

# Biomarkers of Inflammation and Immune Function and Risk of Colorectal Cancer

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**Abstract** A substantial number of prospective epidemiological studies have been conducted to investigate the association between biomarkers of inflammation and immune function and risk of colorectal cancer. Although pre-diagnostic concentrations of these biomarkers, especially C-reactive protein, have been associated with a higher risk of colorectal cancer in some studies, this association does not seem to have a robust support without hints of bias. Future prospective studies should evaluate multiple inflammatory biomarkers with longitudinal measures over the follow-up taking advantage of new multiplex cytokine quantification arrays and use more sophisticated joint or biomarker pattern statistical approaches to capture the complex and dynamic interplay between biomarkers and risk of colorectal cancer. Large collaborative consortia and Mendelian randomization studies should be encouraged to diminish the threat of biases and improve the reliability of this literature.

**Keywords** Inflammation · Colorectal cancer · Biomarkers · C-reactive protein · Interleukin 6 · Tumor necrosis factor  $\alpha$  · Genetic polymorphisms · Cohort studies · Risk of bias

## Introduction

Colorectal cancer (CRC) is the third most commonly diagnosed cancer worldwide that accounted for 1.36 million new cases and 694,000 deaths in 2012 [1]. CRC develops through a gradual accumulation of genetic, both inherited and somatic, and epigenetic changes, leading to the transformation of normal colonic mucosa into invasive cancer. Approximately 70 to 90 % of CRCs arise from adenomatous polyps (adenomas) [2]. The risk of CRC increases with age especially after the age of 50 years, and the risk is also increased by certain high-penetrance inherited genetic mutations (familial adenomatous polyposis and hereditary non-polyposis colorectal cancer), several low penetrance mutations [3], a personal or family history of colorectal neoplasia, or a personal history of inflammatory bowel diseases (IBD) [4]. Several modifiable factors are also associated with increased risk of CRC, including obesity, physical inactivity, smoking, heavy alcohol consumption, type II diabetes, and a diet high in red or processed meat, while the use of aspirin or other non-steroidal anti-inflammatory drugs (NSAIDs) lowers risk [5].

Systemic inflammation is a pathway through which several of the aforementioned risk factors may lead to colorectal neoplasia. For example, obesity is associated with a state of chronic inflammation induced by excessive production of storage lipids and high circulating concentrations of glucose, both of which create a proinflammatory oxidative environment that has been associated with colorectal carcinogenesis [6, 7]. Adipocytes constitutively express the proinflammatory cytokine tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), and TNF- $\alpha$

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expression in adipocytes is markedly increased in obese rodents [8].

However, the link between inflammation and colorectal neoplasia is not only substantiated at a systemic level, and it has been further hypothesized that sporadic colorectal neoplasia arises from colonic areas with chronic subclinical inflammation. Several lines of evidence implicate chronic inflammation as a key predisposing factor to colorectal cancer in IBD [9]. First, the risk for developing colorectal cancer increases with longer duration of colitis. Second, the more colonic surface is involved with colitis, the greater the colon cancer risk. Third, evidence is mounting to suggest that anti-inflammatory medications, especially 5-aminosalicylates, can reduce the development of colorectal dysplasia and cancer in IBD. The evidence directly implicating colonic inflammation as a key predisposing factor to sporadic colorectal neoplasia is weaker compared to the evidence for colitis-associated colorectal neoplasia. Some, but not all, clinical studies have shown that colorectal adenoma and cancer patients had higher fecal calprotectin levels, a direct measure of gastrointestinal mucosal inflammation, than normal subjects [10–12]. Colonic microbiota are being increasingly recognized as important contributors to the health of the gastrointestinal tract and may not only lead to CRC development via modulation of the gut barrier function and initiation of inflammation at the colonic epithelium but could also promote systemic inflammation via leakage of bacterial endotoxins into the circulation [13, 14].

The aim of this review is to examine and critically appraise the associations in the literature between biomarkers of inflammation and immune function and risk of CRC and through which infer about the strength of the evidence linking systemic and/or chronic low-grade colorectal inflammation to colorectal carcinogenesis.

## Circulating Concentrations of Inflammatory Biomarkers and Risk of CRC

### C-Reactive Protein

C-reactive protein (CRP) is a sensitive but non-specific systemic marker of inflammation that is produced mainly in the liver in response to cytokines released by phagocytes during infection, trauma, surgery, burns, tissue infarction, advanced cancer, and chronic inflammatory conditions [15]. The circulating concentrations of CRP in healthy individuals are low, usually below 1 mg/L, but they sharply increase in acute infection reaching concentrations well above 10 mg/L. The half-life of CRP is 19 h [16].

A total of 18 prospective epidemiological studies have been conducted to date to investigate the association of pre-diagnostic high-sensitivity CRP concentrations with risk of CRC (Table 1) and have been summarized in three

meta-analyses [25, 37, 38•]. Fourteen out of 18 studies included in the most recent meta-analysis showed a higher risk of CRC with higher concentrations of CRP, but only six of them yielded statistically significant results [38•]. The summary random effects relative risk of CRC per one unit change in the natural logarithm (ln) of CRP was 1.12 (95 % CI, 1.05–1.21), but substantial heterogeneity was evident among the studies ( $P$  heterogeneity, 0.006;  $I^2$ , 52 %) [38•]. The summary association was statistically significant for colon cancer (RR, 1.13; 95 % CI, 1.05–1.21) but not for rectal cancer (RR, 1.03; 95 % CI, 0.90–1.17), although there was not a statistically significant difference between the estimates by anatomical location. Moreover, the summary association for CRP and CRC appeared stronger in men (RR, 1.18; 95 % CI, 1.06–1.30) than in women (RR, 1.05; 95 % CI, 0.96–1.16), but again the confidence intervals of the associations by sex overlapped. The results of this meta-analysis were consistent to the results of prior meta-analyses, including the one from our group [37].

