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Review Article

Age-dependent regenerative mechanisms in the brain

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Repairing the adult mammalian brain represents one of the greatest clinical challenges in medicine. Injury to the adult brain often results in substantial loss of neural tissue and permanent functional impairment. In contrast with the adult, during development, the mammalian brain exhibits a remarkable capacity to replace lost cells. A plethora of cell-intrinsic and extrinsic factors regulate the age-dependent loss of regenerative potential in the brain. As the developmental window closes, neural stem cells undergo epigenetic changes, limiting their proliferation and differentiation capacities, whereas, changes in the brain microenvironment pose additional challenges opposing regeneration, including inflammation and gliosis. Therefore, studying the regenerative mechanisms during development and identifying what impairs them with age may provide key insights into how to stimulate regeneration in the brain. Here, we will discuss how the mammalian brain engages regenerative mechanisms upon injury or neuron loss. Moreover, we will describe the age-dependent changes that affect these processes. We will conclude by discussing potential therapeutic approaches to overcome the age-dependent regenerative decline and stimulate regeneration.

Introduction

Regeneration is the process of replacing missing or damaged cells and structures to restore the integrity of the tissue and its function. However, regenerative capacity varies greatly across species, tissues and stages. In contrast with fish and salamanders, which can regenerate their nervous systems throughout life [1,2], the mammalian brain shows a dramatic decline in its regenerative capacity after development, eventually leading to limited repair in the adult brain. The molecular and cellular mechanisms that facilitate or inhibit regeneration in the mammalian brain are poorly understood. This significant gap in our knowledge impairs the design of effective therapeutic strategies that stimulate and promote regeneration in the brain.

The mammalian brain was long considered incapable of generating new neurons after development, a limitation described by Ramon Cajal as a 'harsh decree' [3]. However, the identification of adult neurogenesis in specific regions of the brain, termed the canonical neurogenic niches, has broken this dogma [4,5]. Despite the presence of neural stem cells (NSCs), the adult brain exhibits a limited capacity to regenerate upon cell loss due to injury or degeneration, outside of the canonical neurogenic niches and adult neurogenesis. Interestingly, during development, regenerative processes in the mammalian brain are observed in a region-specific manner.

Neuron loss due to injury or neurodegeneration triggers context-dependent acute or chronic changes that stimulate neural stem/stem-like cells and other injury-responsive cells in the brain. For example, cell death upon injury induces up-regulation of damage-associated molecular patterns, including reactive oxygen species (ROS), which stimulate NSCs to initiate regenerative programmes [6]. However, these alarm signals could alternatively trigger anti-regenerative mechanisms through astrocytes and microglia, leading to neuroinflammation and glial scarring, similar to fibrotic scars in other tissues [7,8].

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Most of our knowledge of the cellular and molecular mechanisms of central nervous system regeneration derives from studies on axon regeneration in the spinal cord of zebrafish, axolots, larval *Xenopus* and rodents [9–11]. Axon regeneration refers to the process of restoring neural connectivity. On the other hand, cell regeneration, albeit rare in the adult mammalian brain, is the process of replacing or restoring damaged or missing cells via new cell production. For example, oligodendrocyte progenitors have been extensively studied for their cellular regenerative potential in the mammalian brain [12]. Although scarce, studies exploring regenerative potential during brain development and/or outside the canonical niches provide crucial insights into the regeneration of new cells in the mammalian brain. Here, we summarise the endogenous repair mechanisms to replace cells that occur upon acute cell loss such as stroke, traumatic brain injury or experimental ablation in the developing and adult mammalian brain from a stem cell biology and astroglia perspective. We discuss the cell-intrinsic and extrinsic changes that occur once development ends and upon ageing and their impact on the NSCs' regenerative potential. Finally, we conclude by discussing potential therapeutic strategies to facilitate regeneration in the brain.

Regeneration in the canonical neurogenic niches and cerebral cortex

The developing brain has the potential for regeneration, likely facilitated by the plastic nature of its cells and the permissive microenvironment. The regenerative ability of the embryonic neocortex can be demonstrated indirectly through grafting experiments into the adult injured cortex. In these studies, dissociated embryonic neurons that were transplanted into the cortex integrated into the existing cortical circuits of the adult brain after an injury (e.g. stroke) [13,14]. Interestingly when embryonic tissue was grafted, instead of dissociated neurons, engraftment efficiency increased [15]. Although indirect, these studies highlight the regenerative potential of the embryonic brain and suggest that this is a combination of tissue microenvironment and cell-intrinsic properties.

