

Research

Open Access

## A structural interpretation of the effect of GC-content on efficiency of RNA interference

Chi Yu Chan<sup>1</sup>, C Steven Carmack<sup>1</sup>, Dang D Long<sup>1</sup>, Anil Maliyekkel<sup>2,3</sup>, Yu Shao<sup>1</sup>, Igor B Roninson<sup>2</sup> and Ye Ding\*<sup>1</sup>

Address: <sup>1</sup>Wadsworth Center, New York State Department of Health, 150 New Scotland Avenue, Albany, NY 12208, USA, <sup>2</sup>Cancer Center, Ordway Research Institute, Albany, New York 12208, USA and <sup>3</sup>Department of Molecular Genetics, University of Illinois at Chicago, Chicago, IL 60607, USA

Email: Chi Yu Chan - c.clarence@yahoo.com; C Steven Carmack - steve.carmack@wadsworth.org; Dang D Long - dlong@wadsworth.org; Anil Maliyekkel - amaliyekkel@ordwayresearch.org; Yu Shao - yushao04@yahoo.com; Igor B Roninson - Roninson@ordwayresearch.org; Ye Ding\* - yding@wadsworth.org

\* Corresponding author

from The Seventh Asia Pacific Bioinformatics Conference (APBC 2009)  
Beijing, China. 13–16 January 2009

Published: 30 January 2009

BMC Bioinformatics 2009, 10(Suppl 1):S33 doi:10.1186/1471-2105-10-S1-S33

This article is available from: <http://www.biomedcentral.com/1471-2105/10/S1/S33>

© 2009 Chan et al; licensee BioMed Central Ltd.

This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background:** RNA interference (RNAi) mediated by small interfering RNAs (siRNAs) or short hairpin RNAs (shRNAs) has become a powerful technique for eukaryotic gene knockdown. siRNA GC-content negatively correlates with RNAi efficiency, and it is of interest to have a convincing mechanistic interpretation of this observation. We here examine this issue by considering the secondary structures for both the target messenger RNA (mRNA) and the siRNA guide strand.

**Results:** By analyzing a unique homogeneous data set of 101 shRNAs targeted to 100 endogenous human genes, we find that: 1) target site accessibility is more important than GC-content for efficient RNAi; 2) there is an appreciable negative correlation between GC-content and RNAi activity; 3) for the predicted structure of the siRNA guide strand, there is a lack of correlation between RNAi activity and either the stability or the number of free dangling nucleotides at an end of the structure; 4) there is a high correlation between target site accessibility and GC-content. For a set of representative structural RNAs, the GC content of 62.6% for paired bases is significantly higher than the GC content of 38.7% for unpaired bases. Thus, for a structured RNA, a region with higher GC content is likely to have more stable secondary structure. Furthermore, by partial correlation analysis, the correlation for GC-content is almost completely diminished, when the effect of target accessibility is controlled.

**Conclusion:** These findings provide a target-structure-based interpretation and mechanistic insight for the effect of GC-content on RNAi efficiency.

### Background

RNA interference (RNAi) [1] is a sequence-specific gene silencing mechanism that can be mediated either by small interfering RNAs (siRNAs) of about 21 nt with two-nucleotide 3' overhang [2], or by stably expressed short hairpin RNAs (shRNAs), which are processed by Dicer into siRNAs [3,4]. The antisense (guide) strand guides Argonaute2 (Ago2), the catalytic component of the RNA-induced silencing complex (RISC), to cleave mRNA by base-pairing with the complementary site in the target. Large variation in the efficiency of siRNAs has been commonly observed [5]. Usually, only a small proportion of randomly selected siRNAs are potent. Thus, there has been a great interest in determining rules for improvement of RNAi design. It has been commonly observed that high GC content negatively correlates with RNAi activity. Thus, a low GC-content is among a number of empirical rules on siRNA duplex features that have been proposed [6]. In addition, the importance of target secondary structure and accessibility has been supported by numerous studies [7-14].

It is tempting to seek a mechanistic interpretation for the effect of GC-content on RNAi efficiency. Because high GC can give rise to stable RNA secondary structure, one possible interpretation is the proposal that self-structure of the siRNA guide strand can be detrimental to RNAi activity [15]. We here investigate this issue by considering the secondary structures for both the target messenger RNA (mRNA) and the siRNA guide strand. From analyses of a unique homogeneous data set of 101 shRNAs targeted to 100 endogenous human genes, the results support a target-structure-based interpretation for the effect of GC-content.

