



LncRNA XIST acts as a ceRNA sponging miR-185-5p to modulate pancreatic cancer cell proliferation via targeting CCND2

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Background: Long non-coding RNAs (lncRNAs) have been proved to be involved in the occurrence and progression of various tumors including pancreatic cancer (PC). Growing evidence shows that lncRNA X inactive-specific transcript (XIST) functions as an oncogene in multiple tumorigenesis. However, the underlying mechanism of lncRNA XIST in the progression of PC remains elusive.

Methods: Expression levels of XIST and miR-185-5p both in PC tissues or PC cells were determined using real-time quantitative PCR (qRT-PCR). Gain and loss-of-function of XIST or miR-185-5p was performed for further exploration. Moreover, colony formation assay was performed to assess cell proliferation. Flow cytometry analysis was performed to measure cell cycle and apoptosis. Dual-luciferase reporter assay was conducted to verify the correlation between XIST, miR-185-5p and CCND2, respectively. Additionally, western blot analysis was conducted to determine the expression pattern of apoptosis-related proteins and cell cycle-associated proteins.

Results: Herein, we found that XIST expression was up-regulated while miR-185-5p was down-regulated both in PC tissues and cell lines, compared with that of controls. Moreover, there was a negative correlation between XIST and miR-185-5p. Following that, functional experiments displayed that knockdown of XIST or overexpression of miR-185-5p inhibited cell proliferation, induced cell cycle arrest and promoted apoptosis in PC cells. Furthermore, mechanistic experiments displayed that XIST could negatively regulate miR-185-5p via direct binding. In addition, CCND2 was shown to be a downstream target of miR-185-5p. Importantly, overexpression or knockdown of XIST significantly increased or decreased the expression of CCND2, while these effects were reversed by miR-185-5p.

Conclusions: Taken together, our study demonstrated that lncRNA XIST functions as an oncogene and exerts its regulation via miR-185-5p/CCND2 axis, promoting proliferation and inhibiting apoptosis in PC.

Keywords: Pancreatic cancer (PC); long non-coding RNAs (lncRNA); X inactive-specific transcript (XIST); miR-185-5p; CCND2; proliferation

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Introduction

Due to a high tendency for aggressive invasion and distant metastasis, the lethality rate of pancreatic cancer (PC) is the

fourth highest among all cancers (1). It is reported that PC has a high incidence with >40,000 cases diagnosed each year and a very low 5-year survival rate estimated to be <5% (2,3). Despite the advancement of surgical technologies including

chemotherapy and radiotherapy, the efficient remission of PC treatment has not been emerged. Therefore, finding novel biomarkers for identifying biological characteristics of PC is urgent to further explore.

Long non-coding RNAs (lncRNAs), the RNA without the function of protein coding, consist of more than 200 of nucleotide units (4). Emerging evidence has shown the crucial roles of lncRNAs in various bioprocesses and diseases, not only participate in epigenetic modification and transcription of normal cell, but also regulate the proliferation, differentiation and metastasis of cancer cells (5). It has been demonstrated that the dysregulation of lncRNAs can be potentially functioned as important regulators in the development and progression of PC (6). For example, lncRNA UCA1 was considered to impact the proliferation, invasion and migration of PC via regulating miR-96/FOXO3 (7). And knockdown of MEG3 enhanced the PC cell proliferation, migration and invasion, as well as induce epithelial-mesenchymal transition (EMT) (4).

X inactive-specific transcript (XIST), one of the lncRNA affecting the expression of gene, can mediate the transcription silence of X chromosome via a complex mechanism (8). Besides, XIST has been proved to be associated with development of multiple cancers including non-small cell lung cancer (NSCLC), gastric cancer, and hematoma (9-11). Furthermore, XIST was also found to be elevated and could enhance the proliferation PC cell via interacting with miR-40/miR-124/iASPP or miR-133a/EGFR (12,13). However, deeper insights for XIST in the development of PC still need to be further explored.

MicroRNAs (miRNAs), which bind to the 3'-untranslated region of mRNAs and regulate mRNA expression, are small non-coding RNAs (14). It has been demonstrated that miR-185-5p could act as a tumor-suppressive gene involved in mediating the formation of various tumor tissues, like hepatocellular carcinoma, colorectal cancer and NSCLC (15,16). Consistently, miR-185-5p was down-regulated in PC and could also inhibit the proliferation of PC cell via targeting transcriptional co-activator (15,17). Taken together, we hypothesized that miR-185-5p may also exert an anti-tumor function in PC.

We provided insights in this study into the promotive function of lncRNA XIST and the regulatory network of XIST, miR-185-5p, CCND2 in PC. Our findings suggested that lncRNA XIST could exert its oncogenic function as a ceRNA for miR-185-5p to modulate CCND2 expression, which might provide some new targets for PC treatment.

Methods

Tissue samples

PC specimens and adjacent normal tissues were collected from 70 patients who received surgery in the Second Xiangya Hospital of Central South University from 2012 to 2016. All tissues were immediately collected after resection and stored under -80°C after washing with sterile phosphate-buffered saline (PBS). All patients were informed to write consent and all the experiments involved in this study were approved by the Ethics Committee of the Second Xiangya Hospital of Central South University (No. 201103301).

Cell culture and transfection

PC cell lines PANC-1, ASPC-1, HPAC, BxPC-3 cells and normal pancreatic cell line HPDE cells were all purchased from ATCC (Manassas, VA, USA). Briefly, cells were cultured in RPMI 1640 medium (Thermo Fisher Scientific, USA) containing 10% Gibco[®] fetal bovine serum (FBS) and 100 $\mu\text{g}/\text{mL}$ penicillin-streptomycin (Sigma-Aldrich Co, USA) at 37°C and 5% CO_2 . Lentivirus for shRNA for XIST (sh-XIST), negative control (NC) and overexpression plasmids for XIST (pcDNA3.1-XIST) were purchased from Shanghai GenePharma Co., Ltd. (Shanghai, China). miR-185-5p mimics, miR-185-5p inhibitor and their negative controls were purchased from RiboBio (Guangzhou, China). After cultured to 70–80% confluence, cells were transfected with sh-XIST, pcDNA3.1-XIST, or miR-185-5p mimics, miR-185-5p negative control at a concentration of 50 nmol/L using Lipofectamine 3000 (Invitrogen, USA) according to the manufacturer's instruction.