In the postnatal brain, canonical neurogenic niches maintain regenerative capacity once development ends and throughout adulthood. One observed regenerative mechanism is the increased compensatory proliferation of endogenous NSCs (Figure 1A,B). In the postnatal brain, the two canonical neurogenic regions: the subventricular zone (SVZ) lining the lateral ventricle and the subgranular zone (SGZ) of the hippocampal dentate gyrus, house NSCs that generate the olfactory bulb and hippocampal neurons, respectively [4,5,16,17]. Studies have reported that, upon injury, undifferentiated neuroblasts from the SVZ proliferate and migrate to the site of injury. In a model of cortical stroke, this migration has been shown to occur along an astrocytic tunnel controlled by several factors including chemoattractants, growth factors (e.g. VEGF) and neurotrophins (e.g. BDNF) that connect the SVZ to the injured non-regenerative cortex [18,19]. Indeed, migration of immature neuroblasts is associated with the expression of Doublecortin-positive (DCX⁺) cells, a marker for immature neurons of the SVZ and SGZ, to the site of injury. However, the origin of DCX+ cells in the cortex following injury remains elusive. Evidence of endogenous NSC-mediated regeneration has also been described in the SGZ of the hippocampus where newly generated neurons were observed in situ after induced traumatic brain injury [20] and, in an oligodendrocytes cell death model, DCX+ proliferating progenitors were associated with the generation of mature oligodendrocytes during hippocampus remyelination [21]. However, the poor survival of the newly generated neurons raises the question of whether such injury responses can be considered as successful regeneration [22,23].

Besides NSCs, many studies highlight the glia, specifically astrocytes, as potential cellular sources for regeneration by taking on stem cell-like capacities upon damage (Figure 1A,B). For example, upon injury to the cerebral cortex, DCX expression could be a result of neuroblast migration to the site of injury [24]. However, other studies suggest that the astrocytes present at the site of injury start expressing DCX [25] and adopt stem-like characteristics. Reactive astrocytes isolated from the acutely injured brain were able to take on NSC characteristics *in vitro* in response to sonic hedgehog (SHH) signalling [26]. Similarly, some specialised astrocytes outside of the cerebral cortex such as Bergmann glia of the cerebellum are transcriptionally similar to the radial glia [27], raising the question of whether they maintain any neurogenic potential. Collectively, these studies provide examples of regeneration via compensatory proliferation within the NSC lineage or acquisition of stemness and neurogenic potential of astroglia around the injury site (Figure 1A,B).

Regeneration outside the cerebral cortex

Following cell loss, efficient repair has been observed outside of the canonical neurogenic niches (SVZ and SGZ) in an age-dependent manner. The neonatal mouse cerebellum exhibits high regenerative potential upon loss of at least two of its cell types via distinct mechanisms [28–30].



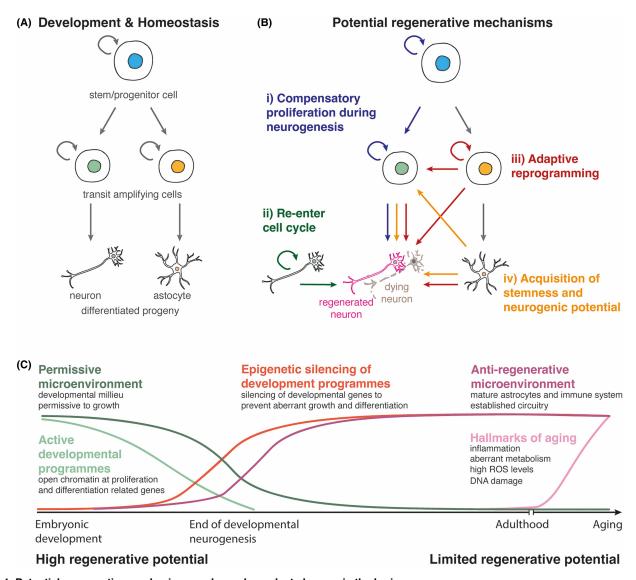


Figure 1. Potential regenerative mechanisms and age-dependent changes in the brain.

(A) Hypothetical schematic of neural stem cell (NSC) lineages during development and homeostasis (oligodendrocytes were omitted). (B) Potential mechanisms of neuron regeneration mediated by (i) compensatory proliferation of NSCs or transit amplifying cells [18–20] (blue), (ii) immature neuron progenitors or neurons re-entering the cell cycle [29] (green), (iii) adaptive reprogramming of NSCs, gliogenic progenitors or astroglia [28,32–35] (red), (iv) astrocytes acquiring stemness or neurogenic potential upon injury [25,26] (yellow). (C) Cell-intrinsic and -extrinsic changes that occur during development and ageing in the brain.