The first prospective study was published in 2004 from the CLUE II cohort of Washington County, MD, with 172 CRC cases and 342 matched controls and found a twofold statistically significant increase in the risk of colorectal cancer comparing persons with the highest versus lowest fourth of C-reactive protein [17]. A subsequent study by Zhang and colleagues was performed within the Women's Health Study, which recruited female health professionals, and found an almost statistically significant inverse association between CRP and colorectal cancer [18]. The two largest studies in the field were published from the European Prospective Investigation into Cancer and Nutrition (EPIC) in 2010 and the Women's Health Initiative observational cohort in 2013 with approximately 1000 CRC cases each, but observed weak and borderline statistically significant associations [28, 34]. A new study was published in 2014, not included in the published meta-analyses, using the NHANES III data and found an imprecise but statistically significant increased risk of colorectal cancer mortality comparing CRP above 1 mg/dL versus undetectable concentrations, but the study did not use a high-sensitivity CRP assay and observed only 59 CRC deaths [39].

The evidence linking CRP concentrations with colorectal adenoma development, a precursor in the natural history of colorectal neoplasia, is weaker. Pre-diagnostic CRP concentrations were not associated with colorectal adenoma in our study in the CLUE II cohort [40]. A subsequent case-control study nested in the Prostate, Lung, Colorectal, and Ovarian Cancer Screening Trial observed a statistically significant 15 % reduction in risk per unit increase in ln CRP [41], which could reflect an attempt by the host to suppress tumor formation. Other studies observed mixed results from null

**Table 1** Characteristics of studies investigating the association between circulating concentrations of inflammatory biomarkers (per one unit change in the ln-transformed concentrations of CRP, IL-6, and TNF- $\alpha$ ) and the risk of colorectal cancer

Author, year	Country	Population	Study design	No. of cases	No. of controls/ population	Inflammatory biomarkers	RR (95 % CI)
Erlinger, 2004 [17]	USA	CLUE II	NCC	172	342	CRP	Colorectum 1.27 (1.03–1.57) Colon 1.37 (1.08–1.74) Rectum 0.99 (0.61–1.59)
Zhang, 2005 [18]	USA	WHS	Cohort	169	27,913	CRP	Colorectum 0.89 (0.76–1.04) Colon 0.80 (0.39–1.67) Rectum 0.52 (0.28–0.99)
Ito, 2005 [19]	Japan	JACC	NCC	141	327	CRP	Colorectum 1.11 (0.94–1.31) Colon 1.11 (0.90–1.37) Rectum 1.13 (0.86–1.49)
Il'yasova, 2005 [20]	USA	HABCS	Cohort	41	2438	CRP IL-6 TNF- $\alpha$	Colorectum 1.44 (1.03–2.02) Colorectum 1.44 (0.90–2.31) Colorectum 0.90 (0.42–1.94)
Trichopoulos, 2006 [21]	Greece	EPIC-Greece	NCC	48	996	CRP	Colorectum 1.19 (0.81–1.74) Colon 1.32 (0.84–2.08) Rectum 1.41 (0.94–2.11)
Siemes, 2006 [22]	Netherlands	Rotterdam Study	Cohort	189	6273	CRP	Colorectum 1.04 (0.88–1.22) Colon 1.15 (0.71–1.88) Rectum 0.70 (0.50–0.99)
Otani, 2006 [23]	Japan	JPHC	NCC	375	750	CRP	Colorectum 1.11 (0.95–1.30) Colon 1.06 (0.87–1.29) Rectum 1.19 (0.90–1.58)
Gunter, 2006 [24]	Finland	ATBC	NCC	130	260	CRP	Colorectum 1.33 (1.05–1.69) Colon 1.32 (0.97–1.79) Rectum 1.49 (1.02–2.17)
Heikkila, 2009 [25]	UK	BWHHS	Cohort	32	3274	CRP IL-6	Colorectum 0.97 (0.70–1.34) Colorectum 0.92 (0.53–1.60)
Heikkila, 2009 [25]	UK	Caerphilly	Cohort	41	1144	CRP IL-6	Colorectum 0.89 (0.66–1.22) Colorectum 0.71 (0.41–1.23)
Allin, 2009 [27]	Denmark	CCHS	Cohort	191	10,082	CRP	Colorectum 1.52 (0.86–2.67)
Aleksandrova, 2010 [28]	Europe	EPIC	NCC	1096	1096	CRP	Colorectum 1.06 (0.99–1.13) Colon 1.09 (1.01–1.18) Rectum 0.99 (0.88–1.10)
Chan, 2011 [29]	USA	NHS	NCC	280	555	CRP IL-6	

**Table 1** (continued)

Author, year	Country	Population	Study design	No. of cases	No. of controls/ population	Inflammatory biomarkers	RR (95 % CI)
							Colorectum 0.85 (0.67–1.09) Colorectum 1.09 (0.77–1.55)
Lee, 2011 [30]	Korea	Health exam records	Cohort	158	80,781	CRP	Colorectum 1.59 (1.13–2.22)
Prizment, 2011 [31]	USA	ARIC	Cohort	166	9836	CRP	Colorectum 1.36 (1.06–1.75) Colon 1.38 (1.05–1.82) Rectum 1.12 (0.64–2.00)
Van Hemelrijck, 2011 [32]	Sweden	AMORIS	Cohort	480	102,749	CRP	Colon 1.00 (0.92–1.07)
Ho, 2012 [33]	USA	WHI-OS	NCC	457	841	IL-6 TNF- $\alpha$	Colorectum 0.91 (0.60–1.38) Colorectum 0.77 (0.51–1.17) <sup>a</sup>
Toriola, 2013 [34]	USA	WHI-OS	NCC	988	988	CRP	Colorectum 1.09 (0.97–1.24) Colon 1.13 (0.99–1.30) Rectum 0.86 (0.62–1.20)
Song, 2013 [35]	USA	HPFS	NCC	274	532	CRP IL-6	Colorectum 1.25 (0.94–1.67) Colon 0.97 (0.68–1.39) Rectum 0.88 (0.49–1.56) Colorectum 1.45 (1.05–2.00)
Wu, 2013 [36]	China	SMHS	NCC	288	576	CRP	Colorectum 1.65 (1.10–2.48) Colon 1.90 (1.12–3.23) Rectum 1.29 (0.66–2.54)

