A. Computel: computation of mean telomere length from whole-genome next-generation sequencing data. *PLoS One*, **2015**, *10*(4), e0125201.

- [6] Meyne, J.; Ratliff, R.L.; Moyzis, R.K. Conservation of the human telomere sequence (TTAGGG)n among vertebrates. *Proc. Natl. Acad. Sci. USA*, **1989**, *86*(18), 7049-7053.
- [7] Wright, W.E.; Tesmer, V.M.; Huffman, K.E.; Levene, S.D.; Shay, J.W. Normal human chromosomes have long G-rich telomeric overhangs at one end. *Genes Dev.*, **1997**, *11*(21), 2801-2809.
- [8] Hänsel, R.; Löhr, F.; Foldynová-Trantírková, S.; Bamberg, E.; Trantírek, L.; Dötsch, V. The parallel G-quadruplex structure of vertebrate telomeric repeat sequences is not the preferred folding topology under physiological conditions. *Nucleic Acids Res.*, 2011, 39(13), 5768-5775.
- [9] Gomez, D.E.; Armando, R.G.; Farina, H.G.; Menna, P.L.; Cerrudo, C.S.; Ghiringhelli, P.D.; Alonso, D.F. Telomere structure and telomerase in health and disease (review). *Int. J. Oncol.*, 2012, 41(5), 1561-1569. [review].
- [10] Palm, W.; de Lange, T. How shelterin protects mammalian telomeres. Annu. Rev. Genet., 2008, 42, 301-334.
- [11] Martínez, P.; Blasco, M.A. Telomeric and extra-telomeric roles for telomerase and the telomere-binding proteins. *Nat. Rev. Cancer*, 2011, 11(3), 161-176.
- [12] Smogorzewska, A.; van Steensel, B.; Bianchi, A.; Oelmann, S.; Schaefer, M.R.; Schnapp, G.; de Lange, T. Control of human telomere length by TRF1 and TRF2. *Mol. Cell. Biol.*, 2000, 20(5), 1659-1668.
- [13] van Steensel, B.; de Lange, T. Control of telomere length by the human telomeric protein TRF1. *Nature*, **1997**, 385(6618), 740-743.
- [14] Lee, C.C.; Huang, T.S. A novel topoisomerase II poison GL331 preferentially induces DNA cleavage at (C/G)T sites and can cause telomere DNA damage. *Pharm. Res.*, 2001, 18(6), 846-851.
- [15] Tejera, A.M.; Stagno d'Alcontres, M.; Thanasoula, M.; Marion, R.M.; Martinez, P.; Liao, C.; Flores, J.M.; Tarsounas, M.; Blasco, M.A. TPP1 is required for TERT recruitment, telomere elongation during nuclear reprogramming, and normal skin development in mice. *Dev. Cell*, **2010**, *18*(5), 775-789.
- [16] Schoeftner, S.; Blasco, M.A. Developmentally regulated transcription of mammalian telomeres by DNA-dependent RNA polymerase II. *Nat. Cell Biol.*, 2008, 10(2), 228-236.
- [17] Nergadze, S.G.; Farnung, B.O.; Wischnewski, H.; Khoriauli, L.; Vitelli, V.; Chawla, R.; Giulotto, E.; Azzalin, C.M. CpG-island promoters drive transcription of human telomeres. *RNA*, 2009, *15*(12), 2186-2194.
- [18] Hsiao, S.J.; Smith, S. Tankyrase function at telomeres, spindle poles, and beyond. *Biochimie*, 2008, 90(1), 83-92.
- [19] Chen, Y.; Yang, Y.; van Overbeek, M.; Donigian, J.R.; Baciu, P.; de Lange, T.; Lei, M. A shared docking motif in TRF1 and TRF2 used for differential recruitment of telomeric proteins. *Science*, 2008, 319(5866), 1092-1096.
- [20] Zhou, X.Z.; Lu, K.P. The Pin2/TRF1-interacting protein PinX1 is a potent telomerase inhibitor. *Cell*, 2001, 107(3), 347-359.
- [21] Kibe, T.; Osawa, G.A.; Keegan, C.E.; de Lange, T. Telomere protection by TPP1 is mediated by POT1a and POT1b. *Mol. Cell. Biol.*, **2010**, *30*(4), 1059-1066.