One such mechanism is the re-entry of immature neurons into the cell cycle to replenish their lost neighbours (Figure 1A,B). Purkinje cells are GABAergic neurons that converge all the information that enters the cerebellar cortex to the primary output neurons of the cerebellum, the excitatory cerebellar nuclei. While Purkinje cells are produced between embryonic days (E) 10–13 in mice, they remain immature until completion of postnatal neurogenesis in the cerebellum, two weeks after birth. In one study, ablation of ~50% of the Purkinje cells at postnatal day (P) 1 resulted in a subpopulation of Purkinje cells, immature Purkinje cell progenitors, re-entering the cell cycle. These cells subsequently replenished their lost neighbours within a day, allowing development to progress normally [29]. Although previously, forcing neurons to proliferate led to cell death [31], this suggests that, within critical developmental windows, injury or cell death could stimulate other immature neurons to re-enter the cell cycle for regeneration.



Another mechanism involves adaptive reprogramming, which comprises the generation of a temporary pool of progenitors/cell states that replenishes the lost cells upon injury or ablation and could exhibit lineage plasticity (Figure 1A,B). During neonatal cerebellar development, molecularly distinct progenitor subpopulations that reside in different regions within the cerebellar cortex produce the requisite cells of the cerebellum. The rhombic lip-derived granule cell progenitors produce the excitatory granule cells, while the ventricular zone-derived Nestin-expressing progenitors (NEPs) generate Bergmann glia, astrocytes and inhibitory neurons via molecular distinct subpopulations that reside in different regions within the cerebellar cortex. Upon death of the granule cell progenitors via irradiation or genetic ablation, a subpopulation of the gliogenic Hopx-expressing NEPs that normally generate Bergmann glia, regenerate granule cell progenitors via adaptive reprogramming in a multistep response that involves proliferation, fate-switch, migration and differentiation [28,32]. Single-cell transcriptomics on control and injured NEPs, fate mapping experiments and loss of function studies reveal that an Ascl1⁺ transitory cell state is required for adaptive reprogramming, likely by mediating the gliogenic-to-neurogenic fate switch [32]. While it is still unknown why the loss of different neurons in the cerebellum triggers different regenerative responses, the inherent regenerative capacity of the neonatal cerebellum makes it a powerful model for studying the molecular and cellular mechanisms of regeneration in the brain.

Interestingly, although not spontaneous, the mammalian retina can also show regenerative potential. Manipulation of cell cycle or cellular reprogramming through ASCL1-mediated transcriptional changes and subsequent neuron production has also been observed in the injured mammalian retina. Here, Müller glia act as a source of progenitor cells capable of de-differentiation to a progenitor state, proliferation and neuronal differentiation upon acute retina damage in an age-dependent manner [33–36] (Figure 1A,B). In this instance, Müller glia share a similar transcriptional profile with retinal progenitors, however, key pro-neural genes are not expressed upon injury. As a result, in contrast with the neonatal cerebellum, the retina doesn't undergo spontaneous reprogramming and requires exogenous manipulation of transcription factors such as ASCL1 to initiate their neural differentiation and pro-regenerative program [33,35].

Recently, new putative neurogenic sites have been speculated to exist in the mammalian brain. Although neurogenic potential is low compared with the canonical niches, the hypothalamus, a region important for homeostatic regulation of physiological processes, is an emerging neurogenic region. The tanycytes, radial glialike cells in the hypothalamus, exhibit stem/progenitor potential both during homeostasis and upon physiological perturbations such as changes in dietary, environmental and hormonal signals [37–39]. However, hypothalamic neurogenesis in response to injury remains to be confirmed but, overall, these observations indicate another cell type with regenerative potential in the brain. In summary, these results demonstrate that different brain regions adopt various regenerative mechanisms to overcome cell loss and highlight the importance of investigating outside the canonical neurogenic niches.

Age-dependent decline in the regenerative potential of the brain

While distinct context-dependent regenerative responses have been observed in different brain regions, one uniform factor across all these responses is that they are tightly regulated by age. While the developing brain has compensatory mechanisms upon injury or neuron loss, the adult brain has limited regenerative potential outside the neurogenic zones. In addition, adult NSCs become quiescent and exhausted with age, as they exhibit hallmarks of ageing such as accumulated DNA damage and oxidative stress (Figure 1C) [40]. In turn, these collectively dampen the regenerative potential of NSCs. In this section, we will summarise and speculate on the cell-intrinsic and -extrinsic mechanisms of regeneration blockage in the brain and how age influences them. Understanding these molecular and cellular mechanisms is the crucial first step to facilitate repair.

