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# Performance Verification of CYP2C19 Enzyme Abundance Polymorphism Settings within the Simcyp Simulator v21

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Abstract: Physiologically based pharmacokinetic (PBPK) modeling has a number of applications, including assessing drug-drug interactions (DDIs) in polymorphic populations, and should be iteratively refined as science progresses. The Simcyp Simulator is annually updated and version 21 included updates to hepatic and intestinal CYP2C19 enzyme abundance, including addition of intermediate and rapid metabolizer phenotypes and changes to the ultra-rapid metabolizer enzyme abundance, with implications for population clearance and DDI predictions. This work details verification of the updates with sensitive CYP2C19 substrates, omeprazole and lansoprazole, using available clinical data from literature. Multiple assessments were performed, including recovery of areas under the concentration-time curve (AUC) and  $C_{max}$  from compiled datasets for each drug, recovery of victim DDI ratios with CYP2C19 and/or CYP3A4 inhibition and recovery of relative exposure between phenotypes. Simulated data were within respective acceptance criteria for >80% of omeprazole AUC values, >70% of lansoprazole AUC and  $C_{max}$ , >60% of AUC and  $C_{max}$  DDI ratios and >80% of exposure ratios between different phenotypes. Recovery of omeprazole C<sub>max</sub> was lower (>50-70% within 2-fold) and possibly attributed to the variety of formulations used in the clinical dataset. Overall, the results demonstrated that the updated data used to parameterize CYP2C19 phenotypes reasonably described the pharmacokinetics of omeprazole and lansoprazole in genotyped or phenotyped individuals.

Keywords: PBPK; CYP2C19; polymorphism; population; Simcyp Simulator

# 1. Introduction

Evaluation of the drug–drug interaction (DDI) potential of a drug candidate is a vital part of drug development to properly inform posology in situations where multiple medications may be co-administered. DDI assessment can become even more complex when considering drugs whose elimination is governed by polymorphic drug metabolizing enzymes or transporters, where subgroups in the population, as often defined by allelic variation, can demonstrate different magnitudes of DDI [1]. DDIs are typically evaluated with designated clinical studies to explore the perpetrator or victim potential of a drug candidate; however, regulatory authorities have recognized the utility of computational modeling techniques in this process [2,3]. A recent evaluation of FDA submissions revealed that almost 50% of new drug approvals included physiologically based pharmacokinetic (PBPK) modeling with the majority used for DDI assessment [4].

PBPK modeling is an ideal tool in exploring DDI potential in genotyped or phenotyped individuals as information on the frequency and enzyme abundance associated with different genotypes or phenotypes can be incorporated in the physiological systems information of the simulations. Compiling physiological systems information can be a resource-intensive process requiring periodic updates based upon the latest scientific



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**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). knowledge and extensive verification to ensure data quality and model performance [5]. However, once obtained, the information can be used in future modeling efforts.

The cytochrome P450 (CYP) enzyme family includes the main enzymes responsible for catalyzing oxidative biotransformation of numerous exogenous substrates [6]. They are located throughout the body; however, higher abundance is noted in organs associated with drug metabolism, namely the liver and intestine [7]. CYP enzymes are of particular importance in drug development because their inhibition, induction, genetic polymorphism or disease-related changes in abundance can have significant impact on the pharmacokinetics of administered drugs [8].

CYP2C19 is a polymorphic drug-metabolizing enzyme belonging to the CYP 2C enzyme family and is the major enzyme responsible for the biotransformation of proton pump inhibitors, such as omeprazole and lansoprazole [9]. Up to 5 enzyme phenotypes have been identified: poor (PM, \*2/\*2, \*3/\*3 and \*2/\*3), intermediate (IM, \*1/\*2, \*1/\*3, \*2/\*17), normal (NM, \*1/\*1, wild type), rapid (RM, \*1/\*17) and ultra-rapid (UM, \*17/\*17) metabolizers [10]. Individuals with the PM phenotype carry two loss-of-function alleles (\*2 or \*3) resulting in significantly reduced metabolism of CYP2C19 substrates, while those with the UM phenotype carry two increased function alleles (\*17) resulting in increased metabolism compared to the wild type (\*1). The distribution of these phenotypes differs amongst ethnic groups, with Asian populations demonstrating a higher percentage of PMs compared to Caucasians and African Americans [11].

The Simcyp Simulator v21 (Certara UK Ltd., Sheffield, UK) was released in December 2021, and implemented updates to the adult population characteristics for CYP2C19. These changes included updated values for hepatic and intestinal CYP2C19 enzyme abundance, the addition of IM and RM phenotypes, and changes to the existing UM population (Table 1). The purpose of this work was to verify the newly updated CYP2C19 abundance values in healthy adult populations currently implemented in Simcyp v21 software. To facilitate this investigation, proton pump inhibitors, omeprazole and lansoprazole, were selected as probe substrates because they are largely metabolized by CYP2C19 with minor contributions from CYP3A4 [12]. Both drugs are recognized as sensitive clinical index CYP2C19 substrates for use as victim drugs in clinical DDI studies by the FDA [13], making them ideal probe drugs for assessing CYP2C19 enzyme abundance. The verification was performed in a stepwise manner and included recovery of omeprazole and lansoprazole plasma exposure from compiled datasets, recovery of victim DDI ratios with CYP2C19 and/or CYP3A4 inhibition, and recovery of the relative change in exposure between phenotypes from select studies. The information obtained in this verification was used to assess the reliability of the updated CYP2C19 enzyme abundances described within Simcyp Simulator v21 software for use in prospective PBPK modeling involving CYP2C19 polymorphisms.

