



Complete Mitochondrial Genome of *Echinostoma hortense* (Digenea: Echinostomatidae)

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Abstract: *Echinostoma hortense* (Digenea: Echinostomatidae) is one of the intestinal flukes with medical importance in humans. However, the mitochondrial (mt) genome of this fluke has not been known yet. The present study has determined the complete mt genome sequences of *E. hortense* and assessed the phylogenetic relationships with other digenean species for which the complete mt genome sequences are available in GenBank using concatenated amino acid sequences inferred from 12 protein-coding genes. The mt genome of *E. hortense* contained 12 protein-coding genes, 22 transfer RNA genes, 2 ribosomal RNA genes, and 1 non-coding region. The length of the mt genome of *E. hortense* was 14,994 bp, which was somewhat smaller than those of other trematode species. Phylogenetic analyses based on concatenated nucleotide sequence datasets for all 12 protein-coding genes using maximum parsimony (MP) method showed that *E. hortense* and *Hypoderaeum conoideum* gathered together, and they were closer to each other than to Fasciolidae and other echinostomatid trematodes. The availability of the complete mt genome sequences of *E. hortense* provides important genetic markers for diagnostics, population genetics, and evolutionary studies of digeneans.

Key words: *Echinostoma hortense*, complete mitochondrial DNA, echinostomatidae, phylogenetic analysis

INTRODUCTION

About 500 species of echinostomatid flukes (Echinostomatidae) had been reported in the world [1], and at least 20 species belonging to 8 genera can infect humans [2]. Among these genera, *Echinostoma* is the largest one, which constitutes 7 species, including *E. hortense*, *E. angustitestis*, *E. cinetorchis*, *E. echinatum*, *E. ilocanum*, *E. macrorchis*, and *E. revolutum* [2]. Although the majority of people infected by echinostomes have no obvious symptoms, severe infections may cause anorexia, lower extremity edema, anemia, weight loss, and dysplasia [3].

The adult worms of *E. hortense* often inhabit the small intestine of humans and many animals [3]. The parasitic fluke *E. hortense* was originally described from rats in Japan [4], and since then, it has been reported in South Korea and China [2,3,5,6]. In addition to rats, it has also been found from cats, dogs, and pigs [6-8], and more importantly, there are many infected human cases in China, Japan, and South Korea [2,3,9-

11]. A high prevalence of human infection by *E. hortense* was reported in Koje-myon, Kochang-gun, Kyongsangnam-do (Province), South Korea, up to 9.5% [11].

Mitochondrial (mt) genome sequences provide valuable genetic markers for investigating population genetics, systematics, and phylogenetics of animal species [12,13]. So far, there are about 18,000 species of digenean trematodes found in the world [14], but only 32 species have complete mt genomes sequenced. In the family Echinostomatidae, only 1 species *Hypoderaeum conoideum* (KM111525) had complete mt genome reported, and no mt genome of genus *Echinostoma* has been available. Previous studies on *E. hortense* have mainly focused on their morphology, life history, and epidemiological investigations [3-11]. However, the mitochondrial genome of this fluke has not been known yet. Therefore, in the present study, we have determined the complete mt genome sequences of *E. hortense* and inferred its phylogenetic relationship with other digenean trematodes.

MATERIALS AND METHODS

Parasites and total genomic DNA isolation

Adult *E. hortense* worms were collected from the small intestine of a naturally infected dog in Daqing, Heilongjiang Prov-

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ince of China. The trematodes were washed in physiological saline, identified morphologically as *E. hortense* based on existing keys and descriptions [15], fixed in 70% (v/v) ethanol (Yixin Biological Technology Co., Shanghai, China), and stored at -20°C until use. Total genomic DNA was extracted from individual fluke using a TIAnamp Genomic DNA Kit (TIANGEN, Beijing, China) according to the manufacturer's instructions, and DNA sample was stored at -20°C until use. In order to independently verify the identity of the specimens, the internal transcribed spacer 2 (ITS2) was amplified from the extracted genomic DNA by PCR and sequenced according to the conventional method. The ITS2 sequences obtained were perfectly matched with that of *E. hortense* available in GenBank (U58101.1).