Flow cytometry assay

Flow cytometry was performed to assess the cell cycle and apoptosis. For the detection of cell cycle, briefly, cells were trypsinized and fixed with 70% absolute ethyl alcohol overnight at 4°C . And then 100 μL RNase A Reagent (Keygen Biotech, Nanjing, China) was added into cells. Propidium iodide reagent (Keygen Biotech) was used to stain the cells at the concentration of 20 $\mu\text{g}/\text{mL}$ for 20 minutes. Then DNA content was analyzed by FACScalibur (BD Bioscience, USA). For the detection of apoptosis, cells were harvested and then re-suspended with buffer. The FITC Annexin V Apoptosis Detection

Kit I (Ruibo, Guangzhou, China) was applied for staining the cells. A FACScan flow cytometer was performed for analysis cells apoptosis. All the experiments were in triplicated.

Colony formation assay

Cell proliferation was detected using clonogenic formation assay. Briefly, cells were seeded into a 6-well plate at a density of 500 cells per well. The cells were fixed with 4% paraformaldehyde and stained by crystal violet (1 mg/mL) after 14 days of culture. Then the formed number of colonies in culture plate seeded with different cells was counted (>50 cells) under microscope.

Dual luciferase reporter assay

To verify the prediction, dual luciferase reporter assay was conducted as reported elsewhere (18). The predicted binding mode was obtained using bioinformatic prediction by software targetscan 5.1 (<http://www.targetscan.org>; Whitehead Institute for Biomedical Research, Cambridge, MA, USA). PCR was performed to amplify the 3'-end fragment of lncRNA XIST containing the predicted miR-185-5p-binding site and to subclone into a pmirGLOluciferase target expression vector (Promega, USA) as the XIST wild-type vector. And the mutated miR-185-5p-binding site was constructed as XIST Mutant vector. 293T cells were co-transfected with 200 ng of either pmirGLO-XIST-wide type (WT) or pmirGLO-XIST-mutant (MUT) vector and 80 ng of miR-185-5p mimics or inhibitor and their negative controls with Lipofectamine 3000 (Invitrogen). The verification of combination between miR-185-5p and CCND2 was realized via co-transfection with pmirGLO-CCND2-WT or pmirGLO-CCND2-MUT and miR-185-5p mimics or negative control. And the relative luciferase activity was measured using Dual-luciferase Reporter Assay Kit (Progenia, USA) after 48 h of transfection.

Quantitative real time PCR (qRT-PCR)

The expression of XIST and miR-185-5p was determined using qRT-PCR. Briefly, total RNA was extracted using Trizol reagent (Takara, Otsu, Japan) according to the manufacturer's instructions. And then, 1 µg RNA was

converted into cDNA using Reverse Transcription Kit (Takara). SYBR Premix Ex Taq (Takara) was applied for qRT-PCR assays. The PCR reactions were conducted in an ABI 7500 Fast RealTime PCR System (Life Technologies).

Primers used in PCR were listed below: XIST: sense: 5'-AGCTCCTCGGACAGCTGTAA-3', anti-sense: 5'-CTCCAGATAGCTGGCAACC-3'; miR-185-5p: sense: 5'-TGGAGAGAAAGGCAGTTCCTGA-3', anti-sense: 5'-GCTTCGGCAGCACATATACTAAAAT-3'; U6: sense: 5'-CTCGCTTCGGCAGCACACA-3', anti-sense: 5'-AGATAGGATTACTACAC-3' GAPDH: sense: 5'-CAAGGTCATCCA TGACAACCTTTG-3', anti-sense: 5'-GTCCACCACCCT GTTGCTGTAG-3'. Relative RNA levels were calculated by the $2^{-\Delta\Delta Cq}$ method. Relative expression was normalized to GAPDH and U6, respectively.

Western blot analysis

Briefly, total protein extracted from the cells was loaded on 10 % SDS-PAGE and transferred to PVDF membranes (Millipore, USA). The membranes were probed with the specific primary antibodies (All purchased from Abcam, USA) as listed: CCND2 (ab207604, 1:1,000), CDC2 (ab32444, 1:1,000), CDC4 (ab12292, 1:500), CDC6 (ab109315, 1:1,000), CCND1 (ab134175, 1:10,000), Caspase-3 (ab13585, 1:500), Bcl-2 (ab32124, 1:1,000), Bax (ab32503, 1:1,000), GAPDH (ab8245, 1:500). After incubation with primary antibodies at 4 °C overnight, membranes were incubated with corresponding secondary horseradish peroxidase-conjugated anti-rabbit IgG antibody (ab6721, 1:5,000) at room temperature for 2 h. The target bands were then scanned using Super Signal West Pico Chemiluminescent Substrate Kit (Pierce, Rockford, IL, USA). GAPDH was served as an internal control.

Statistical analysis

SPSS 18.0 and Graph Pad Prism6.0 software were used for statistical analysis. All data was presented by mean ± Standard deviation (SD). Comparison between two groups was performed using the Student's *t*-test. Comparison among three or more groups was conducted using one-way analysis of variance (ANOVA). The correlation between XIST and miR-185-5p was determined using Spearman's analysis. It was considered to be statistically significant when P-value was less than 0.05.