	Hepatic Abundance (pmol/mg protein) (CV%)							
Simcyp	PM	IM	NM	RM	UM			
V21	0	2.85 (52)	4.4 (52)	7.01 (89)	10.23 (79)			
V20	0	NA	4.4 (71)	NA	8.7 (71)			
	]	Intestinal Abunda	ince (nmol/smal	l intestine) (CV	%)			
Simcyp	PM	IM	NM	RM	UM			
V21	0	1.29 (52)	2 (52)	3.18 (89)	4.65 (79)			
V20	0	NA	2 (77)	NA	4 (77)			

**Table 1.** Updated Hepatic and Intestinal CYP2C19 Enzyme Abundance Implemented in the SimcypSimulator v21 Compared to v20.

NA = not available.

### 2. Materials and Methods

## 2.1. Literature Search

A literature search was conducted using the University of Washington Drug Interaction Database [14] and PubMed to identify CYP2C19 pharmacogenomic studies with omeprazole and lansoprazole, including DDI studies.

Plasma concentration-time data, where available, were digitized using MATLAB R2021a digitize2.m code [15]. Only clinical data from healthy adults were included in the analysis; data from pediatrics, elderly or those with severe gastrointestinal conditions (e.g., peptic ulcers) were excluded.

#### 2.2. Modeling Strategy

Default settings of Sim-NEurCaucasian, Sim-Chinese and Sim-Japanese virtual populations, as well as the SV-Omeprazole and SV-Lansoprazole compound files within the Simcyp Simulator v21 were used in the verification. Enzyme abundance for each CYP2C19 phenotype is listed in Table 1. Noteworthily, most available clinical studies investigating CYP2C19 polymorphism effects on omeprazole and lansoprazole PK were conducted in Asian subjects, who not only have differences in body height and weight but also characteristically have a higher percentage of PMs [11]. Within the Simcyp Simulator v21 the same enzyme abundance data for Japanese, Chinese and Caucasian populations are assigned while maintaining the different phenotype frequencies between these populations, since PBPK model performance in Japanese and Chinese populations was improved by adjusting CYP2C19 abundance to the same level as Caucasians [16].

Details of omeprazole and lansoprazole parameters used for the simulations are presented in Supplementary Table S1. Briefly, both models incorporate first-order oral absorption. Lansoprazole utilized a full PBPK model while omeprazole incorporated a simplified distribution model which includes liver, small intestine, portal vein and a systemic compartment. Elimination for both drugs utilized intrinsic clearances for CYP2C19 and CYP3A4. Omeprazole is known to inhibit its own metabolism via mechanism-based inhibition of CYP2C19 [17]. Relevant parameters have been incorporated into the SV-Omeprazole model and verified to recover multiple dose omeprazole exposure by the software vendor prior to release. CYP2C19 inhibition parameters for omeprazole were assumed to remain the same across CYP2C19 phenotypes such that any CYP2C19 enzyme abundance differences between phenotypes would be reflected in simulated outcomes.

Model performance verification of CYP2C19 enzyme abundance using omeprazole and lansoprazole consisted of 3 steps, namely:

- Recovery of area under the concentration-time curve (AUC) and maximum concentration (C<sub>max</sub>) from compiled datasets
- 2. Recovery of victim DDI ratios with CYP2C19 and/or CYP3A4 inhibition
- 3. Recovery of the relative change in exposure between phenotypes compared to NM, where available, from select studies

#### 2.3. Omeprazole and Lansoprazole PK Prediction in CYP2C19 Phenotypes

Plasma concentration-time data from studies where subjects received a single oral or IV administration were compiled to create a single dataset for each compound per CYP2C19 phenotype [18–54]. All PK parameters and plasma concentration data were normalized to a 20 mg omeprazole dose or a 30 mg lansoprazole dose assuming dose proportionality around these doses. Theoretically, because SV-Omeprazole and SV-Lansoprazole were described with first-order absorption, no transporter involvement in disposition and linear metabolism ( $CL_{int}$ ), the simulated PK and plasma concentration data can be directly compared to the normalized omeprazole or lansoprazole observations. In practice, lansoprazole demonstrated linear pharmacokinetics with single doses from 15 to 60 mg [55]. Omeprazole single dose exposure was generally proportional between 10 and 40 mg [56], which encompassed the range of doses from the investigated studies. Due to the variety of the study populations from the data collated, simulations were performed using the

Sim-NEurCaucasian, Sim-Chinese and Sim-Japanese virtual populations with an age range of 20–50 years and a proportion of 50% female (10 trials with 10 subjects in each trial). Simulated data from each population was compared to the appropriate observed study population, where possible. Due to the absence of specific population data for Korean or Pakistani populations in Simcyp Simulator v21, clinical data was compared to simulated Chinese [57] and Caucasian data, respectively.

Model performance was evaluated by visual inspection of plasma concentration-time curves and recovery of the simulated mean PK parameters (AUC and  $C_{max}$ ) within 2-fold of mean clinical values. Literature references did not specify if means reported were arithmetic or geometric means, therefore arithmetic means were assumed. Because the compiled dataset represented multiple trials and populations, the median and range of the 30 simulated trial means (n = 10 per simulated ethnic group) were plotted against the median and range of the compiled clinical studies to visually assess performance.

