Research Article

A Systems Pharmacology Approach for Identifying the Multiple Mechanisms of Action of the Wei Pi Xiao Decoction for the Treatment of Gastric Precancerous Lesions

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The Wei Pi Xiao (WPX) decoction, based on the theory of traditional Chinese medicine, has been widely used for the treatment of gastric precancerous lesions (GPL). Although WPX is known to be effective for the treatment of GPL, its active ingredients, cellular targets, and the precise molecular mechanism of action are not known. This study aimed to identify the multiple mechanisms of action of the WPX decoction in the treatment of GPL. The active compounds, drug targets, and the key pathways involved in the therapeutic effect of WPX in the treatment of GPL were analyzed by an integrative analysis pipeline. The information pertaining to the compounds present in WPX and their disease targets was obtained from TCMSP and GeneCards, respectively. The mechanisms underlying the therapeutic effect of WPX were investigated with gene ontology (GO) enrichment analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis. A total of 82 bioactive compounds and 146 related targets were identified in this study. Following target analyses, the targets were further mapped to 26 key biological processes and 21 related pathways to construct a target-pathway network and an integrated GPL pathway. The study demonstrated that the WPX formula primarily treats the dysfunctions of GPL and revealed the molecular mechanism underlying the therapeutic effects of the WPX formula in GPL. This study offers a novel approach for the systematic investigation of the mechanisms of action of herbal medicines, which will provide an impetus to the GPL drug development pipeline.

1. Introduction

Gastric cancer is the fifth common malignancy in the world and continues to be the most prevalent cancer in Eastern Asia, especially China [1]. Although several therapeutic strategies have been developed for the treatment of gastric carcinoma over the years, the long-term survival rate is low [2]. It is therefore essential to explore novel molecular targets for the treatment of gastric carcinoma. Gastric carcinogenesis is a multistep and continuous process arising from nonatrophic gastritis, which proceeds to atrophic gastritis, metaplasia, and dysplasia and finally to adenocarcinoma [3]. The high mortality rate of patients with gastric cancer can be reduced by the early detection of precancerous lesions [4]. Therefore, the early intervention of gastric precancerous lesions (GPL), which primarily comprise intestinal metaplasia and dysplasia [5], is an effective strategy for preventing the development of gastric cancer.

In China, traditional Chinese medicine (TCM) has been used for more than 4000 years for the treatment of various diseases [6]. Typically, a combination of plants/minerals is incorporated in the formula, which has immense therapeutic potential for the treatment of complex diseases in a multitargeted manner [7]. The Wei Pi Xiao (WPX) decoction is widely used for the treatment of gastrointestinal diseases, owing to its ability to fortify the spleen, enrich the blood, and dissolve

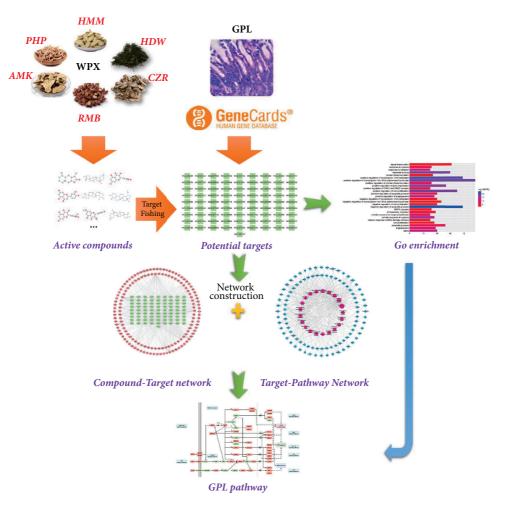


FIGURE 1: The protocol of the systems pharmacology approach used in this study.

stasis. WPX is a complex TCM prescription comprising 6 different herbs, namely, Hedysarum multijugum Maxim (HMM, Huang-qi), Pseudostellaria heterophylla (Miq.) Pax (PHP, Tai-zi-shen), Atractylodes macrocephala Koidz (AMK, Baizhu), Salvia miltiorrhiza Bunge (RMB, Dan-shen), Curcuma zedoaria (Christm.), Roscoe (CZR, E-zhu), and Hedyotis diffusa Willd (HDW, Bai-hua-she-she-cao). HMM, PHP, and AMK, widely used for the treatment of general weakness, have anti-inflammatory and anticarcinogenic activities [8-10] and promote epithelial restitution [11, 12]. RMB and CZR are known to promote blood flow and remove blood stasis according to the theory of TCM, and studies have proven their anti-inflammatory and anticarcinogenic activities [13, 14]. As a traditional heat-clearing and detoxicating herb, HDW is frequently used for the treatment of cancers such as gastric cancer, colorectal cancer, and breast cancer by mediating tumor angiogenesis, proliferation, and apoptosis [15-18]. A previous study demonstrated that WPX can favorably reverse gastric intestinal metaplasia and gastric epithelial dysplasia by blocking the Wnt/ β -catenin pathway [19]. Although the therapeutic efficacy of the WPX formula in the treatment of GPL has been established and the 6 herbs constituting the WPX decoction are known to treat tumors,

the active ingredients, the cellular targets, and the precise molecular mechanism(s) of action of WPX are yet to be known.

A new advanced analytical technique called systems pharmacology has been used in TCM research [20], which has received much attention in recent years. By employing a network-based approach, systems pharmacology is able to systematically identify the effect and mechanism of action of medications used for the treatment of complex diseases at the molecular, cellular, tissue, and organismic levels [21]. This research strategy is being extensively applied in recent years for studying numerous TCM formulas such as the Bushen-Yizhi formula and the Qishen-Yiqi dripping pill, and the efficacies of the formulations have been experimentally verified [22, 23].

This study attempted to investigate the mechanism of action underlying the therapeutic effect of WPX by employing a systems pharmacology approach. The protocol of the integrated systems pharmacology approach used herein is depicted in Figure 1. The active compounds of WPX were first selected on the basis of their oral bioavailability (OB), drug-likeness (DL), and Caco-2 cell permeability, which were evaluated at the molecular level. The targets were then identified by mapping the drug targets with the therapeutic targets of GPL. The targets thus predicted were further mapped to a compound-target network and validated by gene ontology (GO) enrichment analysis. The targets were subsequently used as baits to fish the corresponding pathways from the Kyoto Encyclopedia of Genes and Genomes (KEGG) database. A target-pathway (T-P) network was constructed for further analyses. Finally, an integrated "GPL pathway" was generated for elucidating the molecular mechanism of action of WPX in the treatment of GPL, which offered a novel approach for furthering understanding of TCM.

2. Materials and Methods

2.1. Screening Active Compounds. The compounds present in WPX were identified from the Traditional Chinese Medicines for Systems Pharmacology Database and Analysis Platform (TCMSP; http://lsp.nwu.edu.cn/tcmsp.php), which is a database of Chinese herbal medicines providing information on the relationships between drugs, targets, and diseases [24]. The database contains information on 499 herbs and 12144 compounds, obtained through pharmacological studies and clinical knowledge. The compounds present in the WPX decoction were identified from TCMSP. Barring the 55 identical compounds, a total of 432 compounds were identified, which included the 87 compounds present in HMM, 25 in PHP, 55 in AMK, 202 in RMB, 81 in CZR, and 37 in HDW. In order to identify the potential active compounds present in WPX, the compounds were screened on the basis of OB, DL, and Caco-2 cell permeability.

2.1.1. Evaluation of OB. OB is defined as the percentage of unmodified drug that is absorbed into the circulatory system following oral administration. OB is a reliable indicator of the efficacy of an oral administration for drug delivery [25], and bioactive molecules with high OB often have the potential of being developed into drugs [26]. In this study, the OB of the compounds was calculated by the in-house software, OBioavail1.1, which is reasonably capable of accelerating the prediction of OB [27]. Finally, the herbal components having OB \geq 30% were selected as the candidate molecules for further analyses.