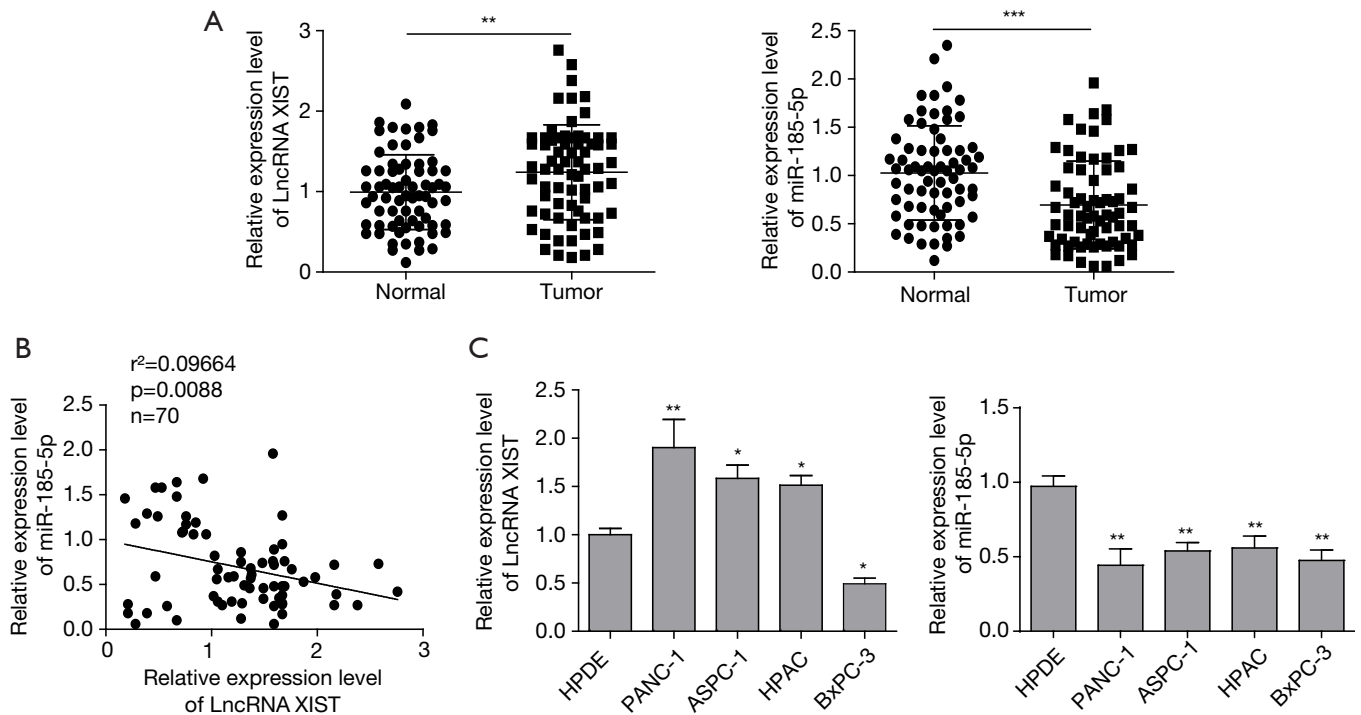


Figure 1 The expression of XIST and miR-185-5p in PC tissues and cell lines. (A) Expression of XIST and miR-185-5p in PC tissues and normal tissues. ** $P < 0.01$ and *** $P < 0.001$ vs. normal. (B) Correlation between XIST and miR-185-5p in PC tissues by Spearman's analysis. (C) Expression of XIST and miR-185-5p in four different PC cell lines and normal pancreatic cell line HPDE cells. * $P < 0.05$ and ** $P < 0.01$ vs. HPDE. XIST, X inactive-specific transcript; PC, pancreatic cancer.

Results

Negative correlation between lncRNA XIST and miR-185-5p in PC tissues and cell lines

To determine the expression of XIST and miR-185-5p both in PC and normal tissues, firstly, we collected PC tissues and paired normal tissues from 70 patients and then the expression of XIST and miR-185-5p was measured using qRT-PCR. The results showed that lncRNA XIST was up-regulated in PC tissues, while miR-185-5p was down-regulated, compared with adjacent normal tissues (Figure 1A). Spearman's analysis also displayed negative correlation between XIST and miR-185-5p (Figure 1B). Meanwhile, four PC cell lines PANC-1, ASPC-1, HPAC, BxPC-3 cells and normal pancreatic cell line HPDE cells were applied for further confirmation. Similarly, as shown in Figure 1C, increasing expression of XIST and decreasing of miR-185-5p were also displayed in PC cell lines. Among all PC cells, PANC-1 cells with the highest expression of XIST and BxPC-3 cells with the lowest expression of XIST

were selected for further experiments.

Effects of XIST on cell proliferation, cell cycle and apoptosis in PC cells

To further explore whether XIST acts as an oncogene in PC, we established PANC-1 cells in which XIST was stably knocked down with sh-XIST transfection and BxPC cells in which XIST was stably overexpressed with pcDNA3.1-XIST transfection (Figure 2A), indicating the successful establishment of lncRNA XIST knockdown or overexpression cell model. Then, we observed that cell cycle was arrested in G0/G1 phase with knocking down XIST using flow cytometry assay. Whereas, cell cycle of BxPC-3 cells transfected with pcDNA3.1-XIST suggested a opposite result (Figure 2B). In parallel, colony formation assay also indicated that knockdown of XIST significantly suppressed colony formation of PANC-1 cells, while overexpression of XIST promoted cell proliferation of BxPC-3 cells (Figure 2C). Moreover, flow cytometry