#### 2.4. Victim DDI Prediction in CYP2C19 Phenotypes

Simulations of DDI studies where omeprazole and lansoprazole were the victim of CYP2C19 and/or CYP3A4 interactions were performed using the same demographic information as the clinical study (i.e., age, sex), if available, otherwise the same settings as described in the previous section were used. Default compound files within the Simcyp Simulator for perpetrator drugs SV-Clarithromycin, SV-Ticlopidine and SV-Fluvoxamine were used. Trial study designs were simulated 10 times with study-matched subject numbers per phenotype to evaluate the range of results from simulations. Model performance was evaluated by comparing simulated mean AUC and  $C_{max}$  ratios to clinical results using criteria suggested by Guest et al. [58].

#### 2.5. Prediction of CYP2C19 Phenotype Ratios

Simulations were performed as described in Section 2.4. Phenotype ratios were compared to NM, where available. If NM data was not available within a study, IM data was used as the comparator because enzyme abundance was the most similar to NM. Model performance was evaluated as described in Section 2.4 where criteria suggested by Guest et al. [58] was applied.

#### 3. Results

#### 3.1. Literature Search

Results of the literature search for pharmacogenomic data available for CYP2C19 substrates, omeprazole and lansoprazole, are reported in Supplementary Table S2. The literature search identified 26 studies for omeprazole and 11 studies for lansoprazole administered as single oral or intravenous doses.

S-mephenytoin, another CYP2C19 substrate, was included in the search however only 1 study was identified that measured plasma concentrations across PM, IM and NM CYP2C19 phenotypes [59], while the remaining identified studies measured urinary metabolic ratio and did not report plasma pharmacokinetics (PK). Thus, further evaluation including S-mephenytoin was not conducted.

#### 3.2. Omeprazole and Lansoprazole PK Prediction in CYP2C19 Phenotypes

The simulated mean AUC for omeprazole recovered >80% of clinical AUC within the 2-fold acceptance criteria per phenotype (Table 2, Figure 1, Supplementary Table S3). Simulated mean  $C_{max}$  did not perform as well, recovering approximately 50–70% of the clinical  $C_{max}$  within the 2-fold acceptance criteria. Simulated omeprazole  $C_{max}$  tended to be underpredicted across phenotypes. As noted in the discussion, different formulations were used in the studies, which may contribute to the observed differences. The simulated mean AUC for lansoprazole recovered >70% and >90% of clinical AUC and  $C_{max}$ , respectively, within the 2-fold acceptance criteria per phenotype (Table 2, Figure 1, Supplementary Table S3).

Simulated and observed mean plasma concentration-time curves for omeprazole and lansoprazole are presented in Supplementary Figure S1 through Supplementary Figure S3.

**Table 2.** Summary of Simulated versus Observed AUC and C<sub>max</sub> Means Across CYP2C19 Phenotypes for Omeprazole and Lansoprazole.

No. of Simulated within 2-Fold of Observed/Total No. Data Points							
Omeprazole <sup>1</sup>	PM	IM	NM	RM	UM		
AUC	25/26	22/26	24/27	2/2	3/3		
C <sub>max</sub>	12/17	8/16	10/17	0/1	1/2		
Lansoprazole <sup>1</sup>							
AUC	7/10	10/11	11/11	1/1	NA		
C <sub>max</sub>	9/10	9/10	9/10	NA	NA		

NA = Not available, <sup>1</sup> Omeprazole includes both oral and IV data; lansoprazole is oral data only.



**Figure 1.** Simulated versus observed median and range of mean AUC and  $C_{max}$  values across CYP2C19 phenotypes for omeprazole and lansoprazole. Median of simulated trial means plotted against median of clinical study means. The red, blue, green and magenta vertical lines are range of simulated trial means; horizontal lines are range of clinical study data (see Table 2 for number of studies per phenotype). The solid black line is the line of unity; black dashed lines indicate 2-fold upper and lower acceptance criteria.

# 3.3. Victim DDI Prediction in CYP2C19 Phenotypes

Two DDI studies with CYP2C19 phenotyping information for omeprazole were available using either clarithromycin or ticlopidine as a CYP3A4 or CYP2C19 inhibitor, respectively [21,29]. Three DDI studies were available for lansoprazole using either clarithromycin (n = 1) or fluvoxamine (n = 2) as a CYP3A4 or combined CYP3A4/CYP2C19 inhibitor, respectively [34,41,52].

Mean AUC DDI ratios for 4/5 studies were within the Guest et al. [58] acceptance criteria for CYP2C19 PMs and 3/5 studies for CYP2C19 IMs and NMs (Figure 2). Mean C<sub>max</sub> DDI ratios for 4/5 studies were within acceptance criteria for CYP2C19 PMs, 2/5 studies for CYP2C19 IMs and 3/5 studies for CYP2C19 NMs. Of the 2 studies investigating DDI between lansoprazole and fluvoxamine, simulated DDI ratios consistently under-predicted data from Miura et al. 2005 [34] while accurately predicting data from Yasuri-Furukori et al., 2004 [52]. The reason for this is unknown as both studies used similar subject groups (age and population) and lansoprazole formulation although the Miura study used a higher dose (60 versus 40 mg).

![](_page_5_Figure_3.jpeg)

**Figure 2.** Simulated versus observed mean AUC and  $C_{max}$  ratios across CYP2C19 phenotypes for omeprazole and lansoprazole. CLA = clarithromycin; FLU = fluvoxamine; LAN = lansoprazole; OME = omeprazole; TIC = ticlopidine. The solid black line is the line of unity, black dashed lines indicate 2-fold upper and lower acceptance criteria and red dashed lines indicate acceptance criteria suggested by Guest et al. [58].

