

Role of artesunate in TGF- β 1-induced renal tubular epithelial-mesenchymal transdifferentiation in NRK-52E cells

YAQIAN ZHANG^{1,2*}, HUANHUAN LI^{1,2*}, JIAJUN ZHU^{3*}, TIAN TIAN WEI^{1,2*},
YINGXIAN PENG^{1,2}, RAN LI^{1,2}, RUI XU⁴, MEI LI¹ and ANZHOU XIA¹

¹Jiangsu Key Laboratory of New Drug Research and Clinical Pharmacy; ²Department of Postgraduate Academy, Xuzhou Medical University, Xuzhou, Jiangsu 221004; ³Department of Anesthesiology, Guanyun County People's Hospital, Lianyungang, Jiangsu 222200; ⁴Department of Pharmacy, Suining County People's Hospital, Xuzhou, Jiangsu 221200, P.R. China

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Abstract. The implications of epithelial-mesenchymal transdifferentiation (EMT) have extended beyond the confines of renal fibrosis to renal tubulointerstitial fibrosis. It has been proposed that EMT may be one of the mechanisms involved in the pathogenesis of renal fibrosis. However, the underlying mechanisms remain unknown. Transforming growth factor (TGF)- β 1 is considered to be an important cytokine which regulates the transdifferentiation of tubular epithelial cells into myofibroblasts in renal tubulointerstitial fibrosis. In the present study, normal rat kidney tubular epithelial cells (NRK-52E) were treated for 48 h with TGF- β 1 (5 ng/ml) and different concentrations of artesunate (ART; 0.01, 0.1 and 1 μ g/ml). Western blotting, reverse transcription-semi quantitative polymerase chain reaction analysis and immunofluorescence staining were used to evaluate the expression of bone morphogenetic protein (BMP)-7, uterine sensitization-associated gene (USAG)-1, E-cadherin, α -smooth muscle actin (α -SMA) and extracellular matrix collagen type I (Col I) mRNA. ART was able to attenuate renal injury in a unilateral ureteral obstruction model. However, its anti-fibrotic effect remains to be elucidated. In the present study, it was observed that ART was able to ameliorate the TGF- β 1-induced alterations in cellular morphology. In addition, ART inhibited the TGF- β 1-induced USAG-1 increase and the decrease in BMP-7. Treatment with ART markedly attenuated the TGF- β 1-induced

upregulation of α -SMA and downregulation of E-cadherin. Additionally, ART was able to significantly attenuate the deposition of interstitial collagens, including Col I. The results of the present study further verified the therapeutic efficacy of ART in TGF- β 1-induced renal interstitial fibrosis. These findings indicated that ART may hold the potential to prevent chronic kidney diseases via the suppression of USAG-1 expression or by increasing BMP-7 expression.

Introduction

Renal tubulointerstitial fibrosis is a prominent pathological characteristic of chronic kidney disease, and is characterized by extensive interstitial myofibroblast activation and excessive matrix protein accumulation, which are the common attributes of numerous types of chronic kidney disease leading to end-stage renal disease (1,2). Previous studies have demonstrated that transforming growth factor (TGF)- β 1 serves an important role in mediating chronic inflammation, myofibroblast activation and extracellular matrix (ECM) accumulation (3-5). At present, there are no effective treatments for renal interstitial fibrosis or ways to impede the progress of the associated chronic kidney disease. Therefore, it is of importance to investigate potential strategies to prevent the development of renal interstitial fibrosis in the diseased kidneys.

Although studies have focused on renal interstitial fibrosis, the mechanism underlying its prevalence remains to be elucidated. Among the putative mechanisms, epithelial-mesenchymal transition (EMT) has become a popular hypothesis (6). The primary events of EMT include the adoption of a mesenchymal-like cellular phenotype and migration through the tubular basement membrane towards the tubular interstitium. It has been established that a number of factors are involved in the process of EMT, which include the profibrotic factor TGF- β 1 and the antifibrotic cytokine bone morphogenetic protein-7 (BMP-7). These are the important molecules that determine the fate of the kidney as they hold the potential to initiate and complete the process of EMT (7,8).

It was previously demonstrated that the product of uterine sensitization-associated gene 1 (USAG-1) acts as a

Correspondence to: Professor Anzhou Xia, Jiangsu Key Laboratory of New Drug Research and Clinical Pharmacy, Xuzhou Medical University, 209 Tongshan Road, Xuzhou, Jiangsu 221004, P.R. China
E-mail: 534163783@qq.com

*Contributed equally

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kidney-specific BMP antagonist, which binds to and inhibits the biological activity of BMP-7 (9-14). A previous study revealed USAG-1-deficient mice to be resistant to *in vivo* kidney injury, while genetic ablation of USAG-1 in the chronic renal injury model of unilateral uretral obstruction (UO) resisted kidney injury and therefore prolonged survival (15). However, following administration of USAG-1^{-/-} mouse anti-BMP-7 neutralizing antibody, the USAG-1^{-/-} renal protective effect in mice was inhibited. Since USAG-1 may serve a notable role in the regulation of BMP-7 renal protection, inhibiting USAG-1 expression may represent a promising therapeutic measure for the treatment or management of renal interstitial fibrosis.

Artemisinin, extracted and isolated from *Artemisia annua compositae* leaves, is a sesquiterpene compound with a peroxide bridge. In addition to exhibiting anti-malarial effects, artemisinin has anti-inflammatory, anti-tumor and anti-fibrotic effects (16,17). Artesunate (ART) is an artemisinin derivative, and has been demonstrated to exhibit a therapeutic effect in the treatment of pulmonary fibrosis, liver fibrosis and myocardial fibrosis (18,19). Additionally, a previous study confirmed that ART was able to effectively alleviate renal fibrosis caused by UO (20); however, whether ART inhibits the occurrence of EMT by inhibiting the activity of USAG-1 or activating the activity of BMP-7 remains unclear.