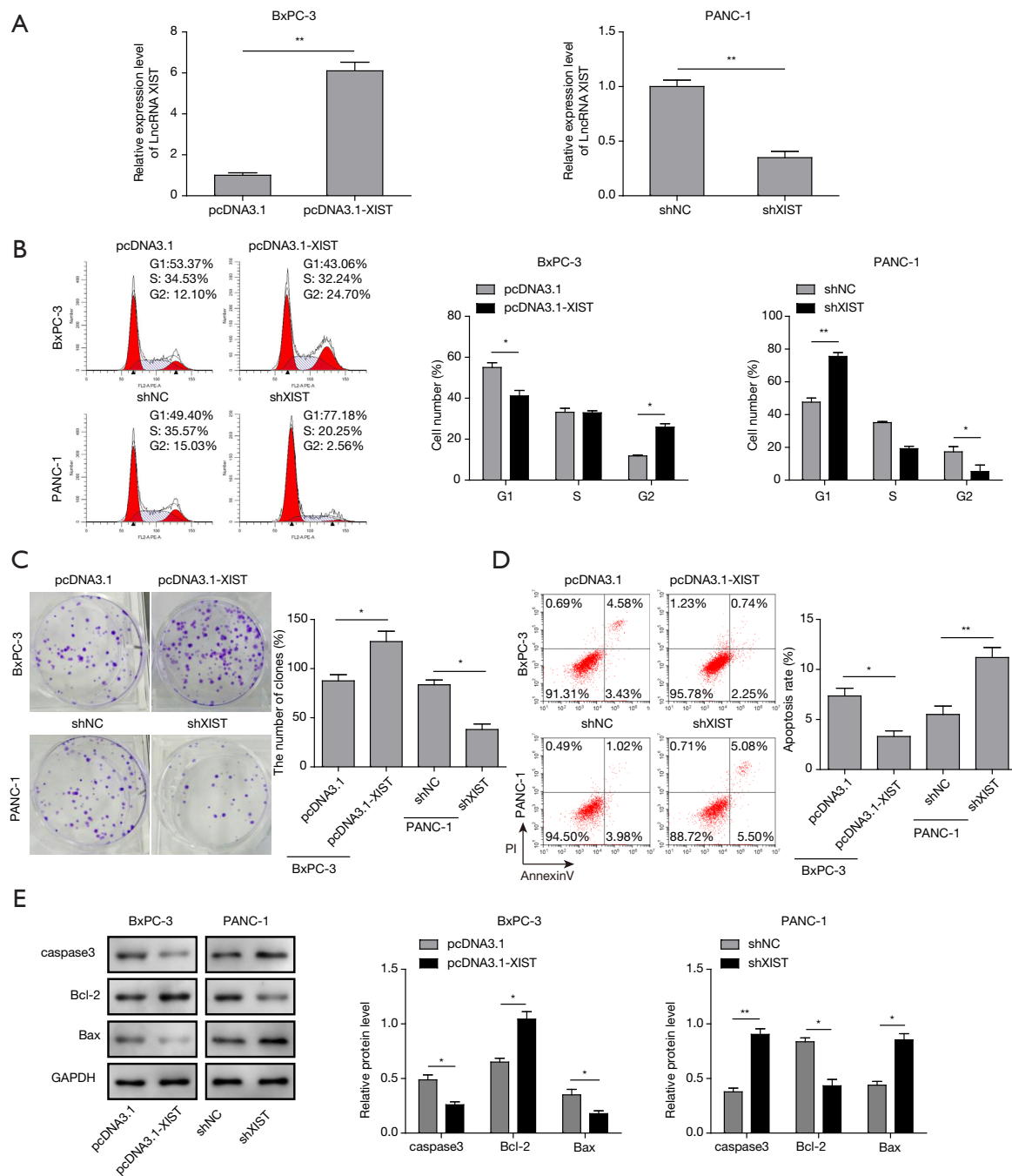


Figure 2 Effects of XIST on cells proliferation and apoptosis in BxPC-3 and PANC-1. (A) Expression of XIST in BxPC-3 cells transfected with pcDNA3.1-XIST or NC (pcDNA3.1) and in PANC-1 cells transfected with sh-XIST or shNC. ****P<0.01 vs. pcDNA3.1 and **P<0.01 vs. shNC.** (B) Cell cycle analysis of BxPC-3 cells transfected with pcDNA3.1-XIST or NC (pcDNA3.1) and in PANC-1 cells transfected with sh-XIST or shNC by the flow cytometry assay. ***P<0.05 vs. pcDNA3.1, *P<0.05 and **P<0.01 vs. shNC.** (C) Cell proliferation of BxPC-3 cells transfected with pcDNA3.1-XIST or NC (pcDNA3.1) and PANC-1 cells transfected with sh-XIST or shNC by colony formation assay. ***P<0.05 vs. pcDNA3.1 and *P<0.05 vs. shNC.** (D) Cell apoptosis of BxPC-3 cells transfected with pcDNA3.1-XIST or NC (pcDNA3.1) and PANC-1 cells transfected with sh-XIST or shNC by the flow cytometry assay. ***P<0.05 vs. pcDNA3.1 and **P<0.01 vs. shNC.** (E) Western blot analysis was performed to measure the expression of Caspase-3, Bcl-2 and Bax. ***P<0.05 vs. pcDNA3.1, *P<0.05 and **P<0.01 vs. shNC.** XIST, X inactive-specific transcript.