### Results

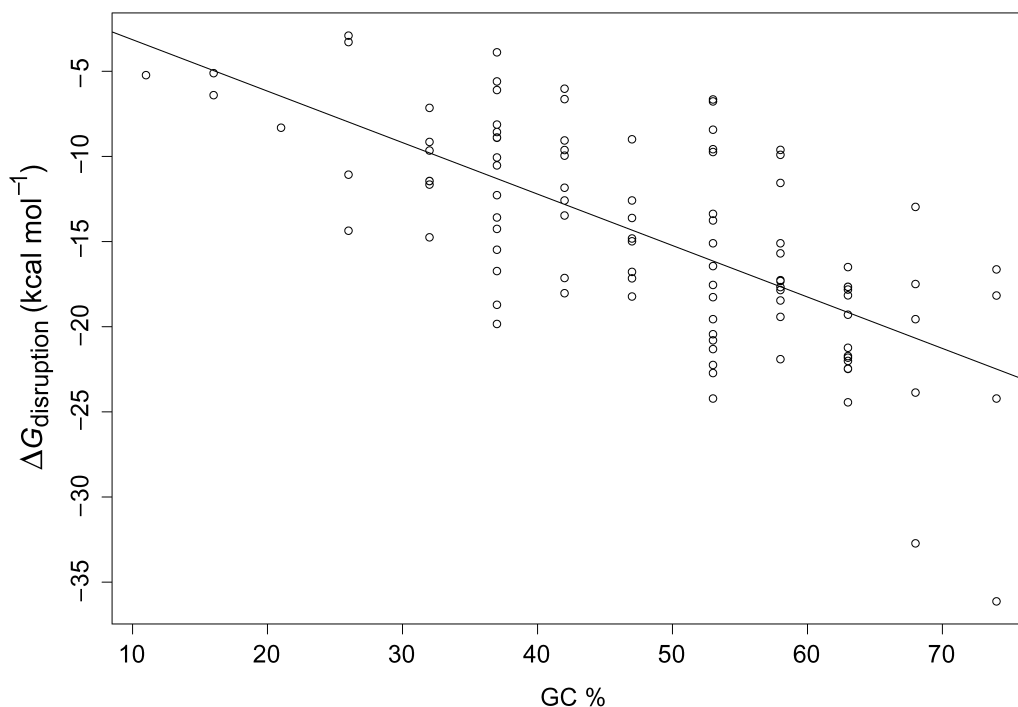
For the shRNA dataset, we first computed Pearson's correlation coefficient and the significance of the correlation for the RNAi activity and a structural parameter or GC%. These calculations were performed by using the R statistical package [16], and the results were summarized in Table 1. We find that, with the highest and significant correlation, target site accessibility is more important than GC-content. We also observe an appreciable negative correlation of -0.1444 between GC-content and RNAi activity, albeit with a *p*-value of 0.1497. Surprisingly, for the predicted optimal structure for the siRNA guide strand, there is a lack of correlation between RNAi activity and either the stability (siRNA MFE in Table 1) or the number of free dangling nucleotides at an end of the structure.

Because the effect of GC-content cannot be explained by the structure of the siRNA guide strand, we hypothesized that, to some extent, GC-content is indicative of target site accessibility. Indeed, there is a highly significant correlation between target site accessibility and GC-content (Table 1; Figure 1). Furthermore, for the representative set of structural RNAs, 62.6% of paired bases are GC, significantly higher than 38.7% for unpaired bases (*p*-value = 5.67E-15 for the Wilcoxon signed rank test). Thus, for a structured RNA, a region with higher GC content is likely to have more stable secondary structure.