CRP C-reactive protein; IL-6 interleukin-6; TNF- $\alpha$  tumor necrosis factor alpha; NCC nested case-control study; RR relative risk; WHS Women's Health Study; JACC Japan Collaborative Cohort Study; JPHC The Japan Public Health Center-based Prospective Study; ATBC Alpha-Tocopherol Beta-Carotene Cancer Prevention Study; CCHS Copenhagen City Heart Study; EPIC European Prospective Investigation into Cancer and Nutrition; AMORIS Apolipoprotein Mortality RISK study; ARIC Atherosclerosis Risk in Communities study; NHS Nurses' Health Study; BWHHS British Women's Heart and Health Study; HPFS Health Professionals Follow-up Study; HABCS Health, Aging and Body Composition cohort; WHI-OS Women's Health Initiative Observational Study; SMHS Shanghai Men's Health Study

<sup>a</sup> Association reported for top vs. bottom quartile

association in a case-control study in Hawaii to a higher risk of large adenomas for participants in the highest category of CRP concentrations in a case-control study in Japan [26, 42].

Given that high-sensitivity CRP is a circulating marker for inflammation, the lack of a consistent association with colorectal neoplasia raises questions on its use as a marker of the inflammatory process associated with colorectal cancer. When we investigated whether increased CRP is a marker of colonic inflammation, we found no such association in a colonoscopy-based study [43•], which implies that CRP may be marking systemic factors (e.g., obesity) that influence colorectal cancer risk or that the observational associations between CRP and colorectal cancer may be afflicted by biases.

In 2012, our group critically appraised the literature of 98 biomarkers and cancer risk evaluating 847 individual studies to shed light on the existence of potential biases and identify associations with robust evidence for their presence [44•, 45]. We concluded that the association between CRP and CRC was not among the robust associations without suggestion of bias, as evidence of large between-study heterogeneity, small-study effects (i.e., smaller studies tended to give larger estimates of effect size compared with larger studies), and excess statistical significance (i.e., the observed number of studies with nominally statistically significant results was greater than the expected number, calculated based on the statistical power to detect a plausible effect) was identified [44•]. It is thus

probable that the statistically significant studies in this field are subject to selective reporting or other biases. Selective reporting bias may emerge when there are many analyses that can be performed (using, for example, different adjustments for confounders or models with different statistical terms for CRP and confounders), but only the analysis with the most statistically significant result is presented [46, 47]. Studies demonstrating an association between inflammatory markers and colorectal neoplasia may also have reverse causation bias due to clinically undetected neoplasia at the time of blood collection. Inadequate control for potential confounders may also influence the findings of observational studies, as obesity, physical activity, and use of aspirin or non-steroidal anti-inflammatory drugs are each strongly related to both inflammatory activity and CRC risk.

Further evidence suggesting a lack of a causal link between CRP and colorectal neoplasia comes from candidate gene association studies that have observed mixed results for the association of genetic variants in the *CRP* gene and colorectal cancer risk [22, 48–52] and genome-wide association studies that have not identified *CRP* variants to be associated with CRC risk [3]. Mendelian randomization analyses have used single-nucleotide polymorphisms in *CRP* and/or genetic scores thereof as instruments of the circulating CRP concentrations in order to correct for potential biases of the observational association and have generally reported null results [50, 53, 54], except for a study in EPIC where genetically twofold higher CRP concentrations were associated with higher risk of CRC, but this result lost nominal statistical significance using alternative definitions of instrumental variables [49]. However, all these Mendelian randomization studies have used sub-optimal sample sizes with number of cancer cases below 900 [55]. Future studies should take advantage of recent developments in this field, i.e., the use of efficient designs and methods to adjust for violation of assumptions [56–58], to elucidate whether potential causal associations exist between inflammatory biomarkers and cancer risk.

### Interleukin-6

Interleukin-6 (IL-6) is a multifunctional cytokine produced by a variety of hematopoietic and non-hematopoietic cells [59] and exerts a proinflammatory role by acting either on a transmembrane type 1 cytokine receptor or by binding to a soluble IL-6 receptor (sIL6R). IL-6 upregulates several acute-phase proteins such as CRP, fibrinogen,  $\alpha$ 1-antitrypsin, and serum amyloid A [60]. There is ample mechanistic evidence suggesting an involvement of IL-6 in CRC development. In vivo experiments on wild-type mice have shown that IL-6 is significantly augmented at the colonic tumor environment, and the growth of colon tumors was suppressed when the mice were treated with antibodies against IL6R [61]. However, results from epidemiological studies regarding the association of

circulating IL-6 concentrations and risk of subsequent colorectal cancer development have been sparse, as only six prospective studies exist (Table 1) [20, 25, 29, 33, 35].

Il'yasova and colleagues found that serum IL-6 concentrations were not statistically significantly associated with the risk of colorectal cancer in the Health Aging and Body Composition Cohort [20]. Similarly, no association was observed in two prospective studies in the UK published in 2009 [25], but the latter three studies included only 30 to 40 colorectal cancer cases. Chan et al. used 279 cases from the Nurses' Health Study but also did not observe a statistically significant association [29], and another large publication with 413 cases from the Women's Health Initiative observational cohort yielded a similar not significant finding [33]. Only a recent paper from the Health Professionals Follow-up Study observed a borderline significant increased risk of colorectal cancer with higher IL-6 concentrations, but this association did not remain significant when the first 2 years of follow-up were excluded [35]. When the results of the latter six studies were synthesized in a meta-analysis, a non-significant summary relative risk was observed (RR, 1.10; 95 % CI, 0.88–1.36) per one unit change in ln IL-6 without substantial between-study heterogeneity ( $P$  heterogeneity, 0.18;  $I^2$ , 35 %) [38•].