Amplification, sequencing, and assembling of mt DNA fragments

Amplification, sequencing, and assembly of mtDNA fragments were performed according to the methods previously described [12,16]. The 7 pairs of oligonucleotide primers were designed based on the conserved regions from published complete mtDNA sequences of *Fasciola hepatica*, *Opisthorchis felinus*, *Paramphistomum cervi*, and *H. conoideum* (Table 1). The cycling conditions were 94°C for 5 min (initial denaturation); then 94°C for 1 min (denaturation), 50°C for 1 min (annealing), 72°C for 1-3 min (extension) for 35 cycles, and a final extension at 72°C for 7 min. Each PCR reaction yielded a single band, detected in a 1.0% (w/v) agarose gel stained with ethidium-bromide. PCR products were directly sequenced on an ABI 3370 DNA sequencer at Shanghai Sangon Biotech Company (Shanghai, China) using a primer walking strategy. The

complete mtDNA sequences of *E. hortense* were assembled using DNASTar software as a sequence editor [17].

Sequence analysis of *E. hortense* mt genome

Gene annotation, genome organization, translation initiation, translation termination codons, and the boundaries between protein-coding genes of mtDNA were identified based on comparison with mt genomes of other trematodes reported previously [18]. Open reading frames and gene boundaries were confirmed by comparing with *H. conoideum* mt genome nucleotide sequences (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). Translation initiation and translation termination codons were identified using genetic codon table for mitochondrion in MEGA 5 [19] and also based on comparison with the mt genomes of the trematodes reported previously [16,18]. The codon usage profiles of 12 protein-coding genes (PCGs) and their nucleotide composition were calculated using Geneious 6.1.5 program (Biomatters Co., Auckland, New Zealand) [20]. For the analyses of transfer RNA genes, putative secondary structures of 22 tRNA genes were identified using the program tRNAscan-SE (<http://lowelab.ucsc.edu/tRNAscan-SE>), or by visual identification combined with manual proofreading through anticodon and tRNA secondary structures of trematodes which are available in GenBank.

Phylogenetic analysis

To assess the phylogenetic relationship of *E. hortense* with other digenean trematodes, the complete mt genome sequences of 24 trematode species were retrieved from GenBank and employed for phylogenetic analysis. The mtDNA sequences were as follows: *Clinostomum complanatum* (KM923964.1), *Calicophoron microbothrioides* (KR337555.1), *Clonorchis sinensis* (JF729303), *Dicrocoelium chinensis* (NC_025279), *Dicrocoelium dendriticum* (NC_025280), *Fasciola gigantica* (KF543342), *F. hepatica* (NC_002546), *Haplorchis taichui* (KF214770), *H. conoideum* (KM111525), *Metagonimus yokogawai* (KC330755), *Og-mocotyle sikae* (KR006934.1), *Opisthorchis felinus* (EU921260), *O. viverrini* (JF739555), *P. cervi* (KF475773), *Paramphistomum leydeni* (KP341657.1), *Paragonimus westermani* (NC_002354), *Schistosoma haematobium* (DQ157222), *S. japonicum* (NC_002544), *S. mansoni* (NC_002545), *S. mekongi* (NC_002529), *S. spindale* (DQ157223), *S. turkestanicum* (HQ283100), *Trichobilharzia re-genti* (NC_009680), and using Monogenea species *Gyrodactylus thymalli* (NC_009682) as the outgroup. The nucleotide alignments involved PCGs were generated on the basis of the pro-

Table 1. Sequences of primers used to amplify PCR fragments of mitochondrial genome from *Echinostoma hortense*

