



Draft Whole-Genome Sequences of 10 *Aeromonas* Strains from Clinical and Environmental Sources

Katie Gray,^a Luke R. Green,^a Roy R. Chaudhuri,^b Jonathan G. Shaw^a

^aDepartment of Infection, Immunity and Cardiovascular Disease, University of Sheffield, Sheffield, United Kingdom ^bDepartment of Molecular Biology and Biotechnology, University of Sheffield, Sheffield, United Kingdom

ABSTRACT Aeromonas bacteria are able to cause disease in a wide range of animals from humans to fish. In this article, we report the draft whole-genome sequences of 10 Aeromonas strains from clinical and environmental sources. These genome sequences will provide a repository of information for further investigations into the pathogenicity of this enigmatic pathogen.

eromonas species are aquatic bacteria found in brackish water and freshwater and are able to cause disease in both cold-blooded and warm-blooded animals (1). In humans, they can give rise to numerous clinical manifestations but are mostly associated with gastrointestinal disease and wound infections (2). A plethora of factors have been linked with aeromonad pathogenicity, including toxigenic factors, such as aerolysin and type III secretion systems (T3SS), and adhesins, such as type IV pili and both polar and lateral flagella (3–5). However, aeromonad pathogenicity is thought to be multifactorial and not reliant upon one virulence determinant. Aeromonas caviae strains Sch3 and Sch29 were isolated from children presenting with gastroenteritis at Sheffield Children's Hospital, and Aeromonas veronii biovar sobria BC88 was isolated from a child with dysentery in Western Australia (6); all the other aeromonads were environmental strains isolated from the River Don at various locations within South Yorkshire, United Kingdom. A. caviae Sch3 has been extensively studied due to its production of lateral flagella (4) and its ability to glycosylate its polar flagella (7, 8). A. veronii biovar sobria BC88 is a model for aeromonad adhesion, as it produces a type IV bundle-forming pilus (5). The analysis of the genomes presented here will allow a deeper understanding of the biology of aeromonads in relation to both their physiology and pathogenicity.

All environmental strains were isolated from river water by inoculation onto cystine lactose electrolyte-deficient (CLED) agar plus Andrade's indicator (Oxoid) and Brilliance UTI clarity agar (Oxoid), as these were nonselective and provided a differential presumptive identification of organisms. Human clinical strains were isolated by the inoculation of feces onto Aeromonas isolation agar (Fluka) containing ampicillin. All strains were identified to the species level by multisequence alignment against the sequences of known aeromonad species. All strains were grown in tryptic soy broth (TSB), and genomic DNA was extracted using a QIAamp DNA minikit (Qiagen). For each of the 10 Aeromonas strains, genomic DNA libraries were prepared using the Nextera XT library kit (Illumina, San Diego, CA). The genomes were nucleotide sequenced using an Illumina HiSeq 2500 platform at MicrobesNG (University of Birmingham, UK). Sequencing used 2 \times 250-bp paired-end reads that gave from 41 to 150 \times depth (Table 1). Trimmomatic v0.38 (9) was used to trim the reads with a sliding window quality cutoff of Q15, and read quality analyses were performed with FastQC software v0.11.8 (http://www.bioinformatics.babraham.ac.uk/projects/fastqc/). The genomes were assembled and annotated using SPAdes v3.7 (10) and Prokka v1.12, respectively, using the standard default settings (11). Genome assembly metrics were calculated with

Citation Gray K, Green LR, Chaudhuri RR, Shaw JG. 2019. Draft whole-genome sequences of 10 *Aeromonas* strains from clinical and environmental sources. Microbiol Resour Announc 8:e00170-19. https://doi.org/10.1128/ MRA.00170-19.

Editor David Rasko, University of Maryland School of Medicine

Copyright © 2019 Gray et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license

Address correspondence to Jonathan G. Shaw, J.G.Shaw@sheffield.ac.uk.