The epidemiological studies have not in general reported results by tumor location, except for the Health Professionals Follow-up Study that observed a statistically significant increased risk of colon cancer with higher IL-6 concentrations, but the estimate was attenuated and not statistically significant anymore after excluding cases that occurred within 2 years of the baseline blood draw [35]. No association was observed for rectal cancer risk. Very recently, we finalized an analysis of pre-diagnostic plasma IL-6 concentrations and risk of colorectal cancer in the CLUE II cohort using 173 incident colorectal cancer cases and 345 matched controls. We observed that participants in the highest tertile of IL-6 concentration had a statistically significant and greater than twofold higher risk of colon cancer compared to participants in the bottom tertile (Tsilidis KK, personal communication). This association remained statistically significant after excluding cases that occurred within 2 or 5 years from the start of follow-up.

The association between *IL6* genetic polymorphisms and risk of CRC has been evaluated in several candidate gene studies with largely null results [48, 62–64]. The -174G>C polymorphism (rs1800795) has been widely evaluated, and a recent meta-analysis of 11 studies incorporating 6481 cases and 7935 controls did not find a statistically significant association [65].

### TNF-Alpha

TNF- $\alpha$  is another pleiotropic cytokine that is elevated in response to infection and tissue damage and has been shown to be involved in the pathogenesis of IBD [66]. However, scarce epidemiological evidence exists about a potential association between

pre-diagnostic TNF- $\alpha$  concentrations and risk of colorectal cancer (Table 1). No statistically significant associations have been observed in two North American prospective studies [33, 67]. Consistent with this evidence, Vaughn and colleagues reported a null association between TNF- $\alpha$  and colon adenoma in a colonoscopy-based case-control study [68]. A surrogate marker for TNF- $\alpha$ , soluble tumor necrosis factor receptor 2 (sTNFR-2), that is more stable in stored frozen samples and less influenced by diurnal variation was significantly associated with increased risk of CRC in women in the Nurses' Health Study [29], but no association was observed in two subsequent studies either in women or in men [35, 69]. Concentrations of sTNFR-2 were not associated with colorectal adenoma recurrence in patients with a prior adenoma [70]. Candidate gene association studies have not observed statistically significant findings between SNPs in *TNFA*, especially rs1800629, and risk of colorectal cancer [48, 62–64].

### Other Inflammatory Biomarkers

The literature evidence regarding the association of circulating concentrations of inflammatory biomarkers, other than CRP, IL-6, and TNF- $\alpha$ , and the risk of colorectal cancer is very sparse. Serum amyloid A (SAA) is a non-specific inflammatory marker produced in the liver in response to infection, trauma, and other inflammatory states and has been hypothesized to be a more sensitive marker of inflammation over CRP for certain diseases [15, 71]. However, pre-diagnostic concentrations of SAA were not associated with CRC in the Women's Health Initiative Observational Study [34]. Macrophage inhibitory cytokine 1 (MIC-1) is another novel inflammatory biomarker, which was positively associated with colorectal cancer risk in the Nurses' Health Study and the Health Professionals Follow-up Study [72]. Fibrinogen concentrations were positively associated with risk of colon cancer in another prospective study in the USA, but white blood cell count, von Willebrand factor, factor VIII, and albumin did not yield statistically significant associations [31]. However, white blood cell count was associated with an increased risk of colon cancer in a Swedish and a Korean prospective study [32, 73]. A recent nested case-control study in the EPIC cohort studied the association of neopterin, a biomarker of cellular immune activity, with the risk of colorectal cancer and found evidence of a U-shaped association, but neopterin has been shown to exert pleiotropic effects on cellular aging, oxidative stress, and inflammation [74]. Another pleiotropic biomarker, advanced glycation end products (AGE), that accumulates during aging and induces oxidative stress and inflammation has been investigated in the prospective Alpha-Tocopherol, Beta-Carotene Cancer Prevention (ATBC) study. High pre-diagnostic concentrations of AGE were not associated with colorectal cancer risk, but high concentrations of the soluble AGE receptor yielded a lower risk [75].

The adipose tissue is an active endocrine organ that secretes several cytokines and hormones, collectively termed adipokines (e.g., leptin, adiponectin, and resistin), which have been demonstrated to have pro- or anti-inflammatory functions. The association between adipokines and colorectal cancer risk has been investigated in several epidemiological studies, the results of which have been mixed [76]. The description and critical appraisal of this evidence are out of scope for the current review, as these adipokines have many pleiotropic effects (e.g., energy homeostasis, insulin signaling, and vascular reactivity) and cannot be considered primarily inflammatory biomarkers.

Calprotectin is a calcium- and zinc-binding protein released by activated innate immunity cells. It is found in cells, tissues, and fluids throughout the body and has been studied in a variety of inflammatory conditions [77]. Fecal calprotectin has been hypothesized to be a direct measure of gastrointestinal mucosal inflammation [78, 79]. Some clinical studies have shown that colorectal adenoma and cancer patients had higher fecal calprotectin levels than normal subjects, but a recent meta-analysis reported a non-statistically significant association between fecal calprotectin and colorectal neoplasia [12].

### Conclusions and Future Directions

The overall literature evidence for an association between inflammatory biomarkers and risk of colorectal cancer is inconclusive. Although concentrations of inflammatory biomarkers, especially CRP, have been associated with colorectal cancer risk in some studies, this association does not seem to have a robust support without hints of bias (e.g., selective reporting, reverse causation, residual confounding). This does not necessarily mean that these biomarkers are not truly associated with colorectal cancer, but there is still substantial uncertainty about them. If any potential causal association is identified in the future between an inflammatory biomarker and risk of colorectal cancer, then this might lead to the potential development or repurposing of drugs targeting receptors of inflammatory markers.