## 3.4. Prediction of CYP2C19 Phenotype Ratios

Of the studies compiled, five were selected to evaluate phenotype ratios based on their capturing a range of CYP2C19 phenotypes [18,19,26,33,40]. Approximately 80% of simulated mean AUC and 100% of simulated mean  $C_{max}$  relative change in exposure between CYP2C19 phenotypes were within the Guest et al. [58] acceptance criteria (Figure 3).

![](_page_6_Figure_1.jpeg)

**Figure 3.** Simulated versus observed mean relative changes in exposure between CYP2C19 phenotypes for select omeprazole (OME) and lansoprazole (LAN) studies [18,19,26,33,40]. The solid black line is the line of unity, black dashed lines indicate 2-fold upper and lower acceptance criteria and red dashed lines indicate acceptance criteria suggested by Guest et al. [58].

## 4. Discussion

PBPK models have proven to be a useful tool in understanding DDIs to the extent that in some situations, simulations can be used in place of clinical studies [2]. As such, it is important that models remain updated with the newest scientific developments and information. Model reproducibility remains a concern with some of the inconsistency observed attributed to changes in software versions [60]. While proper verification of model performance is necessary for any software update, modelers also need to understand the differences that exist between software versions. PBPK modeling software, Simcyp Simulator, is updated annually and version 21 included updates to CYP2C19 phenotype population data.

S-mephenytoin, omeprazole and lansoprazole are CYP2C19 substrates recommended by the FDA for use in clinical DDI studies [13]. While S-mephenytoin is considered as a sensitive CYP2C19 substrate, it was excluded from this analysis due to scarcity of clinical data with differing CYP2C19 phenotypes. This is likely the reason why only omeprazole and lansoprazole are noted as clinical index substrates for CYP2C19 by the FDA [13]. Using the default settings of healthy volunteer virtual Caucasian, Chinese and Japanese populations, as well as SV-Omeprazole, SV-Lansoprazole and perpetrator drug compound files within the Simcyp Simulator v21, the performance of CYP2C19 enzyme abundance updates was verified.

Verification was carried out in a stepwise manner. First, results from a single simulation were compared to a compiled data set per phenotype for single dose omeprazole or lansoprazole as an assessment of general population performance. Secondly, victim DDI of omeprazole with clarithromycin or ticlopidine and lansoprazole with clarithromycin or fluvoxamine were simulated. Finally, selected studies from the compiled dataset were modeled replicating individual study designs in terms of demographics to assess relative changes in exposure between CYP2C19 phenotypes normalized to NM simulations, where data allowed or to IM subjects if no NM data was available.

Overall, the simulations demonstrated reasonable performance of the updated Simcyp CYP2C19 enzyme abundance information in capturing omeprazole and lansoprazole exposure, DDI effects across CYP2C19 phenotypes and relative change in exposure between phenotypes. Where simulated predictions of omeprazole and lansoprazole exposure were outside of acceptance criteria, the PBPK model had a tendency to underpredict the exposure. Of note, the model under-predicted omeprazole exposure for the two studies that included a Pakistani population regardless of whether clinical data was compared to simulated Caucasian, Japanese or Chinese populations [35,36]. This may reflect unique population characteristics currently lacking in the virtual populations available in Simcyp Simulator v21. Limited data was available for CYP2C19 RM and UM subjects having received either omeprazole or lansoprazole. Further verification of these CYP2C19 phenotypes is warranted as additional data becomes available in the public domain.

Simulated data for lansoprazole was more accurate in recovering clinical  $C_{max}$  data than omeprazole. Different formulations of omeprazole were noted between studies, including enteric coated and MUPS (multi-unit pellet system) which are designed to have alternate PK profiles. The omeprazole compound file within v21 of the Simcyp Simulator was designed to recover the exposure of enteric coated omeprazole formulations and it is likely that the  $C_{max}$  for other formulations could be more accurately predicted using alternative absorption rate values.

CYP2C19-mediated DDI of omeprazole by ticlopidine was better predicted than CYP3A4-mediated DDI of clarithromycin across CYP2C19 phenotypes which may reflect future opportunities for improvement in either the clarithromycin or omeprazole compound files. Furthermore, CYP2C19 PM enzyme abundance is defined as no enzyme activity and simulation performance may reflect where the compound file models themselves could be further refined. This verification only included two CYP2C19 substrates; additional verification with other CYP2C19 substrates will further inform system parameter performance.

In conclusion, the latest updates to CYP2C19 enzyme abundances within the Simcyp Simulator v21 reasonably recaptured observed data with sensitive CYP2C19 substrates to warrant its use in PBPK simulations for DDI and polymorphism evaluation.

**Supplementary Materials:** The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/metabo12101001/s1, Figure S1: Simulated and observed mean omeprazole plasma concentration-time data in single oral dose studies per CYP2C19 phenotype; Figure S2: Simulated and observed mean omeprazole plasma concentration-time data in single intravenous dose studies per CYP2C19 phenotype; Figure S3: Simulated and observed mean lansoprazole plasma concentration-time data in single dose studies per CYP2C19 phenotype; Table S1: PBPK model parameters for omeprazole and lansoprazole; Table S2: CYP2C19 literature search results for omeprazole, lansoprazole and S-mephenytoin; Table S3: Simulated versus observed PK parameters for omeprazole and lansoprazole.

