



Draft Genome Sequences for Five *Photorhabdus* Bacterial Symbionts of Entomopathogenic *Heterorhabditis* Nematodes Isolated from India

 **Vishal Singh Somvanshi,^a Bhumika Dubay,^a Jyoti Kushwah,^a Sivakumar Ramamoorthy,^b Udayakumar S. Vishnu,^b Jagadesan Sankarasubramanian,^b Jeyaprakash Rajendhran,^b Uma Rao^a**

^aDivision of Nematology, ICAR-Indian Agricultural Research Institute, New Delhi, India

^bDepartment of Genetics, School of Biological Sciences, Madurai Kamaraj University, Madurai, Tamil Nadu, India

ABSTRACT *Photorhabdus* bacteria exhibit contrasting lifestyles; they are virulent insect pathogens but symbionts of the entomopathogenic *Heterorhabditis* nematodes. *Photorhabdus* genomes encode several secondary metabolites and insecticidal protein toxins. Here, we present the draft genome sequences for five *Photorhabdus* strains isolated from *Heterorhabditis* nematodes collected from various geographical regions of India.

P*hotorhabdus* spp. are Gram-negative gammaproteobacteria found in nature in association with the entomopathogenic nematodes of the genus *Heterorhabditis* (1, 2). The first *Photorhabdus* genome was sequenced in 2003 (3), and at present, 33 genome sequences of various *Photorhabdus* spp. are available in NCBI GenBank (<https://www.ncbi.nlm.nih.gov/genome?term=Photorhabdus>). The genus *Photorhabdus* was revised to include 15 species on the basis of whole-genome, biochemical, chemotaxonomic, and ribosomal protein fingerprinting information, i.e., *P. bodei*, *P. australis*, *P. akhurstii*, *P. caribbeanensis*, *P. hainanensis*, *P. kayaii*, *P. kleinii*, *P. namnaonensis*, *P. noenieputensis*, *P. laumontii*, *P. cinerea*, *P. khanii*, *P. stackebrandtii*, *P. tasmaniensis*, and *P. thracensis* (4). We previously isolated the symbiont bacteria from the infective juveniles (IJs) of the entomopathogenic nematodes isolated from various geographical locations in India (5, 6). Preliminary biochemical and virulence characterization suggested genetic variations between different isolates (6). The 16S rRNA gene marker identified them to be a member of erstwhile *Photorhabdus luminescens* species (Table 1). To ascertain the identity of these isolates and to investigate the reasons for the differences in biochemical characters and virulence (6), we sequenced the genomes of these isolates.

A single colony of each strain was inoculated in 5 ml of Luria-Bertani (LB) broth and grown at 28°C with agitation (200 rpm) for 12 h. The genomic DNA was isolated by using a DNeasy kit (Qiagen, Hilden, Germany). For sequencing library preparation, 100 ng of genomic DNA was sheared enzymatically for 3 to 4 min using an Ion Shear Plus kit. The sheared DNA was purified using AMPure beads (Beckman Coulter Life Sciences, Indianapolis, IN, USA) and ligated with barcoded adapters. Subsequently, the adaptor-ligated fragments were resolved on a 2% E-Gel (Thermo Fisher Scientific, Waltham, MA, USA), and ~330-bp fragments were collected. The size-selected fragments were PCR amplified using adaptor-specific primers for five cycles using high-fidelity Platinum supermix provided in the Ion Plus fragment library kit (Thermo Fisher Scientific, Waltham, MA, USA). The amplified product was purified using AMPure beads, and this final library was used for template generation for sequencing. The whole-genome sequencing was performed by the Semiconductor sequencing using the Ion Torrent Personal Genome Machine (PGM) system.

Citation Somvanshi VS, Dubay B, Kushwah J, Ramamoorthy S, Vishnu US, Sankarasubramanian J, Rajendhran J, Rao U. 2019. Draft genome sequences for five *Photorhabdus* bacterial symbionts of entomopathogenic *Heterorhabditis* nematodes isolated from India. *Microbiol Resour Announc* 8:e01404-18. <https://doi.org/10.1128/MRA.01404-18>.