Cell-intrinsic mechanisms

The state of the epigenome could dramatically influence the proliferation and differentiation potential and hence the repair capacity of the stem/progenitor-like cells after an injury or cell loss. As neural development progresses, NSCs undergo epigenetic changes and switch their fate from neurogenic to gliogenic [41]. The proliferation and differentiation pathways are silenced in the differentiated progeny, potentially to prevent aberrant reprogramming as a protective mechanism against overgrowth or cancers. In addition to the chromatin remodelling that marks the end of differentiation, upon ageing, alterations in the epigenome correlate with an increase in inflammatory pathways [42]. Furthermore, studies comparing the epigenome and transcriptome of adult and



aged NSCs in the SVZ show the silencing of genes that are fundamental for stem cell proliferation and differentiation [43–46]. However, it is important to note that these epigenetic features are cell type- and context-dependent and most of these studies are done *in vitro* using primary NSC cultures or they are restricted to neurogenic zones. Therefore, comparative epigenomic analyses in the emerging regenerative sites discussed earlier, such as the cerebellum, as well as *in vivo* functional analyses are required to further elucidate the age-dependent epigenetic mechanisms affecting the regenerative potential of NSC and NSC-like cells. An open question is to what extent the developmental reprogramming of the epigenome during differentiation vs. the effects of ageing on the epigenome influences regeneration.

Among the factors affecting NSC function, and hence regeneration, is cellular metabolism [47,48]. Mitochondria are known to play a pivotal role in energy metabolism and stress response. Importantly, mitochondrial dysfunction is considered another hallmark of ageing [40]. During differentiation, NSCs undergo a metabolic switch from glycogenic to oxidative phosphorylation and the mitochondria morphologically change to accompany this transition [49,50]. Products of mitochondrial metabolism such as endogenous ROS are shown to regulate NSC self-renewal and differentiation in a transgenic mouse model that reduces ROS levels [6]. However, in the aged brain, accumulation of mitochondrial DNA mutations increases ROS levels, impairs the oxidative phosphorylation machinery and causes DNA damage. These alterations significantly impair neurogenesis and support the change in NSC lineage decisions from a neuronal fate to an astroglia one [51–54]. Collectively, metabolic changes in NSCs during differentiation or ageing could indirectly affect their regenerative potential. Interestingly, although ROS is a damage-associated molecular pattern and is increased in the microenvironment upon cell death, whether the endogenous ROS levels of NSCs or other injury-responsive cells factor into the regenerative response remains to be studied.

Cell-extrinsic mechanisms

Inflammation and scar formation are known to be inhibitory to regeneration in many complex tissues. Upon injury and acute cell loss, brain parenchyma also initiates inflammatory responses. In addition, reactive astrocytes change morphology and transcriptional profile, and importantly, activate proliferation programmes in response to damage-associated molecular patterns, leading to gliosis. In most cases, inflammation and reactive gliosis are thought to be inhibitory to regeneration, while context-dependent nuances may exist [55,56].

Microglia, the resident macrophages of the brain, are derived from the embryonic yolk sac [57]. Microglia are commonly classified into a pro- or anti-inflammatory phenotype. However, recent studies have shown that microglia have vast molecular and cellular heterogeneity across different brain regions and, importantly, between the developing and the adult brain [58]. Furthermore, forebrain organoid models that include microglia showed modulation of NSC behaviour, highlighting the cross-talk between microglia and NSCs during development [59]. Finally, IFNy and cGAS-STING DNA sensing pathways have been shown to drive neuroin-flammation degeneration and ageing, further confirming the importance of the immune system on nervous system pathophysiology [60]. However, the functional relevance of microglial heterogeneity and the cross-talk between microglia and NSC/NSC-like cells during regeneration and repair remain to be understood. Upon injury to the brain, an inflammatory response is initiated by the microglia in a context-dependent manner which, in turn, secrete pro-inflammatory molecules such as cytokines and chemokines to the site of injury [61–63]. While microglia are required for regeneration in the zebrafish spinal cord [64], inflammation could lead to reactive gliosis and a glial scar formation in the adult brain, which are traditionally thought to be inhibitory to regeneration.

Circulating immune cells are also known to have an inflammatory role upon injury with long-term effects. At homeostasis, the blood-brain barrier regulates infiltration into the brain. However, upon injury or ageing, the barrier could break down, allowing the infiltration of immune cells such as T cells into the brain parenchyma leading to inflammation [65,66]. Furthermore, meningeal immune cells also release cytokines to stimulate cells in the brain [67]. Studies on the SVZ NSCs showed antiproliferative effects of IFNy, suggesting implications for NSC function during regeneration [66]. These findings are strikingly different to what happens in the neonatal brain where the presence of an immature immune system promotes hippocampal neurogenesis and spatial learning processes, following CD4⁺ T cell infiltration [68,69].

The astrocytes represent the most abundant glial cell population in the brain. Under physiological conditions, they are involved in synapse formation, neuronal protection and blood brain barrier support [70,71]. Injury or damage in the brain leads to immune cell infiltration and an increase in oxidative stress, which in turn triggers a signalling cascade that changes the molecular and morphological profile of the local astrocytes,



leading to reactive gliosis and glial scars [72,73]. Whilst being anti-regenerative, the scar represents a defensive mechanism against the expansion of the damage to the surrounding healthy tissue [74].