Primer name	Sequence of primer (5'-3')	Size (kb)
EH-1	F: AATATGCAAACACACCCGACTG R: CGATGTTGCGTTCTGTAGAGTCTAAGG	~3.8
EH-2	F: GCCTCCTGTGTTTGTCTCTCGCTTTA R: CTCGCACCATATCCCAATTAGCC	~2.2
EH-3	F: GGTTTATTACAGAGGTTT R: ATAACCATAGTAACAGAA	~2.5
EH-4	F: TCTTGTTCTTGCTATGTTGGCTATT R: TTTAGCGGACCATCTCTTACAC	~2.3
EH-5	F: AGCCAGGTCGGTTCCTATC R: ACCACACAACCTCCGTACAATAAC	~1.4
EH-6	F: CGTTAGGGTGTGTCCAACCTGT R: AGCAACAATCTTCTTTAAGTCCATA	~1.7
EH-7	F: GTGGGAGTATTAGGTCTTGTCCAGG R: GAACACCACTAGAATAAGGAATTAC	~4.7

tein alignment using codon alignment amino acid sequences for 12 PCGs, which were individually aligned utilizing Clustal X 1.83 under default parameters [21]. Phylogenetic analyses were performed with the maximum parsimony (MP) method. Phylogenetic reconstructions were carried out using PAUP* Version 4.0b10 [22]. Phylograms were drawn using the Tree View program version 1.65 [23].

RESULTS

Features of the mt genome of *E. hortense*

The circular complete mt genome of *E. hortense* (GenBank accession no. KR062182) was 14,994 bp in length (Fig. 1). The mt genome encoded 36 genes: 12 PCGs (*cox1-3*, *nad1-6*, *nad4L*, *atp6*, *cytb*), 22 transfer RNA genes, and 2 ribosomal RNA genes (*rrnL* and *rrnS*). All genes were transcribed in the same direction. The relative positions and lengths of each gene are given in Table 2. The nucleotide contents in the mt genome were 41.77% (T), 21.26% (A), 24.79% (G), and 12.18% (C), and the overall A+T content of the mt genome was 63.03%.

A total of 3,349 amino acids were encoded by the *E. hortense*

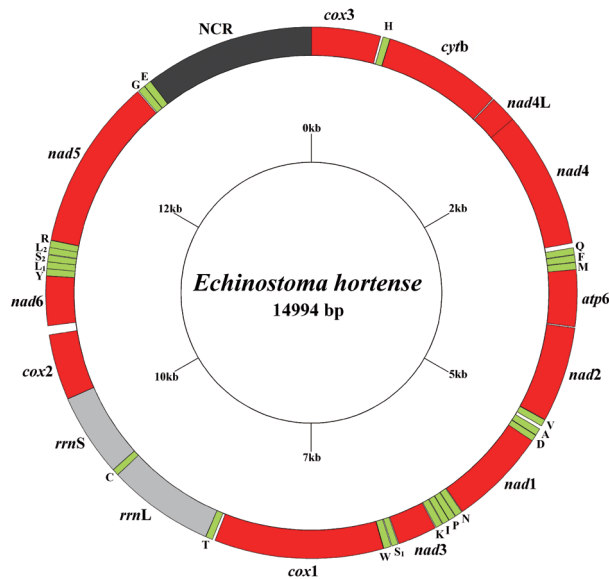


Fig. 1. Arrangement of the mitochondrial genome for *Echinostoma hortense*. Gene scaling is approximate only. All genes have standard nomenclature, including the 22 tRNA genes, which are designated by the one-letter code for the corresponding amino acid, with numerals differentiating each of the 2 leucine and serine specifying tRNAs (L1 and L2 for codon families CUN and UUR, respectively; S1 and S2 for codon families AGC and UCA, respectively).