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## References

- 1. Bahar, M.A.; Setiawan, D.; Hak, E.; Wilffert, B. Pharmacogenetics of drug-drug interaction and drug-drug-gene interaction: A systematic review on CYP2C9, CYP2C19 and CYP2D6. *Pharmacogenomics* **2017**, *18*, 701–739. [CrossRef] [PubMed]
- US Food and Drug Administration. Guidance for Industry: Clinical Drug Interaction Studies—Cytochrome P450 Enzyme- and Transporter-Mediated Drug Interactions. Available online: https://www.fda.gov/media/134581/download (accessed on 14 June 2022).
- 3. European Medicines Agency. Guideline on the Investigation of Drug Interactions. Available online: https://www.ema.europa. eu/en/documents/scientific-guideline/guideline-investigation-drug-interactions-revision-1\_en.pdf (accessed on 14 June 2022).
- Zhang, X.; Yang, Y.; Grimstein, M.; Fan, J.; Grillo, J.A.; Huang, S.M.; Zhu, H.; Wang, Y. Application of PBPK Modeling and Simulation for Regulatory Decision Making and Its Impact on US Prescribing Information: An Update on the 2018-2019 Submissions to the US FDA's Office of Clinical Pharmacology. J. Clin. Pharmacol. 2020, 60 (Suppl. 1), S160–S178. [CrossRef]
- 5. Rostami-Hodjegan, A. Reverse translation in PBPK and QSP: Going backwards in order to go forward with confidence. *Clin. Pharmacol. Ther.* **2018**, *103*, 224–232. [CrossRef]
- 6. Zhao, M.; Ma, J.; Li, M.; Zhang, Y.; Jiang, B.; Zhao, X.; Huai, C.; Shen, L.; Zhang, N.; He, L.; et al. Cytochrome P450 enzymes and drug metabolism in humans. *Int. J. Mol. Sci.* **2021**, *22*, 12808. [CrossRef] [PubMed]
- 7. Fanni, D.; Pinna, F.; Gerosa, C.; Paribello, P.; Carpiniello, B.; Faa, G.; Manchia, M. Anatomical distribution and expression of CYP in humans: Neuropharmacological implications. *Drug Dev Res.* **2021**, *85*, 628–667. [CrossRef] [PubMed]
- Manikandan, P.; Nagini, S. Cytochrome P450 structure, function and clinical significance: A review. *Curr Drug Targets* 2018, 19, 38–54. [CrossRef] [PubMed]
- 9. Zanger, U.M.; Schwab, M. Cytochrome P450 enzymes in drug metabolism: Regulation of gene expression, enzyme activities, and impart of genetic variation. *Pharmacol. Ther.* **2013**, *138*(1), 103–141. [CrossRef]
- Klein, M.D.; Williams, A.K.; Lee, C.R.; Stouffer, G.A. Clinical Utility of CYP2C19 Genotyping to Guide Antiplatelet Therapy in Patients With an Acute Coronary Syndrome or Undergoing Percutaneous Coronary Intervention. *Arterioscler. Thromb. Vasc. Bio.* 2019, 39(4), 647–652. [CrossRef]
- Ozawa, S.; Soyama, A.; Saeki, M.; Fukushima-Uesaka, H.; Itoda, M.; Koyano, S.; Sai, K.; Ohno, Y.; Saito, Y.; Sawada, J. Ethnic differences in genetic polymorphisms of CYP2D6, CYP2C19, CYP3As and MDR1/ABCB1. *Drug Metab. Pharmacokinet.* 2004, 19, 83–95. [CrossRef]
- Kita, T.; Sakaeda, T.; Baba, T.; Aoyama, N.; Kakumoto, M.; Kurimoto, Y.; Kawahara, Y.; Okamura, N.; Kirita, S.; Kasuga, M.; et al. Different contribution of CYP2C19 in the in vitro metabolism of three proton pump inhibitors. *Biol. Pharm. Bull.* 2003, 26, 386–390. [CrossRef]
- 13. US Food and Drug Administration. Drug Development and Drug Interactions | Table of Substrates, Inhibitors and Inducers. Available online: https://www.fda.gov/drugs/drug-interactions-labeling/drug-development-and-drug-interactions-table-substrates-inhibitors-and-inducers#table2-1 (accessed on 14 June 2022).
- 14. University of Washington Drug Interaction Solutions. Available online: https://www.druginteractionsolutions.org/ (accessed on 16 November 2021).
- 15. Anil. Digitize2.m. MATLAB Central File Exchange. 2022. Available online: https://www.mathworks.com/matlabcentral/fileexchange/928-digitize2-m (accessed on 29 June 2021).
- 16. Zhou, L.; Sharma, P.; Yeo, K.R.; Higashimori, M.; Xu, H.; Al-Huniti, N.; Zhou, D. Assessing pharmacokinetic differences in Caucasian and East Asian (Japanese, Chinese and Korean) populations driven by CYP2C19 polymorphism using physiologically-based pharmacokinetic modelling. *Eur. J. Pharm. Sci.* **2019**, *139*, 105061. [CrossRef] [PubMed]
- Zvyaga, T.; Chang, S.Y.; Chen, C.; Yang, Z.; Vuppugalla, R.; Hurley, J.; Thorndike, D.; Wagner, A.; Chimalakonda, A.; Rodrigues, A.D. Evaluation of six proton pump inhibitors as inhibitors of various human cytochromes P450: Focus on cytochrome P450 2C19. Drug Metab. Dispos. 2012, 40, 1698–1711. [CrossRef]
- Baldwin, R.M.; Ohlsson, S.; Pedersen, R.S.; Mwinyi, J.; Ingelman-Sundberg, M.; Eliasson, E.; Bertilsson, L. Increased omeprazole metabolism in carriers of the CYP2C19\*17 allele; a pharmacokinetic study in healthy volunteers. *Br. J. Clin. Pharmacol.* 2008, 65, 767–774. [CrossRef] [PubMed]
- Chang, M.; Tybring, G.; Dahl, M.L.; Gotharson, E.; Sagar, M.; Seensalu, R.; Bertilsson, L. Interphenotype differences in disposition and effect on gastrin levels of omeprazole–suitability of omeprazole as a probe for CYP2C19. *Br. J. Clin. Pharmacol.* 1995, 39, 511–518. [CrossRef]
- 20. Cho, J.Y.; Yu, K.S.; Jang, I.J.; Yang, B.H.; Shin, S.G.; Yim, D.S. Omeprazole hydroxylation is inhibited by a single dose of moclobemide in homozygotic EM genotype for CYP2C19. *Br. J. Clin. Pharmacol.* **2002**, *53*, 393–397. [CrossRef] [PubMed]
- Furuta, T.; Ohashi, K.; Kobayashi, K.; Iida, I.; Yoshida, H.; Shirai, N.; Takashima, M.; Kosuge, K.; Hanai, H.; Chiba, K.; et al. Effects of clarithromycin on the metabolism of omeprazole in relation to CYP2C19 genotype status in humans. *Clin. Pharmacol. Ther.* 1999, *66*, 265–274. [CrossRef]
- Furuta, T.; Ohashi, K.; Kosuge, K.; Zhao, X.J.; Takashima, M.; Kimura, M.; Nishimoto, M.; Hanai, H.; Kaneko, E.; Ishizaki, T. CYP2C19 genotype status and effect of omeprazole on intragastric pH in humans. *Clin. Pharmacol. Ther.* 1999, 65, 552–561. [CrossRef]
- 23. Furuta, T.; Shirai, N.; Xiao, F.; Ohashi, K.; Ishizaki, T. Effect of high-dose lansoprazole on intragastic pH in subjects who are homozygous extensive metabolizers of cytochrome P4502C19. *Clin. Pharmacol. Ther.* **2001**, *70*, 484–492. [CrossRef]

