

Muscarinic Acetylcholine Receptors in the Retina Therapeutic Implications

Yue Ruan ^{1,*}, Andreas Patzak ^{2,*}, Norbert Pfeiffer ¹ and Adrian Gericke ¹

- ¹ Department of Ophthalmology, University Medical Center, Johannes Gutenberg University Mainz, Langenbeckstr. 1, 55131 Mainz, Germany; norbert.pfeiffer@unimedizin-mainz.de (N.P.); adrian.gericke@unimedizin-mainz.de (A.G.)
- Institute of Vegetative Physiology, Charité-Universitätsmedizin Berlin, Charitéplatz 1, 10117 Berlin, Germany
- * Correspondence: yruan@uni-mainz.de (Y.R.); andreas.patzak@charite.de (A.P.); Tel.: +49-6131-17-8276 (Y.R.); +49-3045-052-8220 (A.P.)

Abstract: Muscarinic acetylcholine receptors (mAChRs) belong to the superfamily of G-proteincoupled receptors (GPCRs). The family of mAChRs is composed of five subtypes, M₁, M₂, M₃, M₄ and M₅, which have distinct expression patterns and functions. In the eye and its adnexa, mAChRs are widely expressed and exert multiple functions, such as modulation of tear secretion, regulation of pupil size, modulation of intraocular pressure, participation in cell-to-cell signaling and modula-tion of vascular diameter in the retina. Due to this variety of functions, it is reasonable to assume that abnormalities in mAChR signaling may contribute to the development of various ocular diseases. On the other hand, mAChRs may offer an attractive therapeutic target to treat ocular diseases. Thus far, non-subtype-selective mAChR ligands have been used in ophthalmology to treat dry eye disease, myopia and glaucoma. However, these drugs were shown to cause various side-effects. Thus, the use of subtype-selective ligands would be useful to circumvent this problem. In this review, we give an overview on the localization and on the functional role of mAChR subtypes in the eye and its adnexa with a special focus on the retina. Moreover, we describe the pathophysiological role of mAChRs in retinal diseases and discuss potential therapeutic approaches.

Keywords: muscarinic acetylcholine receptors; glaucoma; retina; retinal ganglion cells; therapeutic strategy

1. Introduction

In 1914, Henry Dale first observed that the actions of acetylcholine (ACh) could be divided into nicotine-like and muscarine-like effects, respectively [1]. Preganglionic parasympathetic neurons release neurotransmitters, such as acetylcholine, from synaptic vesicles of axon terminals into the synaptic cleft. From there, neurotransmitters can bind to receptors in postganglionic parasympathetic cell membranes [2]. Muscarinic acetylcholine receptors (mAChRs) are widely localized on postganglionic parasympathetic neurons and are widely expressed in the central nervous system [3]. Apart from neurons, mAChRs are expressed on many other cell types [4]. In the eye and its adnexa, mAChRs were found to be expressed in the cornea, lens, uvea, conjunctiva, sclera, retina and the lacrimal gland [5–8]. Hence, it is not surprising that mAChRs are involved in diverse important physiological functions in the eye, such as tear fluid production, goblet cell secretion, keratocyte migration and proliferation, pupil size regulation, ocular drainage, lens cell signaling and ocular growth as well as cell-to-cell signaling and vascular reactivity in the retina [9–13]. An increasing number of studies demonstrates that mAChRs are potential pharmacological targets for the treatment of various ocular diseases, such as glaucoma and myopia [14,15].

Glaucoma is a neurodegenerative disease, which is characterized by the impairment and loss of retinal ganglion cells (RGCs) and optic nerve fibers, which may lead to irre-



Citation: Ruan, Y.; Patzak, A.; Pfeiffer, N.; Gericke, A. Muscarinic Acetylcholine Receptors in the Retina—Therapeutic Implications. *Int. J. Mol. Sci.* 2021, *22*, 4989. https://doi.org/10.3390/ijms22094989

Academic Editor: Silvia C. Finnemann

Received: 1 April 2021 Accepted: 4 May 2021 Published: 8 May 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). versible blindness [16]. It has been estimated that there are around 80 million people with glaucoma worldwide [17]. According to another estimate, the global number of people affected by glaucoma will increase to 111.8 million in 2040 [18]. Although elevated intraocular pressure is a major risk factor for glaucoma, several population-based studies reported that intraocular pressure is within the normal range in a large portion of individuals with glaucoma [19,20]. In clinical practice, lowering intraocular pressure (IOP) is essential to prevent development and progression of the disease and to preserve the patients' quality of life [16]. However, in almost half of the patients the disease continues to progress despite normalization of IOP [21]. Hence, novel complementary retinal neuroprotection strategies would be valuable to reduce progressive neurodegeneration in the retina [22]. Pathologic myopia with characteristic degenerative changes in the sclera, choroid, and retinal pigment epithelium is a major cause of visual impairment and blindness worldwide by increasing the risk for ocular complications, such as macular degeneration, retinal detachment and glaucoma [23]. The global prevalence of pathologic myopia is rising rapidly, especially in the younger Asian population [24]. Holden et al. projected that by 2050 the number of people with myopia will increase to 4.758 billion (49.8% of the global population), and 938 million people (9.8% of the world's population) will have high myopia [25]. There is still no effective therapy in the clinical routine to prevent the progression of pathologic myopia, which makes the disease an increasing global health concern.

MAChRs belong to the class of G-protein-coupled receptors (GPCRs) containing seven transmembrane segments, which transfer signals into the cell via coupling with G-proteins. The G-proteins modulate the activity of a number of different effectors, such as ion channels and enzymes [26,27]. The mAChR family is composed of five subtypes, M1, M2, M3, M4 and M_5 with different molecular and signaling properties [3,28,29]. For example, M_1 , M_3 and M_5 have been reported to typically couple to G proteins of the Gq/11 family. However, M_2 and M_4 receptors have been shown to preferentially couple to G proteins of the Gi and Go family [3,29]. Some studies reported that mAChR agonists reduce IOP and exert neuroprotective effects in glaucoma [30–32]. The non-subtype-selective mAChR antagonist, atropine, has been shown to inhibit scleral proliferation and matrix synthesis, and to prevent axial elongation of the eyeball providing a novel therapeutic approach for myopia control [14,33]. Unfortunately, non-subtype-selective mAChR agonists may exert ocular adverse effects in clinical practice, which are related to its constricting effects on the ciliary and pupillary sphincter muscle and systemic adverse effects including increased salivation and sweating, vomiting, diarrhoea and tachycardia [34,35]. For example, the non-subtypeselective mAChR agonist, pilocarpine, which has been used for long-term IOP control can cause blurred vision, brow ache from ciliary spasm and rarely retinal detachment [35]. In addition, the non-specific mAChR antagonist, atropine, acutely induces cycloplegia and photophobia and on the long term might cause premature presbyopia, cataract, and light damage in the retina [36].

To design more specific mAChR-based therapeutic approaches with less side-effects, it is crucial to identify the distribution and physiological function of individual mAChR subtypes and to test the use of highly subtype-selective ligands in laboratory and clinical studies. In this review, we summarize and discuss the localization, the functional and the pathophysiological role of individual mAChR subtypes in the retina. Additionally, we discuss potential therapeutic strategies targeting individual mAChR subtypes.