mt genome, and the length of PCGs was in the order: *nad5* > *cox1* > *nad4* > *cytb* > *nad1* > *nad2* > *cox3* > *cox2* > *atp6* > *nad6* > *nad3* > *nad4L*. The concatenated nucleotide sequences of 12 PCGs were 10,047 bp in length and composed of 45.13% T, 18.54% A, 24.73% G, and 11.60% C, with A+T accounting for 63.67% of the total nucleotides encoding the 12 PCGs. The initiation and termination codons of the above amino acid sequences were identified by sequence comparison with homologs in the mt genomes of other trematodes. The ATG codon was used in 8 PCGs (*cox3*, *cytb*, *nad4*, *nad2*, *nad1*, *nad3*, *cox2*, and *nad5*), and the GTG codon in 4r genes (*nad4L*, *atp6*, *cox1*, and *nad5*) as start codons. The TAG was used as stop codon in 5 genes (*cytb*, *atp6*, *nad1*, *nad3*, and *cox2*) and the TAA termination codon in the remaining 7 genes (*cox3*, *nad4L*, *nad4*, *nad2*, *cox1*, *nad6*, and *nad5*).

Total 22 tRNA genes, which ranged from 59 to 69 bp in length, were predicted from the mt genomes. Among the 22 tRNA genes, tRNA-Ser^{AGC} and tRNA-Ser^{UCA} have unpaired D-arms which are replaced by the loops of 7-15 bp, but other 20 tRNA genes could be folded into the conventional cloverleaf structure (data not shown). The 2 ribosomal RNA genes were inferred, *rrnL* was located between tRNA-Thr and tRNA-Cys, and *rrnS* was located between tRNA-Cys and *cox2*. The *rrnL* and *rrnS* of *E. hortense* were 973 bp and 759 bp in length, respectively. The A+T contents of the *rrnL* and *rrnS* of *E. hortense* mt genome were 61.63% and 61.00%, respectively. Only 1 AT-rich non-coding region was inferred in the mt genome, and the AT-rich region located between *nad5* and *cox3*, which is 1,553 bp long with A+T of 64.58%.

Phylogenetic analysis

Phylogenetic analysis of concatenated nucleotide sequence datasets for all 12 PCGs were performed using MP method, and the result showed that Opisthorchiidae, Heterophyidae, Paragonimidae, Dicrocoeliidae, Fasciolidae, Echinostomatidae, Paramphistomidae, Notocotylidae, and Clinostomidae formed 1 group, whereas Schistosomatidae formed another group (Fig. 2). In former group, besides the bird fluke *Clinostomum complanatum*, other trematodes gathered together and formed 1 clade. In this clade, Fasciolidae, Echinostomatidae, Paramphistomidae, Notocotylidae, and Clinostomidae trematodes belonged to order Echinostomida all formed monophyly. *E. hortense* and *H. conoideum* gathered together, and they were closer to each other than to the Fasciolidae and order Echinostomida trematodes with high nodal support (Fig. 2).

Table 2. The organization of mitochondrial genome of *Echinostoma hortense*

Gene/Region	Positions		No. of		Codons	
	Start (5')	End (3')	Nucleotides	Aminoacids	Initiation	Termination
<i>cox3</i>	1	630	630	209	ATG	TAA
<i>tRNA-His</i>	652	716	65			
<i>cytb</i>	718	1,791	1,074	357	ATG	TAG
<i>nad4L</i>	1,802	2,089	288	95	GTG	TAA
<i>nad4</i>	2,050	3,297	1,248	415	ATG	TAA
<i>tRNA-Gln</i>	3,342	3,403	62			
<i>tRNA-Phe</i>	3,408	3,471	64			
<i>tRNA-Met</i>	3,475	3,539	65			
<i>atp6</i>	3,540	4,055	516	171	GTG	TAG
<i>nad2</i>	4,066	4,935	870	289	ATG	TAA
<i>tRNA-Val</i>	4,939	5,001	63			
<i>tRNA-Ala</i>	5,032	5,096	65			
<i>tRNA-Asp</i>	5,100	5,162	63			
<i>nad1</i>	5,158	6,057	900	299	ATG	TAG
<i>tRNA-Asn</i>	6,063	6,129	67			
<i>tRNA-Pro</i>	6,134	6,196	63			
<i>tRNA-Ile</i>	6,200	6,261	62			
<i>tRNA-Lys</i>	6,267	6,332	66			
<i>nad3</i>	6,340	6,696	357	118	ATG	TAG
<i>tRNA-Ser^{AGC}</i>	6,704	6,762	59			
<i>tRNA-Trp</i>	6,771	6,838	68			
<i>cox1</i>	6,842	8,383	1,542	513	GTG	TAA
<i>tRNA-Thr</i>	8,409	8,473	65			
<i>rrL</i>	8,474	9,446	973			
<i>tRNA-Cys</i>	9,447	9,506	60			
<i>rrS</i>	9,507	10,265	759			
<i>cox2</i>	10,266	10,865	600	199	ATG	TAG
<i>nad6</i>	10,948	11,397	450	149	ATG	TAA
<i>tRNA-Tyr</i>	11,399	11,462	64			
<i>tRNA-Leu^{GUA}</i>	11,463	11,525	63			
<i>tRNA-Ser^{UCA}</i>	11,526	11,587	62			
<i>tRNA-Leu^{UUA}</i>	11,593	11,656	64			
<i>tRNA-Arg</i>	11,655	11,719	65			
<i>nad5</i>	11,722	13,293	1,572	523	GTG	TAA
<i>tRNA-Gly</i>	13,304	13,370	67			
<i>tRNA-Glu</i>	13,373	13,441	69			
NC (64.58%)	13,442	14,994	1,553			