2.1.2. Evaluation of DL. DL is a comparative measure of the functional or physical properties of compounds with those of the majority of known drugs [28]. DL is extensively used for filtering compounds with undesirable properties [29]. Based on the molecular descriptors and the Tanimoto coefficient [30], a self-constructed model was established for calculating the DL index of the compounds in the WPX decoction. The DL was calculated using the following equation:

$$T(\mathbf{A}, \mathbf{B}) = \frac{\mathbf{A} \cdot \mathbf{B}}{\|\mathbf{A}\|^{2} + \|\mathbf{B}\|^{2} - \mathbf{A} \cdot \mathbf{B}}$$
(1)

where **A** is the molecular property of the herbal compound and **B** represents the average molecular property of all the molecules in the DrugBank database (http://www .drugbank.ca/), which were calculated on the basis of Dragon soft descriptors [31]. The compounds with $DL \ge 0.18$ were selected as candidate compounds for subsequent analyses. The threshold value of 0.18 was selected on the basis of the average DL index of all the compounds in DrugBank, which was 0.18.

2.1.3. Evaluation of Caco-2 Cell Permeability. Caco-2 cell permeability is another vital parameter frequently used as a model for studying the passive diffusion of drugs across the intestinal epithelial barrier [32]. In this study, a Caco-2 permeability prediction platform, preCaco-2, was employed for evaluating the drug absorption rate [33], and the baseline Caco-2 cell permeability was set at 0. The threshold values selected for the integrative screening system were OB \geq 30%, DL \geq 0.18, and Caco-2 permeability \geq 0, and the compounds meeting all the three criteria were selected as active compounds for further analyses.

2.2. Identifying the Molecular Targets of WPX. Owing to the diversity of the compounds present in the herbal constituents of WPX, the formulation is capable of targeting multiple proteins, thus making target identification a crucial step in understanding the molecular mechanism underlying the therapeutic properties of WPX. In this study, target prediction was achieved using TCMSP, which uses the SysDT model and the Herb Ingredients' Targets (HIT) database [24]. The SysDT model is developed from two powerful methods, Random Forest (RF) and Support Vector Machine (SVM), and the performance of this model in predicting drug-target interactions is outstanding, with a concordance of 82.83%, a sensitivity of 81.33%, and a specificity of 93.62% [34]. In instances where the targets could not be identified, the Swiss Target Prediction database was used. The Swiss Target Prediction database allows the target prediction of bioactive small molecules based on a combination of 2D and 3D similarity measures with known ligands [35]. In this study, molecular information was retrieved in the form of simplified molecular input line entry specification (SMILES) or Structure from PubChem (https://pubchem.ncbi.nlm.nih.gov/), which is an open chemistry database containing information on the chemical structures, identifiers, chemical and physical properties, biological activities, and other molecular properties of compounds. The compounds were subsequently submitted to the Swiss Target Prediction database for identifying the potential targets, and the confidence score of the prediction probability of a target protein was more than 40%.

Owing to the noncanonical description of the targets thus identified, the UniProt Knowledgebase (UniProtKB; www.uniprot.org/) was used. The identified candidate targets were treated as the query, and the hits categorized under *'Homo sapiens'* were selected from UniProtKB. The molecular targets of the compounds of WPX, along with their gene symbols, were retrieved from the database.

2.3. Disease Targets of GPL. The genes associated with GPL were identified from the Human Gene Database, known as GeneCards (http://www.genecards.org/), which is a searchable, integrative, user-friendly database that provides

comprehensive information on all predicted and annotated human genes, proteins, and diseases [36]. The GeneCards database comprises information from 125 different databases such as HGNC, NCBI, ENSEMBL, and UniProtKB, apart from numerous other related databases, and the information content is considered to be reliable [37]. In order to retrieve information about the related targets from the database, a keyword-based search was performed using the keywords 'gastric precancerous lesions' or 'precancerous lesion of gastric cancer'.

2.4. GO Analysis and KEGG Pathway Enrichment. The GO defines concepts related to gene function and the interrelationships among the functions of different genes. It describes the functions of herbal components in terms of the molecular function(s), the cellular component(s) involved, and the biological process affected [38]. In this study, a GO analysis was performed for understanding the concerned biological processes. KEGG pathway enrichment was additionally performed, for studying the biological effects of the WPX decoction at the pathway level. GO analysis and pathway enrichment were conducted by linking the targets to DAVID (database for annotation, visualization, and integrated discovery; http://david.abcc.ncifcrf.gov). The enriched GO terms and pathways having a false discovery rate (FDR) of less than 0.01 according to Fisher's exact test were selected and subjected to further analyses.

2.5. Network Construction. In order to elucidate the molecular mechanisms underlying the complex therapeutic property of WPX in the treatment of GPL, a compound-target network (C-T network) and a T-P network were generated for studying the relationships among the candidate compounds and the potential disease targets. (1) C-T network: The compound-target interactions were visualized by the C-T network, in which all the active ingredients were connected to their corresponding targets. (2) T-P network: The potential targets identified by preliminary analyses were mapped to the DAVID database for conducting pathway enrichment. The relationships between these potential pathways and GPL were subsequently elucidated by literature mining.

The bipartite graphs were visualized and analyzed by Cytoscape version 3.2.1 [39], which is a robust software for data visualization and integration in Bioinformatics. In this network, the nodes represented the drug compounds and targets, while the edges represented the interactions between them. On the other hand, a vital topological parameter, named degree, was analyzed by the Network Analyzer plugin of Cytoscape [40]. The degree of a node referred to the number of edges connected to the node.

2.6. Construction of the GPL Pathway. For a better understanding of the mechanisms underlying the therapeutic effects of WPX against GPL, an integrated "GPL pathway" was manually constructed based on the T-P network previously generated. The pathways in the T-P network which had no direct and close connections with the disease were removed.

3. Results

3.1. Screening Active Compounds. In order to identify the active ingredients in the WPX decoction, three classical ADME parameters, namely OB, DL, and Caco-2 cell permeability, were employed for screening the compounds. Following screening, 88 potential active compounds, representing 20.37% of the total number of compounds present in WPX, were identified, which included 16 compounds from HMM, 7 from PHP, 7 from AMK, 54 from RMB, 3 from CZR, and 7 from HDW. Since it was possible that the compounds which did not satisfy the screening criteria could also have therapeutic effects in humans, certain compounds were retained as active components on the basis of available information pertaining to their pharmaceutical activities, even if they did not match the screening criteria. For instance, although astragaloside IV (MOL000407, OB = 22.50, DL = 0.15, and Caco-2 permeability = -2.11) has poor OB, DL, and Caco-2 permeability, it was retained as an active compound since it is the major constituent of HMM [41]. Studies have demonstrated that astragaloside IV induces antiinflammatory effects in gastric tissues by suppressing the expression of inflammatory cytokines, such as TNF- α and IL-1 β [42], and restrains epithelial-mesenchymal transitions by inhibiting the PI3K/AKT/NF- kappa B pathway in gastric cancer cells [43]. Additionally, a bioactive constituent of RMB, known as danshensu (MOL007134, OB = 36.91, DL = 0.06, and Caco-2 permeability = -0.27) is a potential antithrombotic and antiplatelet agent, owing to its highly selective inhibition of COX-2 [44]. Atractylenolide I, curcumol, and oleanolic acid were similarly retained as active compounds although they did not satisfy the screening criteria. These 5 compounds of WPX were additionally retained as active compounds. In conclusion, a total of 88 compounds were identified from the WPX decoction on the basis of their biological activities, and a total of 93 compounds were selected as active herbal constituents in this study (represented in Table S1). Table 1 showed parts of compounds from WPX and their corresponding predicted OB, DL, and Caco-2 scores and structure.