Received 19 February 2019 Accepted 1 July 2019 Published 25 July 2019

TABLE	1 Genome accession num	bers and charac	teristics of 1	0 Aeromoi	<i>nas</i> strains tak	en from eithe	er a clinical or	environme	ntal source				
		Mean	No. of	No. of	Largest	Total	GC			L ₅₀	L ₇₅	No. of	ENA accession
Strain	Species (source)	coverage (×)	reads	contigs	contig (bp)	length (bp)	content (%)	N ₅₀ (bp)	N ₇₅ (bp)	(contigs)	(contigs)	CDS	and assembly no.
Sch29	A. caviae (human	49.94	874,346	195	182,866	4,425,373	61.34	86,948	50,669	21	37	3,953	ERS3090042,
	clinical diarrhea)												GCA_901202955
Sch3N	<i>A. caviae</i> (human	84.83	1,155,915	192	240,555	4,737,922	61.39	90,385	43,401	18	37	4,313	ERS3090043,
	clinical diarrhea)												GCA_901212305
BC88	A. <i>veronii</i> (human	111.91	1,381,586	155	1,023,078	4,604,788	58.54	215,763	118,836	9	13	4,065	ERS3090044,
	clinical dysentery)												GCA_901212295
KLG1	A. hydrophila	53.88	730,744	154	260,972	4,908,673	61.22	103,738	64,788	15	30	4,373	ERS3090045,
	(environmental water)												GCA_901212375
KLG2	A. allosaccharophila	150.39	1,670,538	85	626,394	4,513,594	58.87	292,420	141,273	5	11	4,059	ERS3090046,
	(environmental water)												GCA_901212385
KLG5	A. veronii (environmental	70.63	974,322	103	818,983	4,740,061	58.51	280,270	130,857	9	12	4,269	ERS3090047,
	water)												GCA_901212355
KLG6	<i>A. media</i> (environmental	104.31	1,255,304	454	159,134	4,542,840	61.20	36,002	19,277	40	80	4,070	ERS3090048,
	water)												GCA_901212365
KLG7	A. veronii (environmental	53.29	687,242	104	345,454	4,552,893	58.80	139,212	84,196	11	21	4,069	ERS3090049,
	water)												GCA_901212345
KLG8	A. <i>veronii</i> (environmental	54.65	738,573	76	568,763	4,590,381	58.62	198,583	115,053	7	15	4,137	ERS3090050,
	water)												GCA_901212395
KLG9	A. <i>veronii</i> (environmental	40.98	578,468	74	399,442	4,605,689	58.71	180,084	120,803	6	17	4,160	ERS3090051,
	water)												GCA_901212405

QUAST, and the number of annotated coding sequences (CDS) for each aeromonad genome is shown in Table 1.

The sequences will provide a great resource for further investigations into the physiology and pathogenicity of the *Aeromonas* genus.

Data availability. The reads used for assembly of the 10 annotated aeromonad genomes were deposited in the European Nucleotide Archive (ENA) at the European Molecular Biology Laboratory (EMBL) under the accession number PRJEB31025. The specific accession numbers for each sample are supplied in Table 1. The versions described in this paper are the first versions.

ACKNOWLEDGMENT

MicrobesNG provided genome sequencing, which was supported by the BBSRC (grant number BB/L024209/1).

REFERENCES

- Janda JM, Abbott SL. 2010. The genus Aeromonas: taxonomy, pathogenicity and infection. Clin Microbiol Rev 23:35–73. https://doi.org/10 .1128/CMR.00039-09.
- Parker JL, Shaw JG. 2011. Aeromonas spp. clinical microbiology and disease. J Infect 62:109–118. https://doi.org/10.1016/j.jinf.2010.12.003.
- Rabaan AA, Gryllos I, Tomas JM, Shaw JG. 2001. Motility and the polar flagellum are required for *Aeromonas caviae* adherence to HEp-2 cells. Infect Immun 69:4257–4267. https://doi.org/10.1128/IAI.69.7.4257-4267 .2001.
- Gavin R, Rabaan AA, Merino S, Tomas JM, Gryllos I, Shaw JG. 2002. Lateral flagella of *Aeromonas* species are essential for epithelial cell adherence and biofilm formation. Mol Microbiol 43:383–397. https://doi.org/10 .1046/j.1365-2958.2002.02750.x.
- Hadi N, Yang Q, Barnett TC, Tabei SMB, Kirov SM, Shaw JG. 2012. Bundle-forming pilus locus of *Aeromonas veronii* bv sobria. Infect Immun 80:1351–1369. https://doi.org/10.1128/IAI.06304-11.
- Kirov SM, Sanderson K. 1996. Characterization of a type IV bundle forming pilus (SFP) from a gastroenteritis strain of *Aeromonas veronii* biovar sobria. Microb Pathog 21:23–34. https://doi.org/10.1006/mpat .1996.0039.

- Tabei SMB, Hitchen PG, Day-Williams MJ, Merino S, Vart R, Pang P-C, Horsburgh GJ, Viches S, Wilhelms M, Tomás JM, Dell A, Shaw JG. 2009. An *Aeromonas caviae* genomic island is required for both O-antigen lipopolysaccharide biosynthesis and flagellin glycosylation. J Bacteriol 191:2851–2863. https://doi.org/10.1128/JB.01406-08.
- Parker JL, Lowry R, Couto NAS, Wright PC, Stafford GS, Shaw JG. 2014. Maf-dependent bacterial flagellin glycosylation occurs before chaperone binding and flagellar T3SS export. Mol Microbiol 92:258–272. https://doi .org/10.1111/mmi.12549.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.