DISCUSSION

The complete mt genome of *E. hortense* (14,994 bp) is smaller than those of *Eurytrema pancreaticum* (15,031 bp), *H. taichui* (15,130 bp), *M. yokogawai* (15,258 bp), *S. haematobium* (15,003 bp), and *S. spindale* (16,901 bp), but slightly larger than those of other digenean species available in GenBank to date. The mt genome of *E. hortense* lacks *atp8* gene, which is similar to the mt genomes of other trematodes [12,13,16,18]. All genes were transcribed in the same direction, which is consistent with the mt genomes of other digeneans [12,13,16,18,24].

The gene arrangement of 36 genes in *E. hortense* was the same as that of the *H. conoideum* (KM111525), *P. cervi* (KF475773), *D. dendriticum* (NC_025280), and *D. chinensis* (NC_025279), but different from that of other digenean species. Notably, there was a 28-64 bp long overlap between *nad4L* and *nad4* in fluke mt genomes, but mostly was 28 bp or 40 bp in length. In *E. hortense*, *nad4L* and *nad4* overlapped by 40 bp in size which is consistent with that of *O. felinus*, *C. sinensis*, *F. hepatica*, *P. westermani*, *P. cervi*, and *H. taichui* [13,16,18,24,25]. Though *E. hortense* and *H. conoideum* belong to the same family of Echinostomatidae, the lengths of their *cox3* genes were very differ-

