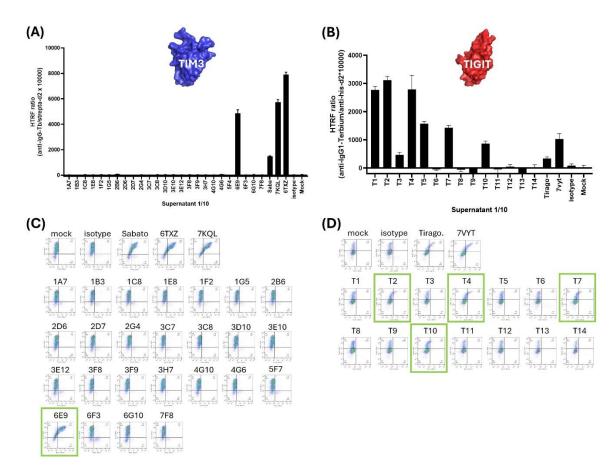
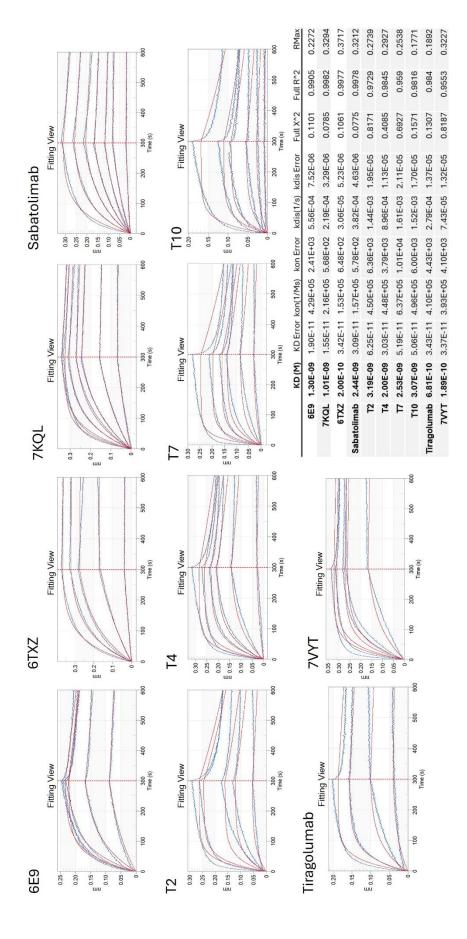


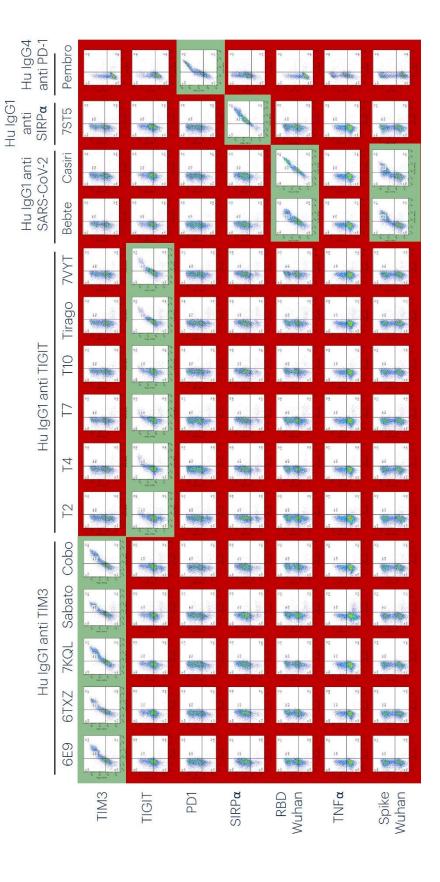
Supplementary Material



Supplementary Figure 1. Binding validation of the IgG1-reformatted scFv identified during phage display screening campaign. The scFv identified by ELISA during the phage display campaign were reformatted as human IgG1 and produced in HEK293 supernatant. (A) and (B) HTRF-based binding validation. The recombinant extracellular domains of TIM3 (A) or TIGIT (B) fused to a biotinylated Avitag, were incubated with the antibodies (supernatant diluted to 1/10). Antibodies and targets were detected with fluorophore-coupled sensors: d2 acceptor coupled to an anti-IgG and terbium donor coupled to the streptavidin. The binding was assessed as the energy transfer between the donor and acceptor and computed as the HTRF ratio: 665nm acceptor emission/620 nm donor emission x 10,000. Curves were fitted mathematically with GraphPad Prism software. (C) and (D) Cytometry-based binding validations. HEK293 cells were transiently transfected with the Flag-tagged full-length TIM3 (C) or TIGIT (D) and incubated with antibodies. The target expression was monitored with a PE-coupled anti-Flag antibody (signal plotted on the y axis), and the binding of the antibodies was followed with an APC-coupled anti-IgG (signal plotted on the x axis). Sabatolimab, 6TXZ, 7KQL, Tiragolumab and 7VYT were used as positive references, and a human anti-IgG1 isotype as a negative control. Green frames highlight binder candidates.