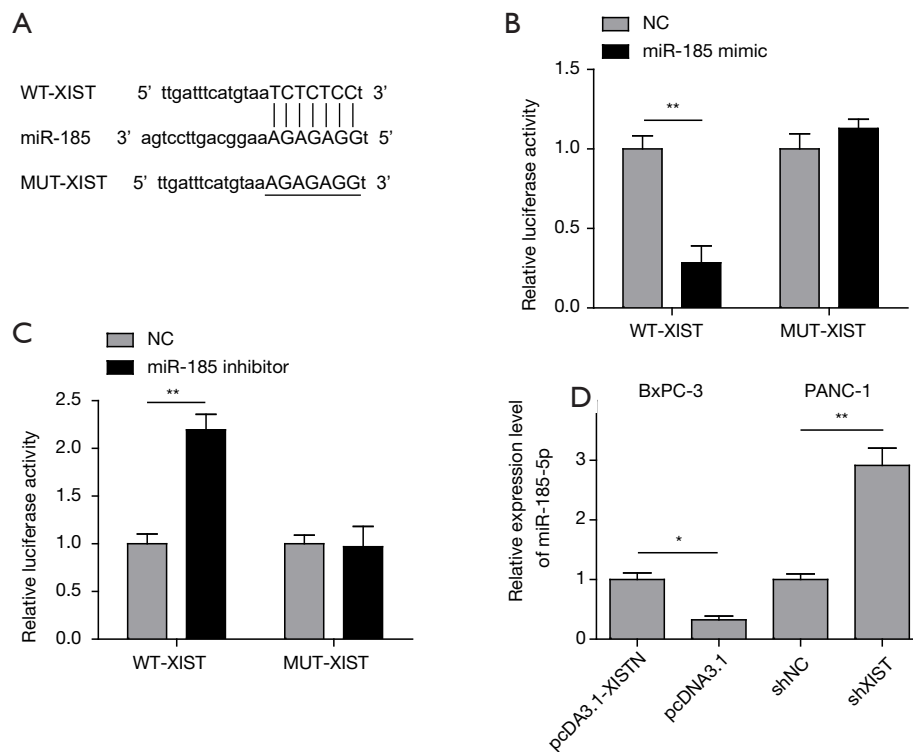


Figure 3 miR-185-5p is a target of XIST. (A) Predicted binding site of XIST and miR-185-5p. (B) and (C) Luciferase activity of WT-XIST and MUT-XIST transfected with miR-185-5p mimics, inhibitor or their negative controls (NC). ** $P < 0.01$. (D) Expression of miR-185-5p in BxPC-3 cells transfected with pcDNA3.1-XIST or NC (pcDNA3.1) and in PANC-1 cells transfected with sh-XIST or shNC. * $P < 0.05$ vs. pcDNA3.1 and ** $P < 0.01$ vs. shNC. XIST, X inactive-specific transcript.

assay showed a significant increase of cell apoptosis after knockdown of XIST in PANC-1 cells, while cell apoptosis showed a reduction with overexpression of XIST in BxPC-3 cells (Figure 2D). In addition, western blot analysis also indicated that overexpression of XIST induced the up-regulation of Bcl-2 and down-regulation of Caspase-3, Bax, while XIST knockdown showed the opposite tendency (Figure 2E). Taken together, these data indicated that lncRNA XIST might function as an oncogene to enhance the tumorigenesis of PC.

miR-185-5p is a direct target of XIST

Recently, accumulating evidence has proved that lncRNAs may participate in the ceRNA regulatory network. Bioinformatic analysis suggested that there is the complementary base pairing between XIST and miR-185-5p (Figure 3A). To confirm the direct binding relationship between XIST and miR-185-5p, a luciferase reporter assay was performed. We found that miR-185-5p mimic markedly

reduced the luciferase activities of pmirGLO-XIST-WT, while there was no obvious difference in cells transfected with miR-185-5p mimic and pmirGLO-XIST-MUT (Figure 3B). Consistently, the luciferase activities in cells transfected with miR-185-5p inhibitor and pmirGLO-XIST-WT or pmirGLO-XIST-MUT showed the same result (Figure 3C). Additionally, an inverse correlation of XIST and miR-185-5p in PC cells was also revealed. As shown in Figure 3D, up-regulation of XIST significantly suppressed miR-185-5p expression in BxPC-3 cells, while down-regulation of XIST markedly increased miR-185-5p expression. These findings implied that XIST directly targets and negatively regulates miR-185-5p.

Effects of miR-185-5p on cell proliferation, cell cycle and apoptosis in PC cells

To further investigate the potential role of miR-185-5p in PC cells, miR-185-5p was overexpressed or knocked down in both BxPC-3 and PANC-1 cells. As shown in Figure 4A,