3.2. Target Identification and Analysis. Based on the aforementioned target fishing approach, a total of 306 targets were predicted to interact with the 93 compounds identified from the 6 herbs in the WPX decoction. However, the targets specific to GPL as well as their mechanisms of interaction were not known. In order to validate the relevance of these proteins in the development of GPL, the GeneCards database was employed for identifying the disease-related genes. Since GeneCards is a gene-centric database, the disease-associated genes are presented in an integrated web card, representing nearly 90% of the human protein-coding genes [45]. A total of 1261 GPL-related genes were identified from the database by employing a keyword-based search. Upon combining the compound targets of WPX with the disease targets, a total of 146 overlapping genes were selected as the potential targets for the treatment of GPL. By this process, 82 compounds were ultimately selected as the active herbal ingredients after discarding 11 candidate compounds that had no relevant

No.	Mol ID	Molecule name	OB	DL	Caco-2	Structure	Herb
1	MOL000006	Luteolin	36.16	0.25	0.19	HO HO O H	PHP, RMB
2	MOL000043	Atractylenolide I	37.37	0.15	1.30		АМК
3	MOL000098	Quercetin	46.43	0.28	0.05		HMM, HDW
4	MOL000211	Mairin	55.38	0.78	0.73		НММ
5	MOL000239	Jaranol	50.83	0.29	0.61		НММ
6	MOL000263	Oleanolic acid	29.02	0.76	0.59		RMB, HDW
7	MOL000296	Hederagenin	36.91	0.75	1.32	HO	HMM, CZR
8	MOL000354	Isorhamnetin	49.60	0.31	0.31		НММ

TABLE 1: 24 representative components from WPX and their corresponding predicted OB, DL, Caco-2 scores and structure.

	TABLE 1: Continued.								
No.	Mol ID	Molecule name	OB	DL	Caco-2	Structure	Herb		
9	MOL000358	Beta-sitosterol	36.91	0.75	1.32		PHP, HDW		
10	MOL000409	Astragaloside IV	17.74	0.15	(2.22)		НММ		
11	MOL000417	Calycosin	47.75	0.24	0.52		НММ		
12	MOL000422	kaempferol	41.88	0.24	0.26	н о о н о н о н о н о н	HMM		
13	MOL000449	Stigmasterol	43.83	0.76	1.44		HDW		
14	MOL000902	Curcumol	103.55	0.13	1.12		CZR		
15	MOL000906	Wenjine	47.93	0.27	0.30		CZR		
16	MOL001659	Poriferasterol	43.83	0.76	1.44		RMB, HDW		

	TABLE 1: Continued.									
No.	Mol ID	Molecule name	OB	DL	Caco-2	Structure	Herb			
17	MOL001689	Acacetin	34.97	0.24	0.67	o o o o o o o n o n	PHP			
18	MOL006554	Taraxerol	38.40	0.77	1.37		PHP			
19	MOL006756	Schottenol	37.42	0.75	1.33		РНР			
20	MOL007111	Isotanshinone II	49.92	0.40	1.03		RMB			
21	MOL007134	Danshensu	36.91	0.06	-0.27		RMB			
22	MOL007151	Tanshindiol B	42.67	0.45	0.05	H OF OF	RMB			
23	MOL007154	Tanshinone IIA	49.89	0.40	1.05		RMB			
24	MOL007156	Tanshinone VI	45.64	0.30	0.48	OH OH OH OH	RMB			

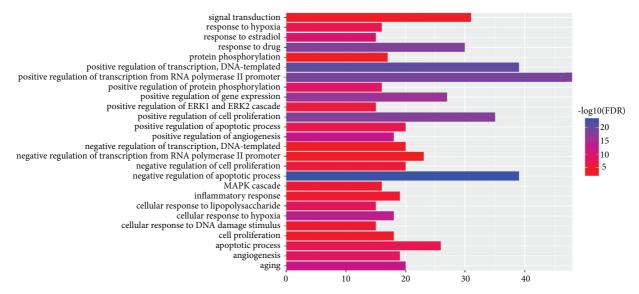


FIGURE 2: Gene ontology (GO) analysis of the target genes associated with GPL. The X-axis represents the significant enrichment counts of these terms, while the Y-axis represents the categories of 'biological process' in the GO of the target genes (FDR ≤ 0.01).

targets. Since the mechanisms underlying the therapeutic effects of TCM formulas are due to the synergistic effects of multiple compounds and targets [46], and since the pivotal targets are the cores of the network from the point of network topology [47], the 146 genes identified herein were considered to be effective therapeutic targets (refer to Table S2).

In order to validate whether the 146 targets are associated with GPL, a GO analysis was performed for elucidating the concerned biological processes. Figure 2 represents the first 26 significantly enriched GO terms (FDR \leq 0.01) for these targets. The *p*-values, FDR, and counts are provided in Table S3. The results indicated that numerous targets are involved in the process of tumorigenesis, including those involved in positive transcriptional regulation by RNA polymerase II promoter (GO:0045944), negative regulation of apoptosis process (GO:0043066), apoptotic process (GO:0006915), positive regulation of cell proliferation (GO:0008284), and inflammatory response (GO:0006954).

3.3. Network Construction and Analysis

3.3.1. Construction and Analysis of the C-T Network. In order to understand the relationships among the herbal constituents in the WPX decoction, the compound targets, and the GPL targets, a C-T network was constructed. The active compounds, targets, and the interactions among them are represented in Figure 3, with 228 nodes, representing the 82 potential compounds and 146 potential targets, and 677 edges. The yellow and green nodes represent the targets and the compounds, while the edges represent the interactions between them. In general, the degree of target interactions is an indicator of the potential significance of the compound the average degree was 4.64, and the average number of edges was 8.26, indicating that the compounds with a high degree might be crucial for the treatment of GPL.

3.3.2. Construction and Analysis of the T-P Network. In this study, 146 targets were mapped to 115 pathways following KEGG pathway enrichment. After combining the pathological data obtained from literature mining with the FDR scores (FDR \leq 0.01), the pathways that had no relationships with GPL were discarded. Finally, 21 remarkably enriched pathways that were likely to be the major pathways in the treatment of GPL were selected (refer Table S4). The T-P network was subsequently generated by mapping 97 of the 146 targets to the major target pathways (Figure 4).

As demonstrated in Figure 4, these targets closely interacted with the pathways involved in cancer (hsa05200, degree = 57), the PI3K-Akt signaling pathway (hsa04151, degree = 37), the MAPK signaling pathway (hsa04010, degree = 24), and the Ras signaling pathway (hsa04014, degree = 23), among others. These pathways are regarded as the key pathways responsible for the progression of GPL.

3.4. The GPL Pathway. Based on the aforementioned results, an integrated "GPL pathway" was constructed by integrating the key pathways, which included the pathways involved in cancer (hsa05200), the PI3K-Akt signaling pathway (hsa04151), the MAPK signaling pathway (hsa04010), the Ras signaling pathway (hsa04014), the FoxO signaling pathway (hsa04068), the HIF-1 signaling pathway (hsa04066), the TNF signaling pathway (hsa04668), the p53 signaling pathway (hsa04064). As shown in Figure 5, the GPL pathway can be separated into three representative therapeutic modules, namely, the cell proliferation module, the cell apoptosis module, and the inflammation module.