The identification of literature was carried out via a search on PubMed. The PubMed database search included the following keywords: (("muscarinic acetylcholine receptors" OR "mAChR" OR "muscarinic acetylcholine receptor subtypes" OR "muscarinic receptors" OR "mAChR" OR "mAChR subtypes" OR "M₁ muscarinic acetylcholine receptor" OR "M₂ muscarinic acetylcholine receptor" OR "M₄ muscarinic acetylcholine receptor" OR "M₄ muscarinic acetylcholine receptor" OR "M₄ muscarinic acetylcholine receptor" OR "M₅ muscarinic acetylcholine receptor" OR "M₄ muscarinic receptor antagonist" OR "Muscarinic receptor agonist") AND ("retina" OR "ocular" OR "RGC" OR "glaucoma" OR "IOP" OR "diabetes" OR "retinal models")). The search was conducted from 13 March to 28 March 2021 with the following inclusion criteria: all studies,

muscarinic acetylcholine receptors in the retina, written in English and published after 1976. In total, the study search resulted in 268 publications. Studies reporting the roles of muscarinic acetylcholine receptors in other organs except for eyes were excluded. Studies in conjunctiva, cornea, iris and lens were excluded. Moreover, expert opinions, abstracts, and letters were excluded. The reference lists of all retrieved articles were reviewed for further identification of potentially relevant studies.

2. Expression and Distribution of mAChRs in the Retina

Based on studies with the labeled radioligands, [³H]propylbenzilylcholine mustard ([³H]PrBCM), [³H]N-methylscopolamine ([³H]NMS) and [³H]quinuclidinyl benzilate ([³H]QNB), a high density of muscarinic binding sites has been demonstrated in rat, bovine and chick retinas, whereas relatively few binding sites have been detected in frog and salamander retinas [37–39]. Autoradiographic experiments in embryo and adult chicken retinas revealed specific mAChR binding sites in the inner synaptic retinal layer [40]. In 1985, Polans et al. found a high density of muscarinic binding sites in the inner plexiform layer (IPL) and the outer plexiform layer (OPL) of the salamander retina [41]. In the same year, Hutchins and Hollyfield presented evidence for a population of mAChRs in the human retina, apparently expressed in the IPL, by using the irreversible ligand, [³H]PrBCM [42].

Based on autoradiographic experiments, it has been suggested in 1988 that mAChR subtype number and distribution change during retinal development [43]. Later, experiments in the ferret retina suggested that the subtypes, number and distribution of mAChRs changes during retinal synaptogenesis [44]. In the study, mAChR-like immunoreactivity was found at amacrine–amacrine cell contacts by electron microscopy and immunohistochemical techniques [44]. Townes-Anderson and Vogt found that mAChRs in the salamander retina are located on amacrine/ganglion, bipolar, and horizontal cells [45]. In 1988, Moroi-Fetters found that stimulation of muscarinic receptors by the subtype-preferring M₁ receptor antagonist, pirenzepine, in the rat retina causes phosphoinositide hydrolysis, which indicated that these receptors appear to be of the M₁ subtype [46].

In 1989, all five mAChR subtypes (M_1 , M_2 , M_3 , M_4 and M_5) were identified [47,48]. Molecular cloning techniques provided a new molecular basis to characterize expression, location and physiological function of all five mAChRs [49]. In 1997, McKinnon et al. examined regulation of mAChR expression in the chicken embryonic retina by using immunoblot, immunoprecipitation and solution hybridization analyses [50]. The authors reported that the M_4 receptor is the main subtype expressed at an early stage in embryonic development, while M_2 and M_3 receptor expression increases at a later stage [50].

However, the precise anatomical location of mAChRs in the retina remained unknown at that time. One year later, Fischer et al. used purified and subtype-specific antibodies directed against M_2 , M_3 and M_4 subtypes to detect receptor immunoreactivity in the retina. The study revealed that in the chick retina the M₂ receptor was expressed in amacrine and ganglion cells, the M₃ receptor was expressed in many bipolar cells and small subsets of amacrine cells and the M_4 receptor was found in amacrine and ganglion cells [51]. In an in vitro and in vivo study, Belmonte et al. demonstrated that retinal Müller glial cells can secrete a factor called MARIA (muscarinic acetylcholine receptor-inducing activity) that can regulate M_2 expression in vitro and in vivo [52]. Another in vitro study demonstrated the presence of the M_1 receptor in cultured human retinal pigment epithelium (RPE) at both the mRNA and the protein level [53]. The expression of M_1 receptor mRNA was also observed in the guinea pig retina, and immunohistochemical findings revealed that the M_1 receptor was expressed in all layers of the retina [54]. Strang et al. used RT-PCR, Western blot analysis and immunohistochemistry to identify the expression and distribution of mAChR subtypes in the rabbit retina [55]. The authors detected mRNA expression for all five mAChR subtypes in the whole neural retina by RT-PCR and Western blotting, and they confirmed that all five mAChR subtypes were expressed by subpopulations of bipolar, amacrine, and ganglion cells by immunohistochemical analyses [55]. According to

a study by Gericke et al. in 2011, only mRNA for the M_3 receptor was detected in murine retinal arterioles [56]. In contrast, mRNA for all five mAChR subtypes was detected in ophthalmic arteries, but mRNA levels for the odd-numbered subtypes, M_1 , M_3 and M_5 , were higher than those for the even-numbered subtypes, M_2 and M_4 [57]. Figure 1 shows the distribution of mAChRs within the retina.

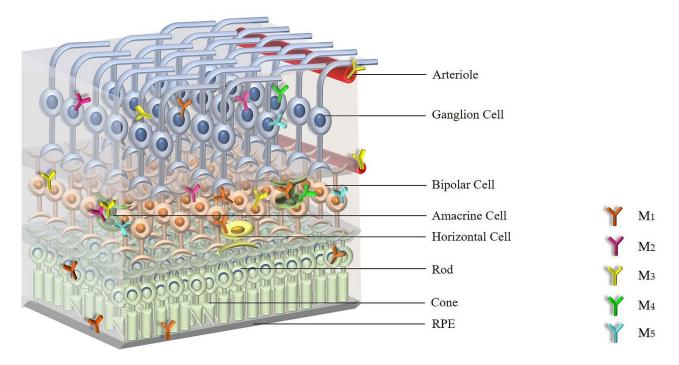


Figure 1. The distribution of individual mAChR subtypes in the retina. Abbreviations: mAChRs: muscarinic acetylcholine receptors; RPE: retinal pigment epithelium; M_1 : M_1 muscarinic acetylcholine receptor; M_2 : M_2 muscarinic acetylcholine receptor; M_3 : M_3 muscarinic acetylcholine receptor; M_4 : M_4 muscarinic acetylcholine receptor; M_5 : M_5 muscarinic acetylcholine receptor.

Based on the expression studies, all five mAChR subtypes have been detected in the retina with the individual subtypes showing an overlapping expression pattern.

3. Cellular Signaling of mAChRs

Peralta et al. and Bonner et al. first cloned and sequenced human mAChRs, which are encoded by the *CHRM1* to *CHRM5* genes [58,59]. The genes give rise to the five subtypes, M₁, M₂, M₃, M₄ and M₅ [3]. The mAChR family belongs to the superfamily of seven-transmembrane receptors, which mediates cellular signal transduction pathways via G-proteins [60]. The M₁, M₃ and M₅ subtypes, which efficiently couple to Gq/11 subtype G proteins, can mobilize phosphoinositides to generate inositol 1, 4, 5-triphosphate (IP3) and 1, 2-diacylglycerol (DAG) via activation of phosphoinositide-specific phospholipase C β (PLC β), leading to an increase in intracellular cytosolic calcium (Ca²⁺) levels and protein kinase C (PKC) activity [61]. This may help to stimulate nitric oxide (NO) production, since neuronal nitric oxide synthase (nNOS) is calcium/calmodulin-dependent [62]. The M₂ and M₄ receptors preferentially couple to pertussis toxin-sensitive Gi and Go proteins, causing an inhibition of the cAMP-dependent pathway via suppression of adenylyl cyclase [63]. Furthermore, both Gq/11-, Gi/o-coupled with mAChRs may exert effects through activation of small GTPases, such as Rho and Ras, and downstream effectors, such as phosphoinositide-3 kinases and mitogen-activated protein kinases [60].