- [22] Fu, D.; Collins, K. Purification of human telomerase complexes identifies factors involved in telomerase biogenesis and telomere length regulation. *Mol. Cell*, **2007**, 28(5), 773-785.
- [23] Rubtsova, M.P. Vasilkova, D.P.; Malyavko, A.N.; Naraikina, Yu.V., Zvereva, M.I.; Dontsova. O.A. Telomere Lengthening and Other Functions of Telomerase. Acta Naturae., 2012, 4(2), 44-61.
- [24] Olivero, O.A.; Poirier, M.C. Preferential incorporation of 3'-azido-2',3'-dideoxythymidine into telomeric DNA and Z-DNA-containing regions of Chinese hamster ovary cells. *Mol. Carcinog.*, **1993**, 8(2), 81-88.
- [25] Gomez, D.; Kassim, A.; Olivero, O. Preferential incorporation of 3'-azido-2',3'-dideoxythymidine (azt) in telomeric sequences of cho cells. *Int. J. Oncol.*, **1995**, 7(5), 1057-1060.
- [26] Strahl, C.; Blackburn, E.H. Effects of reverse transcriptase inhibitors on telomere length and telomerase activity in two immortalized human cell lines. *Mol. Cell. Biol.*, **1996**, *16*(1), 53-65.
- [27] Yegorov, Y.E.; Chernov, D.N.; Akimov, S.S.; Bolsheva, N.L.; Krayevsky, A.A.; Zelenin, A.V. Reverse transcriptase inhibitors suppress telomerase function and induce senescence-like processes in cultured mouse fibroblasts. *FEBS Lett.*, **1996**, *389*(2), 115-118.
- [28] Gomez, D.E.; Tejera, A.M.; Olivero, O.A. Irreversible telomere shortening by 3'-azido-2',3'-dideoxythymidine (AZT) treatment. *Biochem. Biophys. Res. Commun.*, **1998**, 246(1), 107-110.

- [29] Multani, A.S.; Furlong, C.; Pathak, S. Reduction of telomeric signals in murine melanoma and human breast cancer cell lines treated with 3'-azido-2'-3'-dideoxythymidine. *Int. J. Oncol.*, **1998**, *13*(5), 923-925.
- [30] Murakami, J.; Nagai, N.; Shigemasa, K.; Ohama, K. Inhibition of telomerase activity and cell proliferation by a reverse transcriptase inhibitor in gynaecological cancer cell lines. *Eur. J. Cancer*, **1999**, 35(6), 1027-1034.
- [31] Johnston, J.S.; Johnson, A.; Gan, Y.; Wientjes, M.G.; Au, J.L. Synergy between 3'-azido-3'-deoxythymidine and paclitaxel in human pharynx FaDu cells. *Pharm. Res.*, 2003, 20(7), 957-961.
- [32] Brown, T.; Sigurdson, E.; Rogatko, A.; Broccoli, D. Telomerase inhibition using azidothymidine in the HT-29 colon cancer cell line. Ann. Surg. Oncol., 2003, 10(8), 910-915.
- [33] Tejera, A.M.; Alonso, D.F.; Gomez, D.E.; Olivero, O.A. Chronic in vitro exposure to 3'-azido-2', 3'-dideoxythymidine induces senescence and apoptosis and reduces tumorigenicity of metastatic mouse mammary tumor cells. Breast Cancer Res. Treat., 2001, 65, 93-99. Jeng et al., 2011
- [34] Datta, A.; Bellon, M.; Sinha-Datta, U.; Bazarbachi, A.; Lepelletier, Y.; Canioni, D.; Waldmann, T.A.; Hermine, O.; Nicot, C. Persistent inhibition of telomerase reprograms adult T-cell leukemia to p53-dependent senescence. *Blood*, 2006, 108(3), 1021-1029.
- [35] Falcone, A.; Lencioni, M.; Brunetti, I.; Pfanner, E.; Allegrini, G.; Antonuzzo, A.; Andreuccetti, M.; Malvaldi, G.; Danesi, R.; Del Tacca, M.; Conte, P.F. Maximum tolerable doses of intravenous zidovudine in combination with 5-fluorouracil and leucovorin in metastatic colorectal cancer patients. Clinical evidence of significant antitumor activity and enhancement of zidovudine-induced DNA single strand breaks in peripheral nuclear blood cells. *Ann. Oncol.*, **1997**, 8(6), 539-545.