One of the obvious differences between the developing and adult brains is the composition and the molecular identity of microglia and astrocytes. One hypothesis is that the changes in numbers, morphology, functional and molecular identity could be an underlying reason behind the regenerative developing and non-regenerative adult brain. In addition to the immune and glial microenvironment, brain parenchyma undergoes many other changes after development. For example, changes in extracellular matrix composition and increased stiffness were linked to reduced regeneration by oligodendrocyte progenitor cells [75], suggesting other stem populations could also be affected by such changes. Furthermore, although examples of the integration of new neurons into the brain exist [76,77], once the circuitry is established and mature, there could be inhibitory cues that prevent the integration of new neurons into the circuitry.

In summary, a combination of cell-intrinsic and -extrinsic changes influence the repair capacity of the brain (Figure 1C). Importantly, the age-dependent decline in the regenerative potential is multifactorial and requires innovative combinatorial approaches to overcome various effectors. Heterochronic transplantation and parasymbiosis [78] experiments could provide further insights into the contribution of different cell-intrinsic and extrinsic factors.

Potential therapeutic approaches to facilitate brain regeneration

There are no regenerative therapeutic approaches after an injury or acute cell loss in the brain. Current treatments mainly include therapies that target the after-effects of the injury and therefore do not replace/regenerate the lost neurons [79].

While cell replacement therapies are promising for some degenerative diseases such as Parkinson's Disease [80], transplantation approaches may not always be feasible when the injury spans multiple cell types and large areas. Therefore, direct reprogramming in vivo is emerging as a promising therapeutic approach to stimulate new neuron production. Astrocytes represent the ideal target for in vivo reprogramming, given their potential stem-like properties upon injury [26]. Furthermore, since they drive reactive gliosis upon brain damage, altering astrocyte function and lineage propensity could facilitate new neuron production and alleviate the antiregenerative microenvironment. This has been achieved through overexpression of proneural transcription factors such as NeuroD1, Ascl1 and neurogenin2 which are necessary and sufficient to trigger a neuronal fate in experimental models [81-85]. Overexpression of these transcription factors changes the astrocytic chromatin landscape by opening the silenced proneural genetic loci and therefore activates neuronal genetic programs. For example, ASCL1 induces a rapid global change in the transcriptional program of reprogrammed astrocytes. This is achieved by directly binding to its target genes Klf10, Myt1, Neurod4 and Chd7, required for the efficient conversion of astrocytes into neurons [83]. A crucial consideration following direct reprogramming is to assess whether the correct neural identity is achieved and the safety of the gene delivery approaches. Importantly, while direct reprogramming could short-circuit neuron production, it does not overcome the potential antiregenerative effects of tissue microenvironment. For example, in one study, microglia in the microenvironment have been shown to inhibit the Ascl1-induced retinal regeneration in adult mice [86].

Alternatively, stimulation of endogenous stem/progenitor cells through transient manipulations represents another potential therapeutic approach to overcome the cell-intrinsic and extrinsic regeneration blockages. However, we hypothesise that such approaches should be combinatorial, targeted and transient. After traumatic brain injury, injections of growth factors such as EGF and FGF2 in the ventricle of adult rodents increased the proliferation of NSCs, supporting neuron formation and cognitive recovery [87,88]. Whether providing developmental signals and mitogens could be sufficient to overcome the epigenetic silencing of neural differentiation genes and reprogramme the cells to make neurons remains to be determined. Collectively, these findings highlight the need for combinatorial approaches that tackle cell-intrinsic and -extrinsic factors. Therefore, understanding the molecular and cellular mechanisms regulating injury responses and the effects of age is crucial to fully harnessing the potential of endogenous stem cells.

Conclusions and outlook

Although the adult brain is inefficient at making new neurons after cell loss due to an injury or degeneration, the developing brain and active neurogenic niches exhibit regenerative potential. Regenerative mechanisms



could involve compensatory proliferation of NSCs or adaptive reprogramming of injury-responsive progenitors or astroglia. Nevertheless, these processes are tightly regulated with age, with the majority not observed after development. The decline in regenerative potential is likely an evolutionary trade-off to achieve complex circuitry and higher-order function and to prevent aberrant growth and tumorigenesis.

Our current understanding highlights the need for combinatorial approaches that modulate the adverse effects of the injury microenvironment and reverse the cell-intrinsic changes that accumulate over time. Investigating the regenerative processes within the neurogenic niches and elsewhere in the brain, and, finally, contrasting mechanisms observed in highly regenerative organisms, could provide invaluable insights into how to stimulate neuron regeneration following injury.

Perspectives

- The adult mammalian brain exhibits limited regeneration outside the neurogenic niches.
- Developing brain exhibits repair capacity highlighting the age-dependent decline in the regenerative potential in the brain.
- A combination of cell-intrinsic and -extrinsic factors regulate the regeneration in the brain and understanding these factors is the crucial first step for the design of potential future therapies to facilitate regeneration.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

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Abbreviations

NEP, *Nestin*-expressing progenitor; NSC, neural stem cell; ROS, reactive oxygen species; SGZ, subgranular zone; SHH, sonic hedgehog; SVZ, subventricular zone.