Muscarinic receptors play an important role in the development of the retina and in processing visual information [64]. For example, mAChRs regulate the function of bipolar cells in the ON/OFF channel, and the input and output of amacrine, bipolar, ganglion and horizontal cells [52,64]. Additionally, Jardon et al. suggested that a cholinergic loop of

amacrine cells could be involved in the inhibitory pathway from the ON channel to the OFF channel in the frog retina carrying "light on" and "light off" information from the retina to the brain [65,66]. Muscarinic cholinergic transmission exerts a substantial contribution in the retina [67]. In an in vitro study, the muscarinic antagonist, QNB, enhanced the amplitude of the electroretinogram (ERG) b-wave (a measure of ON bipolar cell activation), and induced moderate vasoconstriction in the cat retina [67].

Although mAChRs regulate many important cellular signaling pathways, it is difficult to assign specific functional roles for individual mAChR subtypes in the retina. Jositsch et al. tested the specificity of mAChRs antibodies under different conditions in immunohistochemical labelling on tissue sections by analyzing specimens from respective gene-deficient mice and wild-type mice [68]. The data indicated that immunohistochemical detection of mAChR subtypes in tissue sections is limited to the M₂ subtype [68]. It has been reported that cells frequently co-express more than one mAChR subtype that increases the difficulty of assigning a functional response to a single receptor subtype [3]. The lack of highly selective pharmacological ligands and antibodies for individual mAChR subtypes has hampered conclusions regarding the physiological role of individual muscarinic receptor subtypes. The development of genetically modified mice devoid of M₁- to M₅ receptors helped to circumvent the problem of assigning a specific function to an individual mAChR subtype [69–75]. M₁-M₅ receptor knockout mouse models (M₁-M₅R-/-) have also been used in studies of ocular tissues.

For example, Barathi et al. studied the role of each of the mAChR subtypes in the development of myopia by using $M_1-M_5R-/-$ mice [76]. The authors found that M_2 receptors play a crucial role in myopia development by hindering scleral fibroblast cell proliferation and further scleral remodeling [76]. Based on studies in $M_3R-/-$ mice, Gericke et al. showed that the M_3 receptor is responsible for mediating cholinergic responses in retinal arterioles and the ophthalmic artery [56,57,77]. Laspas examined the amount of cells in the retinal ganglion cell (RGC) layer and the amount of axons in the optic nerve in 5-month-old $M_1R-/-$ and wild-type mice and found no significant difference between both groups [78]. More recently, the same laboratory conducted experiments in 5- and 15-month-old $M_1R-M_5R-/-$ mice to examine whether one of the mAChRs and age have an influence on neuron survival in the retina [30]. Based on these studies, the M_1 receptor was found to be critical for RGC survival in the aging mouse retina [30]. These examples show how genetically modified mice may help to better understand the physiological roles of individual muscarinic receptor subtypes in the retina.

4. Functional Roles of Individual mAChR Subtypes in the Retina

According to their differential coupling to intracellular signaling cascades, the mAChR subtypes have been divided into two subfamilies, the "M₁-like" mAChR subfamily and the "M₂-like" mAChR subfamily [79]. The odd-numbered subtypes, M₁, M₃, and M₅, belong to the "M₁-like" family, which couple to the Gq protein and activate phospholipase C (PLC)-dependent signaling pathways. In neuronal tissue, activation of this signaling cascade increases neuronal excitability through activation of nonspecific cation channels, release of Ca²⁺ from intracellular stores, or inhibition of Ca²⁺-activated K⁺ channels [80]. On the contrary, members of the "M₂-like" family, the even-numbered M₂ and M₄ receptor subtypes, are generally linked to inhibition of adenylyl cyclase activity [79]. Activation of M₂ and M₄ receptors decreases neuronal activity via activation of a subset of K⁺ channels, the inhibition of Ca²⁺ channels, or the inhibition of the Ca²⁺ priming of K⁺ channels [55,81,82]. This suggests that the different subtypes may subserve different functions in the retina.

A plethora of studies demonstrated that the M_1 muscarinic receptor subtype may be crucial for neuron survival in the retina [30]. For example, M_1 receptor activation protected retinal neurons from glutamate-induced cytotoxicity [83]. It has been proposed that activation of M_1 receptors reduces Ca^{2+} influx into the cell and the expression of Bcl-2 and Caspase-3 [62,83,84]. This effect can be blocked by the M_1 -preferring muscarinic receptor antagonist, pirenzepine [84]. Additionally, activation of M_1 receptors significantly increases the survival of RGCs in vitro [85]. Pereira et al. indicated that the mechanism involved in M_1 receptor activity and the survival of RGCs is by the release of polypeptides and activation of insulin receptor kinase receptors [85]. An in vitro study demonstrated that activation of mAChRs effectively protects against hypoxia-induced apoptosis in RGCs via modulation of the hypoxia-inducible factor 1-alpha (HIF-1 α) pathway [86]. Based on these in vitro data, Laspas et al. conducted a study in 5-month-old $M_1R_{-}/_{-}$ mice and age-matched wild-type mice to test the neuroprotective role of the M_1 receptor in vivo. However, the authors found no differences in the number of retinal neurons and the amount of optic nerve axons between $M_1R - / -$ and wild-type mice [30]. In a more recent study in 2019, Laspas et al. examined the potential role of all five muscarinic receptor subtypes on neuroprotection in the RGC layer in congenic mAChR-/- mice of different age categories [30]. Intriguingly, the authors observed that the lack of the M_1 receptor was associated with a reduced RGC density in aged mice. Aged $M_1R-/$ mice also displayed elevated ROS levels in the RGC layer and increased retinal mRNA expression for the prooxidative NADPH oxidase 2 (NOX2) and reduced mRNA levels for the antioxidative enzymes, superoxide dismutase 1 (SOD1), hemeoxygenase-1 (HO-1) and anti-glutathione peroxidase 1 (GPx1) [30]. The findings provided the first direct evidence that the lack of the M₁ receptor leads to accelerated RGC loss in mice via changing in the oxidative/antioxidative balance in favor of oxidative in the retina [30]. L-satropane was reported to be effective in preventing retinal neuron damage, which may be attributed to decreasing cell apoptosis and amyloid- β (A β) production via activation of M₁ receptor subtype [87]. Moreover, in an in vivo study, the M_1 muscarinic receptor was reported to exert protective effects on RGCs via activation of insulin growth factor 1 (IGF-1) and insulin growth factor 1 receptor (IGF-1R) [88]. Additionally, activation of PKC delta was suggested to regulate neurotrophin levels by M_1 muscarinic receptor activation ultimately leading to an increase in RGCs' survival in vitro in the retina [89,90]. In 2021, an in vivo study in rats showed that huperzine A lowers intraocular pressure via the M_3 receptor and exerts neuroprotective effects in the retina by increasing endogenous ACh levels and activating M₁ receptors and their downstream AKT/MAPK signaling pathways [31].

Braga et al. first analyzed the levels of M_3 receptors in retinal cell cultures treated with 50 ng/mL phorbol 12-myristate 13-acetate (PMA, a PKC activator) for 48 h. PMA induced a marked increase in M_3 receptor levels [91]. Based on pharmacological studies employing muscarinic subtype-preferring antagonists, Borda et al. observed that carbachol can stimulate NOS activity and increase the expression of nNOS and iNOS mRNA in the rat retina via activation of M_1/M_3 receptor subtypes [92,93].