3.4.1. The Cell Proliferation Module. The disruption in the balance between the proliferation and apoptosis of gastric

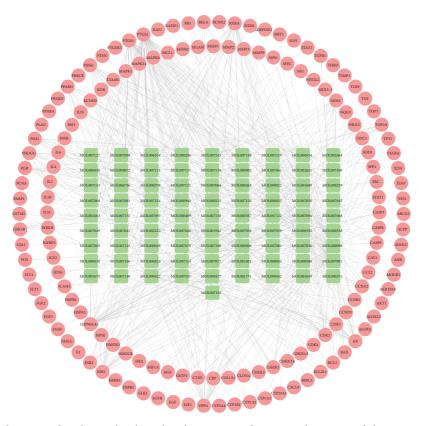


FIGURE 3: The C-T network generated in this study. The red nodes represent the potential targets, and the green nodes represent the herbal compounds, while the lines represent the interactions between them.

epithelial cells is linked to the progression from chronic gastritis, to atrophy, intestinal metaplasia, dysplasia, to ultimately cancer [48]. As demonstrated in Figure 5, the PI3K-Akt and MAPK signaling pathways are closely related to cell proliferation and play an important role in gastric tumourigenesis [49-51]. For instance, the serine/threonine protein kinase, Akt, is involved in regulating a plethora of cellular processes triggered by a wide diversity of extracellular signals and is thus considered to be a key molecule in the PI3K-Akt signaling pathway [52]. Certain compounds in WPX, such as quercetin (MOL000098) from HMM and HDW and luteolin (MOL000006) from PHP and RMB, have been shown to be effective in downregulating the phosphorylation of Akt, which leads to the inhibition of cell proliferation [53, 54]. Additionally, tanshinone IIA (MOL007154), kaempferol (MOL000422), and beta-sitosterol (MOL000358) target JUN, which ultimately interferes with the MAPK signaling pathway, thus inducing apoptosis [55-57]. All these data indicate that the therapeutic effect of WPX in treating GPL could be mediated by regulating cell proliferation.

3.4.2. The Cell Apoptosis Module. Apoptosis is a crucial mechanism leading to cell death, and the failure to inhibit apoptosis induces the formation of certain gastrointestinal malignancies [58]. It has been further demonstrated that apoptosis plays a vital role in the morphogenesis of GPL [59]. As shown in Figure 5, certain targets in the p53 signaling

pathway, the TNF signaling pathway, the Ras signaling pathway, and the FoxO signaling pathway are involved in the process of necrocytosis. For instance, caspase 3 (CASP3), a member of the interleukin-1 beta-converting enzyme family that participates in the TNF and p53 pathways, induces apoptosis [60] and is identified as one of the key effector caspases in the apoptotic machinery [61]. Our results also demonstrated that CASP3 can be regulated by acacetin (MOL001689) from PHP, oleanolic acid (MOL000263) from RMB and HDW, and kaempferol (MOL000422) from HMM. In the Ras and FoxO pathways, the Fas ligand (FasL) that belongs to the TNF family leads to apoptosis upon binding to its receptor and significantly influences the progression of cancer [62]. The aforementioned observations suggest that the therapeutic effect of WPX in the treatment of GPL could be mediated by the induction of apoptosis in the epithelial cells of the gastric mucosa.

3.4.3. The Inflammation Module. Chronic mucosal inflammation is associated with a high risk of progression from chronic gastritis to gastric cancer [63]. Chronic inflammation following infection with *Helicobacter pylori* is recognized as a risk factor for atrophic gastritis, intestinal metaplasia, and adenocarcinoma [64]. As indicated in Figure 5, the inflammatory cytokines including TNF, IL1, IL6, and COX-2 are involved in the TNF signaling pathway, the NF-kappa B signaling pathway, and the MAPK signaling pathway. For

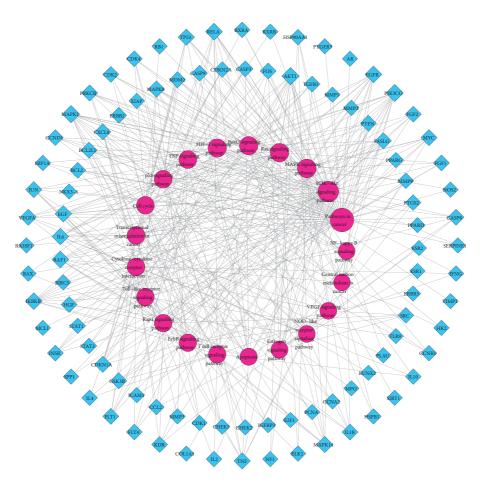


FIGURE 4: The T-P network generated in this study. The blue nodes represent potential targets and the red nodes represent the related pathways. The sizes of the nodes are in proportion to their degree.

instance, COX-2, which is known to induce inflammation and cause tumorigenesis via the NF- kappa B pathway, also participates in the invasion and metastasis of cancer cells [65]. A study demonstrated that danshensu (MOL007134), which is obtained from *RMB*, has COX-2-dependent anticancer properties [66]. These data suggest that the agents that regulate cytokines can suppress inflammation and thereby inhibit the progression of GPL.

4. Discussion

Gastric cancer is one of the most common types of cancer, and the mortality rate has markedly improved [67, 68]. Early diagnosis and treatment of GPL are crucial to reduce the morbidity and mortality of gastric cancer [5, 69]. However, there remain some medical controversies of GPL not solved satisfactorily with current western allopathic therapy[70]. As a useful alternate medicine, TCM is attracting more and more attention across the world for its remarkable effects in clinical practice. WPX, a Chinese herb formula, has been used to treat GPL effectively. More and more evidence shows that herbs and their active compounds in this decoction have biological effects on GPL. Thus, it is imperative to use the systems pharmacology approach combining the screening active components, drug targeting, network, and pathway analysis to explore the therapeutic mechanism of WPX in the treatment of GPL.

The results show that 82 active compounds were obtained from WPX, and 146 potential targets were found to be linked to multiple compounds from different herbs. These indicate that WPX exerts therapeutic effects on GPL through multiple compounds and targets. Among the active compounds linked to the network, quercetin (MOL000098, degree = 104), luteolin (MOL000006, degree = 46), kaempferol (MOL000422, degree = 37), isorhamnetin (MOL000354, degree = 37), acacetin (MOL001689, degree = 20), and astragaloside IV (MOL000409, degree = 4) are well-known bioactive compounds in the treatment of gastric cancer [71-76]. For instance, the compound quercetin from HMM and HDW exhibited the highest degree number of interactions with various protein targets. Being the main dietary flavonoid, quercetin not only functions as a radical-scavenging antioxidant, but also suppresses inflammation for inhibiting carcinogenesis [77]. According to a population-based study, the dietary intake of quercetin can reduce the incidences of stomach cancer, particularly in women who have been exposed to tobacco smoke [78]. Furthermore, the antitumor effect of quercetin is associated with the activation of autophagy

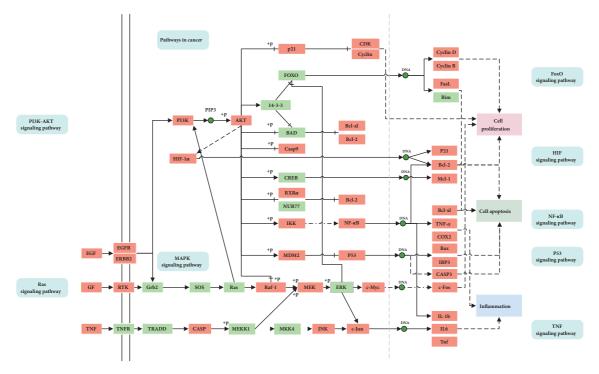


FIGURE 5: The GPL pathway constructed in this study. The orange nodes represent the potential disease protein targets, while the green nodes represent the relevant targets in the pathway.

via modulation of the signaling pathways mediated by AKTmTOR and hypoxia-induced factor 1α (HIF- 1α) [79].