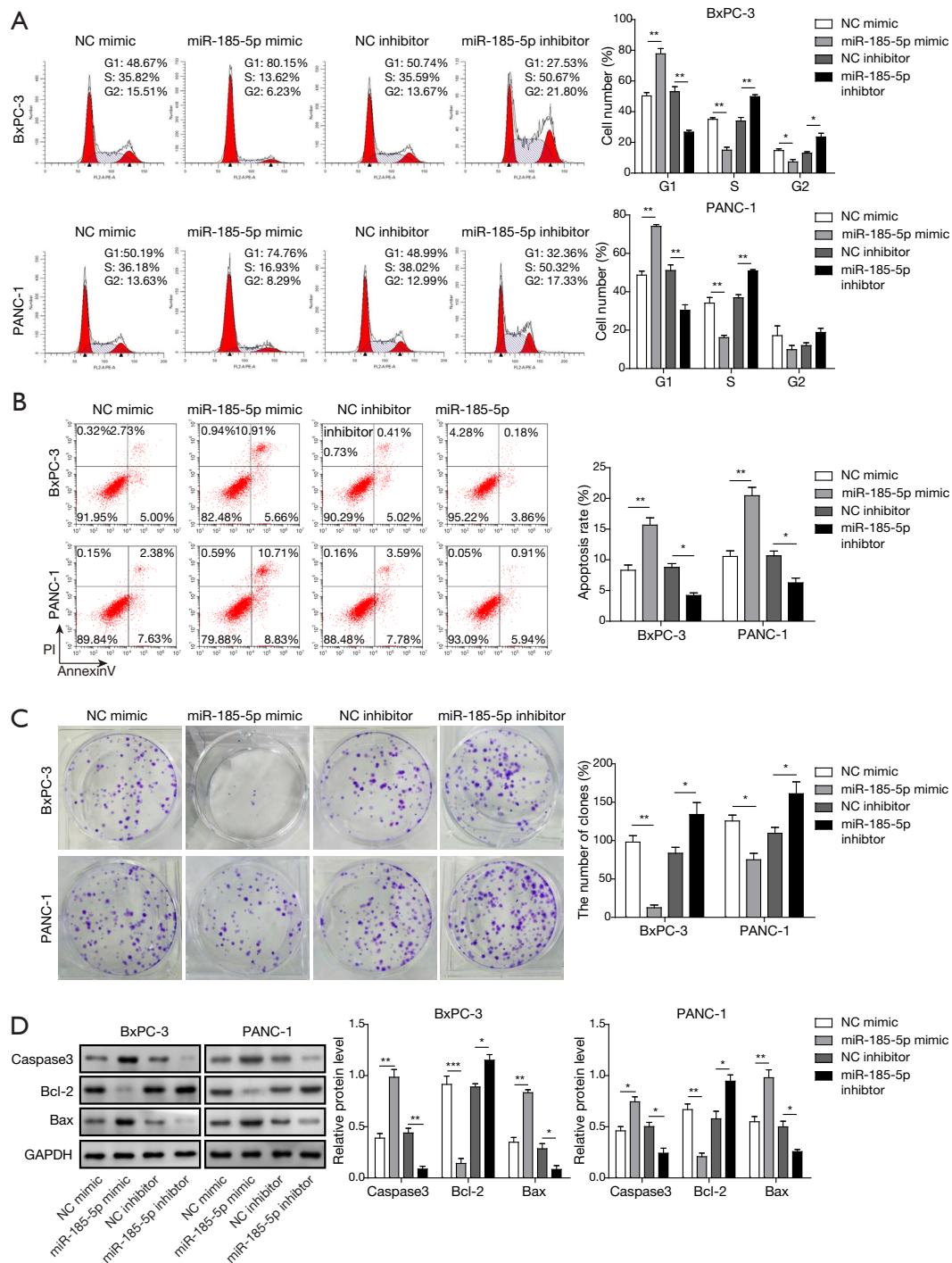


Figure 4 Effects of miR-185-5p on cell proliferation and apoptosis in BxPC-3 and PANC-1. (A) Cell cycle analysis of BxPC-3 and PANC-1 cells transfected with miR-185-5p mimics, inhibitor or NC by flow cytometry. * $P < 0.05$ and ** $P < 0.01$ vs. NC mimic, * $P < 0.05$ and ** $P < 0.01$ vs. NC inhibitor. (B) Cell proliferation of BxPC-3 cells and PANC-1 cells transfected with miR-185-5p mimics, inhibitor or their negative controls (NC) by colony formation assay. * $P < 0.05$ and ** $P < 0.01$ vs. NC mimic and * $P < 0.05$ vs. NC inhibitor. (C) Cell apoptosis of BxPC-3 cells and PANC-1 cells transfected with miR-185-5p mimics, inhibitor and their negative controls (NC) by the flow cytometry. ** $P < 0.01$ vs. NC mimic and * $P < 0.05$ vs. NC inhibitor. (D) Western blot analysis was performed to measure the expression of Caspase-3, Bcl-2 and Bax. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ vs. NC mimic. * $P < 0.05$ and ** $P < 0.01$ vs. NC inhibitor.

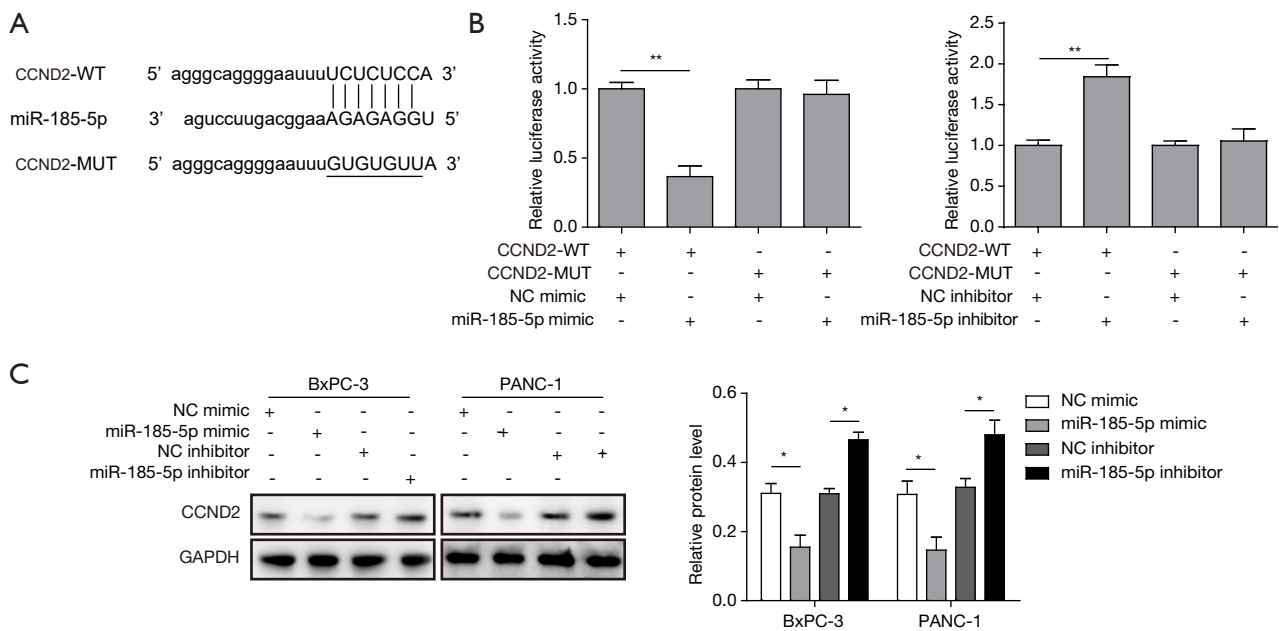


Figure 5 CCND2 is a direct target of miR185-5p. (A) Predicted binding sequence of miR-185-5p and CCND2. (B) Luciferase activity of WT-CCND2 and MUT-CCND2 transfected with miR-185-5p mimics, inhibitor or NC. ** $P < 0.01$. (C) Expression of CCND2 in BxPC-3 and PANC-1 cells transfected with miR-185-5p mimics, inhibitor or their negative controls (NC). * $P < 0.05$ vs. NC mimic and * $P < 0.05$ vs. NC inhibitor.

in both BxPC-3 and PANC-1 cells, when miR-185-5p was overexpressed, cells in G0/G1 were significantly increased and cells in G2/M and S were significantly decreased, while cells transfected with miR-185-5p inhibitor showed opposite results. In addition, high expression of miR-185-5p suppressed cell proliferation and induced apoptosis using the colony formation and flow cytometry assays, respectively. On the contrary, inhibition of miR-185-5p presented the opposite results (Figure 4B,C). Besides, the expression of Bcl-2 was significantly increased within miR-185-5p inhibitor, but Caspase-3 and Bax expression were repressed. Similarly, miR-185-5p mimic displayed the same trend (Figure 4D). All these results indicated miR-185-5p acted as a tumor-suppressive mediator in the development of PC.