There are several ways to improve this evidence in the future. First, the measurement and estimation of a single biomarker and colorectal cancer association in most of the existing studies do not consider the complex synergisms among the many inflammatory biomarkers. Recent developments in serum cytokine quantification technology include multiplex arrays that can measure multiple biomarkers in each sample and better evaluate the complex and dynamic nature of inflammatory responses with substantial cost and sample savings [80]. When many biomarkers are available per participant, then a more sophisticated joint or biomarker pattern statistical approach or an exposure-wide association study could be used to assist in capturing the complete picture of the biomarker and cancer association. Some of these methods have been used in the

existing literature, but their use is still sparse and their results are inconsistent and imprecise [31, 32, 34, 69, 81]. Second, the single measurement of an inflammatory biomarker in time does not represent an average concentration over time and may lead to regression dilution bias. Future studies should better ascertain how long-term exposure to concentrations of inflammatory biomarkers may be related to colorectal cancer risk, as reports have shown that the long-term stability of inflammatory biomarkers is good but not perfect [82, 83].

Third, large collaborative consortia should be encouraged in this field. The use of standardized definitions and protocols for exposures, outcomes, confounders, and statistical analyses may diminish the threat of biases and improve the reliability of this literature. Many of the biases may be further lessened if studies were more completely and transparently reported according to published guidelines, such as the Strengthening the Reporting of Observational Studies in Epidemiology statement and its extension for Molecular Epidemiology [84–86]. Statistical significance testing should not be used as a criterion, either from authors or editors, for publication of biomarker studies. Future Mendelian randomization studies could also assist in elucidating potential causal associations.

#### Compliance with Ethics Guidelines

**Conflict of Interest** Alicia Garcia-Anguita, Artemisia Kakourou, and Konstantinos K. Tsilidis declare that they have no conflict of interest.

**Human and Animal Rights and Informed Consent** This article does not contain any studies with human or animal subjects performed by any of the authors.

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#### References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
  - Of major importance
1. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*. 2015;136(5):E359–86.
  2. Benson 3rd AB. Epidemiology, disease progression, and economic burden of colorectal cancer. *J Manag Care Pharm*. 2007;13(6 Suppl C):S5–18.
  3. Whiffin N, Hosking FJ, Farrington SM, Palles C, Dobbins SE, Zgaga L, et al. Identification of susceptibility loci for colorectal cancer in a genome-wide meta-analysis. *Hum Mol Genet*. 2014;23(17):4729–37.
  4. Hawk ET, Limburg PJ, Viner JL. Epidemiology and prevention of colorectal cancer. *Surg Clin North Am*. 2002;82(5):905–41.
  5. Chan AT, Giovannucci EL. Primary prevention of colorectal cancer. *Gastroenterology*. 2010;138(6):2029–43. **e10**.
  6. Esposito K, Nappo F, Marfella R, Giugliano G, Giugliano F, Ciotola M, et al. Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: role of oxidative stress. *Circulation*. 2002;106(16):2067–72.
  7. Mohanty P, Hamouda W, Garg R, Aljada A, Ghanim H, Dandona P. Glucose challenge stimulates reactive oxygen species (ROS) generation by leucocytes. *J Clin Endocrinol Metab*. 2000;85(8):2970–3.
  8. Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor- $\alpha$ : direct role in obesity-linked insulin resistance. *Science*. 1993;259(5091):87–91.
  9. Itzkowitz SH. Cancer prevention in patients with inflammatory bowel disease. *Gastroenterol Clin North Am*. 2002;31(4):1133–44.
  10. Kristinsson J, Roseth A, Fagerhol MK, Aadland E, Schjonsby H, Bormer OP, et al. Fecal calprotectin concentration in patients with colorectal carcinoma. *Dis Colon Rectum*. 1998;41(3):316–21.
  11. Kronborg O, Ugstad M, Fuglerud P, Johne B, Hardcastle J, Scholefield JH, et al. Faecal calprotectin levels in a high risk population for colorectal neoplasia. *Gut*. 2000;46(6):795–800.
  12. von Roon AC, Karamountzos L, Purkayastha S, Reese GE, Darzi AW, Teare JP, et al. Diagnostic precision of fecal calprotectin for inflammatory bowel disease and colorectal malignancy. *Am J Gastroenterol*. 2007;102(4):803–13.
  13. Ahn J, Sinha R, Pei Z, Dominianni C, Wu J, Shi J, et al. Human gut microbiome and risk for colorectal cancer. *J Natl Cancer Inst*. 2013;105(24):1907–11.
  14. Louis P, Hold GL, Flint HJ. The gut microbiota, bacterial metabolites and colorectal cancer. *Nat Rev Microbiol*. 2014;12(10):661–72.
  15. Gabay C, Kushner I. Acute-phase proteins and other systemic responses to inflammation. *N Engl J Med*. 1999;340(6):448–54.
  16. Mazlam MZ, Hodgson HJ. Why measure C reactive protein? *Gut*. 1994;35(1):5–7.
  17. Erlinger TP, Platz EA, Rifai N, Helzlsouer KJ. C-reactive protein and the risk of incident colorectal cancer. *JAMA*. 2004;291(5):585–90.
  18. Zhang SM, Buring JE, Lee IM, Cook NR, Ridker PM. C-reactive protein levels are not associated with increased risk for colorectal cancer in women. *Ann Intern Med*. 2005;142(6):425–32.
  19. Ito Y, Suzuki K, Tamakoshi K, Wakai K, Kojima M, Ozasa K, et al. Colorectal cancer and serum C-reactive protein levels: a case-control study nested in the JACC Study. *J Epidemiol*. 2005;15 Suppl 2:S185–9.
  20. Il'yasova D, Colbert LH, Harris TB, Newman AB, Bauer DC, Satterfield S, et al. Circulating levels of inflammatory markers and cancer risk in the health aging and body composition cohort. *Cancer Epidemiol Biomarkers Prev*. 2005;14(10):2413–8.
  21. Trichopoulos D, Psaltopoulou T, Orfanos P, Trichopoulou A, Boffetta P. Plasma C-reactive protein and risk of cancer: a prospective study from Greece. *Cancer Epidemiol Biomarkers Prev*. 2006;15(2):381–4.
  22. Siemes C, Visser LE, Coebergh JW, Splinter TA, Witteman JC, Uitterlinden AG, et al. C-reactive protein levels, variation in the C-reactive protein gene, and cancer risk: the Rotterdam Study. *J Clin Oncol*. 2006;24(33):5216–22.