- [36] Jordheim, L.P.; Durantel, D.; Zoulim, F.; Dumontet, C. Advances in the development of nucleoside and nucleotide analogues for cancer and viral diseases. *Nat. Rev. Drug Discov.*, **2013**, *12*(6), 447-464.
- [37] Hukezalie, K.R.; Thumati, N.R.; Côté, H.C.; Wong, J.M. *In vitro* and ex vivo inhibition of human telomerase by anti-HIV nucleoside reverse transcriptase inhibitors (NRTIs) but not by non-NRTIs. *PLoS One*, **2012**, 7(11), e47505.
- [38] Yamaguchi, T.; Takahashi, H.; Jinmei, H.; Takayama, Y.; Saneyoshi, M. Inhibition of vertebrate telomerases by the triphosphate derivatives of some biologically active nucleosides. *Nucleosides Nucleotides Nucleic Acids*, 2003, 22(5-8), 1575-1577, 1575-1577.
- [39] Yamaguchi, T.; Liu, X.; Ogawara, T.; Inomata, M.; Saneyoshi, M. Telomerase inhibition by 3'-azido-2', 3'-dideoxynucleoside 5'triphosphates and telomere shortening in human cultured cells by the corresponding nucleosides. *Nucleic Acids Symp Ser (Oxf)*, 2006, 50(50), 271-272.
- [40] Tendian, S.W.; Parker, W.B. Interaction of deoxyguanosine nucleotide analogs with human telomerase. *Mol. Pharmacol.*, 2000, 57(4), 695-699.
- [41] Hájek, M.; Matulová, N.; Votruba, I.; Holý, A.; Tloust'ová, E. Inhibition of human telomerase by diphosphates of acyclic nucleoside phosphonates. *Biochem. Pharmacol.*, 2005, 70(6), 894-900.
- [42] Bondarev, I. Pharmacological modulation of telomere length in cancer cells for prevention and treatment of cancer. U.S. Patent 8609623 B2 2006 May 18;
- [43] Liu, X.; Takahashi, H.; Harada, Y.; Ogawara, T.; Ogimura, Y.; Mizushina, Y.; Saneyoshi, M.; Yamaguchi, T. 3'-Azido-2',3'dideoxynucleoside 5'-triphosphates inhibit telomerase activity *in vitro*, and the corresponding nucleosides cause telomere shortening in human HL60 cells. *Nucleic Acids Res.*, 2007, 35(21), 7140-7149.
- [44] Fletcher, T.M.; Cathers, B.E.; Ravikumar, K.S.; Mamiya, B.M.; Kerwin, S.M. Inhibition of human telomerase by 7-deaza-2'deoxyguanosine nucleoside triphosphate analogs: potent inhibition by 6-thio-7-deaza-2'-deoxyguanosine 5'-triphosphate. *Bioorg. Chem.*, 2001, 29(1), 36-55.
- [45] Fletcher, T.M.; Salazar, M.; Chen, S.F. Human telomerase inhibition by 7-deaza-2'-deoxypurine nucleoside triphosphates. *Biochemistry*, **1996**, *35*(49), 15611-15617.
- [46] Feng, J.; Funk, W.D.; Wang, S.S.; Weinrich, S.L.; Avilion, A.A.; Chiu, C.P.; Adams, R.R.; Chang, E.; Allsopp, R.C.; Yu, J.; Allsopp, R.C. The RNA component of human telomerase. *Science*, 1995, 269(5228), 1236-1241.
- [47] Cunningham, A.P.; Love, W.K.; Zhang, R.W.; Andrews, L.G.; Tollefsbol, T.O. Telomerase inhibition in cancer therapeutics: mo-

lecular-based approaches. Curr. Med. Chem., 2006, 13(24), 2875-2888.

- [48] Islam, M.A.; Thomas, S.D.; Murty, V.V.; Sedoris, K.J.; Miller, D.M. c-Myc quadruplex-forming sequence Pu-27 induces extensive damage in both telomeric and nontelomeric regions of DNA. J. Biol. Chem., 2014, 289(12), 8521-8531.
- [49] Pascolo, E.; Wenz, C.; Lingner, J.; Hauel, N.; Priepke, H.; Kauffmann, I. GarinChesa P.; Rettig, W.J.; Damm, K.; Schnapp, A. Mechanism of human telomerase inhibition by BIRB1532, a synthetic, non-nucleosidic drug candidate. *J. Biol. Chem.*, **2002**, *277*, 15566-15572.