In the present study, the effect of ART on TGF- β 1-induced EMT was investigated with the aim of examining the potential underlying mechanism. The mechanism may be used to elucidate the association between the effect of ART and the abnormal expression of BMP-7 and USAG-1.

Materials and methods

Reagents. Dulbecco's modified Eagle's medium (DMEM)/high glucose was purchased from Gibco® (Thermo Fisher Scientific, Inc., Waltham, MA, USA), Ausbian fetal bovine serum (FBS) was purchased from Thermo Fisher Scientific, Inc., and penicillin-streptomycin solution was purchased from Beyotime Institute of Biotechnology (Haimen, China). Trypsin/EDTA solution was purchased from Beijing Solarbio Science and Technology Co., Ltd. (Beijing, China) and the semi-quantitative polymerase chain reaction (sq-PCR) kit was obtained from Tiangen Biotech Co., Ltd. (Beijing, China). For western blotting, BMP-7 (cat. no. ab56023), USAG-1 (cat. no. ab99340), E-cadherin (cat. no. 5409-1) and α -smooth muscle actin (α -SMA) (cat. no. 1184-1) antibodies were all purchased from Abcam (Cambridge, UK), GAPDH antibody was obtained from Zhongshan Jinqiao Biotechnology Co., Ltd. (Beijing, China), and for immunofluorescence staining, USAG-1 (cat. no. sc-162253) antibody was obtained from Santa Cruz Biotechnology, Inc. (Dallas, TX, USA), the secondary antibodies were from Zhongshan Jinqiao Biotechnology Co., Ltd. (Beijing, China).

Cell culture and treatment. Normal rat kidney tubular epithelial cells (NRK-52E) were purchased from the Chinese Academy of Sciences Cell Bank (Shanghai, China) and maintained in DMEM/high-glucose medium supplemented with 1% penicillin-streptomycin and 10% FBS, in a humidified incubator containing 95% air and 5% CO₂ at 37°C atmosphere. For EMT experiments, cells were treated with 5 ng/ml TGF- β 1

(Peprotech, Inc., Rocky Hill, NJ, USA) for 48 h (21). ART (>98% purity; molecular weight, 384.43) was purchased from TCI (Shanghai) Development Co., Ltd. (Shanghai, China). In order to determine the effect of ART on the activity and proliferation of NRK-52E cells induced by TGF- β 1, cells were seeded at 60-70% confluence in complete medium containing 10% FBS for 24 h, and subsequently serum-starved for 12 h. The cells were subsequently induced with TGF- β 1 (5 ng/ml), treated with different ART concentrations (0, 0.01, 0.1 and 1 μ g/ml) containing 1% FBS and incubated for 48 h. The groups for the experiment were as follows: Control group, TGF- β 1 group (normal NRK-52E cells treated with 5 ng/ml TGF- β 1), ART-L group (0.01 μ g/ml ART), ART-M group (0.1 μ g/ml ART), and ART-H group (1 μ g/ml ART). Cells were harvested, following which the expression of associated proteins and genes was determined.

MTT assay for cell viability. NRK-52E cells were seeded into 96-well culture plates and treated with various ART concentrations (0, 0.01, 0.1, 1, 5 and 10 μ g/ml) for 48 h. Cell viability was determined via an MTT assay and cells were incubated with 20 μ l MTT solution (0.5 mg/ml) for 4 h at 37°C. The purple formazan crystals derived from the MTT were dissolved in 150 μ l dimethyl sulfoxide and agitated for 10 min. The absorbance at 490 nm was measured with a microplate reader (Elx808; BioTek Instruments, Inc., Winooski, VT, USA).

Morphology observation. After treatment with TGF- β 1 combined with or without ART for 48 h, cells were washed with PBS twice. The morphological changes of NRK-52E cells were observed under a phase-contrast photomicroscope (Leica Microsystems GmbH, Wetzlar, Germany) and photographed using a digital camera.

Reverse transcription (RT)-sqPCR analysis. Total RNA was extracted using TRIzol reagent (Tiangen Biotech Co., Ltd.), according to manufacturer's instructions, and the cDNA was synthesized using a TIANScript RT kit (Tiangen Biotech Co., Ltd.) (22). Primers were synthesized by Sangon Biotech Co., Ltd. (Shanghai, China). sqPCR was performed as previously described (22). The sequences of the primers used were as follows: α -SMA forward, 5'-GCATCCACGAAACCACCT-3' and reverse, 5'-CGCCGATCCAGACAG AAT-3' (210 bp); E-cadherin forward, 5'-TTCAACCAAGC ACGTACCA-3' and reverse, 5'-CAGAATGCCCTCGTT GGTCT-3' (186 bp); BMP-7 forward, 5'-GCACCTCCAGG AAAAC-3' and reverse, 5'-AAGCCCAGATGGTACGG-3' (443 bp); USAG-1 forward, 5'-AGATTTGATTGCGTGGAA GAC-3' and reverse, 5'-GGTTCGGAGGGATTAGTTTG-3' (311 bp); collagen type I (Col I) forward, 5'-TCAGGGGCG AAGGCAACAGT-3' and reverse, TTGGGATGGAGGGAG TTTACAGA-3' (219 bp); and GAPDH forward, 5'-TGC TGAGTATGTCGTGGAGT-3' and reverse, 5'-AGTCTT CTGAGTGGCAGTGAT-3' (289 bp). PCR amplification was performed with initial denaturation at 94°C for 5 min, followed by 30 consecutive cycles of denaturation at 94°C for 30 sec, annealing at 58-62°C for 30 sec and extension at 72°C for 1 min, with a final extension at 72°C for 7 min. The amplified products were analyzed by electrophoresis on a 1.5% (w/v) agarose gel and stained with fluorescence staining dye

Goldview (Beijing Solarbio Science and Technology Co., Ltd.) alongside a DNA marker (Tiangen Biotech Co., Ltd.) The relative mRNA levels of various genes were calculated following normalization to GAPDH. The signal intensity of the images was analyzed using ImageJ version 1.48 software (National Institutes of Health, Bethesda, MD, USA).