On the other hand, numerous targets were found to be linked to multiple compounds from different herbs, which indicated the synergistic property of the compounds in the WPX decoction in the treatment of GPL. The target PTGS2 (Prostaglandin-Endoperoxide Synthase 2) for instance, also known as COX-2, was connected to 64 active compounds of the WPX formula. COX-2 plays an important role in the progression of GPL [80], and a reduction in the levels of COX-2 is related to the regression of precancerous lesions [81]. It has been revealed that celecoxib, a selective COX-2 inhibitor, has therapeutic effects on the regression of advanced gastric lesions [82]. By analyzing the constituents and targets, the network revealed that the WPX formula may treat GPL via numerous pathways and cellular processes. According to GO enrichment and KEGG pathway enrichment analyses, we infer that WPX may exert a therapeutic effect by interfering with apoptosis and cell proliferation, and mucosal inflammation. For instance, the GO term like RNA polymerase II, one of the RNAP enzymes found in the nucleus of eukaryotic cells [83], is associated with the survival of gastric cancer cells [84]. It has been additionally confirmed that other GO terms such as negative regulation of apoptosis process, apoptotic process, and positive regulation of cell proliferation are closely associated with gastric carcinogenesis [85]. Among the enriched pathways linked to the network, the PI3K-Akt signaling pathway could sense cell growth factors and is frequently activated by genome amplification in gastric cancer [86]. The MAPK signaling pathway, which was first discovered in cancer cells, plays

an important role in transducing the external signals from mitogens into intracellular signaling events that promote cell growth, proliferation, and differentiation [87] and acts as a primary mediator of inflammation during tumor progression [88]. Some pathways such as the TNF signaling pathway (hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa04668, degree = 19), the p53 signaling pathway (hsa04115, hsa0468, degree = 19), the p53 signaling pathway (hsa046868, degree = 19), the p53 signaling pathway (hsa04688, degree = 19), the p53 signaling pathway (hsa048868degree = 18), and pathways involved in cell cycle progression (hsa04110, degree = 18) and apoptosis (hsa04210, degree = 14) have been established as target pathways for the treatment of GPL. Our previous study demonstrated that WPX can suppress the progression of GPL by inducing apoptosis via stimulation of p53 expression [89]. Additionally, certain pathways related to cell proliferation and apoptosis such as the Ras signaling pathway (hsa04014, degree = 23), the VEGF signaling pathway (hsa04370, degree = 12), and the NF-kappa B signaling pathway (hsa04064, degree = 11) have been established as potential target pathways for the treatment of gastric carcinogenesis [90]. It is well known that inflammation plays decisive role at different stages of tumor development, including initiation, promotion, malignant conversion, invasion, and metastasis[91]. WPX can reduce inflammation and reverse the progression of GPL by regulating the expression of TNF[92], which involves in the MAPK signaling pathway, TNF signaling pathway, NODlike receptor signaling pathway, and NF-kappa B signaling pathway. On the basis of the results obtained in this study, we speculate that the WPX decoction targets multiple signaling pathways in an integrated manner to regulate cell proliferation, evade apoptosis, and suppress inflammation.

Although numerous researches have demonstrated that TCM formulations are effective and safe in treating GPL [93], the mechanisms of action are not fully revealed. It is imperative to illustrate the molecular mechanism of WPX in GPL treatment. The result indicated that the pharmacological mechanisms of WPX on GPL might be strongly associated with its synergic modulation on cell proliferation, cell apoptosis, and inflammation. The systems pharmacology approach applied in this work provides a novel way to decipher the underlying mechanisms of WPX and contribute to a better understanding of herbal medicines. However, there was no experimental verification in this work, such as quantitative real-time PCR or western blot analysis to estimate the predictions. Further works are required to confirm the results of this study and to reveal the key mechanisms.

5. Conclusions

GPL is related to the development of gastric cancer that progressively advances through the formation of sequential lesions, and the treatment of precancerous lesions offers an effective measure for decreasing morbidity in the initial phase of gastric carcinogenesis. In this study, we employed a systems pharmacology approach by integrating active compound screening, target prediction, network analysis, and pathway analysis to explore the molecular mechanisms underlying the therapeutic effect of the WPX decoction in the treatment of GPL. The key results of this study are as follows:

- A total of 82 bioactive compounds were identified from WPX, which provided potential clues for investigating the molecular mechanism underlying the therapeutic effect of WPX.
- (2) A total of 146 targets were predicted by comparing the targets of the WPX compounds with the GPL targets, which further demonstrated the characteristic multitargeting property of WPX. The results of GO enrichment and C-T network analysis revealed that WPX exerts a therapeutic effect by interfering with apoptosis and cell proliferation.
- (3) The results of GO enrichment and the integrated GPL pathways revealed that the WPX decoction targets 26 key biological processes and 21 pathways involved in the pathogenesis of GPL, which further demonstrated the three dysfunctions of GPL, namely, cell proliferation, apoptosis, and mucosal inflammation.
- (4) On the whole, this study investigated the mechanisms of action of WPX in the treatment of GPL and provides a novel approach for exploring TCM formulations. The results will advance the cognitive understanding of TCM formulations and promote the therapeutic application of traditional medicines for the treatment of diseases in the present day scenario.

Data Availability

The compounds information from WPX, the targets information about GPL, the GO terms, and the KEGG pathways used to support the findings of this study are included within the supplementary information files.

Conflicts of Interest

There are no conflicts of interest to declare.

Authors' Contributions

Huafeng Pan conceived and designed the study. Liangjun Yang, Wei Liu, Maoyi Yang, Jiali Li, and Xiangzhen Fan carried out the research. Zhipeng Hu and Liangjun Yang approved the final version of the submitted manuscript.

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Supplementary Materials

Table S1: 93 compounds from WPX and their corresponding predicted OB, DL, and Caco-2 scores and structures. 88 compounds that meet the parameters OB \geq 30%, DL \geq 0.18, and Caco-2 \geq 0 were preserved as active compounds. 5 compounds which do not meet the criterion but have been validated with various pharmaceutical activities were also reserved as the active components. Table S2: The information of GPL-related targets. By combining the compound targets of WPX and the disease-related targets, 146 overlapping ones were selected as the key targets in the treatment of GPL. Table S3: The GO terms of therapy target genes and their corresponding count, *p*-values, and FDR. Through the GO enrichment of the key targets, 26 top GO terms were obtained which indicate that large numbers of targets were involved in the process of tumorigenesis. Table S4: The KEGG pathways of therapy target genes and their corresponding count, pvalues, and FDR. Through the KEGG enrichment of the key targets, 21 remarkably enriched pathways involved in cell proliferation, apoptosis, and inflammation were obtained. (Supplementary Materials)

References

- J. Ferlay, I. Soerjomataram, R. Dikshit et al., "Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012," *International Journal of Cancer*, vol. 136, no. 5, pp. E359–E386, 2015.
- [2] F. Sun, H. Sun, X. Mo et al., "Increased survival rates in gastric cancer, with a narrowing gender gap and widening socioeconomic status gap: A period analysis from 1984 to 2013," *Journal of Gastroenterology and Hepatology*, vol. 33, no. 4, pp. 837–846, 2018.
- [3] P. Correa, "Human gastric carcinogenesis: a multistep and multifactorial process—first American Cancer Society Award lecture on cancer epidemiology and prevention," *Cancer Research*, vol. 52, no. 24, pp. 6735–6740, 1992.
- [4] T.-H. Chen, C.-T. Chiu, C. Lee et al., "Circulating microRNA-22-3p Predicts the Malignant Progression of Precancerous

Gastric Lesions from Intestinal Metaplasia to Early Adenocarcinoma," *Digestive Diseases and Sciences*, vol. 63, no. 9, pp. 2301– 2308, 2018.