- [50] Tahtouh, R.; Azzi, A-S.; Alaaeddine, N.; Chamat, S.; Bouharoun-Tayoun, H.; Wardi, L.; Raad, I.; Sarkis, R.; Antoun, N.A.; Hilal, G. Telomerase inhibition decreases alpha-fetoprotein expression and secretion by hepatocellular carcinoma cell lines: *in vitro* and *in vivo* study. *PLoS One*, **2015**, *10*(3), e0119512.
- [51] Wardi, L.; Alaaeddine, N.; Raad, I.; Sarkis, R.; Serhal, R.; Khalil, C.; Hilal, G. Glucose restriction decreases telomerase activity and enhances its inhibitor response on breast cancer cells: possible extra-telomerase role of BIBR 1532. *Cancer Cell Int.*, **2014**, *14*, 60.
- [52] Kim, M-Y.; Vankayalapati, H.; Shin-Ya, K.; Wierzba, K.; Hurley, L.H. Telomestatin, a potent telomerase inhibitor that interacts quite specifically with the human telomeric intramolecular g-quadruplex. *J. Am. Chem. Soc.*, **2002**, *124*(10), 2098-2099.
- [53] Gomez, D.; O'Donohue, M-F.; Wenner, T.; Douarre, C.; Macadré, J.; Koebel, P.; Giraud-Panis, M.J.; Kaplan, H.; Kolkes, A.; Shin-ya, K.; Riou, J.F. The G-quadruplex ligand telomestatin inhibits POT1 binding to telomeric sequences *in vitro* and induces GFP-POT1 dissociation from telomeres in human cells. *Cancer Res.*, 2006, 66(14), 6908-6912.
- [54] Taetz, S.; Baldes, C.; Mürdter, T.E.; Kleideiter, E.; Piotrowska, K.; Bock, U.; Haltner-Ukomadu, E.; Mueller, J.; Huwer, H.; Schaefer, U.F.; Klotz, U.; Lehr, C.M. Biopharmaceutical characterization of the telomerase inhibitor BRACO19. *Pharm. Res.*, **2006**, *23*(5), 1031-1037.
- [55] Granzhan, A.; Monchaud, D.; Saettel, N.; Guédin, A.; Mergny, J-L Marie-Paule Teulade-Fichou, M-P. "One Ring to Bind Them All"—Part II: Identification of Promising G-Quadruplex Ligands by Screening of Cyclophane-Type Macrocycles Journal of Nucleic Acids. 2010, [2010], 460561, 1-19. doi:10.4061/2010/525862.
- [56] Chung, W.J.; Heddi, B.; Tera, M.; Iida, K.; Nagasawa, K.; Phan, A.T. Solution structure of an intramolecular (3 + 1) human telomeric G-quadruplex bound to a telomestatin derivative. J. Am. Chem. Soc., 2013, 135(36), 13495-13501.
- [57] Kim, S.J.; McAlpine, S.R. Solid phase versus solution phase synthesis of heterocyclic macrocycles. *Molecules*, 2013, 18(1), 1111-1121.
- [58] Cocco, M.J.; Hanakahi, L.A.; Huber, M.D.; Maizels, N. Specific interactions of distamycin with G-quadruplex DNA. *Nucleic Acids Res.*, 2003, 31(11), 2944-2951.
- [59] Zaffaroni, N.; Lualdi, S.; Villa, R.; Bellarosa, D.; Cermele, C.; Felicetti, P.; Rossi, C.; Orlandi, L.; Daidone, M.G. Inhibition of telomerase activity by a distamycin derivative: effects on cell proliferation and induction of apoptosis in human cancer cells. *Eur. J. Cancer*, 2002, 38(13), 1792-1801.
- [60] Moore, M.J.; Cuenca, F.; Searcey, M.; Neidle, S. Synthesis of distamycin A polyamides targeting G-quadruplex DNA. Org. Biomol. Chem., 2006, 4(18), 3479-3488.
- [61] Drygin, D.; Siddiqui-Jain, A.; O'Brien, S.; Schwaebe, M.; Lin, A.; Bliesath, J.; Ho, C.B.; Proffitt, C.; Trent, K.; Whitten, J.P.; Lim, J.K.; Von Hoff, D.; Anderes, K.; Rice, W.G. Anticancer activity of CX-3543: a direct inhibitor of rRNA biogenesis. *Cancer Res.*, 2009, 69(19), 7653-7661.