Western blot analysis. Cells were lysed with radioimmunoprecipitation assay buffer (Beyotime Institute of Biotechnology), supplemented with phenylmethylsulfonyl fluoride. The protein samples were quantified using bicinchoninic acid protein assay reagents (Beyotime Institute of Biotechnology). Western blot analysis was performed as previously described (23). Following boiling for 10 min in a 5X loading buffer (Beyotime Institute of Biotechnology), protein samples (45 μg) were separated by SDS-PAGE on an 8% gel and transferred onto nitrocellulose membranes (EMD Millipore, Billerica, MA, USA). The membranes were subsequently blocked in PBS containing 3% BSA (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) and incubated with primary antibodies at a 1:1,000 dilution overnight at 4°C. The primary antibodies used were as follows: Anti- α -SMA, anti-E-cadherin, anti-BMP-7 and anti-USAG-1. Following washing with PBS-Tween-20, the membrane was incubated with alkaline phosphatase-conjugated goat anti-rabbit and horse anti-mouse secondary antibodies at a 1:1,000 dilution at room temperature for 2 h. Signal detection was performed using an enhanced chemiluminescence kit (Shanghai Tianneng Technology Co., Ltd., Shanghai, China). The band density was measured using ImageJ software.

Immunofluorescent staining. The expressions of E-cadherin, α -SMA, BMP-7 and USAG-1 in NRK-52E cells were analyzed by immunofluorescence staining. Cells (2×10^4 cells/ml) were cultured on glass coverslips and treated with TGF- β 1 with or without 1 $\mu\text{g}/\text{ml}$ ART for 48 h. Following treatment for 48 h, cells in the basement layer were washed three times with PBS and fixed in 4% paraformaldehyde for 15 min at room temperature. Following three extensive washes with PBS, the cells were treated with 0.5% Triton X-100 for 10 min and blocked with 2% normal donkey serum (Sigma-Aldrich; Merck KGaA) for 1 h at room temperature, and subsequently incubated with primary antibodies (1:200) at 4°C overnight. Cells were washed with PBS for 3-5 min and incubated with goat anti-rabbit and rabbit anti-goat secondary antibodies (Zhongshan Jinqiao Biotechnology Co., Ltd., Beijing, China) at a dilution of 1:200 for 2 h. Following washing in PBS, cells were stained with DAPI (Beyotime Institute of Biotechnology) at room temperature to visualize the nuclei. Following washing in PBS, the device was mounted with anti-fluorescence quenching agent (Beyotime Institute of Biotechnology) and cover-slipped; digital images were captured using an inverted fluorescent microscope (magnification, $\times 400$).

Statistical analysis. All data were reported as the mean \pm standard deviation. Statistical analysis was performed using the statistical package SPSS for Windows, version 13.0 (SPSS, Inc., Chicago, IL, USA) and GraphPad Prism v. 5.0 (GraphPad Software, Inc., La Jolla, CA, USA). The comparisons among different groups were made using one-way analysis of variance followed by least significant

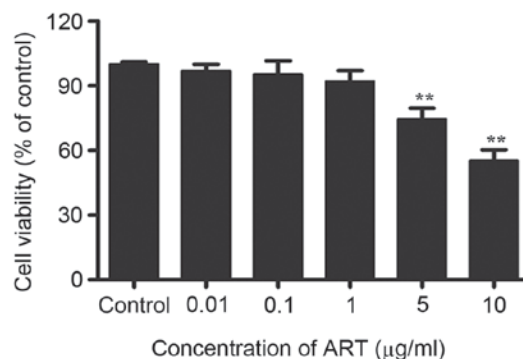


Figure 1. Effect of ART on the cell viability of NRK-52E. NRK-52E cells were incubated with different concentrations of ART for 48 h. All data are presented as the mean \pm standard deviation. $n=3$. ** $P<0.01$ vs. control. ART, artesunate.

difference/Dunnett-T3 tests. $P<0.05$ was considered to indicate a statistically significant difference. All experiments were repeated ≥ 3 times.

Results

Effect of TGF- β 1 on NRK-52E cells. In order to investigate the effect of TGF- β 1 on NRK-52E cells, cells were treated with different concentrations of TGF- β 1 (2, 5 and 10 $\mu\text{g}/\text{ml}$) for 48 h. The results demonstrated that 5 and 10 $\mu\text{g}/\text{ml}$ TGF- β 1 was able to significantly increase the expression of α -SMA mRNA and protein. In addition, TGF- β 1 (5 and 10 $\mu\text{g}/\text{ml}$) was able to downregulate the expression of E-cadherin mRNA, while a concentration of 2 $\mu\text{g}/\text{ml}$ TGF- β 1 had no significant effect on α -SMA mRNA and protein and E-cadherin mRNA compared with the control group. Additionally, TGF- β 1 was able to decrease cell-cell contact and the cells adopted a more elongated morphology at a concentration of 5 ng/ml. However, TGF- β 1 did not influence cell morphology at a concentration of 2 ng/ml. Therefore, 5 ng/ml TGF- β 1 was selected as the concentration to be used in the following experiment (data not shown). Subsequently, NRK-52E cells were treated with 5 ng/ml TGF- β 1 for 24, 48 and 72 h, and the results demonstrated that the expression of α -SMA mRNA was significantly upregulated compared with the control group when treated for 48 h, while TGF- β 1 had no marked effect on α -SMA mRNA expression when cells were treated for 24 h. In addition, the cellular morphology gradually altered from oval to a long spindle shape over time, and this alteration was apparent at 48 h; however, cellular morphology was almost unaltered compared with the control group at 24 h (data not shown). According to the above results, 5 ng/ml TGF- β 1 for 48 h was selected as the treatment to be used for the remainder of the experiments.