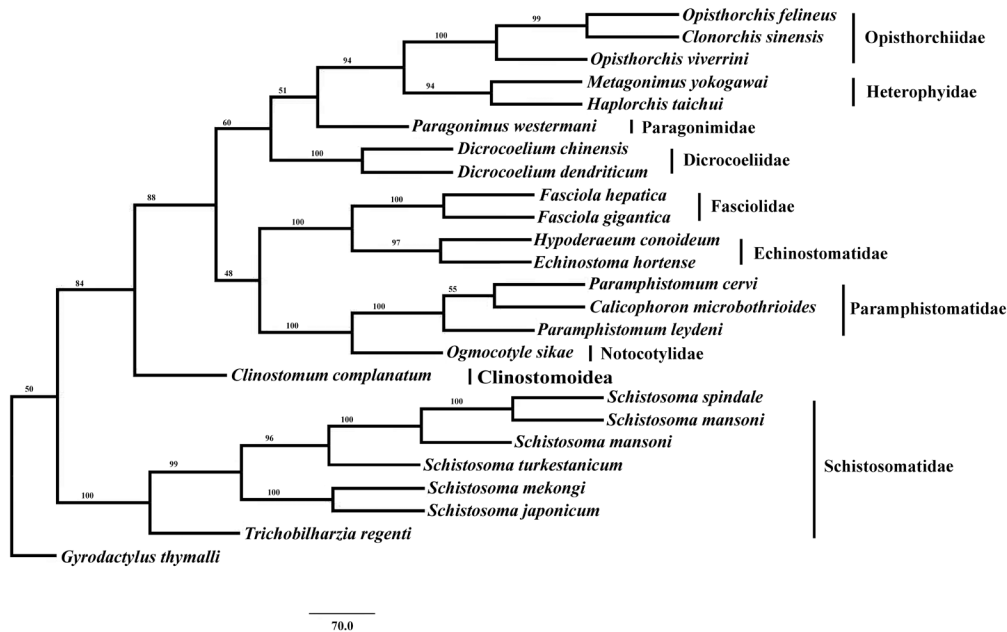


Fig. 2. Phylogenetic relationship of *Echinostoma hortense* with other digenean trematodes based on mitochondrial genome sequence data. The concatenated amino acid sequences of 12 protein-coding genes were analyzed with maximum parsimony (MP) method, using Monogenean species *Gyrodactylus thymalli* (NC_009682) as the outgroup.

ent, with that of *E. hortense* shorter than *H. conoideum* by 312 bp.

The mt genome of *E. hortense* had 22 tRNA genes, like the majority of trematode mt genome sequences, except for that of *P. westermani* Korean isolate (NC_002354) and Indian isolate which have 23 and 24 tRNAs [24], respectively. The *rmlL* of *E. hortense*, at 973 bp, was the shortest among trematodes recorded yet, and the *rmlS* (759 bp) within the range of typical sizes for digenean mt genomes (700-800 bp). Usually, the size of non-coding region of trematode mt genome is between 55 bp and 2,500 bp, some of the trematode mt genomes have 2 non-coding regions (*F. hepatica*, *F. gigantica*, *P. cervi*, *D. dendriticum*, and *D. chinensis*), and a few flukes only contain 1 region (*T. regenti*, *S. mekongi*, *H. conoideum*, and *H. taichui*). Similarly, the mt genome of *E. hortense* only contains 1 non-coding region. In most trematode mt genomes, between genes or between tRNAs, there are many intergenic sequences with their lengths ranging usually from 1 bp to 50 bp, and the longest intergenic region in *E. hortense* mt genome reached 82 bp, but it was not AT-rich (45.12%) region.

There were many studies about phylogenetic status in the family Echinostomatidae have been done by some researchers. For example, Kudlai et al. [26] used 28S rDNA to study the phylogeny of 25 species of the superfamily Echinostomatoidea

by Bayesian analysis. The results showed that 6 clades were clustered, and family Echinostomatidae was the largest group. *Echinostoma* and *Echinoparyphium* clustered together in a weakly-supported clade while the members of the other genera formed a 100% supported grouping. *Echinostoma* had a closer relationship with *Echinoparyphium* than to other genera of the family Echinostomatidae. In order to assess the relationships of *Echinostoma* with other closely related echinostomatids, 37 *nad1* sequences and 15 ITS sequences of the genera *Echinostoma*, *Echinoparyphium*, *Isthmiophora*, and *Hypoderaeum* were analyzed by Kostadinova et al. [27]. The results showed that the genus *Echinostoma* is not a monophyletic group, and that *E. hortense* is most distantly related with other *Echinostoma* trematodes. This result was reconfirmed by Saijuntha et al. [28], who studied the phylogenetic relationships of *Echinostoma* and *Hypoderaeum* based on the *cox1* sequences, and the results revealed that *E. revolutum* and *E. malayanum* of genus *Echinostoma* did not form a monophyletic clade and/or sister taxa. In contrast, a study of the phylogenetic relationships of North American echinostomes indicated that the genus *Echinostoma* is monophyletic, and the genera *Hypoderaeum* and *Echinoparyphium* are closely related as sister genera [29]. These contradictory results may be related to differences in geographical localities, sampled numbers, and the snail hosts of the echino-

stomes.