CCND2 is a direct target of miR-185-5p

To study the role of miR-185-5p in the mechanisms of PC, bioinformatics analysis was performed to predict the target of miR-185-5p and found that CCND2 may be related to miR-185-5p (Figure 5A). And then the luciferase reporter assay was subjected to validate the effect of miR-185-5p on CCND2 expression. The results showed the luciferase

activity was significantly decreased when cells were co-transfected with miR-185-5p mimic and CCND2 wild-type vector, but not CCND2 mutant vector, while the luciferase activity was increased within miR-185-5p inhibitor co-transfected with CCND2 wild-type vector (Figure 5B). In addition, western blot analysis was performed to measure the expression of CCND2. And the results showed that transfection with miR-185-5p mimic decreased CCND2 protein expression, while transfection with miR-185-5p inhibitor increased CCND2 expression level (Figure 5C).

lncRNA XIST acts as a ceRNA sponging miR-185-5p to regulate CCND2 modulating cell proliferation and apoptosis

In order to explore whether XIST exerted its oncogenic role through miR-185-5p/CCND2 axis in PC, cells were transfected with pcDNA3.1-XIST and miR-185-5p mimic or sh-XIST and miR-185-5p inhibitor in combination. Results showed that in BxPC-3 cells, when transfected with pcDNA3.1-XIST, cell proliferation was significantly increased, while this effect was blocked with miR-185-5p mimic (Figure 6A). On the contrary, in PANC-1 cells transfected with sh-XIST, cell proliferation was significantly

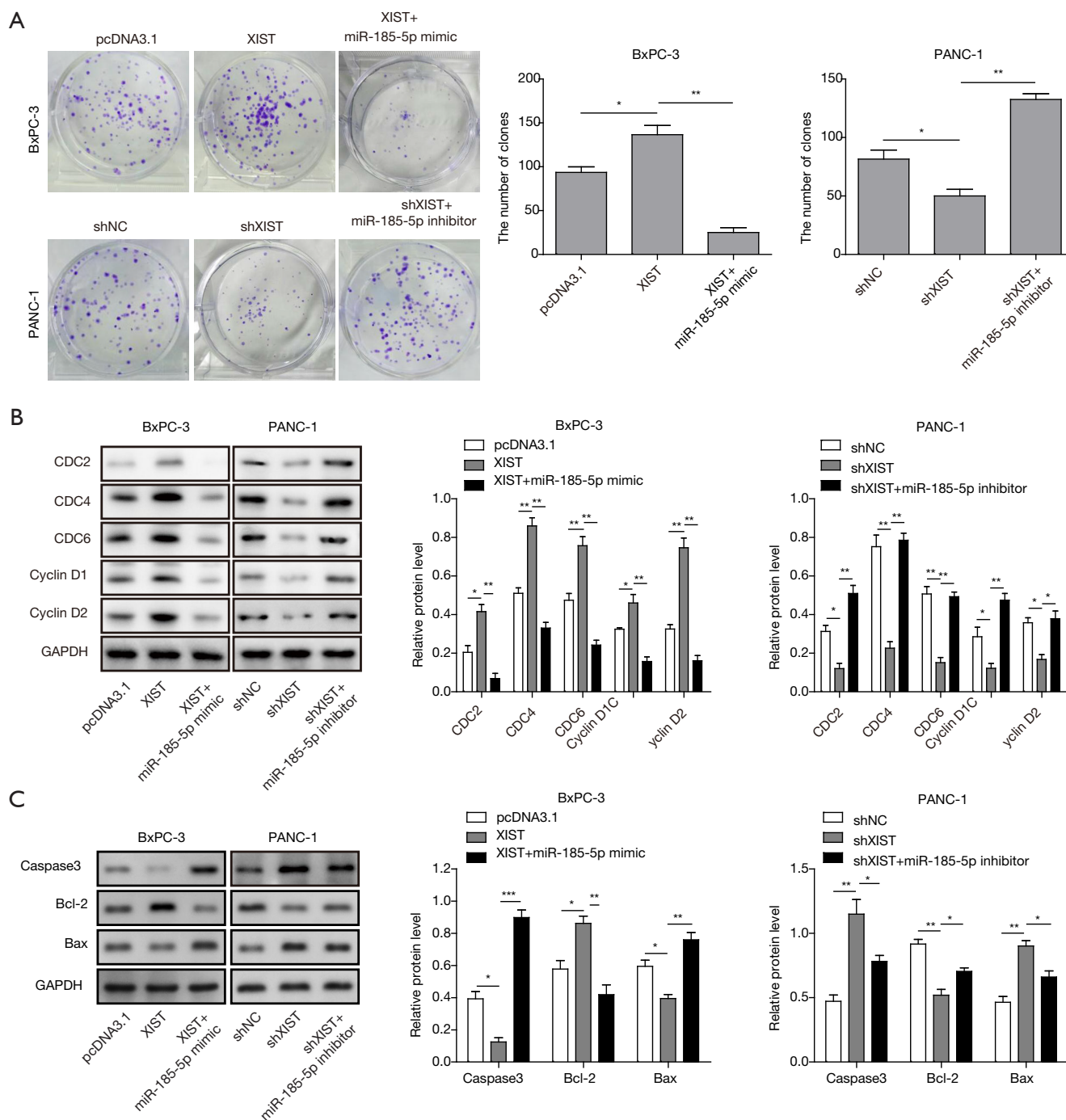


Figure 6 XIST acts as a ceRNA to regulate CCND2 via sponging miR-185-5p. (A) Cell proliferation of BxPC-3 cells transfected with pcDNA3.1, pcDNA3.1-XIST, pcDNA3.1-XIST and miR-181-5p mimics, and PANC-1 cells transfected shNC, sh-XIST, sh-XIST and miR-181-5p inhibitor by colony formation assay. * $P < 0.05$ vs. pcDNA3.1, * $P < 0.05$ vs. shNC, ** $P < 0.01$ vs. pcDNA3.1-XIST and ** $P < 0.01$ vs. sh-XIST. (B) Expression of CCND2 and cell cycle related proteins CDC2, CDC4, CDC6, CCND1 in BxPC-3 and PANC-1 cells. * $P < 0.05$ and ** $P < 0.01$ vs. pcDNA3.1, * $P < 0.05$ and ** $P < 0.01$ vs. shNC, ** $P < 0.01$ vs. pcDNA3.1-XIST, * $P < 0.05$ and ** $P < 0.01$ vs. sh-XIST. (C) Proteins expression of Caspase-3, Bcl-2 and Bax was determined using western blot analysis. * $P < 0.05$ vs. pcDNA3.1 and ** $P < 0.01$ vs. shNC. ** $P < 0.01$ and *** $P < 0.001$ vs. pcDNA3.1-XIST. * $P < 0.05$ vs. shNC. XIST, X inactive-specific transcript.