Effect of ART on cellular viability. In order to determine the effect of ART on cell viability, NRK-52E cells (2.5×10^4 cells/ml) were seeded into 96-well culture plates and treated with different concentrations of ART (0.01, 0.1, 1, 5 and 10 $\mu\text{g}/\text{ml}$) for 48 h, following which cell viability was determined. The results demonstrated that there was little or no effect on cell viability low-doses of ART (0.01, 0.1 and

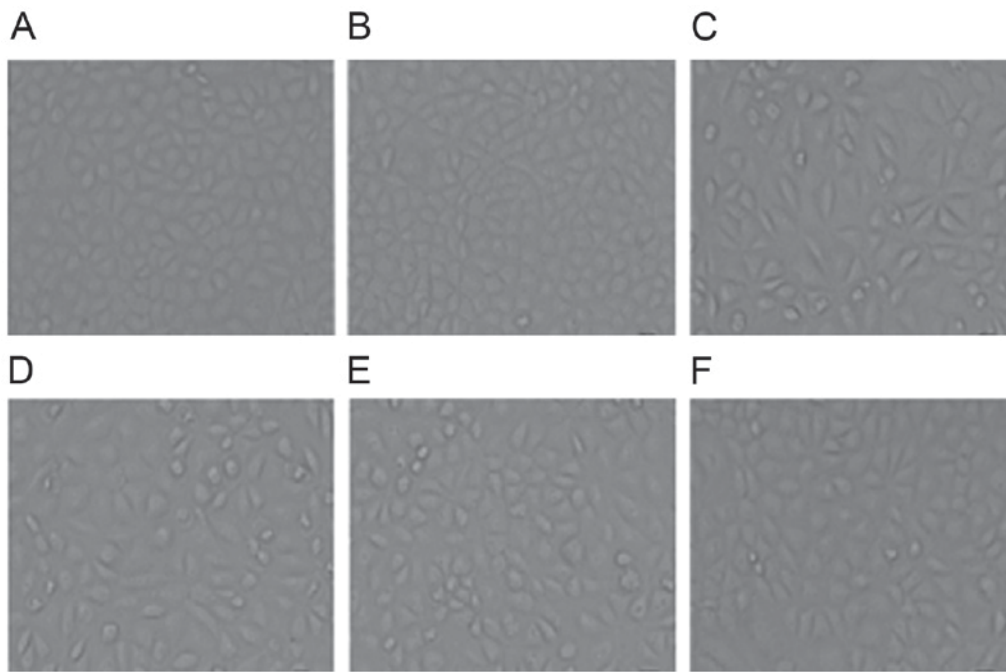


Figure 2. Effect of ART on the morphology of NRK-52E cells. Incubation of NRK-52E cells with different concentrations of ART with or without transforming growth factor- β 1 for 48 h. Representative images are presented: (A) Control group; (B) ART group; (C) TGF- β 1 group; (D) ART low dose group; (E) ART moderate dose group; (F) ART high dose group. Magnification, $\times 100$. ART, artesunate.

1 $\mu\text{g/ml}$). However, treatment with 5 or 10 $\mu\text{g/ml}$ ART resulted in a significant decrease in cell viability (Fig. 1). Therefore, 0.01, 0.1 and 1 $\mu\text{g/ml}$ were used for the following experiments.

Effect of ART on cell morphology. In order to observe the alterations in cellular morphology caused by treatment with ART, the morphological alterations in NRK-52E cells were observed under a phase-contrast photomicroscope (Leica Microsystems GmbH) and photographed using a digital camera. The control cells exhibited a typical epithelial cuboidal shape with cobblestone morphology (Fig. 2A). Treatment with ART alone (ART group) had no effect on morphology compared with the control group (Fig. 2B); therefore, this group was not included in the following experiment. By contrast, NRK-52E cells exposed to TGF- β 1 (5 ng/ml) exhibited a decrease in cell-cell contacts and adopted a more elongated morphological shape. Notably, treatment with TGF- β 1 resulted in a morphology which was fibroblast-like in nature and identifiable by the presence of elongated lamellipodia and a spindle shape, as presented in Fig. 2C. However, ART was able to improve the morphology of cells to different degrees and a dose of ART at 1 $\mu\text{g/ml}$ rendered the cellular morphology close to the normal cellular morphology (Fig. 2D-F). These results indicated that ART was able to improve TGF- β 1-induced morphological alterations in cells.

Effects of ART on the levels of E-cadherin, α -SMA and Col I in NRK-52E cells. In order to evaluate the regulatory effects of ART in TGF- β 1-induced EMT, the expression of E-cadherin and α -SMA was examined at the mRNA and protein levels in epithelial cells. The level of α -SMA expression was low, while E-cadherin expression was high in normal renal tubular epithelial cells. The results suggested that TGF- β 1-induced

cells exhibited a significant decrease in the gene expression of E-cadherin (Fig. 3A; $P < 0.01$), and ART-L had no significant effect on the expression of E-cadherin. The expression of E-cadherin was significantly increased in the ART-M and ART-H groups, particularly at the highest concentration of ART (Fig. 3A). The mRNA expression of α -SMA in TGF- β 1-induced EMT was significantly upregulated compared with the control group. Treatment with ART significantly attenuated the TGF- β 1-induced increase in the mRNA expression of α -SMA, and this was more apparent at moderate and high doses of ART (Fig. 3B). These results confirmed that EMT served a role in tubulointerstitial fibrosis, and that ART may improve renal fibrosis by inhibiting the process of EMT. The occurrence of EMT ultimately leads to deposition of the ECM, and Col I is the ECM deposit which results from EMT occurrence. To evaluate the regulatory effects of ART in TGF- β 1-induced ECM protein accumulation, the mRNA expression of Col I was examined. The results demonstrated that the expression of Col I was increased in cells undergoing EMT, which was pathologically associated with fibrosis, while treatment with ART in the low concentration group decreased the level of Col I and the difference was statistically significant with moderate and high concentrations of ART (Fig. 3C). The results demonstrated that the ECM was significantly accumulated in renal fibrosis, and that ART was able to decrease the accumulation of the ECM.