23. Otani T, Iwasaki M, Sasazuki S, Inoue M, Tsugane S, Japan Public Health Center-Based Prospective Study G. Plasma C-reactive protein and risk of colorectal cancer in a nested case-control study: Japan public health center-based prospective study. *Cancer Epidemiol Biomarkers Prev.* 2006;15(4):690–5.
24. Gunter MJ, Stolzenberg-Solomon R, Cross AJ, Leitzmann MF, Weinstein S, Wood RJ, et al. A prospective study of serum C-reactive protein and colorectal cancer risk in men. *Cancer Res.* 2006;66(4):2483–7.
25. Heikkila K, Harris R, Lowe G, Rumley A, Yamell J, Gallacher J, et al. Associations of circulating C-reactive protein and interleukin-6 with cancer risk: findings from two prospective cohorts and a meta-analysis. *Cancer Causes Control.* 2009;20(1):15–26.
26. Otake T, Uezono K, Takahashi R, Fukumoto J, Tabata S, Abe H, et al. C-reactive protein and colorectal adenomas: Self Defense Forces Health Study. *Cancer Sci.* 2009;100(4):709–14.
27. Allin KH, Bojesen SE, Nordestgaard BG. Baseline C-reactive protein is associated with incident cancer and survival in patients with cancer. *J Clin Oncol.* 2009;27(13):2217–24.
28. Aleksandrova K, Jenab M, Boeing H, Jansen E, Bueno-de-Mesquita HB, Rinaldi S, et al. Circulating C-reactive protein concentrations and risks of colon and rectal cancer: a nested case-control study within the European Prospective Investigation into Cancer and Nutrition. *Am J Epidemiol.* 2010;172(4):407–18.
29. Chan AT, Ogino S, Giovannucci EL, Fuchs CS. Inflammatory markers are associated with risk of colorectal cancer and chemopreventive response to anti-inflammatory drugs. *Gastroenterology.* 2011;140(3):799–808. **quiz e11.**
30. Lee S, Choe JW, Kim HK, Sung J. High-sensitivity C-reactive protein and cancer. *J Epidemiol.* 2011;21(3):161–8.
31. Prizment AE, Anderson KE, Visvanathan K, Folsom AR. Association of inflammatory markers with colorectal cancer incidence in the atherosclerosis risk in communities study. *Cancer Epidemiol Biomarkers Prev.* 2011;20(2):297–307.
32. Van Hemelrijck M, Holmberg L, Garmo H, Hammar N, Walldius G, Binda E, et al. Association between levels of C-reactive protein and leukocytes and cancer: three repeated measurements in the Swedish AMORIS study. *Cancer Epidemiol Biomarkers Prev.* 2011;20(3):428–37.
33. Ho GY, Wang T, Gunter MJ, Strickler HD, Cushman M, Kaplan RC, et al. Adipokines linking obesity with colorectal cancer risk in postmenopausal women. *Cancer Res.* 2012;72(12):3029–37.
34. Toriola AT, Cheng TY, Neuhauser ML, Wener MH, Zheng Y, Brown E, et al. Biomarkers of inflammation are associated with colorectal cancer risk in women but are not suitable as early detection markers. *Int J Cancer.* 2013;132(11):2648–58.
35. Song M, Wu K, Ogino S, Fuchs CS, Giovannucci EL, Chan AT. A prospective study of plasma inflammatory markers and risk of colorectal cancer in men. *Br J Cancer.* 2013;108(9):1891–8.
36. Wu J, Cai Q, Li H, Cai H, Gao J, Yang G, et al. Circulating C-reactive protein and colorectal cancer risk: a report from the Shanghai Men's Health Study. *Carcinogenesis.* 2013;34(12):2799–803.
37. Tsilidis KK, Branchini C, Guallar E, Helzlsouer KJ, Erlinger TP, Platz EA. C-reactive protein and colorectal cancer risk: a systematic review of prospective studies. *Int J Cancer.* 2008;123(5):1133–40.
38. Zhou B, Shu B, Yang J, Liu J, Xi T, Xing Y. C-reactive protein, interleukin-6 and the risk of colorectal cancer: a meta-analysis. *Cancer Causes Control.* 2014;25(10):1397–405. **This study is the most recent meta-analysis investigating the association between pre-diagnostic circulating C-reactive protein (CRP) and interleukin-6 (IL-6) concentrations and risk of colorectal cancer. The results suggested that CRP was associated with an increased risk of colorectal cancer, but substantial heterogeneity was evident among the studies. No statistically significant association was observed between IL-6 and colorectal cancer.**