To further investigate the relationships among GC-content, target accessibility and RNAi activity, we performed partial correlation analysis. Partial correlation measures the degree of association between two random variables, with the effect of a set of controlling random variables removed. For the three variables of our interest here, the calculation was performed with a published R script for three variables [17]. By partial correlation analysis, we

**Table 1: Correlation analyses for structural parameters, GC%, and RNAi activity**

Pair of measures		Pearson's correlation coefficient	Significance of correlation (p-value)
$\Delta G_{\text{disruption}}$	RNAi activity	0.2382	0.0165
siRNA GC%	RNAi activity	-0.1444	0.1497
siRNA MFE	RNAi activity	0.0531	0.5982
Number of 3' free nts	RNAi activity	0.0108	0.9146
Number of 5' free nts	RNAi activity	0.0538	0.5933
Number of 3' free nts with $\geq 2$ 5' free nts	RNAi activity	0.1311	0.2465
$\Delta G_{\text{disruption}}$	siRNA GC%	-0.6832	3.519E-15



**Figure 1**

Strong correlation between  $\Delta G_{\text{disruption}}$  and GC-content of siRNA guide strand (linear regression line is shown for predicting  $\Delta G_{\text{disruption}}$  by GC%, with  $R^2 = 0.4668$  and  $P\text{-value} = 3.52E-15$ ).

found that when the effect of target accessibility is controlled (i.e., removed), the correlation between GC-content and RNAi activity is 0.026, with a  $p$ -value of 0.7978. This near complete diminishment of correlation supports the hypothesis that the negative correlation typically observed between GC-content and RNAi efficiency is mainly due to structural inaccessibility often associated with high GC-content of the target site.

Because the protein complexes involved in gene regulation are similar for microRNAs [18] and siRNAs, we folded all of 137 worm microRNAs from microRNA Registry [19]. We found that for 79 (58%) of the microRNAs, the predicted optimal structure has a stability (i.e., free energy) under 0.0 kcal/mol, typically with two to five consecutive base pairs. Furthermore, for 9 (7%) of the microRNAs, either the 5' end or the 3' end is completely involved in intramolecular base-pairing. These suggest that some intramolecular structures can be tolerated for the regulatory functions by animal microRNAs, and these structures are likely to be weakened or completely abolished upon interaction with the RISC.

## Discussion

The results of the analyses suggest that, to a large extent, the effect of GC-content on RNAi is due to the target structure and site accessibility rather than the structure of the siRNA guide strand. However, for the purpose of the rational design of RNAi experiments, GC-content cannot be a substitute for predicted target site accessibility, owing to its substantially lower correlation (Table 1). Similar observations were made in a previous analysis of other RNAi datasets based on alternative structural calculations [9]. The common findings from this study and the previous study support a target-structure-based interpretation for the effect of GC-content on the RNAi efficiency.

Contrasting with the conclusion from a published study [15], we did not observe any effect of potential folding of siRNA guide strand on RNAi efficiency. It is likely that the ability for the siRNA guide strand to fold may be negatively affected by the enzymatic activity by the RNAi machinery including duplex unwinding by helicase. In addition, constrained by RISC, the siRNA guide strand is unlikely to fold freely into a stable structure, regardless of GC-content.

Based on nine siRNAs with 11 common nts for a region of a single target, it was reported that the RNAi activity was strongly correlated with the number of free (unpaired) dangling nts at the ends of the structure predicted for the siRNA guide strand, and there was a poor correlation for other sequence features and target accessibility [15]. With our much larger and more representative dataset, we did not observe any correlation for the number of free dangling nts. For the negative finding on target accessibility, there may be two reasons. First, the accessibility was calculated with probabilities of unpaired individual bases in the Boltzmann ensemble of RNA structures [20]. Although this represents a major improvement over the use of a single structure such as the optimal fold, the accessibility can be arguably better assessed by consideration of free energy changes for siRNA-target hybridization [7]. Second, the assessment of the effect of target accessibility on target recognition and RNAi activity requires controlling for the upstream effect of siRNA duplex asymmetry [7]. The small data set of nine siRNAs is inadequate. The analysis of 137 worm microRNAs suggests that some intramolecular structures can be tolerated for the regulatory functions by microRNAs, and these structures are likely to be weakened or completely abolished upon interaction with the RISC. Target cleavage by RNAi machinery and translation repression by microRNA pathway may have different effects on the structure of the target and the small RNA:target duplex. It is possible that the effects of intramolecular structure can be different between siRNAs and microRNAs, so that the lack of negative effect of structure for siRNAs cannot be simply reasoned by the lack of self-folding effect for microRNAs. Nevertheless, the results of our analyses on the shRNA data do not support the previous conclusion on the significance of the effects of structures of the siRNA guide strands [15].