Editor Julia A. Maresca, University of Delaware

Copyright © 2019 Somvanshi et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Vishal Singh Somvanshi, vssomvanshi@iari.res.in, or Uma Rao, umarao@iari.res.in.

Received 10 October 2018

Accepted 17 December 2018

Published 24 January 2019

TABLE 1 Information and genome statistics for the sequenced *Photorhabdus* bacterial strains and comparison to the already sequenced and annotated reference genome of *Photorhabdus luminescens* subsp. *laumondii* TTO1^a

Feature ^b	Data for strain:			
	<i>P. akhurstii</i> IARI-SGMG3	<i>P. akhurstii</i> IARI-SGHRS2	<i>P. akhurstii</i> IARI-SGHRA4	<i>P. akhurstii</i> IARI-SGMS1
Place of origin	Meghalaya (northeastern Himalayan region), India	Haryana (Trans-Gangetic Plains), India	Haryana (Trans-Gangetic Plains), India	Maharashtra (western plateau and hill region), India
Nematode host	<i>Heterorhabditis</i> sp.	<i>Heterorhabditis indica</i>	<i>Heterorhabditis indica</i>	<i>Heterorhabditis indica</i>
16S rRNA gene accession no.	JX221722	KJ995730	JX221723	JX240394
No. of reads	2,583,080	1,391,368	1,415,756	1,195,633
Total data generated (Mb)	540	288	297	255
Insert size (bp)	72	72	72	72
Genome size (bp)	5,663,704	5,514,710	5,414,651	5,395,311
Coverage (×)	96	51	53	45
No. of contigs	228	220	212	342
GC content (%)	42.9	42.7	42.5	42.6
No. of CDS	5,036	5,055	4,955	4,942
No. of RNAs	128	78	75	73
<i>N</i> ₅₀ (bp)	92,101	82,937	95,541	43,831
Predicted no. of genes	5,016	5,083	4,953	5,040
No. (%) of annotated genes	4,068 (81.1)	4,207 (82.7)	4,083 (82.4)	4,101 (81.4)
No. (%) of genes matched to reference genome	4,188 (83.5)	4,246 (83.5)	4,170 (84.2)	4,246 (84.2)
No. of genes annotated but absent in reference genome	332	333	386	368
No. of genes present in reference genome but not annotated	516	536	499	541
SRA accession no.	SRX3720927	SRX3720926	SRX3720929	SRX3720925
WGS GenBank accession no.	PUWT000000000	PUWU000000000	PUWW000000000	PUWX000000000

^aFor all strains, Semiconductor Sequencing using the Ion Torrent Personal Genome Machine was used, with 200-bp chemistry for library preparation, and MIRA version 4.0.2 with *de novo* assembly.

^bCDS, coding sequences; WGS, whole-genome shotgun.

The good-quality reads were exported using the FileExporter plugin in the Ion Torrent Personal Genome Machine-associated Torrent Suite software, using the default parameters. The *de novo* genome assembly, scaffold construction, and gap closure were done by using MIRA version 4.0.2 (7), with the default parameters and providing the technology as "Iontor." The fold coverage was estimated using the total number of sequence reads divided by the estimated genome size. Gene prediction was made *ab initio* using GeneMarkS (8) with default parameters. These predicted genes were further mapped to the reference genome (*P. luminescens* subsp. *laumondii* TTO1, NCBI RefSeq accession number [NC_005126](#)) using Blast2GO (9), with an E value cutoff of 1.0E-3. The amino acid sequences of the predicted genes were matched with the nonredundant protein database and annotated with InterProScan using Blast2GO.

The sequencing generated 1.2 to 2.5 million reads, generating 255 to 540 Mb of total sequence data (Table 1). The *de novo* assembly resulted in final genome sizes of 5.3 to 5.6 Mb, with a coverage of 45 to 96× (Table 1). A total of 190 to 228 contigs were obtained, with an N_{50} value of 43 to 103 kb. The GC content of the *Photorhabdus* genomes was 42.5 to 42.7%. We predicted 4,953 to 5,933 genes in the sequenced *Photorhabdus* strains, of which 78.6 to 82.7% could be annotated. These genes showed 83.5 to 85.8% match to the reference *P. luminescens* subsp. *laumondii* TTO1 genome (Table 1).