39. Swede H, Hajduk AM, Sharma J, Rawal S, Rasool H, Vella AT, et al. Baseline serum C-reactive protein and death from colorectal cancer in the NHANES III cohort. *Int J Cancer.* 2014;134(8):1862–70.
40. Tsilidis KK, Erlinger TP, Rifai N, Hoffman S, Hoffman-Bolton J, Helzlsouer KJ, et al. C-reactive protein and colorectal adenoma in the CLUE II cohort. *Cancer Causes Control.* 2008;19(6):559–67.
41. Gunter MJ, Cross AJ, Huang WY, Stanczyk FZ, Purdue M, Xue X, et al. A prospective evaluation of C-reactive protein levels and colorectal adenoma development. *Cancer Epidemiol Biomarkers Prev.* 2011;20(3):537–44.
42. Ognjanovic S, Yamamoto J, Saltzman B, Franke A, Ognjanovic M, Yokochi L, et al. Serum CRP and IL-6, genetic variants and risk of colorectal adenoma in a multiethnic population. *Cancer Causes Control.* 2010;21(7):1131–8.
43. Joshu CE, Tsilidis KK, Peskoe SB, Giardiello FM, Dlugniewski PJ, Nelson WG, et al. The association between circulating high-sensitivity C-reactive protein concentration and pathologic measures of colonic inflammation. *Cancer Causes Control.* 2014;25(4):409–18. **This colonoscopy-based study evaluated the association between circulating CRP concentrations and colonic mucosal measures of inflammation. The authors observed that CRP was not associated with colonic inflammation, which implies that CRP may be marking systemic factors (e.g., obesity) that influence colorectal cancer risk or that the observational associations between CRP and colorectal cancer may be afflicted by biases.**
44. Tsilidis KK, Papatheodorou SI, Evangelou E, Ioannidis JP. Evaluation of excess statistical significance in meta-analyses of 98 biomarker associations with cancer risk. *J Natl Cancer Inst.* 2012;104(24):1867–78. **This is an umbrella review that evaluated the epidemiological literature of biomarker concentrations and cancer risk. The authors concluded that the associations between CRP and colorectal cancer had evidence of large between-study heterogeneity, small-study effects, and excess statistical significance suggesting existence of selective reporting or other biases.**
45. Papatheodorou SI, Tsilidis KK, Evangelou E, Ioannidis JP. Application of credibility ceilings probes the robustness of meta-analyses of biomarkers and cancer risk. *J Clin Epidemiol.* 2015;68(2):163–74.
46. Ioannidis JP. Why most discovered true associations are inflated. *Epidemiology.* 2008;19(5):640–8.
47. Chan AW, Altman DG. Identifying outcome reporting bias in randomised trials on PubMed: review of publications and survey of authors. *BMJ.* 2005;330(7494):753.
48. Tsilidis KK, Helzlsouer KJ, Smith MW, Grinberg V, Hoffman-Bolton J, Clipp SL, et al. Association of common polymorphisms in IL10, and in other genes related to inflammatory response and obesity with colorectal cancer. *Cancer Causes Control.* 2009;20(9):1739–51.
49. Nimptsch K, Aleksandrova K, Boeing H, Janke J, Lee YA, Jenab M, et al. Association of CRP genetic variants with blood concentrations of C-reactive protein and colorectal cancer risk. *Int J Cancer.* 2015;136(5):1181–92.
50. Heikkila K, Silander K, Salomaa V, Jousilahti P, Koskinen S, Pukkala E, et al. C-reactive protein-associated genetic variants and cancer risk: findings from FINRISK 1992, FINRISK 1997 and Health 2000 studies. *Eur J Cancer.* 2011;47(3):404–12.
51. Prizment AE, Folsom AR, Dreyfus J, Anderson KE, Visvanathan K, Joshu CE, et al. Plasma C-reactive protein, genetic risk score, and risk of common cancers in the Atherosclerosis Risk in Communities study. *Cancer Causes Control.* 2013;24(12):2077–87.
52. Slattery ML, Curtin K, Poole EM, Duggan DJ, Samowitz WS, Peters U, et al. Genetic variation in C-reactive protein in relation