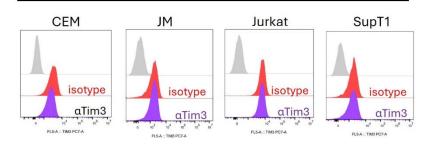


Supplementary figure S2: Complete BLI data. Human IgG1 of the 5 final candidates and reference antibodies were produced in HEK293 supernatant, concentrated and dosed. BLI AHC biosensors were loaded with the candidate antibodies. The association with the target was measured on 3 to 6 concentrations of TIM3 of TIGIT, and the dissociation was passively induced in PBS. KD and other binding kinetic values indicated in the table were obtained from the simultaneous fit of all the shown curves. The binding kinetics (kon, kdis) and affinity constant (KD) for the 5 final candidate antibodies and their references were summarized in the table.

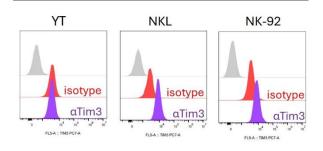


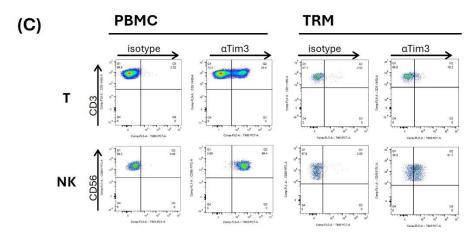
Binding was evaluated as the apparition of a APC+PE+ population (top right quartile). Red frames indicate plots where no binding Supplementary figure S3: Specificity assessment cytometry plots. HEK293 cells were transiently transfected with either one of the Flag-tagged full-length targets (TIM3, TIGIT, PD1, SIRP-α, TNFα, SARS-CoV-2 Spike or its RBD domain) anchored to the membrane via the CD8 transmembrane domain. The target expression was monitored with a PE-coupled anti-Flag antibody (signal plotted on the y axis), and the binding of the antibodies was followed with an APC-coupled anti-IgG (signal plotted on the x axis). was observed between target and antibody while green frames designate plots where antibody binds to the target on HEK293 cells.