## Conclusion

Target accessibility as primarily determined by target secondary structure is an important determinant for RNAi potency. The commonly observed negative effect of high siRNA GC-content on RNAi potency is due to generally poor target accessibility for a high GC target site, rather than the likelihood that the high GC siRNA guide strand may form stable intramolecular secondary structure. These findings provide a target-structure-based interpretation and mechanistic insight for the effect of siRNA GC-content on RNAi efficiency.

## Methods

### Short hairpin data set

We used a data set of 101 shRNA sequences targeting 100 different endogenous human genes (i.e., two shRNAs for only one gene). They were obtained from the analysis of a library of shRNA sequences generated from randomly fragmented cDNA of normalized (reduced-redundancy)

cDNA of all of the genes expressed in the MCF-7 human breast carcinoma cells. The generation and testing of the library will be described in detail elsewhere (Maliyekkel, A., Shao, Y., Warholc, N., Cole, K., Ding, Y., and Roninson, I.B., in preparation). shRNA activity was determined by measuring the levels of each target mRNA by real-time PCR, in triplicate. Percent knockdown was calculated from the ratio of mRNA levels with and without doxycycline. This data set (see Additional file 1) was previously employed to assess the effect of target structure on RNAi efficiency [7], and is provided. The siRNAs resulting from shRNA cleavage by Dicer are mostly 19 bp or 20 bp in length (with additional 2-nt 3' overhang), at comparable yields [21]. Because the computational results are highly similar for both lengths, we here report the results for the length of 19 bp.

For several reasons, we chose not to consider other RNAi datasets available in the literature. For example, the large dataset from a Norvatis study [22] was based on a reporter assay for 34 genes, whereas the assay of our dataset measures RNAi activity in an endogenous context for a much larger number of human genes. The heavy overlapping between target sites for several published siRNA datasets [6,23] can introduce a bias in analysis. Because such bias is difficult to assess, we decided to avoid such problem in data selection in our earlier study [7].

### A target-structure based energetic parameter for assessment of target accessibility

A number of approaches have been published for quantifying target site accessibility for rational design of RNA-targeting nucleic acids. Based on target structures predicted by RNA folding algorithms, these methods are either probabilistic or energetic. Probabilistic methods assess the probability that a base or a block of bases is single stranded [20,24,25], whereas energetic methods model the energy exchanges of the hybridization process [7,26-31], thus arguably providing more refined measures of accessibility. For example, consider two target sites with (nearly) equal probability of being single stranded. If one site has high AU, and the other has high GC, then the energetic costs for disrupting the target structure, and the stabilities of the hybrid could be quite different for the two sites. In data analysis for some of our studies, energy measures were observed to give improved correlations than probabilistic measures. Thus, our efforts in recent years have focused on energetic models. Below, we briefly discuss several major methods.

The Sfold structure sample [32,33] allows computation of both probabilistic measures [24] and energetic measures of target accessibility [7,28-31]. It is well established that a single-stranded block of 4–5 nts can facilitate the nucleation step of the hybridization [34,35]. Thus, a moderate

structure sample is sufficient for revealing potential effective sites by using block size of 4 nts for accessibility profiling [24]. The major advantage of using the structure sampling algorithm is that the time consuming partition-function calculation for the whole target sequence only needs to be computed once. Folding constraints such as maximum nucleotide distance  $L$  for two bases to form a pair can be imposed for "local" folding. Such local folding was found to be significant for prokaryotic applications [29]. For prokaryotes, transcription and translation are tightly coupled events so that the target mRNA is unlikely to be able to fold globally. In contrast, eukaryotic mRNAs are first transcribed in nucleus and then transported to cytoplasm where they can conceivably fold globally before they engage in interactions with other molecules in the cytoplasm for regulation of gene translation. Global folding using Sfold sampling algorithm can reveal highly unstructured sites that are well "conserved" in the likely mRNA structure population. These well-predicted sites can be valuable for the selection of effective target sites.

Target site disruption energy,  $\Delta G_{\text{disruption}}$ , is the energy cost of local disruption of the mRNA structure so that the binding site becomes completely single-stranded [7]. A largely single-stranded (i.e., structurally accessible) site does not require substantial structure alteration for the guide siRNA strand to bind to the target.  $\Delta G_{\text{disruption}}$  is a quantitative measure of the structural accessibility at the target site, and is calculated based on target secondary structures predicted by Sfold [32] to address the likely population of mRNA structures.