The expression of E-cadherin and α -SMA protein was examined in total cell lysates using western blotting. The results demonstrated that TGF- β 1 was able to increase the expression of α -SMA and decrease the E-cadherin expression in NRK-52E cells, demonstrating that TGF- β 1 stimulated EMT in tubular cells, which is consistent with previous reports (6). However, treatment with ART significantly

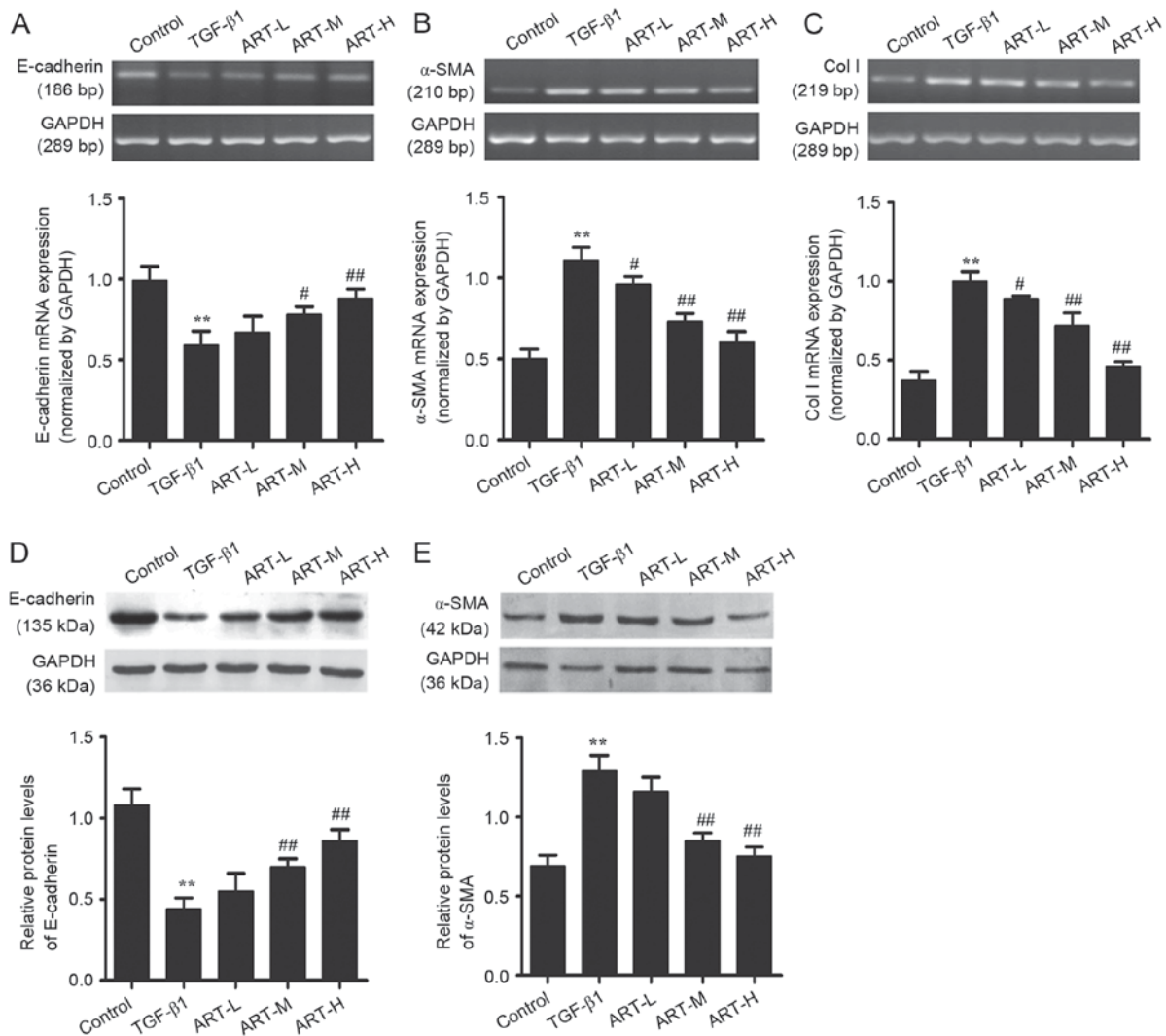


Figure 3. Effect of ART on the expression of E-cadherin, α -SMA and Col I at the mRNA and protein levels in NRK-52E cells. Incubation of NRK-52E cells with different concentrations of ART with or without TGF- β 1 for 48 h. Reverse transcription-semi-quantitative polymerase chain reaction analysis was used to detect the expression of (A) E-cadherin, (B) α -SMA and (C) Col I mRNA. Western blotting was applied to examine the protein levels of (D) E-cadherin and (E) α -SMA. GAPDH was used as the internal loading control. All data are presented as the mean \pm standard deviation. n=3. **P<0.01 vs. control; #P<0.05, ##P<0.01 vs. TGF- β 1. ART, artesunate; ART-L, low-dose ART; ART-M, moderate-dose ART; ART-H, high-dose ART; α -SMA, α -smooth muscle actin; Col I, collagen type I; TGF- β 1, transforming growth factor- β 1.

increased E-cadherin expression and downregulated α -SMA protein expression, which is in line with previous findings in the UUO model (20). In addition, the effect was more marked at moderate and high doses of ART (Fig. 3D and E).

The involvement of α -SMA and E-cadherin in TGF- β 1-induced EMT was further investigated via a fluorescence confocal assay. Under basal condition, the expression of E-cadherin was abundant in renal tubular epithelial cells, and was primarily localized to the plasma membrane (Fig. 4A). When the renal tubular cells were stimulated by TGF- β 1, the expression of E-cadherin was markedly decreased, as exhibited by decreased green fluorescence; when cells were pretreated with ART, the TGF- β 1-induced E-cadherin delocalization was inhibited (Fig. 4A). By contrast, the abundance of the mesenchymal marker α -SMA was low in renal tubular cells under control conditions, as indicated by the weak fluorescence in confocal images (Fig. 4B). When the cells were stimulated by TGF- β 1, the expression of α -SMA was markedly increased, as exhibited by the increased fluorescence

in the confocal images (Fig. 4B). When renal tubular cells were pretreated with ART, TGF- β 1 failed to increase α -SMA expression (Fig. 4B). These results further confirmed that ART may improve TGF- β 1-induced EMT progression.