The expression of nicotinic AChR (nAChR: $\alpha 3$, $\alpha 4$, $\alpha 6$, $\alpha 7$, $\beta 2$ and $\beta 4$ nAChR subunits) and/or mAChR by amacrine and ganglion cells has been described in retinas of Rhesus monkeys and rabbits [94–96]. Retinal nAChRs mediate visual processing and may have effects on refractive development and ocular neovascularization [94]. In the retina, there was an overlap in the expression patterns of M₁, M₄ and M₅ muscarinic receptors with those of non- $\alpha 7$ and $\alpha 7$ nAChRs in presumptive amacrine and ganglion cells [97]. Strang et al. suggest that the determining the role of mAChRs in retinal processing is complicated by the concomitant expression of nAChRs by the same cells [55]. A study in $\alpha 7$ nAChR–/– mice demonstrated that M₂ and M₄ mAChR subtype transcripts were significantly upregulated in the RGC layer [98].

There are not many studies on the functional role of M_2 receptors in the retina. Several pieces of evidence suggest that activation of M_2 and M_4 receptors is involved in visual processing [55]. The M_2 receptor has been reported to increase Ca^{2+} influx, exclusively due to Ca^{2+} mobilization from intracellular stores [99]. The M_2 muscarinic receptor was shown to inhibit adenylyl cyclase activity and to activate inwardly rectifying potassium (K⁺) channels [100]. However, these responses can be rapidly attenuated by receptor desensitization [100,101]. Antal et al. found that activation of M_2 receptors regulates feedforward inhibition following activated RGCs [102]. Cimini et al. indicated that the

production of NO in response to M_2 muscarinic receptor activation may lead to an increase in cGMP, which can modulate the mutual interactions of acetylcholine-glycine-gammaaminobutyric acid (GABA) in the inner retina [103].

The M_4 receptor exerts a direct inhibitory control on dopamine D_1 -like receptor signaling [104]. In a rat glaucoma model, Almasieh et al. demonstrated that activation of M_1 and M_4 receptors promotes RGCs' survival [15]. Moreover, the partially selective M_1/M_4 muscarinic antagonist, pirenzepine, was reported to be successful in preventing myopia progression in animal models [105]. The M_4 -selective antagonist, himbacine, could also prevent myopia in chicken by daily intravitreal injections [106].

All these studies provide evidence that individual mAChRs exert specific functional roles in the retina, which offers new therapeutic perspectives for mAChR ligands. A variety of studies reported associations of endothelial NOS (eNOS) genetic polymorphism or adrenergic receptor gene polymorphisms with retinal diseases [107,108]. Additionally, mAChR subtype gene polymorphisms have been reported [109]. Unfortunately, studies on mAChR genetic polymorphisms with respect to retinal diseases have not been reported so far [109]. Such studies would be appreciated to shed some light on the role of mAChR in the development of specific retinal diseases.

5. Strategies to Target Individual mAChR Subtypes

Individual mAChR subtypes are novel targets for the treatment of various diseases including Alzheimer's disease, Parkinson's disease, type 2 diabetes, schizophrenia and glaucoma [15,30,110–112]. The first-generation muscarinic agonist, pilocarpine, was initially used as a topical glaucoma therapy in the late 1800s and approved by the United States Food and Drug Administration (FDA) in 1974 [113]. Although pilocarpine has been used as an ocular hypotensive agent for 40 years, it causes ocular side effects due to poor selective pharmacokinetic properties. Therefore, clinical approaches are required that modulate individual mAChR subtype activity with high selectivity. However, there are still few ophthalmological studies that focus on subtype-selective mAChR ligands. For example, several randomized double-blind, placebo-controlled studies tested the impact of cevime-line, a specific agonist of the M₃ muscarinic receptor, on dry eye symptoms in patients with Sjögren's syndrome and most of them suggested some beneficial effects [114–117].

Due to high sequence conservation in the orthosteric binding site of mAChR subtypes, it has been difficult to develop mAChR ligands with high subtype selectivity [118]. Subtype-preferring M_1 agonists used for the treatment of central nervous system (CNS) disorders have been reported in the patent or primary literature. However, subsequent studies indicated that previous orthosteric agonists are not highly selective when evaluated across multiple systems [119]. Moreover, previous studies trying to develop highly selective ligands for individual mAChR subtypes have failed because of the difficulty of developing compounds that are truly subtype-selective [119]. In consequence, many researchers are now focusing on developing allosteric activators of mAChRs including both positive allosteric modulators (PAMs) and allosteric agonists, which offer new opportunities to target specific mAChR subtypes for therapeutic purposes [120]. In the treatment of Alzheimer's disease and other CNS disorders, several novel selective M₁ agonists and allosteric potentiators have been identified, providing important new tools to evaluate the potential utility of selective activators of the M_1 receptor [121]. For example, BPB and 77-LH-28-1 have been reported to exert highly selective agonist activity for the M_1 receptor [119,122,123]. Lebois et al. have discovered a novel highly selective M_1 allosteric agonist VU0357017, with a potentially novel allosteric binding site in the third extracellular loop of the M₁ receptor [124]. The novel PAMs for M₁, VU0090157, VU0029767 and benzyl quinolone carboxylic acid (BQCA), compete for binding at the orthosteric ACh-binding site, but have no direct agonist activity. However, they induce a robust leftward shift of the concentration-response relationship of ACh at activating the M_1 receptor [121]. The highly selective PAMs for the M₄ receptor, VU0010010, VU0152099, VU0152100 and LY2033298, were reported to be an important breakthrough for selective activation of the M_4 receptor

exerting no activity at any other mAChR subtype [124]. By novel microwave-assisted chemistry in in vitro and in vivo probe projects, Weaver et al. described the discovery and development of the first highly selective M_1 antagonist, VU0255035, which was shown to be active in vivo and penetrated the blood-brain-barrier [118].

Apart from the well-known therapeutic applications for CNS diseases, mAChRs subtype ligands may find potential applications in ophthalmology, such as in the field of retinal neuroprotection [125–127].

6. Conclusions

All five mAChR subtypes, M_1 through M_5 , were found to be expressed in the retina. Experimental studies over the past decade were focusing on the biology, pharmacology and structure of mAChRs. Knockout animal models have provided clues to the specific functions of mAChR subtypes in various physiological and pathophysiological processes of the retina. The M_1 receptor is suggested to be involved in retinal neuron survival and, therefore, appears to be a promising therapeutic target. Studies of mAChR genetic polymorphisms focusing on retinal diseases as well as studies employing genetically modified animal models and new mAChR ligands with high subtype selectivity will be helpful to shed more light on the physiological and pathophysiological role of individual muscarinic receptor subtypes in the retina. New pharmacologic compounds with high selectivity for individual mAChR subtypes have already been studied in CNS disorders and may also offer attractive tools to treat retinal diseases.