References

- 1 Lust, K. and Tanaka, E.M. (2019) A comparative perspective on brain regeneration in amphibians and teleost fish. Dev. Neurobiol. 79, 424–436 https://doi.org/10.1002/dneu.22665
- 2 Amamoto, R., Huerta, V.G.L., Takahashi, E., Dai, G.P., Grant, A.K., Fu, Z.Y. et al. (2016) Adult axolotls can regenerate original neuronal diversity in response to brain injury. Elife 5, e13998 https://doi.org/10.7554/eLife.13998
- 3 Ramón y Cajal, S. (1928) Degeneration and regeneration of the nervous system. Clarendon Press, Oxford, U.K.
- 4 Altman, J. and Das, G.D. (1966) Autoradiographic and histological studies of postnatal neurogenesis. I. A longitudinal investigation of kinetics migration and transformation of cells incorporating tritiated thymidine in neonate rats with special reference to postnatal neurogenesis in some brain regions.

 J. Comp. Neurol. 126, 337–389 https://doi.org/10.1002/cne.901260302
- 5 Paton, J.A. and Nottebohm, F. (1984) Neurons generated in the adult brain are recruited into functional circuits. Science 225, 1046–1048 https://doi. org/10.1126/science.6474166



- 6 Le Belle, J.E., Orozco, N.M., Paucar, A.A., Saxe, J.P., Mottahedeh, J., Pyle, A.D. et al. (2011) Proliferative neural stem cells have high endogenous ROS levels that regulate self-renewal and neurogenesis in a PI3K/Akt-dependant manner. *Cell Stem Cell* **8**, 59–71 https://doi.org/10.1016/j.stem.2010.11.
- Bujak, M., Ren, G., Kweon, H.J., Dobaczewski, M., Reddy, A., Taffet, G. et al. (2007) Essential role of smad3 in infarct healing and in the pathogenesis of cardiac remodeling. Circulation 116, 2127–2138 https://doi.org/10.1161/CIRCULATIONAHA.107.704197
- 8 Griffin, M.F., Borrelli, M.R., Garcia, J.T., Januszyk, M., King, M., Lerbs, T. et al. (2021) JUN promotes hypertrophic skin scarring via CD36 in preclinical in vitro and in vivo models. *Sci. Transl. Med.* **13**, eabb3312 https://doi.org/10.1126/scitranslmed.abb3312
- 9 Lowry, N., Goderie, S.K., Adamo, M., Lederman, P., Charniga, C., Gill, J. et al. (2008) Multipotent embryonic spinal cord stem cells expanded by endothelial factors and Shh/RA promote functional recovery after spinal cord injury. Exp. Neurol. 209, 510–522 https://doi.org/10.1016/j.expneurol. 2007.09.031
- 10 Luo, F.C., Silver, J. and Luo, Y. (2019) Modulation of receptor protein tyrosine phosphatase sigma promotes neuroregeneration and functional recovery after ischemic stroke. Stroke 50, TMP33 https://doi.org/10.1161/str.50.suppl_1.TMP33
- Walker, S.E. and Echeverri, K. (2022) Spinal cord regeneration the origins of progenitor cells for functional rebuilding. Current Opinion in Genetics & Development 75, 101917 https://doi.org/10.1016/j.gde.2022.101917
- Maki, T., Liang, A.C., Miyamoto, N., Lo, E.H. and Arai, K. (2013) Mechanisms of oligodendrocyte regeneration from ventricular-subventricular zone-derived progenitor cells in white matter diseases. Front Cell Neurosci. 7, 275 https://doi.org/10.3389/fncel.2013.00275
- 13 Krzyspiak, J., Khodakhan, K. and Hébert, J.M. (2022) Potential variables for improved reproducibility of neuronal cell grafts at stroke sites. *Cells* 11, 1656 https://doi.org/10.3390/cells11101656
- 14 Falkner, S., Grade, S., Dimou, L., Conzelmann, K.-K., Bonhoeffer, T., Gotz, M. et al. (2017) Erratum: transplanted embryonic neurons integrate into adult neocortical circuits. *Nature* 541, 122 https://doi.org/10.1038/nature20808
- 15 Krzyspiak, J., Yan, J.Q., Ghosh, H.S.S., Galinski, B., Lituma, P.J.J., Alvina, K. et al. (2022) Donor-derived vasculature is required to support neocortical cell grafts after stroke (vol 59, 102642, 2022). Stem Cell Res. 60, 102700 https://doi.org/10.1016/j.scr.2022.102700
- 16 Kempermann, G., Song, H.J. and Gage, F.H. (2015) Neurogenesis in the adult hippocampus. Cold Spring Harb. Perspect. Biol. 7, a018812 https://doi.org/10.1101/cshperspect.a018812
- Kornack, D.R. and Rakic, P. (1999) Continuation of neurogenesis in the hippocampus of the adult macaque monkey. *Proc. Natl Acad. Sci. U.S.A.* **96**, 5768–5773 https://doi.org/10.1073/pnas.96.10.5768
- Williamson, M.R., Le, S.P., Franzen, R.L., Donlan, N.A., Rosow, J.L., Nicot-Cartsonis, M.S. et al. (2023) Subventricular zone cytogenesis provides trophic support for neural repair in a mouse model of stroke. *Nat. Commun.* **14**, 6341 https://doi.org/10.1038/s41467-023-42138-0