Effect of ART on the expression of USAG-1 and BMP-7 at the mRNA and protein levels in NRK-52E cells. TGF- β 1 is considered to be an important cytokine for the induction of renal interstitial fibrosis, and BMP-7 is a member of the TGF- β 1 superfamily. A previous study reported that BMP-7 may be able to reverse the renal structural alterations and the degree of fibrosis in UUO rats by regulating the downstream molecules against decapentaplegic homolog (Smad)1/5/8 signaling pathway (24). USAG-1 serves as the endogenous antagonist of BMP-7. Therefore, the present study examined the expression of USAG-1 and BMP-7, and the underlying mechanism.

In order to determine the regulatory effects of ART in TGF- β 1-induced BMP-7 and USAG-1 expression, RT-sqPCR analysis and western blotting were performed (Fig. 5). TGF- β 1

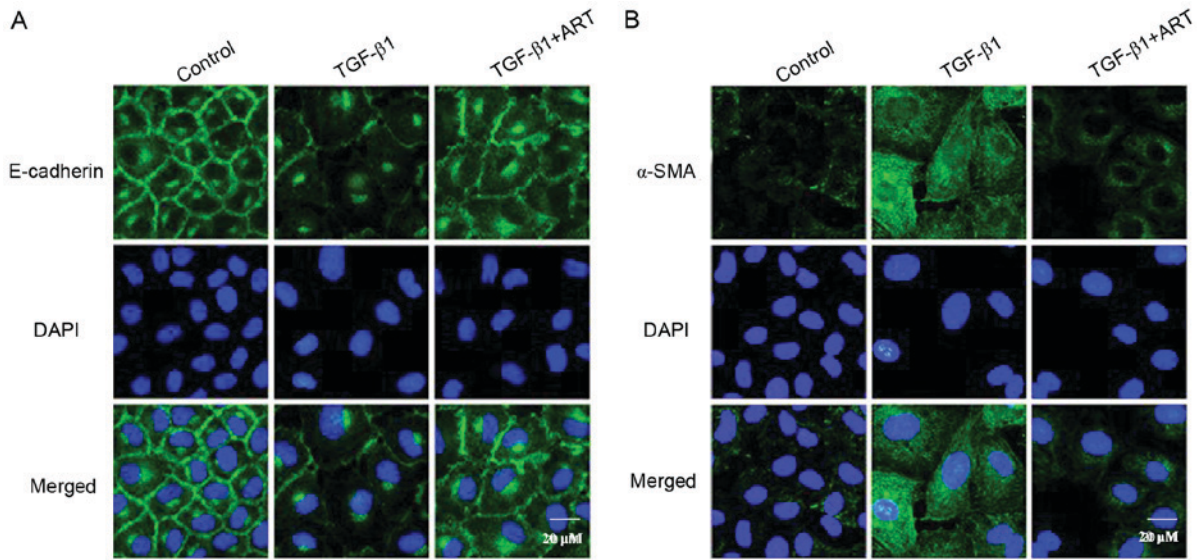


Figure 4. Immunofluorescence staining was performed to determine the expression of (A) E-cadherin and (B) α -SMA. Magnification, x400. α -SMA, α -smooth muscle actin; TGF- β 1, transforming growth factor- β 1; ART, artesunate.