(A) Tlines

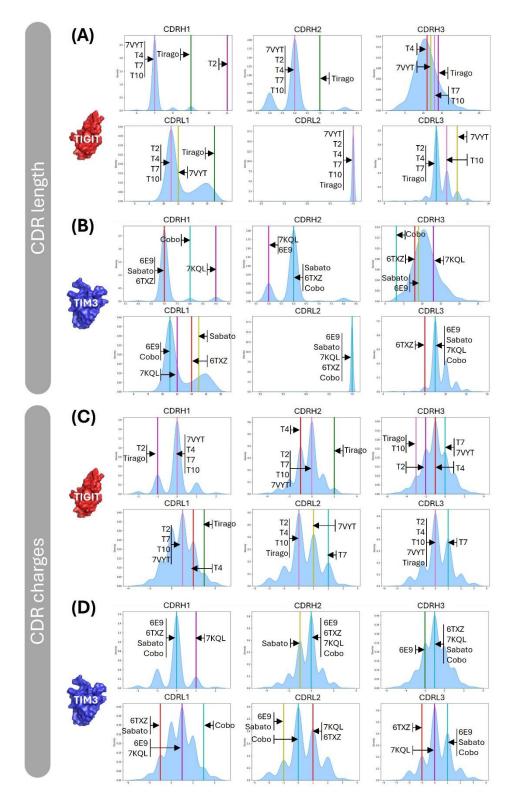


(B) NK lines

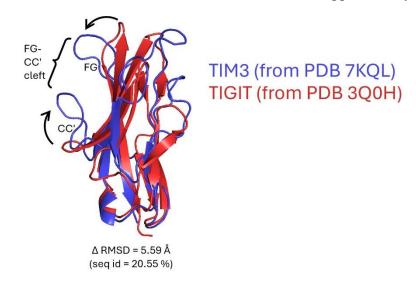


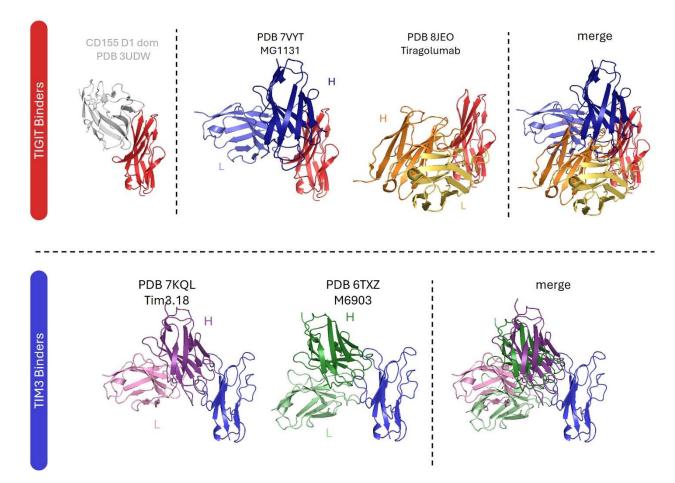


Supplementary figure S4: TIM3 expression on immune-relevant cells. The detection of endogenous TIM3 was performed by flow cytometry with commercial antibody or isotype control. Four T cells lines (CEM, JM, Jurkat and SupT1) (A), three NK cells lines (YT, NKL and NK-92) (B) and primary activated PBMC and TRM cultivated with TGF β and IL15 (C) were investigated. The T and NK subpopulation of the PBMC and TRM were optically isolated by staining CD3 and CD56 (CD3+ T Lymphocytes and CD3- CD56+ NK Lymphocytes).

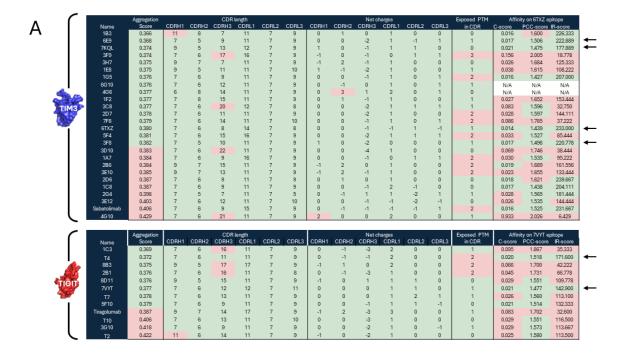


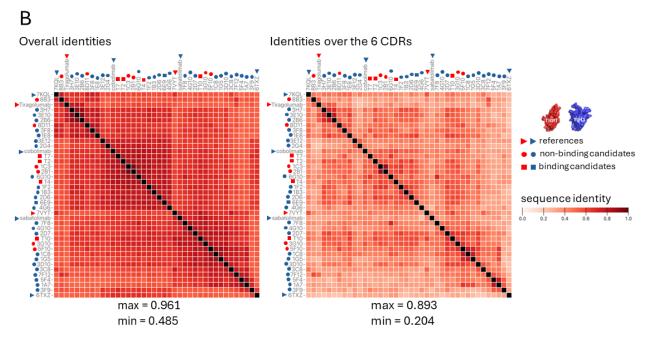
Supplementary figure S5: CDR lengths and charges analysis. The CDR lengths (**A**, **B**) and charges (**C**, **D**) were computed for 735 INN antibodies and their distribution showed in blue. Our anti-TIGIT (**A**, **C**) and anti-TIM3 (**B**, **D**) candidates as well as their references were compared with the INN distribution.





Supplementary figure S6: Visual representation of publicly disclosed anti-TIGIT and anti-TIM3 antibodies structures. The 3D structure overlay of TIGIT (from PDB 3Q0H) and TIM3 (from pdb 7KQL) were represented in red and blue. The FG-CC' cleft is indicated. The 3D structure of CD155 (pdb 3UDW), MG113 (pdb 7VYT), and Tiragolumab (pdb 8JEO) were represented with TIGIT individually or merged together. The 3D structures of Tim3.18 (pdb 7KQL) and M6903 (pdb 6TXZ) were represented with TIM3 individually or merged together.





Supplementary figure S7: Multidimensional analysis of all the candidates identified by phage display. (**A**) Developability and affinity parameters were evaluated for all the anti-TIM3 (upper table) and anti-TIGIT (lower table). Developability analysis includes the predicted aggregation scores, the CDRs lengths and charges analysis, and the number of exposed PTMs motifs in the CDRs. The affinity scores were predicted on 6TXZ epitope on TIM3 or 7VYT epitope on TIGIT, for all but 2 antibodies which revealed aberrant scores (N/A). The values above the threshold are shown in green, the ones below in pink. The arrows show the 6 antibodies exhibited values above the threshold in all the dimensions studied. (**B**) Pairwise sequence identities of the antibodies for their full-length VH and VL sequences (left pannel), and their 6CDRs (right pannels).