Author Contributions: Conceptualization, A.G. and Y.R.; writing—original draft preparation, Y.R.; writing—review and editing, A.G., A.P. and N.P.; visualization, Y.R.; supervision, A.G.; All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Dale, H.H. The Action of Certain Esters and Ethers of Choline, and Their Relation to Muscarine. *J. Pharmacol. Exp. Ther.* **1914**, *6*, 147.
- 2. McCorry, L.K. Physiology of the Autonomic Nervous System. Am. J. Pharm Educ. 2007, 71, 78. [CrossRef]
- Caulfield, M.P.; Birdsall, N.J.M. International Union of Pharmacology. XVII. Classification of Muscarinic Acetylcholine Receptors. *Pharmacol. Rev.* 1998, 50, 279. [PubMed]
- Wessler, I.; Kirkpatrick, C.J. Acetylcholine Beyond Neurons: The Non-neuronal Cholinergic System in Humans. *Br. J. Pharmacol.* 2008, 154, 1558–1571. [CrossRef] [PubMed]
- 5. Liu, S.; Li, J. Expression and Function of Muscarinic Receptor Subtypes on Human Cornea and Conjunctiva. *Investig. Ophthalmol. Vis. Sci.* **2007**, *48*, 2987–2996. [CrossRef]
- 6. Ríos, J.D.; Forde, K. Development of Conjunctival Goblet Cells and Their Neuroreceptor Subtype Expression. *Investig. Ophthalmol. Vis. Sci.* **2000**, *41*, 2127–2137.
- Liu, Q.; Wu, J. Changes in Muscarinic Acetylcholine Receptor Expression in Form Deprivation Myopia in Guinea Pigs. *Mol. Vis.* 2007, 13, 1234–1244.
- 8. Mitchelson, F. Muscarinic Receptor Agonists and Antagonists: Effects on Ocular Function. In *Muscarinic Receptors*; Fryer, A.D., Christopoulos, A., Eds.; Springer: Berlin/Heidelberg, Germany, 2012; pp. 263–298.
- 9. Sloniecka, M. Neuropeptides and Neurotransmitters in Keratocytes: Importance in Corneal Wound Healing Processes. Ph.D. Thesis, Comprehensive Summary, Umeå University, Umeå, Sweden, 2015.
- Smith, E.L., III; Redburn, D.A. Permanent Alterations in Muscarinic Receptors and Pupil Size Produced by Chronic Atropinization in Kittens. *Investig. Ophthalmol. Vis. Sci.* 1984, 25, 239–243.
- 11. McBrien, N.A.; Jobling, A.I. Expression of Muscarinic Receptor Subtypes in Tree Shrew Ocular Tissues and Their Regulation during the Development of Myopia. *Mol. Vis.* **2009**, *15*, 464–475. [PubMed]
- 12. Ríos, J.D.; Zoukhri, D. Immunolocalization of Muscarinic and Vip Receptor Subtypes and Their Role in Stimulating Goblet Cell Secretion. *Investig. Ophthalmol. Vis. Sci* 1999, 40, 1102–1111.
- 13. Duncan, G.; Collison, D.J. Role of the Non-neuronal Cholinergic System in the Eye: A Review. *Life Sci.* 2003, 72, 2013–2019. [CrossRef]
- 14. Lin, H.-J.; Wan, L. Muscarinic Acetylcholine Receptor 3 is Dominant in Myopia Progression. *Investig. Ophthalmol. Vis. Sci.* 2012, 53, 6519–6525. [CrossRef] [PubMed]

- 15. Almasieh, M.; Zhou, Y. Structural and Functional Neuroprotection in Glaucoma: Role of Galantamine-mediated Activation of Muscarinic Acetylcholine Receptors. *Cell Death Dis.* **2010**, *1*, e27. [CrossRef]
- 16. Weinreb, R.N.; Aung, T. The Pathophysiology and Treatment of Glaucoma: A Review. *JAMA* **2014**, *311*, 1901–1911. [CrossRef] [PubMed]
- 17. Quigley, H.A.; Broman, A.T. The Number of People with Glaucoma Worldwide in 2010 and 2020. *Br. J. Ophthalmol.* 2006, *90*, 262–267. [CrossRef] [PubMed]
- 18. Tham, Y.-C.; Li, X. Global Prevalence of Glaucoma and Projections of Glaucoma Burden Through 2040: A Systematic Review and Meta-analysis. *Ophthalmology* **2014**, *121*, 2081–2090. [CrossRef]
- 19. Quigley, H.A.; Addicks, E.M. Optic Nerve Damage in Human Glaucoma. II. The Site of Injury and Susceptibility to Damage. *Arch. Ophthalmol.* **1981**, *99*, 635–649. [CrossRef] [PubMed]
- 20. Weinreb, R.N.; Khaw, P.T. Primary Open-angle Glaucoma. Lancet 2004, 363, 1711–1720. [CrossRef]
- 21. Heijl, A.; Leske, M.C. Reduction of Intraocular Pressure and Glaucoma Progression: Results from the Early Manifest Glaucoma trial. *Arch. Ophthalmol.* **2002**, *120*, *1268–1279*. [CrossRef]
- 22. Nucci, C.; Martucci, A. Neuroprotective Agents in the Management of Glaucoma. Eye 2018, 32, 938–945. [CrossRef]
- 23. Grossniklaus, H.E.; Green, W.R. Pathologic Findings in Pathologic Myopia. Retina 1992, 12, 127–133. [CrossRef] [PubMed]
- 24. Morgan, I.; Rose, K. How Genetic is School Myopia? Prog. Retin. Eye Res. 2005, 24, 1–38. [CrossRef] [PubMed]
- 25. Holden, B.A.; Fricke, T.R. Global Prevalence of Myopia and High Myopia and Temporal Trends from 2000 Through 2050. *Ophthalmology* **2016**, *123*, 1036–1042. [CrossRef] [PubMed]
- 26. Schultz, G.; Rosenthal, W. Role of G Proteins in Calcium Channel Modulation. Annu. Rev. Physiol. 1990, 52, 275–292. [CrossRef]
- 27. Simon, M.I.; Strathmann, M.P. Diversity of G Proteins in Signal Transduction. Science 1991, 252, 802–808. [CrossRef] [PubMed]
- 28. Dreyer, F. Acetylcholine Receptor. Br. J. Anaesth. 1982, 54, 115–130. [CrossRef]
- 29. Kostenis, E.; Zeng, F.-Y. Structure-function Analysis of Muscarinic Receptors and Their Associated G Proteins. *Life Sci.* **1999**, *64*, 355–362. [CrossRef]
- Laspas, P.; Zhutdieva, M.B. The M(1) Muscarinic Acetylcholine Receptor Subtype is Important for Retinal Neuron Survival in Aging Mice. Sci. Rep. 2019, 9, 5222. [CrossRef]
- 31. Yu, P.; Dong, W.-P. Huperzine a Lowers Intraocular Pressure via the M3 mAChR and Provides Retinal Neuroprotection via the M1 mAChR: A Promising Agent for the Treatment of Glaucoma. *Ann. Transl. Med.* **2021**, *9*, 332. [CrossRef]
- 32. Broadley, K.J.; Kelly, D.R. Muscarinic Receptor Agonists and Antagonists. Molecules 2001, 6, 142–193. [CrossRef]
- 33. Barathi, V.A.; Beuerman, R.W. Molecular Mechanisms of Muscarinic Receptors in Mouse Scleral Fibroblasts: Prior to and after Induction of Experimental Myopia with Atropine Treatment. *Mol. Vis.* **2011**, *17*, 680–692. [PubMed]
- Marquis, R.E.; Whitson, J.T. Management of Glaucoma: Focus on Pharmacological Therapy. *Drugs Aging* 2005, 22, 1–21. [CrossRef]
 Lusthaus, J.; Goldberg, I. Current Management of Glaucoma. *Med. J. Aust.* 2019, 210, 180–187. [CrossRef] [PubMed]
- 36. McBrien, N.A.; Stell, W.K. How does Atropine Exert Its Anti-myopia Effects? *Ophthalmic Physiol. Opt.* **2013**, *33*, 373–378. [CrossRef]
- 37. Hutchins, J.B. Review: Acetylcholine as a Neurotransmitter in the Vertebrate Retina. Exp. Eye Res. 1987, 45, 1–38. [CrossRef]
- 38. Hruska, R.E.; White, R. Muscarinic Cholinergic Receptors in Mammalian Retina. Brain Res. 1978, 148, 493–498. [CrossRef]
- 39. Atterwill, C.K.; Mahoney, A. The Uptake and Subcellular Distribution of 3-h-choline by the Retina. Br. J. Pharmacol. 1975, 53, 447P.
- 40. Sugiyama, H.; Daniels, M.P. Muscarinic Acetylcholine Receptors of the Developing Retina. *Proc. Natl. Acad. Sci. USA* **1977**, *74*, 5524–5528. [CrossRef] [PubMed]
- 41. Polans, A.S.; Hutchins, J.B. Muscarinic Cholinergic Receptors in the Retina of the Larval Tiger Salamander. *Brain Res.* **1985**, 340, 355–362. [CrossRef]
- 42. Hutchins, J.B.; Hollyfield, J.G. Acetylcholine Receptors in the Human Retina. Investig. Ophthalmol. Vis. Sci. 1985, 26, 1550–1557.
- Cho, N.J.; Klein, W.L. Muscarinic Acetylcholine Receptors from Avian Retina and Heart Undergo Different Patterns of Molecular Maturation. J. Neurochem. 1988, 50, 1403–1411. [CrossRef]
- 44. Hutchins, J.B. Development of Muscarinic Acetylcholine Receptors in the Ferret Retina. Dev. Brain Res. 1994, 82, 45-61. [CrossRef]
- 45. Townes-Anderson, E.; Vogt, B.A. Distribution of Muscarinic Acetylcholine Receptors on Processes of Isolated Retinal Cells. *J. Comp. Neurol.* **1989**, *290*, 369–383. [CrossRef] [PubMed]
- 46. Moroi-Fetters, S.E.; Neff, N.H. Muscarinic Receptor-mediated Phosphoinositide Hydrolysis in the Rat Retina. *J. Pharmacol. Exp. Ther.* **1988**, 246, 553. [PubMed]
- 47. Bonner, T.I. New Subtypes of Muscarinic Acetylcholine Receptors. Trends Pharmacol. Sci. 1989, (Suppl.), 11–15. [PubMed]
- 48. Wall, S.J.; Yasuda, R.P. The Ontogeny of M1–M5 Muscarinic Receptor Subtypes in Rat Forebrain. *Dev. Brain Res.* **1992**, *66*, 181–185. [CrossRef]
- 49. Bonner, T.I. The Molecular Basis of Muscarinic Receptor Diversity. Trends Neurosci. 1989, 12, 148–151. [CrossRef]
- McKinnon, L.A.; Rosoff, M. Regulation of Muscarinic Receptor Expression and Function in Cultured Cells and in Knock-out Mice. *Life Sci.* 1997, 60, 1101–1104. [CrossRef]
- 51. Fischer, A.J.; McKinnon, L.A. Identification and Localization of Muscarinic Acetylcholine Receptors in the Ocular Tissues of the Chick. *J. Comp. Neurol.* **1998**, *392*, 273–284. [CrossRef]