Therefore, Saijuntha et al. [28] suggested that the traditional morphological taxonomy of echinostomes, which is based on the size, shape, and arrangement of collar spines, the development of circumoral disc and also the testicular shape, should be reconsidered. The phylogenetic analysis in the present study indicated that the 2 fluke species, *E. hortense* and *H. conoideum*, of the family Echinostomatidae gathered together, which were closely related than to trematodes of the family Fasciolidae and other Echinostomida trematodes with high nodal support. This result was similar to the findings of previous reports. Although *E. hortense* and *H. conoideum* mt genomes were sequenced, more sequences of echinostomatid trematodes mt genomes are still needed to assess the phylogenetic relationships of these trematodes.

In conclusion, the present study determined the complete mt genome sequences of *E. hortense*, the first mt genome sequence of the genus *Echinostoma* of Echinostomatidae. The mtDNA dataset provided robust genetic evidence that *E. hortense* and *H. conoideum* were closer to each other than to the Fasciolidae and other Echinostomida trematodes, as well as provided new and useful genetic marker for further studies of the taxonomy, identification, and systematics of echinostomatid flukes.

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CONFLICT OF INTEREST

We have no conflict of interest related to this study.

REFERENCES

1. Yamaguti S. Synopsis of digenetic trematodes of vertebrates. Tokyo, Japan. Keigaku Publishing Co. 1971, pp 277-323.
2. Chai JY, Shin EH, Lee SH, Rim HJ. Foodborne intestinal flukes in Southeast Asia. *Korean J Parasitol* 2009; 47 (suppl): S69-S102.
3. Lee SK, Chung NS, Ko IH, Ko HI, Chai JY. Two cases of natural human infection by *Echinostoma hortense*. *Korean J Parasitol* 1986; 24: 77-81.
4. Asada J. On a new echinostomatid trematode and its life history. *Trans Jpn Soc Pathol* 1926; 16: 293-294.
5. Chai JY, Park JH, Jung BK, Guk SM, Kim JL, Shin EH, Klein TA, Kim HC, Chong ST, Baek LJ, Song JW. Echinostome infections in the striped-field mouse, *Apodemus agrarius*, and the Ussuri white-toothed shrew, *Crociodura lasiura*, caught near the demilitarized zone, Gyeonggi-do (Province), Republic of Korea. *Korean J Parasitol* 2009; 47: 311-314.
6. Jiang XL, Guan GJ, Yan JB. The significance of the *Echinostoma hortense* found in the pig in Sichuan. *Agr Sci Rep* 1985; 2: 82-84 (in Chinese).
7. Cho SY, Kang SY, Ryang YS. Helminthes infections in the small intestine of stray dogs in Eunjung city, Kyunggi do, Korea. *Korean J Parasitol* 1981; 19: 55-59.
8. Chai JY, Bahk YY, Sohn WM. Trematodes recovered in the small intestine of stray cats in the Republic of Korea. *Korean J Parasitol* 2013; 51: 99-106.
9. Fan SQ, Sun MF. A case of *Echinostoma hortense* first found in human body in China and morphological observation of the worm. *J Harbin Med Univ* 1989; 23: 161-163 (in Chinese).
10. Arizono N, Uemoto K, Kondo K, Matsuno K, Yoshida Y, Maeda T, Yoshida H, Muto K, Inoue Z, Takahashi K. Studies on *Echinostoma hortense* Asada, 1926 with special reference to its human infection. *Jpn J Parasitol* 1976; 1: 36-45 (in Japanese).
11. Son WY, Huh S, Lee SU, Woo HC, Hong SJ. Intestinal trematode infections in the villagers in Koje-myon, Kochang-gun, Kyongsangnam-do, Korea. *Korean J Parasitol* 1994; 32: 149-155.
12. Wang Y, Wang CR, Zhao GH, Gao JE, Li MW, Zhu XQ. The complete mitochondrial genome of *Orientobilharzia turkestanicum* supports its affinity with African *Schistosoma* spp. *Infect Genet Evol* 2011; 11: 1964-1970.
13. Lee D, Choe S, Park H, Jeon HK, Chai JY, Sohn WM, Yong TS, Min DY, Rim HJ, Eom KS. Complete mitochondrial genome of *Haplorchis taichui* and comparative analysis with other trematodes. *Korean J Parasitol* 2013; 51: 719-726.
14. Cribb TH, Bray RA, Littlewood DTJ, Pichelin SP, Herniou EA. The Digenea. In Littlewood DTJ, Bray RA, eds, *Interrelationships of the Platyhelminthes*. London, UK. Taylor and Francis. 2001, pp 168-185.
15. Cho CM, Tak WY, Kweon YO, Kim SK, Choi YH, Kong HH, Chung DI. A human case of *Echinostoma hortense* (Trematoda: Echinostomatidae) infection diagnosed by gastroduodenal endoscopy in Korea. *Korean J Parasitol* 2003; 41: 117-120.
16. Le TH, Blair D, McManus DP. Complete DNA sequence and gene organization of the mitochondrial genome of the liverfluke, *Fasciola hepatica* L. (Platyhelminthes: Trematoda). *Parasitology* 2001; 123: 609-621.
17. Burland TG. DNASTAR's Lasergene sequence analysis software. *Methods Mol Biol* 2000; 132: 71-91.
18. Shekhovtsov SV, Katokhin AV, Kolchanov NA, Mordvinov VA. The complete mitochondrial genomes of the liver flukes *Opisthorchis felineus* and *Clonorchis sinensis* (Trematoda). *Parasitol Int*