- 19 Wu, N., Sun, X.C., Zhou, C., Yan, J. and Cheng, C.J. (2023) Neuroblasts migration under control of reactive astrocyte-derived BDNF: a promising therapy in late neurogenesis after traumatic brain injury. Stem Cell Res. Ther. 14, 2 https://doi.org/10.1186/s13287-022-03232-0
- Nakatomi, H., Kuriu, T., Okabe, S., Yamamoto, S., Hatano, O., Kawahara, N. et al. (2002) Regeneration of hippocampal pyramidal neurons after ischemic brain injury by recruitment of endogenous neural progenitors. Cell 110, 429–441 https://doi.org/10.1016/S0092-8674(02)00862-0
- 21 Klein, B., Mrowetz, H., Kreutzer, C., Rotheneichner, P., Zaunmair, P., Lange, S. et al. (2020) DCX+ neuronal progenitors contribute to new oligodendrocytes during remyelination in the hippocampus. *Sci. Rep.* **10**, 20095 https://doi.org/10.1038/s41598-020-77115-w
- 22 Battistini, J., Mastrorilli, V., di Robilant, V., Saraulli, D., Marinelli, S. and Vecchioli, S. (2023) Role of running-activated neural stem cells in the anatomical and functional recovery after traumatic brain injury in p21 knock-out mice. *Int. J. Mol. Sci.* **24**, 2911 https://doi.org/10.3390/jims24032911
- 23 Cameron, H. and McKay, R. (2001) Adult neurogenesis produces a large pool of new granule cells in the dentate gyrus. J. Comp. Neurol. 435, 406–417 https://doi.org/10.1002/cpe.1040
- 24 Jin, K., Sun, Y.J., Xie, L., Peel, A., Mao, X.O., Batteur, S. et al. (2003) Directed migration of neuronal precursors into the ischemic cerebral cortex and striatum. *Mol. Cell. Neurosci.* **24**, 171–189 https://doi.org/10.1016/S1044-7431(03)00159-3
- 25 Kunze, A., Achilles, A., Keiner, S., Witte, O.W. and Redecker, C. (2015) Two distinct populations of doublecortin-positive cells in the perilesional zone of cortical infarcts. BMC Neurosci. 16, 20 https://doi.org/10.1186/s12868-015-0160-8
- 26 Sirko, S., Behrendt, G., Johansson, P.A., Tripathi, P., Costa, M., Bek, S. et al. (2013) Reactive glia in the injured brain acquire stem cell properties in response to sonic hedgehog. [corrected]. *Cell Stem Cell* **12**, 426–439 https://doi.org/10.1016/j.stem.2013.01.019
- Heng, X., Guo, Q., Leung, A.W. and Li, J.Y. (2017) Analogous mechanism regulating formation of neocortical basal radial glia and cerebellar Bergmann glia. Elife 6, e23253 https://doi.org/10.7554/eLife.23253
- Wojcinski, A., Lawton, A.K., Bayin, N.S., Lao, Z.M., Stephen, D.N. and Joyner, A.L. (2017) Cerebellar granule cell replenishment postinjury by adaptive reprogramming of Nestin+ progenitors. *Nat. Neurosci.* **20**, 1361 https://doi.org/10.1038/nn.4621
- 29 Bayin, N.S., Wojcinski, A., Mourton, A., Saito, H., Suzuki, N. and Joyner, A.L. (2018) Age-dependent dormant resident progenitors are stimulated by injury to regenerate Purkinje neurons. Elife 7, e39879 https://doi.org/10.7554/eLife.39879
- 30 Altman, J., Anderson, W.J. and Wright, K.A. (1969) Early effects of X-irradiation of cerebellum in infant rats decimation and reconstitution of external granular layer. *Exp. Neurol.* **24**, 196 https://doi.org/10.1016/0014-4886(69)90015-6
- 31 Feddersen, R.M., Ehlenfeldt, R., Yunis, W.S., Clark, H.B. and Orr, H.T. (1992) Disrupted cerebellar cortical development and progressive degeneration of Purkinje-cells in SV40-T antigen transgenic mice. *Neuron* **9**, 955–966 https://doi.org/10.1016/0896-6273(92)90247-B
- 32 Bayin, N.S., Mizrak, D., Stephen, D.N., Lao, Z.M., Sims, P.A. and Joyner, A.L. (2021) Injury-induced ASCL1 expression orchestrates a transitory cell state required for repair of the neonatal cerebellum. *Sci. Adv.* 7, eabj1598 https://doi.org/10.1126/sciadv.abj1598
- 33 Jorstad, N.L., Wilken, M.S., Todd, L., Finkbeiner, C., Nakamura, P., Radulovich, N. et al. (2020) STAT signaling modifies Ascl1 chromatin binding and limits neural regeneration from muller glia in adult mouse retina. Cell Rep. 30, 2195–2208 https://doi.org/10.1016/j.celrep.2020.01.075
- 34 Karl, M.O., Hayes, S., Nelson, B.R., Tan, K., Buckingham, B. and Reh, T.A. (2008) Stimulation of neural regeneration in the mouse retina. *Proc. Natl Acad. Sci. U.S.A.* **105**, 19508–19513 https://doi.org/10.1073/pnas.0807453105
- Pollak, J., Wilken, M.S., Ueki, Y., Cox, K.E., Sullivan, J.M., Taylor, R.J. et al. (2013) ASCL1 reprograms mouse Muller glia into neurogenic retinal progenitors. *Development* **140**, 2619–2631 https://doi.org/10.1242/dev.091355
- 36 Löffler, K., Schäfer, P., Völkner, M., Holdt, T. and Karl, M. (2015) Age-dependent Muller glia neurogenic competence in the mouse retina. Glia 63, 1809–1824 https://doi.org/10.1002/glia.22846