- [62] Phatak, P.; Burger, A.M. Telomerase and its potential for therapeutic intervention. Br. J. Pharmacol., 2007, 152(7), 1003-1011.
- [63] Guittat, L.; De Cian, A.; Rosu, F.; Gabelica, V.; De Pauw, E.; Delfourne, E.; Mergny, J.L. Ascididemin and meridine stabilise Gquadruplexes and inhibit telomerase in vitro. *Biochim. Biophys. Acta*, 2005, *1724*(3), 375-384.
- [64] Zhang, W.J.; Oua, T-M.; Lua, Y-J.; Huanga, Y-Y.; Wua, W-B.; Huanga, Z-S.; Zhoua, J-L.; Wongc, K-Y.; Gua, L-Q. 9-Substituted berberine derivatives as G-quadruplex stabilizing ligands in telomeric DNA Bioorganic &. Med. Chem., 2007, 15(16), 5493-5501.
- [65] Zhou, W-J.; Deng, R.; Zhang, X-Y.; Feng, G-K.; Gu, L-Q.; Zhu, X.F. G-quadruplex ligand SYUIQ-5 induces autophagy by telo-

mere damage and TRF2 delocalization in cancer cells. *Mol. Cancer Ther.*, **2009**, *8*(12), 3203-3213.

- [66] Grand, C.L.; Han, H.; Muñoz, R.M.; Weitman, S.; Von Hoff, D.D.; Hurley, L.H.; Bearss, D.J. The cationic porphyrin TMPyP4 downregulates c-MYC and human telomerase reverse transcriptase expression and inhibits tumor growth in vivo. *Mol. Cancer Ther.*, 2002, 1(8), 565-573.
- [67] Klebanoff, C.A.; Acquavella, N.; Yu, Z.; Restifo, N.P. Therapeutic cancer vaccines: are we there yet? *Immunol. Rev.*, 2011, 239(1), 27-44.
- [68] Buseman, C.M.; Wright, W.E.; Shay, J.W. Is telomerase a viable target in cancer? *Mutat. Res.*, 2012, 730(1-2), 90-97.
- [69] Vetsika, E.K.; Konsolakis, G.; Aggouraki, D.; Kotsakis, A.; Papadimitraki, E.; Christou, S.; Menez-Jamet, J.; Kosmatopoulos, K.; Georgoulias, V.; Mavroudis, D. Immunological responses in cancer patients after vaccination with the therapeutic telomerase-specific vaccine Vx-001. *Cancer Immunol. Immunother.*, **2012**, *61*(2), 157-168.
- [70] Engell-Noerregaard, L.; Hansen, T.H.; Met, Ö.; Andersen, M.H.; Svane, I.M.; Andersen, M.H. Dendritic Cells Transfected with mRNA for p53, Survivin and hTERT as Treatment for Patients With Malignant Melanoma or Breast Cancer-A Phase I Study. J. Immunother., 2010, 33(8), 876.
- [71] Mocellin, S.; Pooley, K.A.; Nitti, D. Telomerase and the search for the end of cancer. *Trends Mol. Med.*, 2013, 19(2), 125-133.
- [72] Minev, B.; Hipp, J.; Firat, H.; Schmidt, J.D.; Langlade-Demoyen, P.; Zanetti, M. Cytotoxic T cell immunity against telomerase reverse transcriptase in humans. *Proc. Natl. Acad. Sci. USA*, 2000, 97(9), 4796-4801.
- [73] Cortez-Gonzalez, X.; Zanetti, M. Telomerase immunity from bench to bedside: round one. J. Transl. Med., 2007, 5, 12.
- [74] Kotsakis, E-K.; Vetsika, S.; Christou, D.; Hatzidaki, N.; Vardakis, D.; Aggouraki, G.; Konsolakis, V.; Georgoulias, C.; Christo-phyllakis, P. Cordopatis; K. Kosmatopoulos, K.; Mavroudis, D. Clinical outcome of patients with various advanced cancer types vaccinated with an optimized cryptic human telomerase reverse transcriptase [TERT] peptide: results of an expanded phase II study. Ann. Oncol., 2011, 22(8), 1736-1747.