- to colon and rectal cancer risk and survival. *Int J Cancer*. 2011;128(11):2726–34.
53. Allin KH, Nordestgaard BG, Zacho J, Tybjaerg-Hansen A, Bojesen SE. C-reactive protein and the risk of cancer: a Mendelian randomization study. *J Natl Cancer Inst*. 2010;102(3):202–6.
  54. Shinohara RT, Frangakis CE, Platz E, Tsilidis K. Designs combining instrumental variables with case-control: estimating principal strata causal effects. *Int J Biostat* 2012;8(1).
  55. Pierce BL, Ahsan H, Vanderweele TJ. Power and instrument strength requirements for Mendelian randomization studies using multiple genetic variants. *Int J Epidemiol*. 2011;40(3):740–52.
  56. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet Epidemiol*. 2013;37(7):658–65.
  57. Pierce BL, Burgess S. Efficient design for Mendelian randomization studies: subsample and 2-sample instrumental variable estimators. *Am J Epidemiol*. 2013;178(7):1177–84.
  58. Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. *Int J Epidemiol*. 2015;44(2):512–25.
  59. Kishimoto T, Akira S, Narazaki M, Taga T. Interleukin-6 family of cytokines and gp130. *Blood*. 1995;86(4):1243–54.
  60. Naka T, Nishimoto N, Kishimoto T. The paradigm of IL-6: from basic science to medicine. *Arthritis Res*. 2002;4 Suppl 3:S233–42.
  61. Becker C, Fantini MC, Wirtz S, Nikolaev A, Lehr HA, Galle PR, et al. IL-6 signaling promotes tumor growth in colorectal cancer. *Cell Cycle*. 2005;4(2):217–20.
  62. Basavaraju U, Shebl FM, Palmer AJ, Berry S, Hold GL, El-Omar EM, et al. Cytokine gene polymorphisms, cytokine levels and the risk of colorectal neoplasia in a screened population of Northeast Scotland. *Eur J Cancer Prev*. 2015;24(4):296–304.
  63. Theodoropoulos G, Papaconstantinou I, Felekouras E, Nikiteas N, Karakitsos P, Panoussopoulos D, et al. Relation between common polymorphisms in genes related to inflammatory response and colorectal cancer. *World J Gastroenterol*. 2006;12(31):5037–43.
  64. Landi S, Moreno V, Gioia-Patricola L, Guino E, Navarro M, de Oca J, et al. Association of common polymorphisms in inflammatory genes interleukin (IL)6, IL8, tumor necrosis factor alpha, NFKB1, and peroxisome proliferator-activated receptor gamma with colorectal cancer. *Cancer Res*. 2003;63(13):3560–6.
  65. Hu JJ, Wang ZT, Zhong J. Lack of association between the interleukin 6 gene -174G>C polymorphism and colorectal cancer: evidence from a meta-analysis. *Genet Mol Res*. 2013;12(3):2205–14.
  66. Craven B, Zaric V, Martin A, Mureau C, Egan LJ. Effect of genetic deletion or pharmacological antagonism of tumor necrosis factor alpha on colitis-associated carcinogenesis in mice. *Inflamm Bowel Dis* 2015.
  67. Yamaji T, Iwasaki M, Sasazuki S, Tsugane S. Interaction between adiponectin and leptin influences the risk of colorectal adenoma. *Cancer Res*. 2010;70(13):5430–7.
  68. Vaughn CB, Ochs-Balcom HM, Nie J, Chen Z, Thompson CL, Tracy R, et al. No association between circulating levels and genetic variants of IL-6 and TNF-alpha and colon adenoma. *Gastroenterology Res* 2013;6(2).
  69. Ho GY, Wang T, Zheng SL, Tinker L, Xu J, Rohan TE, et al. Circulating soluble cytokine receptors and colorectal cancer risk. *Cancer Epidemiol Biomarkers Prev*. 2014;23(1):179–88.
  70. Ho GY, Xue X, Cushman M, McKeown-Eyssen G, Sandler RS, Ahnen DJ, et al. Antagonistic effects of aspirin and folic acid on inflammation markers and subsequent risk of recurrent colorectal adenomas. *J Natl Cancer Inst*. 2009;101(23):1650–4.
  71. Raynes JG, Cooper EH. Comparison of serum amyloid A protein and C-reactive protein concentrations in cancer and non-malignant disease. *J Clin Pathol*. 1983;36(7):798–803.
  72. Mehta RS, Song M, Bezawada N, Wu K, Garcia-Albeniz X, Morikawa T, et al. A prospective study of macrophage inhibitory cytokine-1 (MIC-1/GDF15) and risk of colorectal cancer. *J Natl Cancer Inst*. 2014;106(4), dju016.
  73. Lee YJ, Lee HR, Nam CM, Hwang UK, Jee SH. White blood cell count and the risk of colon cancer. *Yonsei Med J*. 2006;47(5):646–56.
  74. Aleksandrova K, Chuang SC, Boeing H, Zuo H, Tell GS, Pischon T, et al. A prospective study of the immune system activation biomarker neopterin and colorectal cancer risk. *J Natl Cancer Inst* 2015;107(4).
  75. Jiao L, Taylor PR, Weinstein SJ, Graubard BI, Virtamo J, Albanes D, et al. Advanced glycation end products, soluble receptor for advanced glycation end products, and risk of colorectal cancer. *Cancer Epidemiol Biomarkers Prev*. 2011;20(7):1430–8.
  76. Aleksandrova K, Nimptsch K, Pischon T. Influence of obesity and related metabolic alterations on colorectal cancer risk. *Curr Nutr Rep*. 2013;2(1):1–9.
  77. Smith LA, Gaya DR. Utility of faecal calprotectin analysis in adult inflammatory bowel disease. *World J Gastroenterol*. 2012;18(46):6782–9.
  78. Brydon WG, Campbell SS, Anderson NA, Wilson RG, Ghosh S. Faecal calprotectin levels and colorectal neoplasia. *Gut*. 2001;48(4):579–80.
  79. Poullis A, Foster R, Mendall MA, Fagerhol MK. Emerging role of calprotectin in gastroenterology. *J Gastroenterol Hepatol*. 2003;18(7):756–62.
  80. Leng SX, McElhane JE, Walston JD, Xie D, Fedarko NS, Kuchel GA. ELISA and multiplex technologies for cytokine measurement in inflammation and aging research. *J Gerontol A Biol Sci Med Sci*. 2008;63(8):879–84.
  81. Aleksandrova K, Jenab M, Bueno-de-Mesquita HB, Fedirko V, Kaaks R, Lukanova A, et al. Biomarker patterns of inflammatory and metabolic pathways are associated with risk of colorectal cancer: results from the European Prospective Investigation into Cancer and Nutrition (EPIC). *Eur J Epidemiol*. 2014;29(4):261–75.
  82. Platz EA, Sutcliffe S, De Marzo AM, Drake CG, Rifai N, Hsing AW, et al. Intra-individual variation in serum C-reactive protein over 4 years: an implication for epidemiologic studies. *Cancer Causes Control*. 2010;21(6):847–51.
  83. Pischon T, Hankinson SE, Hotamisligil GS, Rifai N, Rimm EB. Leisure-time physical activity and reduced plasma levels of obesity-related inflammatory markers. *Obes Res*. 2003;11(9):1055–64.
  84. Altman DG, Simera I, Hoey J, Moher D, Schulz K. EQUATOR: reporting guidelines for health research. *Lancet*. 2008;371(9619):1149–50.
  85. von Elm E, Altman DG, Egger M, Pocock SJ, Gotsche PC, Vandenbroucke JP, et al. The Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement: guidelines for reporting observational studies. *PLoS Med*. 2007;4(10), e296.
  86. Gallo V, Egger M, McCormack V, Farmer PB, Ioannidis JP, Kirsch-Volders M, et al. STrengthening the Reporting of OBServational studies in Epidemiology-Molecular Epidemiology (STROBE-ME): an extension of the STROBE statement. *PLoS Med*. 2011;8(10), e1001117.