Data availability. The data generated in this study can be accessed at NCBI under SRA study accession number [SRP133050](#), BioProject number [PRJNA434554](#), and SRA experiment accession numbers [SRX3720925](#) to [SRX3720929](#). The draft genome accession numbers are provided in Table 1.

ACKNOWLEDGMENTS

This work was supported by funding from the Science and Engineering Research Board, Department of Science and Technology, Government of India (grant SB/SO/AS/010/2014 to V.S.S.), and in-house funding from the Division of Nematology, ICAR-Indian Agricultural Research Institute (New Delhi, India).

REFERENCES

- Boemare N, Akhurst R, Mourant R. 1993. DNA relatedness between *Xenorhabdus* spp. (Enterobacteriaceae), symbiotic bacteria of entomopathogenic nematodes, and a proposal to transfer *Xenorhabdus luminescens* to a new genus, *Photorhabdus* gen. nov. Int J Syst Evol Microbiol 43: 249–255. <https://doi.org/10.1099/00207713-43-2-249>.
- Waterfield NR, Ciche T, Clarke D. 2009. *Photorhabdus* and a host of hosts. Annu Rev Microbiol 63:557–574. <https://doi.org/10.1146/annurev.micro.091208.073507>.
- Duchaud E, Rusniok C, Frangeul L, Buchrieser C, Givaudan A, Taourit S, Bocs S, Boursaux-Eude C, Chandler M, Charles JF, Dassa E, Derose R, Derzelle S, Freyssinet G, Gaudriault S, Médigue C, Lanois A, Powell K, Siguier P, Vincent R, Wingate V, Zouine M, Glaser P, Boemare N, Danchin A, Kunst F. 2003. The genome sequence of the entomopathogenic bacterium *Photorhabdus luminescens*. Nat Biotechnol 21:1307–1313. <https://doi.org/10.1038/nbt886>.
- Machado RAR, Wuthrich D, Kuhnert P, Arce CCM, Thonen L, Ruiz C, Zhang X, Robert CAM, Karimi J, Kamali S, Ma J, Bruggmann R, Erb M. 2018. Whole-genome-based revisit of *Photorhabdus* phylogeny: proposal for the elevation of most *Photorhabdus* subspecies to the species level and description of one novel species *Photorhabdus bodei* sp. nov., and one novel subspecies *Photorhabdus laumondii* subsp. *clarkei* subsp. nov. Int J Syst Evol Microbiol 68:2664–2681. <https://doi.org/10.1099/ijsem.0.002820>.
- Kumar P, Ganguly S, Somvanshi VS. 2015. Identification of virulent entomopathogenic nematode isolates from a countrywide survey in India. Int J Pest Manag 61:135–143. <https://doi.org/10.1080/09670874.2015.1023869>.
- Kushwah J, Kumar P, Garg V, Somvanshi VS. 2017. Discovery of a highly virulent strain of *Photorhabdus luminescens* ssp. *akhurstii* from Meghalaya, India. Indian J Microbiol 57:125–128. <https://doi.org/10.1007/s12088-016-0628-y>.
- Chevreux B, Pfisterer T, Drescher B, Driesel AJ, Müller WE, Wetter T, Suhai S. 2004. Using the miraEST assembler for reliable and automated mRNA transcript assembly and SNP detection in sequenced ESTs. Genome Res 14:1147–1159. <https://doi.org/10.1101/gr.1917404>.
- Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. Nucleic Acids Res 29:2607–2618. <https://doi.org/10.1093/nar/29.12.2607>.
- Conesa A, Götz S. 2008. Blast2GO: a comprehensive suite for functional analysis in plant genomics. Int J Plant Genomics 2008:619832. <https://doi.org/10.1155/2008/619832>.