- [75] Georgoulias, V.; Douillard, J.Y.; Khayat, D.; Manegold, C.; Rosell, R.; Rossi, A.; Menez-Jamet, J.; Iché, M.; Kosmatopoulos, K.; Gridelli, C. A multicenter randomized phase IIb efficacy study of Vx-001, a peptide-based cancer vaccine as maintenance treatment in advanced non-small-cell lung cancer: treatment rationale and protocol dynamics. *Clin. Lung Cancer*, **2013**, *14*(4), 461-465.
- [76] Fenoglio, D.; Parodi, A.; Lavieri, R.; Kalli, F.; Ferrera, F.; Tagliamacco, A.; Guastalla, A.; Lamperti, M.G.; Giacomini, M.; Filaci, G. Immunogenicity of GX301 cancer vaccine: Four (telomerase peptides) are better than one. *Hum. Vaccin. Immunother.*, **2015**, *11*(4), 838-850.
- [77] Marian, C.O.; Cho, S.K.; McEllin, B.M.; Maher, E.A.; Hatanpaa, K.J.; Madden, C.J.; Mickey, B.E.; Wright, W.E.; Shay, J.W.; Bachoo, R.M. The telomerase antagonist, imetelstat, efficiently targets glioblastoma tumor-initiating cells leading to decreased proliferation and tumor growth. *Clin. Cancer Res.*, **2010**, *16*(1), 154-163.
- [78] Ruden, M.; Puri, N. Novel anticancer therapeutics targeting telomerase. *Cancer Treat. Rev.*, 2013, 39(5), 444-456.
- [79] Schepelmann, S.; Ogilvie, L.M.; Hedley, D.; Friedlos, F.; Martin, J.; Scanlon, I.; Chen, P.; Marais, R.; Springer, C.J. Suicide gene therapy of human colon carcinoma xenografts using an armed oncolytic adenovirus expressing carboxypeptidase G2. *Cancer Res.*, 2007, 67(10), 4949-4955.
- [80] Nemunaitis, J.; Tong, A.W.; Nemunaitis, M.; Senzer, N.; Phadke, A.P.; Bedell, C.; Adams, N.; Zhang, Y.A.; Maples, P.B.; Chen, S.; Pappen, B.; Burke, J.; Ichimaru, D.; Urata, Y.; Fujiwara, T. A phase I study of telomerase-specific replication competent oncolytic adenovirus (telomelysin) for various solid tumors. *Mol. Ther.*, **2010**, *18*(2), 429-434.
- [81] Picard, D. Intracellular dynamics of the Hsp90 co-chaperone p23 is dictated by Hsp90. *Exp. Cell Res.*, 2006, 312(2), 198-204.
- [82] Ning, X.; Yang, S.; Wang, R.; Zhang, R.; Guo, L.; Tie, J.; Cheng, Y.; Wang, G.; Wan, S.; Fang, D. POT1 deficiency alters telomere length and telomere-associated gene expression in human gastric cancer cells. *Eur. J. Cancer Prev.*, **2010**, *19*(5), 345-351.
- [83] Kiran, K.G.; Palaniswamy, M.; Angayarkanni, J. Human telomerase inhibitors from microbial source. World J. Microbiol. Biotechnol., 2015, 31(9), 1329-1341.

- [84] Wei, C.; Wang, Y.; Zhang, M. Synthesis and binding studies of novel di-substituted phenanthroline compounds with genomic promoter and human telomeric DNA G-quadruplexes. Org. Biomol. Chem., 2013, 11(14), 2355-2364.
- [85] Ueno, T.; Takahashi, H.; Oda, M.; Mizunuma, M.; Yokoyama, A.; Goto, Y.; Mizushina, Y.; Sakaguchi, K.; Hayashi, H. Inhibition of human telomerase by rubromycins: implication of spiroketal system of the compounds as an active moiety. *Biochemistry*, 2000, 39(20), 5995-6002.
- [86] Togashi, K.; Ko, H-R.; Ahn, J.S.; Osada, H. Inhibition of telomerase activity by fungus metabolites, CRM646-A and thielavin B. *Bi*osci. Biotechnol. Biochem., 2001, 65(3), 651-653.
- [87] Tabata, Y.; Ikegami, S.; Yaguchi, T.; Sasaki, T.; Hoshiko, S.; Sakuma, S.; Shin-Ya, K.; Seto, H. Diazaphilonic acid, a new azaphilone with telomerase inhibitory activity. J. Antibiot., 1999, 52(4), 412-414.