- Belmonte, K.E.; McKinnon, L.A. Developmental Expression of Muscarinic Acetylcholine Receptors in Chick Retina: Selective Induction of M2 Muscarinic Receptor Expression in ovo by a Factor Secreted by Muller Glial Cells. J. Neurosci. 2000, 20, 8417–8425. [CrossRef]
- 53. Zhang, L.H.; Yan, D.S. Expression of Muscarinic Acetylcholine Receptor-1 in Human Retinal Pigment Epithelium. *Zhonghua Yan Ke Za Zhi Chin. J. Ophthalmol.* **2006**, *42*, 1109–1112.
- 54. Tao, Y.; Li, X.-L. Effect of Green Flickering Light on Myopia Development and Expression of M1 Muscarinic Acetylcholine Receptor in Guinea Pigs. *Int. J. Ophthalmol.* **2018**, *11*, 1755–1760. [PubMed]
- 55. Strang, C.E.; Renna, J.M. Muscarinic Acetylcholine Receptor Localization and Activation Effects on Ganglion Response Properties. *Investig. Ophthalmol. Vis. Sci.* 2010, *51*, 2778–2789. [CrossRef]
- Gericke, A.; Sniatecki, J.J. Identification of the Muscarinic Acetylcholine Receptor Subtype Mediating Cholinergic Vasodilation in Murine Retinal Arterioles. *Investig. Ophthalmol. Vis. Sci.* 2011, 52, 7479–7484. [CrossRef]
- Gericke, A.; Mayer, V.G.A. Cholinergic Responses of Ophthalmic Arteries in M3 and M5 Muscarinic Acetylcholine Receptor Knockout Mice. *Investig. Ophthalmol. Vis. Sci.* 2009, 50, 4822–4827. [CrossRef] [PubMed]
- Peralta, E.G.; Ashkenazi, A. Distinct Primary Structures, Ligand-binding Properties and Tissue-specific Expression of Four Human Muscarinic Acetylcholine Receptors. *EMBO J.* 1987, 6, 3923–3929. [CrossRef] [PubMed]
- Bonner, T.I.; Young, A.C. Cloning and Expression of the Human and Rat M5 Muscarinic Acetylcholine Receptor Genes. *Neuron* 1988, 1, 403–410. [CrossRef]
- 60. van Koppen, C.J.; Kaiser, B. Regulation of Muscarinic Acetylcholine Receptor Signaling. *Pharmacol. Ther.* **2003**, *98*, 197–220. [CrossRef]
- 61. Rümenapp, U.; Asmus, M. The M3 Muscarinic Acetylcholine Receptor Expressed in HEK-293 Cells Signals to Phospholipase D via G12 but not Gq-type G Proteins: Regulators of G Proteins as Tools to Dissect Pertussis Toxin-resistant G Proteins in Receptor-Effector Coupling. *J. Biol. Chem.* **2001**, *276*, 2474–2479. [CrossRef]
- 62. Mathes, C.; Thompson, S.H. The Nitric Oxide/Cgmp Pathway Couples Muscarinic Receptors to the Activation of Ca2+ Influx. *J. Neurosci.* **1996**, *16*, 1702–1709. [CrossRef]
- 63. Haga, T. Molecular Properties of Muscarinic Acetylcholine Receptors. *Proc. Jpn. Acad. Ser. B Phys. Biol. Sci.* 2013, 89, 226–256. [CrossRef]
- 64. Goin, J.C.; Nathanson, N.M. Subtype-specific Regulation of the Expression and Function of Muscarinic Acetylcholine Receptors in Embryonic Chicken Retinal Cells. J. Neurochem. 2002, 83, 964–972. [CrossRef]
- 65. Jardon, B.; Bonaventure, N. Possible Involvement of Cholinergic and Glycinergic Amacrine Cells in the Inhibition Exerted by the on Retinal Channel on the Off Retinal Channel. *Eur. J. Pharmacol.* **1992**, *210*, 201–207. [CrossRef]
- 66. Dowling, J.E. The Retina: An Approachable Part of the Brain; Harvard University Press: London, UK, 1987.
- 67. Niemeyer, G.; Jurklies, B. Binding and Electrophysiology of the Muscarinic Antagonist QNB in the Mammalian Retina. *Klin. Mon. Augenheilkd.* **1995**, *206*, 380–383. [CrossRef] [PubMed]
- Jositsch, G.; Papadakis, T. Suitability of Muscarinic Acetylcholine Receptor Antibodies for Immunohistochemistry Evaluated on Tissue Sections of Receptor Gene-deficient Mice. *Naunyn Schmiedebergs Arch. Pharmacol.* 2009, 379, 389–395. [CrossRef] [PubMed]
- 69. Hamilton, S.E.; Loose, M.D. Disruption of the M1 Receptor Gene Ablates Muscarinic Receptor-dependent M Current Regulation and Seizure Activity in Mice. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 13311. [CrossRef]
- Gomeza, J.; Shannon, H. Pronounced Pharmacologic Deficits in M2 Muscarinic Acetylcholine Receptor Knockout Mice. *Proc. Natl. Acad. Sci. USA* 1999, 96, 1692–1697. [CrossRef] [PubMed]
- Matsui, M.; Motomura, D. Multiple Functional Defects in Peripheral Autonomic Organs in Mice Lacking Muscarinic Acetylcholine Receptor Gene for the M3 Subtype. Proc. Natl. Acad. Sci. USA 2000, 97, 9579. [CrossRef]
- Gomeza, J.; Zhang, L. Enhancement of d1 Dopamine Receptor-mediated Locomotor Stimulation in M4 Muscarinic Acetylcholine Receptor Knockout Mice. Proc. Natl. Acad. Sci. USA 1999, 96, 10483. [CrossRef] [PubMed]
- Woolley, M.L.; Carter, H.J. Attenuation of Amphetamine-induced Activity by the Non-selective Muscarinic Receptor Agonist, Xanomeline, is Absent in Muscarinic M4 Receptor Knockout Mice and Attenuated in Muscarinic M1 Receptor Knockout Mice. *Eur. J. Pharmacol.* 2009, 603, 147–149. [CrossRef] [PubMed]
- 74. Yamada, M.; Lamping, K.G. Cholinergic Dilation of Cerebral Blood Vessels is Abolished in M(5) Muscarinic Acetylcholine Receptor Knockout Mice. *Proc. Natl. Acad. Sci. USA* 2001, *98*, 14096. [CrossRef]
- Steidl, S.; Yeomans, J.S. M5 Muscarinic Receptor Knockout Mice Show Reduced Morphine-induced Locomotion but Increased Locomotion after Cholinergic Antagonism in the Ventral Tegmental Area. J. Pharmacol. Exp. Ther. 2009, 328, 263. [CrossRef]
- 76. Barathi, V.A.; Kwan, J.L. Muscarinic Cholinergic Receptor (M2) Plays a Crucial Role in the Development of Myopia in Mice. *Dis. Model. Mech.* **2013**, *6*, 1146–1158. [CrossRef]
- 77. Gericke, A.; Steege, A. Role of the M3 Muscarinic Acetylcholine Receptor Subtype in Murine Ophthalmic Arteries after Endothelial Removal. *Investig. Ophthalmol. Vis. Sci.* **2014**, *55*, 625–631. [CrossRef] [PubMed]
- Laspas, P.; Sniatecki, J.J. Effect of the M1 Muscarinic Acetylcholine Receptor on Retinal Neuron Number Studied with Gene-Targeted Mice. J. Mol. Neurosci. 2015, 56, 472–479. [CrossRef]
- Lanzafame, A.A.; Christopoulos, A. Cellular Signaling Mechanisms for Muscarinic Acetylcholine Receptors. *Recept. Channels* 2003, 9, 241–260. [CrossRef] [PubMed]
- 80. Brown, D.A.; Abogadie, F.C. Muscarinic Mechanisms in Nerve Cells. Life Sci. 1997, 60, 1137–1144. [CrossRef]