- He, N.; Huang, S.L.; Zhu, R.H.; Tan, Z.R.; Liu, J.; Zhu, B.; Zhou, H.H. Inhibitory effect of troleandomycin on the metabolism of omeprazole is CYP2C19 genotype-dependent. *Xenobiotica* 2003, 33, 211–221. [CrossRef]
- 25. Hu, X.P.; Xu, J.M.; Hu, Y.M.; Mei, Q.; Xu, X.H. Effects of CYP2C19 genetic polymorphism on the pharmacokinetics and pharmacodynamics of omeprazole in Chinese people. *J. Clin. Pharm. Ther.* **2007**, *32*, 517–524. [CrossRef]
- Hunfeld, N.G.; Mathot, R.A.; Touw, D.J.; van Schaik, R.H.; Mulder, P.G.; Franck, P.F.; Kuipers, E.J.; Geus, W.P. Effect of CYP2C19\*2 and \*17 mutations on pharmacodynamics and kinetics of proton pump inhibitors in Caucasians. *Br. J. Clin. Pharmacol.* 2008, 65, 752–760. [CrossRef] [PubMed]
- Ieiri, I.; Kubota, T.; Urae, A.; Kimura, M.; Wada, Y.; Mamiya, K.; Yoshioka, S.; Irie, S.; Amamoto, T.; Nakamura, K.; et al. Pharmacokinetics of omeprazole (a substrate of CYP2C19) and comparison with two mutant alleles, C gamma P2C19m1 in exon 5 and C gamma P2C19m2 in exon 4, in Japanese subjects. *Clin. Pharmacol. Ther.* **1996**, *59*, 647–653. [CrossRef]
- Ieiri, I.; Kishimoto, Y.; Okochi, H.; Momiyama, K.; Morita, T.; Kitano, M.; Morisawa, T.; Fukushima, Y.; Nakagawa, K.; Hasegawa, J.; et al. Comparison of the kinetic disposition of and serum gastrin change by lansoprazole versus rabeprazole during an 8-day dosing scheme in relation to CYP2C19 polymorphism. *Eur. J. Clin. Pharmacol.* 2001, *57*, 485–492. [CrossRef]
- Ieiri, I.; Kimura, M.; Irie, S.; Urae, A.; Otsubo, K.; Ishizaki, T. Interaction magnitude, pharmacokinetics and pharmacodynamics of ticlopidine in relation to CYP2C19 genotypic status. *Pharmacogenet. Genomics* 2005, 15, 851–859. [CrossRef] [PubMed]
- Ishizawa, Y.; Yasui-Furukori, N.; Takahata, T.; Sasaki, M.; Tateishi, T. The effect of aging on the relationship between the cytochrome P450 2C19 genotype and omeprazole pharmacokinetics. *Clin. Pharmacokinet.* 2005, 44, 1179–1189. [CrossRef] [PubMed]
- 31. Jin, S.K.; Kang, T.S.; Eom, S.O.; Kim, J.I.; Lee, H.J.; Roh, J. CYP2C19 haplotypes in Koreans as a marker of enzyme activity evaluated with omeprazole. *J. Clin. Pharm. Ther.* **2009**, *34*, 437–446. [CrossRef] [PubMed]
- Li, C.Y.; Zhang, J.; Chu, J.H.; Xu, M.J.; Ju, W.Z.; Liu, F.; Jian-Dong, Z. A correlative study of polymorphisms of CYP2C19 and MDR1 C3435T with the pharmacokinetic profiles of lansoprazole and its main metabolites following single oral administration in healthy adult Chinese subjects. *Eur. J. Drug Metab. Pharmacokinet.* 2014, 39, 121–128. [CrossRef]
- Michaud, V.; Kreutz, Y.; Skaar, T.; Ogburn, E.; Thong, N.; Flockhart, D.A.; Desta, Z. Efavirenz-mediated induction of omeprazole metabolism is CYP2C19 genotype dependent. *Pharmacogenomics J.* 2014, 14, 151–159. [CrossRef]
- 34. Miura, M.; Tada, H.; Yasui-Furukori, N.; Uno, T.; Sugawara, K.; Tateishi, T.; Suzuki, T. Enantioselective disposition of lansoprazole in relation to CYP2C19 genotypes in the presence of fluvoxamine. *Br. J. Clin. Pharmacol.* **2005**, *60*, 61–68. [CrossRef]
- 35. Nazir, S.; Iqbal, Z.; Ahmad, L.; Shah, Y.; Nasir, F. Pharmacokinetics of omeprazole and its metabolites in three phases of menstrual cycle. *Eur. J. Drug Metab. Pharmacokinet.* **2015**, *40*, 13–22. [CrossRef]
