



## Draft Genome Sequence of *Plesiomonas shigelloides* Strain zfcc0051 (Phylum *Proteobacteria*)

Zoe K. VanderHoek,<sup>a</sup> Addison M. Browning,<sup>a</sup> Quinn Washburn,<sup>a</sup> Micheal L. Kent,<sup>a</sup> Thomas J. Sharpton,<sup>a,b</sup> (b) Christopher A. Gaulke<sup>a,c,d</sup>

<sup>a</sup>Department of Microbiology, Oregon State University, Corvallis, Oregon, USA <sup>b</sup>Department of Statistics, Oregon State University, Corvallis, Oregon, USA <sup>c</sup>Department of Pathobiology, University of Illinois Urbana-Champaign, Urbana, Illinois, USA <sup>d</sup>Carl R. Woese Institute for Genomic Biology, University of Illinois Urbana-Champaign, Urbana, Illinois, USA

**ABSTRACT** Here, we report a draft genome sequence of *Plesiomonas shigelloides* strain zfcc0051, an isolate derived from zebrafish (*Danio rerio*) feces. The genome consists of 115 contigs (>500 bp) and has a total assembly length of 4,041,537 bases.

P lesiomonas shigelloides is a facultative, anaerobic, non-spore-forming, Gram-negative bacillus frequently found in freshwater ecosystems, freshwater fish guts, and occasionally, aquatic mammals (1, 2). In laboratory and wild zebrafish, *Plesiomonas shigelloides* abundance is high (3), and it has been identified as a potential indicator of health (4, 5). In contrast, other reports indicate that *P. shigelloides* can act as a pathogen of zebrafish (2, 6); however, direct exposure with *P. shigelloides* does not consistently yield infection (7). One possible explanation for the conflicting evidence surrounding the associations between *P. shigelloides* and health is that genetic variation among isolates gives rise to strains that can differentially impact fish physiology. However, the limited number of publicly available genomes for *Plesiomonas* spp. complicates comparative genomic investigations that could shed light on this question. To facilitate such investigations, we sequenced and annotated a microbial isolate from zebrafish gut associated with *Plesiomonas shigelloides*.

A fresh fecal sample was collected from a 9-L tank containing approximately 30 12month-old tropical 5D line laboratory-raised zebrafish (Corvallis, Oregon), aseptically syringe-homogenized, serially diluted, plated on brain heart infusion (BHI) agar (BD), and incubated at 27°C. Individual colonies were picked and further isolated with two rounds of streak plating. One isolated colony was used to inoculate BHI broth and was incubated at 27°C for 24 h. Bacterial DNA was isolated from this pure culture using the UltraClean microbial DNA isolation kit (Qiagen) and subsequently used for 16S amplicon and genome sequencing. A 1,400-bp fragment of the 16S rRNA gene was amplified using 27F and 1492R (8) primers, purified with the UltraClean PCR cleanup kit (Qiagen), sequenced on an ABI 3730 DNA analyzer (Applied Biosystems), and aligned to the NCBI 16S rRNA database using BLAST v2.9.0 (9). The best alignment (97.4% identity) for this culture was associated with Plesiomonas shigelloides (GenBank version number NR\_117763.1). Next, we generated and sequenced a bacterial DNA library using the Nextera XT kit and a MiSeq instrument. The resulting 3,234,209 300-bp paired-end reads were subsequently filtered and trimmed using ea-utils v1.04 (10). SPAdes v3.1.1 (11) was used to assemble a 4,041,537-bp genome in 604 contigs (115 to contigs > 500 bp) with an N<sub>50</sub> value of 187,457 (all contigs), 51.7% GC content, and  $480 \times$  coverage. Genome completeness was assessed as 100% by examining the presence of 40 universal marker genes (CheckM v1.1.2) (12). Similar to other P. shigelloides strains, Prodigal v2.6.3 (13) and the NCBI Prokaryotic Genome Annotation Pipeline (v5.1) identified 3,952 and 3,693 protein-coding genes in our assembly, respectively. Given the identity between the 16S rRNA gene of our isolate and P. shigelloides, we

**Editor** Leighton Pritchard, SIPBS, University of Strathclyde

**Copyright** © 2022 VanderHoek et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Christopher A. Gaulke, cgaulke@illinois.edu.

The authors declare no conflict of interest.