- Brown, D.A.; Selyanko, A.A. Membrane Currents Underlying the Cholinergic Slow Excitatory Post-synaptic Potential in the Rat Sympathetic Ganglion. J. Physiol. 1985, 365, 365–387. [CrossRef]
- 82. Wess, J.; Liu, J. Structural Basis of Receptor/G Protein Coupling Selectivity Studied with Muscarinic Receptors as Model Systems. *Life Sci.* **1997**, *60*, 1007–1014. [CrossRef]
- 83. Tan, P.-P.; Yuan, H.-H. Activation of Muscarinic Receptors Protects Against Retinal Neurons Damage and Optic Nerve Degeneration in Vitro and in Vivo Models. *CNS Neurosci. Ther.* **2014**, *20*, 227–236. [CrossRef]
- 84. Zhou, W.; Zhu, X. Neuroprotection of Muscarinic Receptor Agonist Pilocarpine Against Glutamate-induced Apoptosis in Retinal Neurons. *Cell. Mol. Neurobiol.* 2008, 28, 263–275. [CrossRef] [PubMed]
- 85. Pereira, S.P.F.; Medina, S.V. Cholinergic Activity Modulates the Survival of Retinal Ganglion Cells in Culture: The Role of M1 Muscarinic Receptors. *Int. J. Dev. Neurosci.* 2001, *19*, 559–567. [CrossRef]
- 86. Zhu, X.; Zhou, W. Pilocarpine Protects Cobalt Chloride-induced Apoptosis of rgc-5 Cells: Involvement of Muscarinic Receptors and hif-1 Alpha Pathway. *Cell. Mol. Neurobiol.* **2010**, *30*, 427–435. [CrossRef] [PubMed]
- Yu, P.; Zhou, W. L-satropane Prevents Retinal Neuron Damage by Attenuating Cell Apoptosis and aβ Production via Activation of M1 Muscarinic Acetylcholine Receptor. *Curr. Eye Res.* 2017, 42, 1319–1326. [CrossRef] [PubMed]
- Granja, M.G.; Gomes Braga, L.E. Igf-1 and igf-1r Modulate the Effects of il-4 on Retinal Ganglion Cells Survival: The Involvement of M1 Muscarinic Receptor. *Biochem. Biophys. Res. Commun.* 2019, 519, 53–60. [CrossRef] [PubMed]
- Braga, L.E.G.; Miranda, R.L. Pkc Delta Activation Increases Neonatal Rat Retinal Cells Survival in Vitro: Involvement of Neurotrophins and M1 Muscarinic Receptors. *Biochem. Biophys. Res. Commun.* 2018, 500, 917–923. [CrossRef] [PubMed]
- dos Santos, A.A.; Medina, S.V. Protein Kinase C Regulates the Expression of M1 Receptors and BDNF in Rat Retinal Cells. Neurochem. Res. 2009, 34, 884–890. [CrossRef]
- 91. Braga, L.E.G.; Granja, M.G. Pma Increases M3 Muscarinic Receptor Levels and Decreases Retinal Cells Proliferation Through a Change in the Levels of Cell-cycle Regulatory Proteins. *Neurosci. Lett.* **2013**, 550, 29–34. [CrossRef]
- 92. Borda, E.; Berra, A. Correlations between Neuronal Nitric Oxide Synthase and Muscarinic M3/M1 Receptors in the Rat Retina. *Exp. Eye Res.* **2005**, *80*, 391–399. [CrossRef]
- Berra, A.; Ganzinelli, S. Inducible Nitric Oxide Synthase Subserves Cholinergic Vasodilation in Retina. Vis. Neurosci. 2005, 22, 371–377. [CrossRef]
- 94. Liu, J.; McGlinn, A.M. Nicotinic Acetylcholine Receptor Subunits in Rhesus Monkey Retina. *Investig. Ophthalmol. Vis. Sci.* 2009, 50, 1408–1415. [CrossRef]
- 95. Strang, C.E.; Amthor, F.R. Rabbit Retinal Ganglion Cell Responses to Nicotine can be Mediated by Beta2-containing Nicotinic Acetylcholine Receptors. *Vis. Neurosci.* 2003, 20, 651–662. [CrossRef]
- Strang, C.E.; Andison, M.E. Rabbit Retinal Ganglion Cells Express Functional Aalpha7 Nicotinic Acetylcholine Receptors. Am. J. Physiol. Cell Physiol. 2005, 289, C644–C655. [CrossRef]
- Strang, C.E.; Long, Y. Nicotinic and Muscarinic Acetylcholine Receptors Shape Ganglion Cell Response Properties. J. Neurophysiol. 2015, 113, 203–217. [CrossRef]
- Smith, M.L.; Souza, F.G.O. Acetylcholine Receptors in the Retinas of the α7 Nicotinic Acetylcholine Receptor Knockout Mouse. *Mol. Vis.* 2014, 20, 1328–1356. [PubMed]
- Schmidt, M.; Bienek, C. Differential Calcium Signalling by M2 and M3 Muscarinic Acetylcholine Receptors in a Single Cell Type. Naunyn Schmiedebergs Arch. Pharmacol. 1995, 352, 469–476. [CrossRef] [PubMed]
- 100. Pals-Rylaarsdam, R.; Hosey, M.M. Two Homologous Phosphorylation Domains Differentially Contribute to Desensitization and Internalization of the M2 Muscarinic Acetylcholine Receptor. J. Biol. Chem. **1997**, 272, 14152–14158. [CrossRef]
- 101. Zang, W.J.; Yu, X.J. On the Role of G Protein Activation and Phosphorylation in Desensitization to Acetylcholine in Guinea-Pig Atrial Cells. *J. Physiol.* **1993**, *464*, 649–679. [CrossRef]
- 102. Antal, M.; Acuna-Goycolea, C. Cholinergic Activation of M2 Receptors Leads to Context-dependent Modulation of Feedforward Inhibition in the Visual Thalamus. *PLoS Biol.* **2010**, *8*, e1000348. [CrossRef] [PubMed]
- Cimini, B.A.; Strang, C.E. Role of Acetylcholine in Nitric Oxide Production in the Salamander Retina. J. Comp. Neurol. 2008, 507, 1952–1963. [CrossRef]
- 104. Onali, P.; Olianas, M. Muscarinic M4 Receptor Inhibition of Dopamine d1-like Receptor Signalling in Rat Nucleus Accumbens. *Eur. J. Pharmacol.* 2002, 448, 105–111. [CrossRef]
- 105. Arumugam, B.; McBrien, N.A. Muscarinic Antagonist Control of Myopia: Evidence for M4 and M1 Receptor-based Pathways in the Inhibition of Experimentally-induced Axial Myopia in the Tree Shrew. *Investig. Ophthalmol. Vis. Sci.* 2012, 53, 5827–5837. [CrossRef]
- Cottriall, C.L.; Truong, H.-T. Inhibition of Myopia Development in Chicks Using Himbacine: A Role for M4 Receptors? *Neuroreport* 2001, 12, 2453–2456. [CrossRef]
- 107. Inagaki, Y.; Mashima, Y. Polymorphism of Beta-adrenergic Receptors and Susceptibility to Open-angle Glaucoma. *Mol. Vis.* **2006**, *12*, 673–680.
- 108. Liao, Q.; Wang, D.-H. Association of Genetic Polymorphisms of Enos with Glaucoma. Mol. Vis. 2011, 17, 153–158. [PubMed]
- Michel, M.C.; Teitsma, C.A. Polymorphisms in Human Muscarinic Receptor Subtype Genes. *Handb. Exp. Pharmacol.* 2012, 208, 49–59. [CrossRef]