An alternative to the local disruption assumption is the global disruption model. For this model, as a result of siRNA:mRNA hybridization, the base pairs outside the target site can be rearranged so that the mRNA adopts a new globally altered structure. In this case,  $\Delta G_{\text{after}}$  must be calculated by refolding the mRNA with the binding site constrained to be unpaired. This constraint option has been implemented in Sfold and available through the Sfold web server [32]. However, refolding will cost a hefty computational price. This global model is essentially equivalent to an approach based on exact calculation of ensemble free energies from initial folding and refolding [27]. This approach makes the assumption that the target will re-establish structure equilibrium after siRNA binding. The analysis of siRNA datasets in our study suggests that target cleavage by RNAi machinery appear to be rather rapid so that the target may not have time to refold before cleavage [7]. This issue warrants further investigation.

An extension of the McCaskill algorithm [20] can compute the probability that a block of nucleotides is single stranded [25]. However, for each block, this extension

requires re-computation of the partition functions for the entire RNA and is too time consuming to be efficient for scanning through all possible blocks of a long RNA in the search of best target sites. To handle this problem for RNAi application, a short local RNA folding window of size  $W$  was used, along with  $L$  and block length  $u$  [23]. These treatments introduce substantial uncertainty in computational analysis. Indeed, for  $u$ , the empirically selected optimal values are quite different for two training datasets [23], raising the concern of the general applicability of optimal parameter values learned from one source of data. For a specific mRNA, because it is not possible to have accurate information on its independent folding domains which may be better predicted individually, the overall prediction accuracy would be compromised by a pre-specified local folding window length that does not suit this specific mRNA. The major findings from this study are the same as we previously reported [7], i.e., target accessibility as a down stream factor in the RNAi pathway and duplex asymmetry for facilitating RISC assembly [36,37] are two most important factors for RNAi efficiency. The study also compared predictive performance with other methods, but using only 12 data points from an independent test dataset of 360 siRNAs. There was no comparison involving  $\Delta G_{\text{disruption}}$ , our energetic parameter for measuring target site accessibility [7]. For a complete comparison of methods, correlation analysis and other statistical analyses such regression with significance assessment would need to be performed for the whole test dataset and preferably other RNAi datasets from different experimental systems.

Clearly, more studies and analyses would be needed to compare these methods and to further investigate relevant issues such as the validity of global or local folding, but such studies and analyses are beyond the focus of this work. Here, we will employ our own parameter,  $\Delta G_{\text{disruption}}$ , for calculating target site accessibility.

#### Other parameters

For measuring the stability of structure of the siRNA guide strand, the minimum free energy (MFE) of the optimal folding was computed with mfold [38]. Because the folding space is rather small for a tiny siRNA, we considered the use of the optimal fold adequate. For the optimal fold of the siRNA guide strand, we computed the number of free-dangling nucleotides (nts) at the 3' end, the number of free-dangling nucleotides (nts) at the 5' end. These numbers of free nts and the number of free 3' nts with at least two free 5' nts were reported to be highly correlated with RNAi activity [15]. For our dataset, there were at least two free 5' nts for 80 of the 101 shRNAs. In addition, the GC % of the siRNA guide strand was computed.

### Structural RNA data set

To estimate frequency of GC in paired or unpaired regions of structural RNAs with secondary structures elucidated from comparative analysis, we considered a representative set of 81 RNA sequences that was used in a previous work [39]. The set included 10 tRNAs, 10 5S rRNAs, 10 RNase P RNAs, 10 SRP RNAs, 10 tmRNAs, nine group I introns, two group II introns, 10 16S rRNAs, 10 23S rRNA.

### Competing interests

The authors declare that they have no competing interests.

### Authors' contributions

CYC, CSC, DDL, AM and YS performed computational analyses. AM generated the shRNA data under the supervision of IR and made the first observation of correlation between GC content and target disruption energy. YD supervised the computational work and wrote the computational portions of the manuscript. IR drafted the description of the shRNA dataset. All authors read and approved the final manuscript.

### Acknowledgements

The Computational Molecular Biology and Statistics Core at the Wadsworth Center is acknowledged for providing computing resources for this work. The authors thank Andrew Reily for suggesting partial correlation analysis. This work was supported in part by National Science Foundation grants DMS-0200970, DBI-0650991 and National Institutes of Health grant R01 GM068726 to Y.D., and National Institutes of Health grants R33 CA95996, R01 CA62099 and R01 AG17921 to I.B.R.