Received 2 February 2022 Accepted 13 May 2022 Published 31 May 2022 used fastANI v1.32 (14) to calculate the average nucleotide identity (ANI) of our isolate and a reference *P. shigelloides* strain, NCTC10360 (GenBank version number GCA\_900087055.1). These genomes shared 97.3% ANI, consistent with the determination that our isolate represents a novel strain of *P. shigelloides*.

**Data availability.** This whole-genome shotgun project has been deposited at DDBJ/ ENA/GenBank under the accession number JAFNAA000000000. The version described in this paper is version JAFNAA010000000. The raw sequence reads are deposited at the Sequence Read Archive under number SRP357730. Workflow with parameters is available at https://github.com/chrisgaulke/zfcc.

## **ACKNOWLEDGMENTS**

The Center for Quantitative Life Sciences at Oregon State University made this work possible via Sanger and Illumina sequencing.

Funding for this project was imparted by an Oregon State University Postdoctoral Association professional development award and institutional funds to C.A.G., an NSF DEB award (1557192) to T.J.S., a National Institute of Environmental Health Sciences award (1R01ES030226) to T.J.S., and an NIH ORIP award (R24OD010998) to M.L.K.

## REFERENCES

- Gu W, Gonzalez-Rey C, Krovacek K, Levin RE. 2006. Genetic variability among isolates of Plesiomonas shigelloides from fish, human clinical sources and fresh water, determined by RAPD typing. Food Biotechnol 20:1–12. https://doi.org/10.1080/08905430500522030.
- Janda JM, Abbott SL, McIver CJ. 2016. Plesiomonas shigelloides revisited. Clin Microbiol Rev 29:349–374. https://doi.org/10.1128/CMR.00103-15.
- Roeselers G, Mittge EK, Stephens WZ, Parichy DM, Cavanaugh CM, Guillemin K, Rawls JF. 2011. Evidence for a core gut microbiota in the zebrafish. ISME J 5:1595–1608. https://doi.org/10.1038/ismej.2011.38.
- Gaulke CA, Martins ML, Watral VG, Humphreys IR, Spagnoli ST, Kent ML, Sharpton TJ. 2019. A longitudinal assessment of host-microbe-parasite interactions resolves the zebrafish gut microbiome's link to Pseudocapillaria tomentosa infection and pathology. Microbiome 7:10. https://doi .org/10.1186/s40168-019-0622-9.
- Gaulke CA, Barton CL, Proffitt S, Tanguay RL, Sharpton TJ. 2016. Triclosan exposure is associated with rapid restructuring of the microbiome in adult zebrafish. PLoS One 11:e0154632-20. https://doi.org/10.1371/ journal.pone.0154632.
- Hansen JD, Woodson JC, Deas E, McPherson V, Welch TJ. 2016. An outbreak of Plesiomonas shigelloides in zebrafish. Fish Shellfish Immunol 53: 95–96. https://doi.org/10.1016/j.fsi.2016.04.034.
- Kent ML, Sanders JL, Spagnoli S, Al-Samarrie CE, Murray KN. 2020. Review of diseases and health management in zebrafish Danio rerio (Hamilton 1822) in research facilities. J Fish Dis 43:637–650. https://doi.org/10.1111/ jfd.13165.

- Jiang H, Dong H, Zhang G, Yu B, Chapman LR, Fields MW. 2006. Microbial diversity in water and sediment of Lake Chaka, an athalassohaline lake in Northwestern China. Appl Environ Microbiol 72:3832–3845. https://doi .org/10.1128/AEM.02869-05.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- Aronesty E. 2013. Comparison of sequencing utility programs [Abstract]. Open Bioinforma J 7:1–8. https://doi.org/10.2174/1875036201307010001.
- Prjibelski A, Antipov D, Meleshko D, Lapidus A, Korobeynikov A. 2020. Using SPAdes de novo assembler. Curr Protoc Bioinformatics 70:e102. https://doi.org/10.1002/cpbi.102.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome Res 25:1043–1055. https:// doi.org/10.1101/gr.186072.114.
- Hyatt D, Chen G-L, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. https://doi.org/10.1186/1471-2105 -11-119.
- Jain C, Rodriguez-R LM, Phillippy AM, Konstantinidis KT, Aluru S. 2018. High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. Nat Commun 9:1. https://doi.org/10.1038/s41467 -018-07641-9.