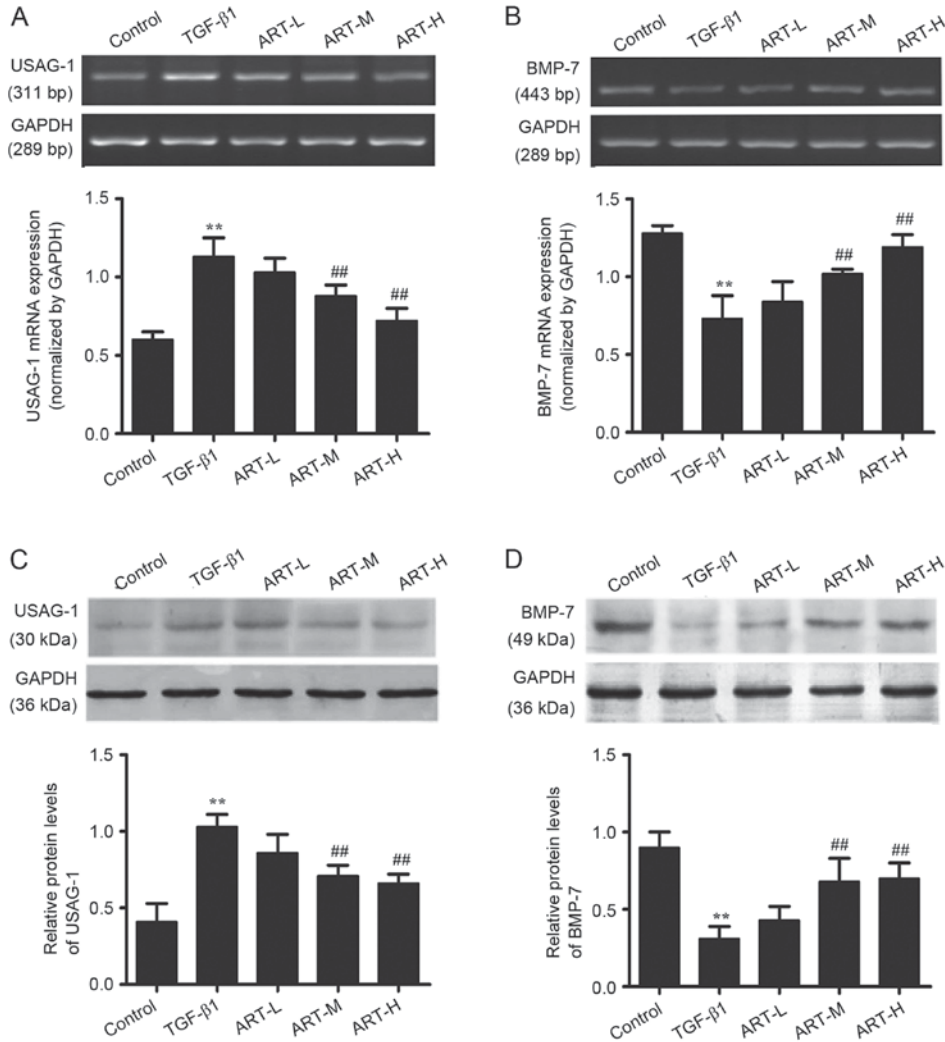


Figure 5. Effect of ART on the expression of USAG-1 and BMP-7 at the mRNA and protein levels in NRK-52E cells. NRK-52E cells were incubated with different concentrations of ART, with or without TGF- β 1, for 48 h. Reverse transcription-semi-quantitative polymerase chain reaction analysis was used to detect the expression of (A) USAG-1 and (B) BMP-7 mRNA. Western blotting was applied to examine the protein levels of (C) USAG-1 and (D) BMP-7. GAPDH was used as the internal loading control. All data are presented as the mean \pm standard deviation. n=3. **P<0.01 vs. control; ##P<0.01 vs. TGF- β 1. ART, artesunate; ART-L, low-dose ART; ART-M, moderate-dose ART; ART-H, high-dose ART; TGF- β 1, transforming growth factor- β 1; USAG-1, uterine sensitization-associated gene 1; BMP-7, bone morphogenetic protein 7.

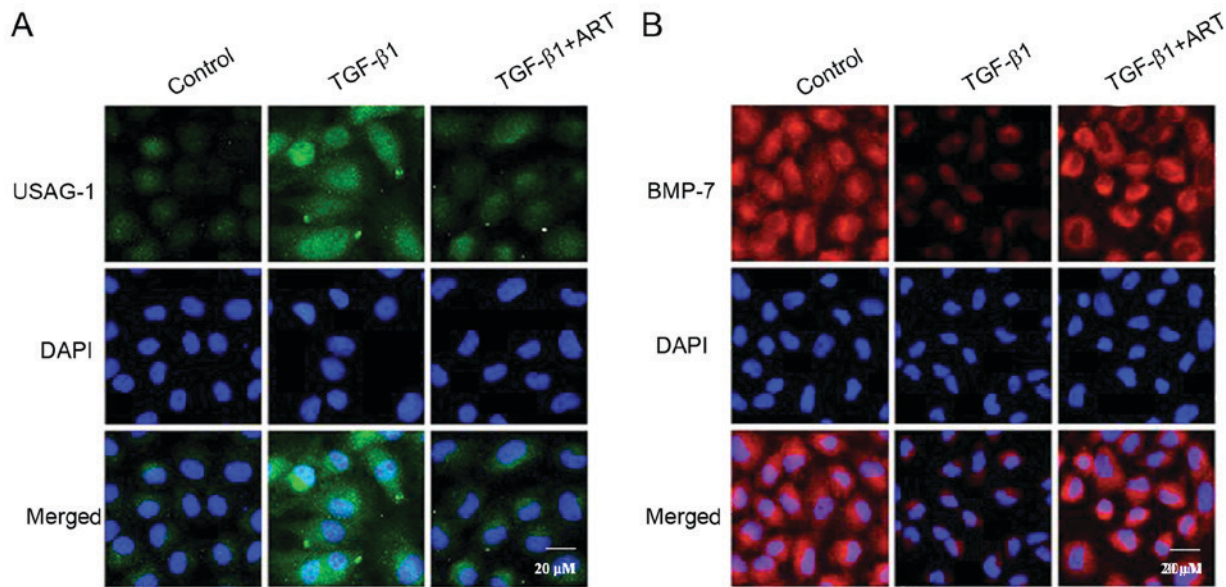


Figure 6. Immunofluorescence staining was performed to determine the expression of (A) USAG-1 and (B) BMP-7. Magnification, $\times 400$. TGF- $\beta 1$, transforming growth factor- $\beta 1$; ART, artesunate; USAG-1, uterine sensitization-associated gene 1; BMP-7, bone morphogenetic protein 7.

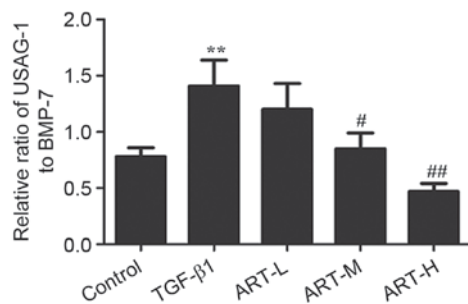


Figure 7. Effects of ART on the protein ratio of USAG-1 to BMP-7. Data are presented as the mean \pm standard deviation. $n=3$. ** $P < 0.01$ vs. control; # $P < 0.05$, ### $P < 0.01$ vs. TGF- $\beta 1$. ART, artesunate; ART-L, low-dose ART; ART-M, moderate-dose ART; ART-H, high-dose ART; TGF- $\beta 1$, transforming growth factor- $\beta 1$; USAG-1, uterine sensitization-associated gene 1; BMP-7, bone morphogenetic protein 7.

upregulated the expression of USAG-1 in NRK-52E cells as presented in Fig. 5A and C. Treatment with ART (0.1 and 1 $\mu\text{g/ml}$) significantly decreased the expression of USAG-1 in a dose-dependent manner compared with the TGF- $\beta 1$ treated group (Fig. 5A and C). The effect of ART on the expression of BMP-7 was additionally detected. Compared with the control group, the expression of BMP-7 significantly decreased when treated with TGF- $\beta 1$; however, ART was able to restore the expression of BMP-7, and the highest dose of ART exerted the most marked effect (Fig. 5B and D). The results of the RT-sqPCR analysis of USAG-1 and BMP-7 were consistent with those of the western blot analysis.