- [88] Mizushina, Y.; Takeuchi, T.; Sugawara, F.; Yoshida, H. Anticancer targeting telomerase inhibitors: β-rubromycin and oleic acid. *Mini Rev. Med. Chem.*, **2012**, *12*(11), 1135-1143.
- [89] Huang, P.R.; Yeh, Y.M.; Wang, T.C. Potent inhibition of human telomerase by helenalin. *Cancer Lett.*, 2005, 227(2), 169-174.
- [90] Warabi, K.; Matsunaga, S.; van Soest, R.W.; Fusetani, N. Dictyodendrins A-E, the first telomerase-inhibitory marine natural products from the sponge Dictyodendrilla verongiformis. *J. Org. Chem.*, **2003**, *68*(7), 2765-2770.
- [91] Lavelle, F.; Riou, J.F.; Laoui, A.; Mailliet, P. Telomerase: a therapeutic target for the third millennium? *Crit. Rev. Oncol. Hematol.*, 2000, 34(2), 111-126.
- [92] Reddel, R.R.; Bryan, T.M.; Colgin, L.M.; Perrem, K.T.; Yeager, T.R. Alternative lengthening of telomeres in human cells. *Radiat. Res.*, 2001, 155(1 Pt 2), 194-200.
- [93] Wu, X.; Kemp, B.; Amos, C.I.; Honn, S.E.; Zhang, W.; Walsh, G.L.; Spitz, M.R. Associations among telomerase activity, p53 protein overexpression, and genetic instability in lung cancer. *Br. J. Cancer*, **1999**, 80(3-4), 453-457.

- [94] Burger, A.M.; Dai, F.; Schultes, C.M.; Reszka, A.P.; Moore, M.J.; Double, J.A.; Neidle, S. The G-quadruplex-interactive molecule BRACO-19 inhibits tumor growth, consistent with telomere targeting and interference with telomerase function. *Cancer Res.*, 2005, 65(4), 1489-1496.
- [95] Schrijvers, D.; Teurfs, W. Clinical Trials at the End-of-life. BAOJ Pall Medicine, 2015, 1(1), 001-007.
- [96] Gunnur Dikmen, Z.; Gellert, G.C.; Jackson, S. Gryaznov, Sergei.; Tressler, R.; Dogan, P. Wright, W.E.; Shay, J.W. *In vivo* Inhibition of Lung Cancer by A Novel Human Telomerase Inhibitor GRN163L. *Cancer Res.*, 2005, 65(17), 7866-7873.
- [97] Goldblatt, E.M.; Erickson, P.A.; Gentry, E.R.; Gryaznov, S.M.; Herbert, B.S. Lipid-conjugated telomerase template antagonists sensitize resistant HER2-positive breast cancer cells to trastuzumab. *Breast Cancer Res. Treat.*, 2009, *118*(1), 21-32.
- [98] Goldblatt, E.M.; Gentry, E.R.; Fox, M.J.; Gryaznov, S.M.; Shen, C.; Herbert, B.S. The telomerase template antagonist GRN163L alters MDA-MB-231 breast cancer cell morphology, inhibits growth, and augments the effects of paclitaxel. *Mol. Cancer Ther.*, 2009, 8(7), 2027-2035.
- [99] Dong, X.; Liu, A.; Zer, C.; Feng, J.; Zhen, Z.; Yang, M.; Zhong, L. siRNA inhibition of telomerase enhances the anti-cancer effect of doxorubicin in breast cancer cells. *BMC Cancer*, 2009, *9*, 133.
- [100] Fujiwara, T.; Kagawa, S.; Kishimoto, H.; Endo, Y.; Hioki, M.; Ikeda, Y.; Sakai, R.; Urata, Y.; Tanaka, N.; Fujiwara, T. Enhanced antitumor efficacy of telomerase-selective oncolytic adenoviral agent OBP-401 with docetaxel: preclinical evaluation of chemovirotherapy. *Int. J. Cancer*, **2006**, *119*(2), 432-440.
- [101] Tamakawa, R.A.; Fleisig, H.B.; Wong, J.M. Telomerase inhibition potentiates the effects of genotoxic agents in breast and colorectal cancer cells in a cell cycle-specific manner. *Cancer Res.*, 2010, 70(21), 8684-8694.