- Fisher, A. Cholinergic Modulation of Amyloid Precursor Protein Processing with Emphasis on M1 Muscarinic Receptor: Perspectives and Challenges in Treatment of Alzheimer's Disease. J. Neurochem. 2012, 120, 22–33. [CrossRef] [PubMed]
- 111. Dencker, D.; Thomsen, M. Muscarinic Acetylcholine Receptor Subtypes as Potential Drug Targets for the Treatment of Schizophrenia, Drug abuse, and Parkinson's Disease. *ACS Chem. Neurosci.* **2012**, *3*, 80–89. [CrossRef]
- Weston-Green, K.; Huang, X.-F. Second Generation Antipsychotic-induced Type 2 Diabetes: A Role for the Muscarinic M3 Receptor. CNS Drugs 2013, 27, 1069–1080. [CrossRef] [PubMed]
- 113. Seibold, L.K.; Wagner, B.D. The Diurnal and Nocturnal Effects of Pilocarpine on Intraocular Pressure in Patients Receiving Prostaglandin Analog Monotherapy. *J. Ocul. Pharmacol. Ther.* **2018**, *34*, 590–595. [CrossRef]
- Leung, K.C.M.; McMillan, A.S. The Efficacy of Cevimeline Hydrochloride in the Treatment of Xerostomia in Sjögren's Syndrome in Southern Chinese Patients: A Randomised Double-blind, Placebo-controlled Crossover Study. *Clin. Rheumatol.* 2008, 27, 429–436. [CrossRef]
- 115. Fife, R.S.; Chase, W.F. Cevimeline for the Treatment of Xerostomia in Patients with Sjögren Syndrome: A Randomized Trial. *Arch. Intern. Med.* **2002**, *162*, 1293–1300. [CrossRef]
- Petrone, D.; Condemi, J.J. A Double-blind, Randomized, Placebo-controlled Study of Cevimeline in Sjögren's Syndrome Patients with Xerostomia and Keratoconjunctivitis Sicca. Arthritis Rheum. 2002, 46, 748–754. [CrossRef]
- 117. Ono, M.; Takamura, E. Therapeutic Effect of Cevimeline on Dry Eye in Patients with Sjögren's Syndrome: A Randomized, Double-blind Clinical Study. *Am. J. Ophthalmol.* **2004**, *138*, 6–17. [CrossRef] [PubMed]
- Weaver, C.D.; Sheffler, D.J. Discovery and Development of a Potent and Highly Selective Small Molecule Muscarinic Acetylcholine Receptor Subtype i (mAChR 1 or M1) Antagonist in Vitro and in Vivo Probe. *Curr. Top. Med. Chem.* 2009, *9*, 1217–1226. [CrossRef] [PubMed]
- 119. Conn, P.J.; Jones, C.K. Subtype-selective Allosteric Modulators of Muscarinic Receptors for the Treatment of CNS Disorders. *Trends Pharmacol. Sci.* **2009**, *30*, 148–155. [CrossRef]
- Digby, G.J.; Shirey, J.K. Allosteric Activators of Muscarinic Receptors as Novel Approaches for Treatment of CNS Disorders. *Mol. Biosyst.* 2010, 6, 1345–1354. [CrossRef] [PubMed]
- 121. Kinney, G.G. Muscarinic Receptor Activation for the Treatment of Schizophrenia. Neuropsychopharmacology 2006, 31, S26.
- 122. Thomas, R.L.; Mistry, R. G Protein Coupling and Signaling Pathway Activation by M1 Muscarinic Acetylcholine Receptor Orthosteric and Allosteric Agonists. *J. Pharmacol. Exp. Ther.* **2008**, *327*, 365–374. [CrossRef] [PubMed]
- 123. Jones, C.K.; Brady, A.E. Tbpb is a Highly Selective M1 Allosteric Mscarinic Receptor Agonist in Vitro and Produces Robust Antipsychotic-like Effects in Vivo. In Proceedings of the Neuropsychopharmacology, Paris, France, 16–20 September 2006; pp. S116–S117.
- Shirey, J.K.; Xiang, Z. An Allosteric Potentiator of M4 mAChR Modulates Hippocampal Synaptic Transmission. *Nat. Chem. Biol.* 2008, 4, 42–50. [CrossRef] [PubMed]
- 125. Scarr, E. Muscarinic Receptors: Their Roles in Disorders of the Central Nervous System and Potential as Therapeutic Targets. *CNS Neurosci. Ther.* **2012**, *18*, 369–379. [CrossRef] [PubMed]
- 126. Matera, C.; Tata, A.M. Pharmacological Approaches to Targeting Muscarinic Acetylcholine Receptors. *Recent Pat. CNS Drug Discov.* 2014, *9*, 85–100. [CrossRef] [PubMed]
- 127. Greig, N.H.; Reale, M. New Pharmacological Approaches to the Cholinergic System: An Overview on Muscarinic Receptor Ligands and Cholinesterase Inhibitors. *Recent Pat. CNS Drug Discov.* 2013, *8*, 123–141. [CrossRef] [PubMed]