This article has been published as part of *BMC Bioinformatics* Volume 10 Supplement 1, 2009: Proceedings of The Seventh Asia Pacific Bioinformatics Conference (APBC) 2009. The full contents of the supplement are available online at <http://www.biomedcentral.com/1471-2105/10?issue=S1>

### References

1. Fire A, Xu S, Montgomery MK, Kostas SA, Driver SE, Mello CC: **Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*.** *Nature* 1998, **391(6669)**:806-811.
2. Elbashir SM, Harborth J, Lendeckel W, Yalcin A, Weber K, Tuschl T: **Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells.** *Nature* 2001, **411(6836)**:494-498.
3. Brummelkamp TR, Bernards R, Agami R: **A system for stable expression of short interfering RNAs in mammalian cells.** *Science* 2002, **296(5567)**:550-553.
4. Paddison PJ, Caudy AA, Bernstein E, Hannon GJ, Conklin DS: **Short hairpin RNAs (shRNAs) induce sequence-specific silencing in mammalian cells.** *Genes Dev* 2002, **16(8)**:948-958.
5. Holen T, Amarzguioui M, Wiiger MT, Babaie E, Prydz H: **Positional effects of short interfering RNAs targeting the human coagulation trigger Tissue Factor.** *Nucleic acids research* 2002, **30(8)**:1757-1766.
6. Reynolds A, Leake D, Boese Q, Scaringe S, Marshall WS, Khvorovova A: **Rational siRNA design for RNA interference.** *Nature biotechnology* 2004, **22(3)**:326-330.
7. Shao Y, Chan CY, Maliyekkel A, Lawrence CE, Roninson IB, Ding Y: **Effect of target secondary structure on RNAi efficiency.** *RNA* 2007, **13(10)**:1631-1640.
8. Westerhout EM, Ooms M, Vink M, Das AT, Berkhout B: **HIV-1 can escape from RNA interference by evolving an alternative structure in its RNA genome.** *Nucleic acids research* 2005, **33(2)**:796-804.
9. Heale BS, Soifer HS, Bowers C, Rossi JJ: **siRNA target site secondary structure predictions using local stable substructures.** *Nucleic acids research* 2005, **33(3)**:e30.
10. Kretschmer-Kazemi Far R, Sczakiel G: **The activity of siRNA in mammalian cells is related to structural target accessibility: a comparison with antisense oligonucleotides.** *Nucleic acids research* 2003, **31(15)**:4417-4424.
11. Schubert S, Grunweller A, Erdmann VA, Kurreck J: **Local RNA target structure influences siRNA efficacy: systematic analysis of intentionally designed binding regions.** *J Mol Biol* 2005, **348(4)**:883-893.
12. Luo KQ, Chang DC: **The gene-silencing efficiency of siRNA is strongly dependent on the local structure of mRNA at the targeted region.** *Biochem Biophys Res Commun* 2004, **318(1)**:303-310.
13. Overhoff M, Alken M, Far RK, Lemaitre M, Lebleu B, Sczakiel G, Robbins I: **Local RNA target structure influences siRNA efficacy: a systematic global analysis.** *J Mol Biol* 2005, **348(4)**:871-881.
14. Ameres SL, Martinez J, Schroeder R: **Molecular basis for target RNA recognition and cleavage by human RISC.** *Cell* 2007, **130(1)**:101-112.
15. Patzel V, Rutz S, Dietrich I, Koberle C, Scheffold A, Kaufmann SH: **Design of siRNAs producing unstructured guide-RNAs results in improved RNA interference efficiency.** *Nature biotechnology* 2005, **23(11)**:1440-1444.
16. **The R Project for Statistical Computing** [<http://www.r-project.org>]
17. **Partial correlation coefficient** [<http://www.yilab.gatech.edu/pcor.R>]
18. Ambros V: **The functions of animal microRNAs.** *Nature* 2004, **431(7006)**:350-355.
19. Griffiths-Jones S, Saini HK, van Dongen S, Enright AJ: **miRBase: tools for microRNA genomics.** *Nucleic acids research* 2008, **D154-158**.
20. McCaskill JS: **The equilibrium partition function and base pairing probabilities for RNA secondary structure.** *Biopolymers* 1990, **29(6-7)**:1105-1119.
21. Rose SD, Kim DH, Amarzguioui M, Heidel JD, Collingwood MA, Davis ME, Rossi JJ, Behlke MA: **Functional polarity is introduced by Dicer processing of short substrate RNAs.** *Nucleic acids research* 2005, **33(13)**:4140-4156.
22. Huesken D, Lange J, Mickanin C, Weiler J, Asselbergs F, Warner J, Meloon B, Engel S, Rosenberg A, Cohen D, et al.: **Design of a genome-wide siRNA library using an artificial neural network.** *Nature biotechnology* 2005, **23(8)**:995-1001.
23. Tafer H, Ameres SL, Obernosterer G, Gebeshuber CA, Schroeder R, Martinez J, Hofacker IL: **The impact of target site accessibility on the design of effective siRNAs.** *Nature biotechnology* 2008, **26(5)**:578-583.
24. Ding Y, Lawrence CE: **Statistical prediction of single-stranded regions in RNA secondary structure and application to predicting effective antisense target sites and beyond.** *Nucleic acids research* 2001, **29(5)**:1034-1046.
25. Muckstein U, Tafer H, Hackermuller J, Bernhart SH, Stadler PF, Hofacker IL: **Thermodynamics of RNA-RNA binding.** *Bioinformatics (Oxford, England)* 2006, **22(10)**:1177-1182.
26. Mathews DH, Burkard ME, Freier SM, Wyatt JR, Turner DH: **Predicting oligonucleotide affinity to nucleic acid targets.** *RNA* 1999, **5(11)**:1458-1469.
27. Lu ZJ, Mathews DH: **Efficient siRNA selection using hybridization thermodynamics.** *Nucleic acids research* 2008, **36(2)**:640-647.
28. Shao Y, Wu S, Chan CY, Klapper JR, Schneider E, Ding Y: **A structural analysis of in vitro catalytic activities of hammerhead ribozymes.** *BMC Bioinformatics* 2007, **8(1)**:469.
29. Shao Y, Wu Y, Chan CY, McDonough K, Ding Y: **Rational design and rapid screening of antisense oligonucleotides for prokaryotic gene modulation.** *Nucleic acids research* 2006, **34(19)**:5660-5669.
30. Long D, Chan CY, Ding Y: **Analysis of microRNA-target interactions by a target structure based hybridization model.** *Pac Symp Biocomput* 2008:64-74.
31. Long D, Lee R, Williams P, Chan CY, Ambros V, Ding Y: **Potent effect of target structure on microRNA function.** *Nat Struct Mol Biol* 2007, **14**:287-294.