inhibited but the effect was significantly recovered with miR-185-5p inhibitor (Figure 6A). Besides, expression levels of CCND2 and cell cycle-related proteins CDC2, CDC4, CDC6, CCND1 were significantly up-regulated when XIST was overexpression in BxPC-3 cells and were markedly down-regulated when XIST was knocked down in PANC-1 cells. And these effects were significantly reversed by overexpression or inhibition of miR-185-5p, respectively (Figure 6B). Meanwhile, overexpression of XIST, decreased the expression of Caspase-3 and Bax, but induced Bcl-2 up-regulation, and when XIST was knocked down, the alteration trends were opposite. Altogether, all these effects were reversed by additional transfection of miR-185-5p mimic or inhibitor (Figure 6C). These results suggested that miR-185-5p could be involved in XIST-mediated biological functions of PC, indicating that XIST could modulate cell proliferation and apoptosis through miR-185-5p/CCND2 axis in the development of PC.

Discussion

Despite the development of treatment and diagnosis, the prognosis of PC is still very poor and thus deeper understanding for molecular mechanisms of PC development is urgent (19). Currently, the dysregulation of non-coding RNAs have been demonstrated in various cancers, including PC (20,21). For example, Huang *et al.* showed that up-regulation of lncRNA PVT1 in PC patients was closely associated with the patients' poor prognosis (22). Gu *et al.* also found lncRNA MEG3 exerted its anti-cancer effects contributing to the suppression on PC development (23). It was also observed lncRNA HOTTIP could modulate cancer stem cell properties by regulating HOXA9 in PC (24). Additionally, lncRNA XIST was also reported to be involved in the progression of PC. A recent study demonstrated that XIST could regulate proliferation of PC cells through regulation of miR-133a/EGFR (13). However deeper insights for how XIST influence PC process are still unknown.

In the present study, we also found XIST was up-regulated in PC cells. Moreover, we found that XIST contributed to cell proliferation and inhibited cell apoptosis of PC cells, and the effects might be through inhibiting miR-185-5p. Roles for XIST in PC and other cancers as an oncogene have been reported in several researches. It has been proved that up-regulation of XIST was associated with the development of PC (12). Sun *et al.* showed XIST exerted oncogenic functions in PC by miR-34a-5p (18). Yao

et al. indicated that the inhibition of XIST showed tumor-suppressive functions in human glioblastoma stem cells by up-regulating miR-152 (25). On the contrary, several studies also revealed the anti-tumor function of XIST in the cancer development. It was found that loss of XIST could promote brain metastasis in breast cancer (26). Huang *et al.* also found knockdown of XIST resulted in enhanced cell viability (27). Thus, these findings suggested that XIST might demonstrate different roles in different cancers.

miR-185-5p has been proven as a tumor suppressor in several cancers. It was found in prostatic cancer tissue and cell lines, miR-185-5p was down-regulated (28). And miR-185-5p could also inhibit the epithelial mesenchymal transition of breast cancer cells (29). LncRNAs such as H19 and PDIA3P were also reported to regulate miR-185-5p in cancer development (30,31). Moreover, a recent study demonstrated XIST could promote progression of gastric cancer by targeting miR-185 (32). In the present study, we verified that miR-185-5p was also a downstream target of XIST in PC. Meanwhile, miR-185-5p was closely associated with XIST-mediated biological functions in PC cells. Our results showed that XIST exerts its oncogenic functions on cell proliferation, cell cycle and apoptosis, thus contributing to PC progression, while these effects were significantly reversed by miR-185-5p. Our finding indicated that miR-185-5p might be a tumor suppressor in the development of PC.

Role of CCND2 in cancer development has been noticed in many studies. Evron *et al.* demonstrated CCND2 expression was down-regulated in the majority of breast cancers (33). Zhang *et al.* showed miR-206 could inhibit gastric cancer proliferation by repressing CCND2 (34). Besides, several studies also showed the relationship of miR-185-5p and CCND2. Sun *et al.* found lncRNA PDIA3P could interact with miR-185-5p by targeting CCND2 in oral squamous cell carcinoma (35). Bibaki *et al.* showed miR-185 could target on CCND2 in lung cancer (36). In the present study, we also found miR-185-5p could inhibit proliferation and migration of PC cells by down-regulation of CCND2. Besides, we confirmed that CCND2 was a downstream target of miR-185-5p, which could directly target CCND2 in PC cells, and the effects were regulated by XIST. The present study also has some limitations. First CCND2 is not the only downstream target of XIST in cancer development, thus the XIST/miR-185-5p/CCND2 axis is also one of the regulation ways for PC. Other signaling pathways and deeper insights are still needed.

In conclusion, our study provided the first evidence that XIST could promote cell proliferation, inhibit cell apoptosis

through modulating miR-185-5p/CCND2 axis. This study might give deeper understanding for role of XIST and miR-185-5p in development of PC, and might provide some new targets for PC treatment. However, our study is limited *in vitro* experiments and the certain molecular mechanisms of XIST in PC need to be further verified *in vivo*.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tcr.2020.01.26>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). All patients were informed to write consent and all the experiments involved in this study were approved by the Ethics Committee of the Second Xiangya Hospital of Central South University (No. 201103301).

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