- Nazir, S.; Iqbal, Z.; Ahmad, L.; Ahmad, S. Variation in Pharmacokinetics of Omeprazole and Its Metabolites by Gender and CYP2C19 Genotype in Pakistani Male and Female Subjects. *Pak. J. Pharm. Sci.* 2016, 29, 887–894. Available online: https://www.ncbi.nlm.nih.gov/pubmed/27166533 (accessed on 1 September 2022). [PubMed]
- Park, S.; Hyun, Y.J.; Kim, Y.R.; Lee, J.H.; Ryu, S.; Kim, J.M.; Oh, W.Y.; Na, H.S.; Lee, J.G.; Seo, D.W.; et al. Effects of CYP2C19 Genetic Polymorphisms on PK/PD Responses of Omeprazole in Korean Healthy Volunteers. J. Korean Med. Sci. 2017, 32, 729–736. [CrossRef] [PubMed]
- 38. Payan, M.; Rouini, M.R.; Tajik, N.; Ghahremani, M.H.; Tahvilian, R. Hydroxylation index of omeprazole in relation to CYP2C19 polymorphism and sex in a healthy Iranian population. *Daru* **2014**, *22*, 81. [CrossRef] [PubMed]
- 39. Qiao, H.L.; Hu, Y.R.; Tian, X.; Jia, L.J.; Gao, N.; Zhang, L.R.; Guo, Y.Z. Pharmacokinetics of three proton pump inhibitors in Chinese subjects in relation to the CYP2C19 genotype. *Eur. J. Clin. Pharmacol.* **2006**, *62*, 107–112. [CrossRef]
- Roman, M.; Ochoa, D.; Sanchez-Rojas, S.D.; Talegon, M.; Prieto-Perez, R.; Rivas, A.; Abad-Santos, F.; Cabaleiro, T. Evaluation of the relationship between polymorphisms in CYP2C19 and the pharmacokinetics of omeprazole, pantoprazole and rabeprazole. *Pharmacogenomics* 2014, 15, 1893–1901. [CrossRef]
- Saito, M.; Yasui-Furukori, N.; Uno, T.; Takahata, T.; Sugawara, K.; Munakata, A.; Tateishi, T. Effects of clarithromycin on lansoprazole pharmacokinetics between CYP2C19 genotypes. *Br. J. Clin. Pharmacol.* 2005, 59, 302–309. [CrossRef]
- Sakai, T.; Aoyama, N.; Kita, T.; Sakaeda, T.; Nishiguchi, K.; Nishitora, Y.; Hohda, T.; Sirasaka, D.; Tamura, T.; Tanigawara, Y.; et al. CYP2C19 genotype and pharmacokinetics of three proton pump inhibitors in healthy subjects. *Pharm. Res.* 2001, 18, 721–727. [CrossRef]
- Shimizu, M.; Uno, T.; Niioka, T.; Yaui-Furukori, N.; Takahata, T.; Sugawara, K.; Tateishi, T. Sensitive determination of omeprazole and its two main metabolites in human plasma by column-switching high-performance liquid chromatography: Application to pharmacokinetic study in relation to CYP2C19 genotypes. J. Chromatogr. B Analyt. Technol. Biomed. Life Sci. 2006, 832, 241–248. [CrossRef]
- Shiohira, H.; Yasui-Furukori, N.; Yamada, S.; Tateishi, T.; Akamine, Y.; Uno, T. Hydroxylation of R(+)- and S(-)-omeprazole after racemic dosing are different among the CYP2C19 genotypes. *Pharm. Res.* 2012, 29, 2310–2316. [CrossRef]
- 45. Shirai, N.; Furuta, T.; Moriyama, Y.; Okochi, H.; Kobayashi, K.; Takashima, M.; Xiao, F.; Kosuge, K.; Nakagawa, K.; Hanai, H.; et al. Effects of CYP2C19 genotypic differences in the metabolism of omeprazole and rabeprazole on intragastric pH. *Aliment. Pharmacol. Ther.* 2001, 15, 1929–1937. [CrossRef]
- 46. Tu, J.H.; Hu, D.L.; Dai, L.L.; Sun, Y.; Fan, L.; Zhang, M.; Tan, Z.R.; Chen, Y.; Li, Z.; Zhou, H.H. Effect of glycyrrhizin on CYP2C19 and CYP3A4 activity in healthy volunteers with different CYP2C19 genotypes. *Xenobiotica* **2010**, *40*, 393–399. [CrossRef] [PubMed]