32. Ding Y, Chan CY, Lawrence CE: **Sfold web server for statistical folding and rational design of nucleic acids.** *Nucleic acids research* 2004:W135-141.
33. Ding Y, Lawrence CE: **A statistical sampling algorithm for RNA secondary structure prediction.** *Nucleic acids research* 2003, **31(24)**:7280-7301.
34. Hargittai MR, Gorelick RJ, Rouzina I, Musier-Forsyth K: **Mechanistic insights into the kinetics of HIV-1 nucleocapsid protein-facilitated tRNA annealing to the primer binding site.** *J Mol Biol* 2004, **337(4)**:951-968.
35. Zhao JJ, Lemke G: **Rules for ribozymes.** *Mol Cell Neurosci* 1998, **11(1-2)**:92-97.
36. Schwarz DS, Hutvagner G, Du T, Xu Z, Aronin N, Zamore PD: **Asymmetry in the assembly of the RNAi enzyme complex.** *Cell* 2003, **115(2)**:199-208.
37. Khvorova A, Reynolds A, Jayasena SD: **Functional siRNAs and miRNAs exhibit strand bias.** *Cell* 2003, **115(2)**:209-216.
38. Zuker M: **Mfold web server for nucleic acid folding and hybridization prediction.** *Nucleic Acids Res* 2003, **31(13)**:3406-3415.
39. Ding Y, Chan CY, Lawrence CE: **RNA secondary structure prediction by centroids in a Boltzmann weighted ensemble.** *RNA* 2005, **11(8)**:1157-1166.

Publish with **BioMed Central** and every scientist can read your work free of charge

*"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."*

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