- 2010; 59: 100-103.
19. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol* 2011; 28: 2731-2739.
 20. Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 2012; 28: 1647-1649.
 21. Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG. The CLUSTAL_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res* 1997; 25: 4876-4882.
 22. Swofford DL. PAUP*. Phylogenetic analysis using parsimony (and other methods). Version 4.0b10. Sunderland MA, Sinauer Associates. 2003.
 23. Page RD. TreeView: an application to display phylogenetic trees on personal computers. *Comput Appl Biosci* 1996; 12: 357-358.
 24. Biswal DK, Chatterjee A, Bhattacharya A, Tandon V. The mitochondrial genome of *Paragonimus westermani* (Kerbert, 1878), the Indian isolate of the lung fluke representative of the family Paragonimidae (Trematoda). *Peer J* 2014; 2: e484.
 25. Yan HB, Wang XY, Lou ZZ, Li L, Blair D, Yin H, Cai JZ, Dai XL, Lei MT, Zhu XQ, Cai XP, Jia WZ. The mitochondrial genome of *Paramphistomum cervi* (Digenea), the first representative for the family Paramphistomidae. *PLoS One* 2013; 8: e71300.
 26. Kudlai O, Tkach VV, Pulis EE, Kostadinova A. Redescription and phylogenetic relationships of *Euparyphium capitaneum* Dietz, 1909, the type-species of *Euparyphium* Dietz, 1909 (Digenea: Echinostomatidae). *Syst Parasitol* 2015; 90: 53-65.
 27. Kostadinova A, Herniou EA, Barrett J, Littlewood DT. Phylogenetic relationships of *Echinostoma Rudolphi*, 1809 (Digenea: Echinostomatidae) and related genera re-assessed via DNA and morphological analyses. *Syst Parasitol* 2003; 54: 159-176.
 28. Saijuntha W, Sithithaworn P, Duenngai K, Kiatsopit N, Andrews RH, Petney TN. Genetic variation and relationships of four species of medically important echinostomes (Trematoda: Echinostomatidae) in South-East Asia. *Infect Genet Evol* 2011; 11: 375-381.
 29. Detwiler JT, Bos DH, Minchella DJ. Revealing the secret lives of cryptic species: examining the phylogenetic relationships of echinostome parasites in North America. *Mol Phylogenet Evol* 2010; 55: 611-620.