- [5] L. Marques-Silva, M. Areia, L. Elvas, and M. Dinis-Ribeiro, "Prevalence of gastric precancerous conditions: A systematic review and meta-analysis," *European Journal of Gastroenterol*ogy & Hepatology, vol. 26, no. 4, pp. 378–387, 2014.
- [6] F. Tang, Q. Zhang, Z. Nie, S. Yao, and B. Chen, "Sample preparation for analyzing traditional Chinese medicines," *TrAC Trends in Analytical Chemistry*, vol. 28, no. 11, pp. 1253–1262, 2009.
- [7] X. Wang, A. Zhang, and H. Sun, "Future perspectives of Chinese medical formulae: chinmedomics as an effector," OMICS: A Journal of Integrative Biology, vol. 16, no. 7-8, pp. 414–421, 2012.
- [8] C.-Q. Li, L.-C. He, H.-Y. Dong, and J.-Q. Jin, "Screening for the anti-inflammatory activity of fractions and compounds from *Atractylodes macrocephala* koidz," *Journal of Ethnopharmacol*ogy, vol. 114, no. 2, pp. 212–217, 2007.
- [9] Z. Chen, S. Li, X. Wang, and C. L. Zhang, "Protective effects of Radix Pseudostellariae polysaccharides against exerciseinduced oxidative stress in male rats," *Experimental and Therapeutic Medicine*, vol. 5, no. 4, pp. 1089–1092, 2013.
- [10] J. Fu, Z. Wang, L. Huang et al., "Review of the botanical characteristics, phytochemistry, and pharmacology of *Astragalus membranaceus* (Huangqi)," *Phytotherapy Research*, vol. 28, no. 9, pp. 1275–1283, 2014.
- [11] H.-P. Song, R.-L. Li, X. Chen et al., "Atractylodes macrocephala Koidz promotes intestinal epithelial restitution via the polyamine - Voltage-gated K+ channel pathway," *Journal of Ethnopharmacology*, vol. 152, no. 1, pp. 163–172, 2014.
- [12] S. Adesso, R. Russo, A. Quaroni, G. Autore, and S. Marzocco, "Astragalus membranaceus extract attenuates inflammation and oxidative stress in intestinal epithelial cells via NF-κB activation and Nrf2 response," *International Journal of Molecular Sciences*, vol. 19, no. 3, 2018.
- [13] G.-L. Xu, D. Geng, M. Xie et al., "Chemical composition, antioxidative and anticancer activities of the essential oil: Curcumae rhizoma-sparganii rhizoma, a traditional herb pair," *Molecules*, vol. 20, no. 9, pp. 15781–15796, 2015.
- [14] H. Gao, L. Huang, F. Ding et al., "Simultaneous purification of dihydrotanshinone, tanshinone I, cryptotanshinone, and tanshinone IIA from Salvia miltiorrhiza and their antiinflammatory activities investigation," *Scientific Reports*, vol. 8, no. 1, p. 8460, 2018.
- [15] Z. Liu, M. Liu, M. Liu, and J. Li, "Methylanthraquinone from *Hedyotis diffusa* WILLD induces Ca²⁺-mediated apoptosis in human breast cancer cells," *Toxicology in Vitro*, vol. 24, no. 1, pp. 142–147, 2010.
- [16] J. Lin, L. Wei, A. Shen et al., "Hedyotis diffusa Willd extract suppresses Sonic hedgehog signaling leading to the inhibition of colorectal cancer angiogenesis," *International Journal of Oncology*, vol. 42, no. 2, pp. 651–656, 2013.
- [17] Y. Niu and Q.-X. Meng, "Chemical and preclinical studies on *Hedyotis diffusa* with anticancer potential," *Journal of Asian Natural Products Research*, vol. 15, no. 5, pp. 550–565, 2013.
- [18] C. Wang, X. Zhou, Y. Wang et al., "The Antitumor Constituents from *Hedyotis Diffusa* Willd," *Molecules*, vol. 22, no. 12, Article ID 2101, 2017.
- [19] J.-H. Zeng, H.-F. Pan, Y.-Z. Liu et al., "Effects of Weipixiao (fkfkffk) on Wnt pathway-associated proteins in gastric mucosal epithelial cells from rats with gastric precancerous lesions,"

Chinese Journal of Integrative Medicine, vol. 22, no. 4, pp. 267–275, 2016.

- [20] C. Huang, C. Zheng, Y. Li, Y. Wang, A. Lu, and L. Yang, "Systems pharmacology in drug discovery and therapeutic insight for herbal medicines," *Briefings in Bioinformatics*, vol. 15, no. 5, pp. 710–733, 2014.
- [21] S. I. Berger and R. Iyengar, "Network analyses in systems pharmacology," *Bioinformatics*, vol. 25, no. 19, pp. 2466–2472, 2009.
- [22] Z. Liu, F. Guo, Y. Wang et al., "BATMAN-TCM: A Bioinformatics Analysis Tool for Molecular mechANism of Traditional Chinese Medicine," *Scientific Reports*, vol. 6, Article ID 21146, 2016.
- [23] H. Cai, Y. Luo, X. Yan et al., "The Mechanisms of Bushen-Yizhi Formula as a Therapeutic Agent against Alzheimer's Disease," *Scientific Reports*, vol. 8, no. 1, Article ID 3104, 2018.
- [24] J. Ru, P. Li, J. Wang et al., "TCMSP: a database of systems pharmacology for drug discovery from herbal medicines," *Journal of Cheminformatics*, vol. 6, p. 13, 2014.
- [25] H. Liu, J. Wang, W. Zhou, Y. Wang, and L. Yang, "Systems approaches and polypharmacology for drug discovery from herbal medicines: an example using licorice," *Journal of Ethnopharmacology*, vol. 146, no. 3, pp. 773–793, 2013.
- [26] M. A. Alam, F. I. Al-Jenoobi, A. M. Al-Mohizea, and R. Ali, "Understanding and managing oral bioavailability: Physiological concepts and patents," *Recent Patents on Anti-Cancer Drug Discovery*, vol. 10, no. 1, pp. 87–96, 2015.
- [27] X. Xu, W. Zhang, C. Huang et al., "A novel chemometric method for the prediction of human oral bioavailability," *International Journal of Molecular Sciences*, vol. 13, no. 6, pp. 6964–6982, 2012.
- [28] W. P. Walters and M. A. Murcko, "Prediction of 'drug-likeness," *Advanced Drug Delivery Reviews*, vol. 54, no. 3, pp. 255–271, 2002.
- [29] S. Tian, J. Wang, Y. Li, D. Li, L. Xu, and T. Hou, "The application of in silico drug-likeness predictions in pharmaceutical research," *Advanced Drug Delivery Reviews*, vol. 86, pp. 2–10, 2015.
- [30] Y. Yamanishi, M. Kotera, M. Kanehisa, and S. Goto, "Drugtarget interaction prediction from chemical, genomic and pharmacological data in an integrated framework," *Bioinformatics*, vol. 26, no. 12, pp. i246–i254, 2010.
- [31] A. Mauri, V. Consonni, M. Pavan, and R. Todeschini, "DRAGON software: An easy approach to molecular descriptor calculations," *Match Communications in Mathematical & in Computer Chemistry*, vol. 56, no. 2, pp. 237–248, 2006.
- [32] P. Artursson and J. Karlsson, "Correlation between oral drug absorption in humans and apparent drug permeability coefficients in human intestinal epithelial (Caco-2) cells," *Biochemical and Biophysical Research Communications*, vol. 175, no. 3, pp. 880–885, 1991.
- [33] L. Li, Y. Li, Y. Wang, S. Zhang, and L. Yang, "Prediction of human intestinal absorption based on molecular indices," *Journal of Molecular Science*, vol. 23, no. 4, pp. 286–291, 2007.
- [34] H. Yu, J. Chen, X. Xu et al., "A systematic prediction of multiple drug-target interactions from chemical, genomic, and pharmacological data," *PLoS ONE*, vol. 7, no. 5, Article ID e37608, 2012.
- [35] D. Gfeller, A. Grosdidier, M. Wirth, A. Daina, O. Michielin, and V. Zoete, "SwissTargetPrediction: A web server for target prediction of bioactive small molecules," *Nucleic Acids Research*, vol. 42, no. 1, pp. W32–W38, 2014.