In order to observe the localization of BMP-7 and USAG-1 in NRK-52E cells, immunofluorescent staining was performed to detect the two proteins (Fig. 6). The results demonstrated that the expression of USAG-1 in NRK-52E cells was minimal, while the expression of USAG-1 was markedly increased when treated with 5 ng/ml TGF- $\beta 1$. Notably, ART inhibited the increased expression of USAG-1 (Fig. 6A). Additionally, BMP-7 was abundantly expressed in NRK-52E cells; however,

the expression of BMP-7 was significantly decreased when induced by TGF- $\beta 1$, and ART was able to reverse this effect (Fig. 6B). The results further clarified that the expression of BMP-7 and USAG-1 may alter when induced by TGF- $\beta 1$, and suggested that ART may reverse these effects.

Effect of ART on the ratio of USAG-1 to BMP-7. In order to demonstrate the role of BMP-7 and USAG-1 in EMT, the effects of ART on the protein ratio of USAG-1 to BMP-7 were evaluated following treatment with TGF- $\beta 1$. The ratio increased in the TGF- $\beta 1$ group compared with the control group (Fig. 7). However, the three doses of ART in the experiment inhibited the elevation of the ratio, and this was significant in the ART-H and ART-M groups, demonstrating that the ameliorative effect of ART in EMT may be achieved by restoring the ratio of USAG-1 and BMP-7.

Discussion

The primary aim of the present study was to investigate the role of ART in EMT induced by TGF- $\beta 1$. In the present study, 2 days of exposure to TGF- $\beta 1$ in renal proximal tubular cells significantly increased the levels of α -SMA and decreased the expression of E-cadherin, suggesting that TGF- $\beta 1$ was able to induce the occurrence of EMT (25). In addition, it was demonstrated that ART was able to ameliorate TGF- $\beta 1$ -induced renal interstitial fibrosis. Notably, ART reversed the TGF- $\beta 1$ -induced increase in USAG-1 and decrease in BMP-7. The downregulation of USAG-1 expression resulted in the inhibition of EMT. These results provided evidence for a possible antifibrotic mechanism in renal interstitial fibrosis.

Renal interstitial fibrosis, which is characterized by the accumulation of the ECM, is the principal underlying pathology in the progression of chronic kidney diseases, and is a common pathway of chronic kidney disease progression to end-stage renal failure (1). The pathogenesis of renal fibrosis is characterized by tubular atrophy, tubular cell loss,

myofibroblast accumulation and excessive ECM protein deposition (26). EMT is a process through which epithelial cells lose their epithelial-specific biomarkers, undergo cytoskeletal remodeling, and gain a mesenchymal phenotype (4). It has been reported that the TGF- β -Smad signaling pathway may promote the occurrence of EMT. Studies have demonstrated that tubular EMT is an important resource of fibrogenic myofibroblasts and serves a central role in tubulointerstitial fibrosis, including in diabetes nephropathy. In the present study, ART-treated cells displayed a significant decrease in the levels of α -SMA and an increase the expression of E-cadherin, and ART was able to improve the morphological alterations induced by TGF- β 1 and restore the morphology of the epithelial cells; these results indicated that ART may improve renal fibrosis induced by TGF- β 1. Additionally, ART reversed the process of EMT induced by TGF- β 1.

BMP-7, a morphogenetic protein, has been hypothesized to serve a role in antifibrosis in renal tubular epithelial cells (27). Under normal circumstances, there is abundant BMP-7 expression in renal tubular epithelial cells, and BMP-7 is able to maintain the normal functioning of renal tubular epithelial cells (28). The expression of BMP-7 has been demonstrated to be significantly downregulated under disease conditions, including hypertension, aristolochic acid nephropathy, diabetic nephropathy and other disease states (29,30). In addition, it was reported that exogenous administration of recombinant human BMP-7 was able to reverse the alterations in renal structure and the degree of fibrosis in UUO rats by regulating the downstream Smad1/5/8 signaling pathway (24). A previous study demonstrated that BMP-7 inhibited TGF- β 1-induced fibrogenesis and EMT, and induced EMT *in vitro* (31). In the present study, the levels of BMP-7 mRNA and protein levels decreased during the process of EMT, which was further confirmed by the immunofluorescent staining results. However, ART was able to inhibit the process of EMT and significantly increase the levels of BMP-7, indicating that ART may inhibit the EMT process via an upregulation of BMP-7 expression.

It has been demonstrated that the local activity of endogenous BMP-7 is controlled by the regulation of its expression, in addition to certain classes of molecules termed BMP antagonists (13). USAG-1, a dominant antagonist of BMP-7, is primarily expressed in renal tubular epithelial cells and contributes to renal injury (9,10). In previous studies, it was demonstrated that USAG-1-knockout mice exhibited reduced tubulointerstitial fibrosis in a UUO model, and improved renal function in acute and chronic kidney injury. The above results illustrated that the upregulation of USAG-1 may serve an important role in the process of renal interstitial fibrosis (12). In the present study, the expression of USAG-1 was upregulated when induced by TGF- β 1, although it significantly decreased when treated with ART, particularly the high dose of ART. In addition, the reduction of USAG-1 expression may improve other indicators of fibrosis. The results stated above suggested that one of the possible mechanisms underlying the antifibrotic effects of ART may be associated with inhibition of the expression of USAG-1 in NRK-52E cells.

In conclusion, the results above demonstrate that ART was able to inhibit EMT induced by TGF- β 1. These effects may be associated with the upregulation of BMP-7 or the inhibition of

USAG-1 expression, although the exact mechanisms require further investigation.

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