- Uno, T.; Yasui-Furukori, N.; Takahata, T.; Sugawara, K.; Tateishi, T. Lack of significant effect of grapefruit juice on the pharmacokinetics of lansoprazole and its metabolites in subjects with different CYP2C19 genotypes. J. Clin. Pharmacol. 2005, 45, 690–694. [CrossRef]
- Uno, T.; Niioka, T.; Hayakari, M.; Yasui-Furukori, N.; Sugawara, K.; Tateishi, T. Absolute bioavailability and metabolism of omeprazole in relation to CYP2C19 genotypes following single intravenous and oral administrations. *Eur. J. Clin. Pharmacol.* 2007, 63, 143–149. [CrossRef]
- 49. Xu, H.R.; Chen, W.L.; Li, X.N.; Chu, N.N. The effect of CYP2C19 activity on pharmacokinetics of lansoprazole and its active metabolites in healthy subjects. *Pharm. Biol.* 2010, *48*, 947–952. [CrossRef] [PubMed]
- 50. Yamada, S.; Shiohira, H.; Yasui-Furukori, N.; Tateishi, T.; Akamine, Y.; Uno, T. The (R)-omeprazole hydroxylation index reflects CYP2C19 activity in healthy Japanese volunteers. *Eur. J. Clin. Pharmacol.* **2013**, *69*, 1423–1428. [CrossRef] [PubMed]
- Yang, L.J.; Fan, L.; Liu, Z.Q.; Mao, Y.M.; Guo, D.; Liu, L.H.; Tan, Z.R.; Peng, L.; Han, C.T.; Hu, D.L.; et al. Effects of allicin on CYP2C19 and CYP3A4 activity in healthy volunteers with different CYP2C19 genotypes. *Eur. J. Clin. Pharmacol.* 2009, 65, 601–608. [CrossRef]
- Yasui-Furukori, N.; Saito, M.; Uno, T.; Takahata, T.; Sugawara, K.; Tateishi, T. Effects of fluvoxamine on lansoprazole pharmacokinetics in relation to CYP2C19 genotypes. J. Clin. Pharmacol. 2004, 44, 1223–1229. [CrossRef]
- Yin, O.Q.; Tomlinson, B.; Chow, A.H.; Waye, M.M.; Chow, M.S. Omeprazole as a CYP2C19 marker in Chinese subjects: Assessment of its gene-dose effect and intrasubject variability. *J. Clin. Pharmacol.* 2004, 44, 582–589. [CrossRef]
- 54. Zhang, D.; Wang, X.; Yang, M.; Wang, G.; Liu, H. Effects of CYP2C19 polymorphism on the pharmacokinetics of lansoprazole and its main metabolites in healthy Chinese subjects. *Xenobiotica* **2011**, *41*, 511–517. [CrossRef]
- 55. Landes, B.D.; Petite, J.P.; Flouvat, B. Clinical pharmacokinetics of lansoprazole. Clin. Pharmacokinet. 1995, 28, 458–470. [CrossRef]
- Andersson, T.; Cederberg, C.; Regardh, C.G.; Skanberg, I. Pharmacokinetics of various single intravenous and oral doses of omeprazole. *Eur. J. Clin. Pharmacol.* 1990, 39, 195–197. [CrossRef]
- Kim, Y.; Hatley, O.; Rhee, S.J.; Yi, S.; Lee, H.A.; Yoon, S.; Chung, J.Y.; Yu, K.S.; Lee, H. Development of a Korean-specific virtual population for physiologically based pharmacokinetic modelling and simulation. *Biopharm. Drug Dispos.* 2019, 40, 135–150. [CrossRef] [PubMed]
- Guest, E.J.; Aarons, L.; Houston, J.B.; Rostami-Hodjegan, A.; Galetin, A. Critique of the two-fold measure of prediction success for ratios: Application for the assessment of drug-drug interactions. *Drug Metab. Dispos.* 2011, 39, 170–173. [CrossRef] [PubMed]
- 59. Wedlund, P.J.; Aslanian, W.S.; Jacqz, E.; McAllister, C.B.; Branch, R.A.; Wilkinson, G.R. Phenotypic Differences in Mephenytoin Pharmacokinetics in Normal Subjects. *J. Pharmacol. Exp. Ther.* **1985**, *234*, 662–669. Available online: https://www.ncbi.nlm.nih. gov/pubmed/4032286 (accessed on 1 September 2022). [PubMed]
- 60. Tiwari, K.; Kananathan, S.; Roberts, M.G.; Meyer, J.P.; Sharif Shohan, M.U.; Xavier, A.; Maire, M.; Zyoud, A.; Men, J.; Ng, S.; et al. Reproducibility in systems biology modelling. *Mol. Syst. Biol.* **2021**, *17*, e9982. [CrossRef] [PubMed]