- [36] M. Rebhan, V. Chalifa-Caspi, J. Prilusky, and D. Lancet, "GeneCards: integrating information about genes, proteins and diseases," *Trends in Genetics*, vol. 13, no. 4, p. 163, 1997.
- [37] M. Safran, I. Dalah, J. Alexander et al., "GeneCards Version 3: the human gene integrator," *Database*, vol. 2010, Article ID baq020, 2010.
- [38] M. Ashburner, C. A. Ball, J. A. Blake et al., "Gene ontology: tool for the unification of biology," *Nature Genetics*, vol. 25, no. 1, pp. 25–29, 2000.
- [39] G. Su, J. H. Morris, B. Demchak, and G. D. Bader, "Biological network exploration with cytoscape 3," *Current Protocols in Bioinformatics*, vol. 47, pp. 8.13.1–8.13.24, 2014.
- [40] P. Shannon, A. Markiel, O. Ozier et al., "Cytoscape: a software Environment for integrated models of biomolecular interaction networks," *Genome Research*, vol. 13, no. 11, pp. 2498–2504, 2003.
- [41] W.-J. Zhang and B. Frei, "Astragaloside IV Inhibits NF-κB Activation and Inflammatory Gene Expression in LPS-Treated Mice," *Mediators of Inflammation*, vol. 2015, Article ID 274314, 11 pages, 2015.
- [42] S. Qin, K. Huang, Z. Fang, J. Yin, and R. Dai, "The effect of Astragaloside IV on ethanol-induced gastric mucosal injury in rats: Involvement of inflammation," *International Immunopharmacology*, vol. 52, pp. 211–217, 2017.
- [43] J. Zhu and K. Wen, "Astragaloside IV inhibits TGF-βl-induced epithelial-mesenchymal transition through inhibition of the PI3K/Akt/NF-κB pathway in gastric cancer cells," *Phytotherapy Research*, 2018.
- [44] C. Yu, D. Qi, W. Lian et al., "Effects of danshensu on platelet aggregation and thrombosis: in vivo arteriovenous shunt and venous thrombosis models in rats," *PLoS ONE*, vol. 9, no. 11, Article ID e110124, 2014.
- [45] S. Fishilevich, S. Zimmerman, A. Kohn et al., "Genic insights from integrated human proteomics in GeneCards," *Database*, vol. 2016, 2016.
- [46] D. C. Hao and P. G. Xiao, "Network pharmacology: A rosetta stone for traditional chinese medicine," *Drug Development Research*, vol. 75, no. 5, pp. 299–312, 2014.
- [47] T. Korcsmáros, M. S. Szalay, C. Böde, I. A. Kovács, and P. Csermelyt, "How to design multi-target drugs: target search options in cellular networks," *Expert Opinion on Drug Discovery*, vol. 2, no. 6, pp. 799–808, 2007.
- [48] W. K. Leung, J. Yu, K. F. To et al., "Apoptosis and proliferation in Helicobacter pylori-associated gastric intestinal metaplasia," *Alimentary Pharmacology & Therapeutics*, vol. 15, no. 9, pp. 1467–1472, 2001.
- [49] Y.-Y. Hsieh, C.-H. Shen, W.-S. Huang et al., "Resistin-induced stromal cell-derived factor-1 expression through Toll-like receptor 4 and activation of p38 MAPK/ NFκB signaling pathway in gastric cancer cells," *Journal of Biomedical Science*, vol. 21, p. 59, 2014.
- [50] E. K. Kim and E.-J. Choi, "Compromised MAPK signaling in human diseases: an update," *Archives of Toxicology*, vol. 89, no. 6, pp. 867–882, 2015.
- [51] S. S. Singh, W. N. Yap, F. Arfuso et al., "Targeting the PI3K/Akt signaling pathway in gastric carcinoma: a reality for personalized medicine?" *World Journal of Gastroenterology*, vol. 21, no. 43, pp. 12261–12273, 2015.
- [52] G. Risso, M. Blaustein, B. Pozzi, P. Mammi, and A. Srebrow, "Akt/PKB: One kinase, many modifications," *Biochemical Journal*, vol. 468, no. 2, pp. 203–214, 2015.

- [53] M. Zhu, D. Chen, D. Li et al., "Luteolin inhibits angiotensin IIinduced human umbilical vein endothelial cell proliferation and migration through downregulation of src and Akt phosphorylation," *Circulation Journal*, vol. 77, no. 3, pp. 772–779, 2013.
- [54] H.-C. Pan, Q. Jiang, Y. Yu, J.-P. Mei, Y.-K. Cui, and W.-J. Zhao, "Quercetin promotes cell apoptosis and inhibits the expression of MMP-9 and fibronectin via the AKT and ERK signalling pathways in human glioma cells," *Neurochemistry International*, vol. 80, pp. 60–71, 2015.
- [55] S. H. Sook, H.-J. Lee, J.-H. Kim et al., "Reactive oxygen species-mediated activation of AMP-activated protein kinase and c-jun N-terminal kinase plays a critical role in betasitosterol-induced apoptosis in multiple myeloma U266 cells," *Phytotherapy Research*, vol. 28, no. 3, pp. 387–394, 2014.
- [56] C.-C. Su, "Tanshinone IIA inhibits gastric carcinoma AGS Cells through increasing p-p38, p-JNK and p53 but Reducing p-ERK, CDC2 and Cyclin B1 expression," *Anticancer Reseach*, vol. 34, no. 12, pp. 7097–7110, 2014.
- [57] J.-H. Choi, S.-E. Park, S.-J. Kim, and S. Kim, "Kaempferol inhibits thrombosis and platelet activation," *Biochimie*, vol. 115, pp. 177–186, 2015.
- [58] F. G. Que and G. J. Gores, "Cell death by apoptosis: Basic concepts and disease relevance for the gastroenterologist," *Gastroenterology*, vol. 110, no. 4, pp. 1238–1243, 1996.
- [59] M. Ishida, Y. Gomyo, S. Tatebe, S. Ohfuji, and H. Ito, "Apoptosis in human gastric mucosa, chronic gastritis, dysplasia and carcinoma: Analysis by terminal deoxynucleotidyl transferasemediated dUTP-biotin nick end labelling," *Virchows Archiv*, vol. 428, no. 4-5, pp. 229–235, 1996.
- [60] S. Amptoulach, A. C. Lazaris, I. Giannopoulou, N. Kavantzas, E. Patsouris, and N. Tsavaris, "Expression of caspase-3 predicts prognosis in advanced noncardia gastric cancer," *Medical Oncology*, vol. 32, no. 1, p. 416, 2015.
- [61] R. Romagnoli, F. Prencipe, L. C. Lopez-Cara et al., "Synthesis and biological evaluation of alpha-bromoacryloylamido indolyl pyridinyl propenones as potent apoptotic inducers in human leukaemia cells," *Journal of Enzyme Inhibition and Medicinal Chemistry*, vol. 33, no. 1, pp. 727–742, 2018.
- [62] M. Wang and P. Su, "The role of the Fas/FasL signaling pathway in environmental toxicant-induced testicular cell apoptosis: An update," *Systems Biology in Reproductive Medicine*, vol. 64, no. 2, pp. 93–102, 2018.
- [63] T. R. Menheniott, L. O'Connor, Y. T. Chionh et al., "Loss of gastrokine-2 drives premalignant gastric inflammation and tumor progression," *The Journal of Clinical Investigation*, vol. 126, no. 4, pp. 1383–1400, 2016.
- [64] P. Correa, W. Haenszel, C. Cuello, S. Tannenbaum, and M. Archer, "A model for gastric cancer epidemiology," *The Lancet*, vol. 2, no. 7924, pp. 58–60, 1975.
- [65] Z. Chen, M. Liu, X. Liu et al., "COX-2 regulates E-cadherin expression through the NF-κB/Snail signaling pathway in gastric cancer," *International Journal of Molecular Medicine*, vol. 32, no. 1, pp. 93–100, 2013.
- [66] L. Tao, S. Wang, Y. Zhao et al., "Phenolcarboxylic acids from medicinal herbs exert anticancer effects through disruption of COX-2 activity," *Phytomedicine*, vol. 21, no. 11, pp. 1473–1482, 2014.
- [67] C. Hamashima, "Current issues and future perspectives of gastric cancer screening," World Journal of Gastroenterology, vol. 20, no. 38, pp. 13767–13774, 2014.

- [68] Y. J. Choi and N. Kim, "Gastric cancer and family history," *Korean Journal of Internal Medicine*, vol. 31, no. 6, pp. 1042–1053, 2016.
- [69] V. Pasechnikov, S. Chukov, E. Fedorov, I. Kikuste, and M. Leja, "Gastric cancer: prevention, screening and early diagnosis," *World Journal of Gastroenterology*, vol. 20, no. 38, pp. 13842– 13862, 2014.
- [70] J. K. Sung, "Diagnosis and management of gastric dysplasia," *Korean Journal of Internal Medicine*, vol. 31, no. 2, pp. 201–209, 2016.
- [71] A. Haghi, H. Azimi, and R. Rahimi, "A Comprehensive Review on Pharmacotherapeutics of Three Phytochemicals, Curcumin, Quercetin, and Allicin, in the Treatment of Gastric Cancer," *Journal of Gastrointestinal Cancer*, vol. 48, no. 4, pp. 314–320, 2017.
- [72] M. Zang, L. Hu, B. Zhang et al., "Luteolin suppresses angiogenesis and vasculogenic mimicry formation through inhibiting Notch1-VEGF signaling in gastric cancer," *Biochemical and Biophysical Research Communications*, vol. 490, no. 3, pp. 913– 919, 2017.
- [73] H. Song, J. Bao, Y. Wei et al., "Kaempferol inhibits gastric cancer tumor growth: An in vitro and in vivo study," *Oncology Reports*, vol. 33, no. 2, pp. 868–874, 2015.
- [74] K. A. Manu, M. K. Shanmugam, L. Ramachandran et al., "Isorhamnetin augments the anti-tumor effect of capeciatbine through the negative regulation of NF-κB signaling cascade in gastric cancer," *Cancer Letters*, vol. 363, no. 1, pp. 28–36, 2015.
- [75] M.-H. Pan, C.-S. Lai, P.-C. Hsu, and Y.-J. Wang, "Acacetin induces apoptosis in human gastric carcinoma cells accompanied by activation of caspase cascades and production of reactive oxygen species," *Journal of Agricultural and Food Chemistry*, vol. 53, no. 3, pp. 620–630, 2005.
- [76] Z. F. Wang, D. G. Ma, Z. Zhu et al., "Astragaloside IV inhibits pathological functions of gastric cancer-associated fibroblasts," *World Journal of Gastroenterology*, vol. 23, no. 48, pp. 8512–8525, 2017.
- [77] C. S. Yang, J. M. Landau, M. T. Huang, and H. L. Newmark, "Inhibition of carcinogenesis by dietary polyphenolic compounds," *Annual Review of Nutrition*, vol. 21, pp. 381–406, 2001.
- [78] A. M. Ekström, M. Serafini, O. Nyrén, A. Wolk, C. Bosetti, and R. Bellocco, "Dietary quercetin intake and risk of gastric cancer: results from a population-based study in Sweden," *Annals of Oncology*, vol. 22, no. 2, pp. 438–443, 2011.
- [79] K. Wang, R. Liu, J. Li et al., "Quercetin induces protective autophagy in gastric cancer cells: involvement of Akt-mTORand hypoxia-induced factor 1α-mediated signaling," *Autophagy*, vol. 7, no. 9, pp. 966–978, 2011.
- [80] B.-S. Sheu, H.-B. Yang, S.-M. Sheu, A.-H. Huang, and J.-J. Wu, "Higher Gastric Cycloxygenase-2 Expression and Precancerous Change in Helicobacter pylori-Infected Relatives of Gastric Cancer Patients," *Clinical Cancer Research*, vol. 9, no. 14, pp. 5245–5251, 2003.
- [81] Y. Zhang, K.-F. Pan, L. Zhang et al., "Helicobacter pylori, cyclooxygenase-2 and evolution of gastric lesions: results from an intervention trial in China," *Carcinogenesis*, vol. 36, no. 12, pp. 1572–1579, 2015.
- [82] B. C. Y. Wong, L. Zhang, J.-L. Ma et al., "Effects of selective COX-2 inhibitor and Helicobacter pylori eradication on precancerous gastric lesions," *Gut*, vol. 61, no. 6, pp. 812–818, 2012.
- [83] R. A. Young, "RNA polymerase II," Annual Review of Biochemistry, vol. 60, pp. 689–715, 1991.

- [84] X. Hu, F. Zhang, D. Luo et al., "URI promotes gastric cancer cell motility, survival, and resistance to adriamycin in vitro," *American Journal of Cancer Research*, vol. 6, no. 6, pp. 1420– 1430, 2016.
- [85] H. H. Xia and N. J. Talley, "Apoptosis in gastric epithelium induced by Helicobacter pylori infection: implications in gastric carcinogenesis," *American Journal of Gastroenterology*, vol. 96, no. 1, pp. 16–26, 2001.
- [86] J. Shi, D. Yao, W. Liu et al., "Highly frequent PIK3CA amplification is associated with poor prognosis in gastric cancer," *BMC Cancer*, vol. 12, p. 50, 2012.
- [87] R. J. Orton, O. E. Sturm, V. Vyshemirsky, M. Calder, D. R. Gilbert, and W. Kolch, "Computational modelling of the receptor-tyrosine-kinase-activated MAPK pathway," *Biochemical Journal*, vol. 392, no. 2, pp. 249–261, 2005.
- [88] A. L. Ray, K. L. Berggren, S. R. Cruz, G. N. Gan, and E. J. Beswick, "Inhibition of MK2 suppresses IL-1β, IL-6, and TNFα-dependent colorectal cancer growth," *International Journal of Cancer*, vol. 142, no. 8, pp. 1702–1711, 2018.
- [89] H. F. Pan, J. L. Ren, Z. M. Zhao, and J. Liu, "Effect of Weipixiao on cell generation cycle distribution and apoptosisrelated gene expression in gastric mucosal epithelial cells of gastric precancerous lesion rats with Spleen-deficiency chronic atrophic gastritis," *Journal of Guangzhou University of Traditional Chinese Medicine (Chin)*, vol. 27, no. 05, pp. 488–491, 2010.
- [90] J. Shi, Y.-P. Qu, and P. Hou, "Pathogenetic mechanisms in gastric cancer," *World Journal of Gastroenterology*, vol. 20, no. 38, pp. 13804–13819, 2014.
- [91] S. I. Grivennikov, F. R. Greten, and M. Karin, "Immunity, Inflammation, and Cancer," *Cell*, vol. 140, no. 6, pp. 883–899, 2010.
- [92] H. Li, H. Pan, Z. Zhao et al., "Effect of Weipixiao on Plasma Tumor Necrosis Factor Alpha and Interleukin-4 Expression in Rats with Gastric Precancerous Lesions," *Journal of Guangzhou University of Traditional Chinese Medicine*, vol. 32, no. 02, pp. 271–274, 2015 (Chinese).
- [93] X. J. Mo, C. H. Wei, and G. Z. Cheng, "Efficacy of Chinese Herbal Medicine in Chronic Atrophic Gastritis: A Systematic Review," *Liaoning Journal of Traditional Chinese Medicine*, vol. 40, no. 05, pp. 840–846, 2013.