- 37 Bielefeld, P., Abbink, M.R., Davidson, A.R., Reijner, N., Abiega, O., Lucassen, P.J. et al. (2021) Early life stress decreases cell proliferation and the number of putative adult neural stem cells in the adult hypothalamus. Stress 24, 189–195 https://doi.org/10.1080/10253890.2021.1879787
- 38 Infantes-López, M.I., Nieto-Quero, A., Chaves-Peña, P., Zambrana-Infantes, E., Cifuentes, M., Márquez, J. et al. (2023) New insights into hypothalamic neurogenesis disruption after acute and intense stress: implications for microglia and inflammation (vol 17, 1190418, 2023). Front. Neurosci. 17, 1335034 https://doi.org/10.3389/fnins.2023.1335034
- 39 Mu, W.H., Li, S., Xu, J.K., Guo, X.Z., Wu, H.D., Chen, Z.H. et al. (2021) Hypothalamic Rax+ tanycytes contribute to tissue repair and tumorigenesis upon oncogene activation in mice. *Nat. Commun.* **12**, 2288 https://doi.org/10.1038/s41467-021-22640-z
- 40 Lopez-Otin, C., Blasco, M.A., Partridge, L., Serrano, M. and Kroemer, G. (2023) Hallmarks of aging: an expanding universe. *Cell* **186**, 243–278 https://doi.org/10.1016/j.cell.2022.11.001
- 41 Lin, Y., Yang, J., Shen, Z., Ma, J., Simons, B.D. and Shi, S.H. (2021) Behavior and lineage progression of neural progenitors in the mammalian cortex. *Curr. Opin. Neurobiol.* **66**, 144–157 https://doi.org/10.1016/j.conb.2020.10.017
- 42 Benayoun, B.A., Pollina, E.A., Singh, P.P., Mahmoudi, S., Harel, I., Casey, K.M. et al. (2019) Remodeling of epigenome and transcriptome landscapes with aging in mice reveals widespread induction of inflammatory responses. *Genome Res.* **29**. 697–709 https://doi.org/10.1101/gr.240093.118
- 43 Kalamakis, G., Brüne, D., Ravichandran, S., Bolz, J., Fan, W.Q., Ziebell, F. et al. (2019) Quiescence modulates stem cell maintenance and regenerative capacity in the aging brain. *Cell* **176**, 1407 https://doi.org/10.1016/j.cell.2019.01.040
- 44 Maybury-Lewis, S.Y., Brown, A.K., Yeary, M., Sloutskin, A., Dhakal, S., Juven-Gershon, T. et al. (2021) Changing and stable chromatin accessibility supports transcriptional overhaul during neural stem cell activation and is altered with age. *Aging Cell* **20**, e13499 https://doi.org/10.1111/acel.13499
- 45 Lupo, G., Nisi, P.S., Esteve, P., Paul, Y.L., Novo, C.L., Sidders, B. et al. (2018) Molecular profiling of aged neural progenitors identifies Dbx2 as a candidate regulator of age-associated neurogenic decline. *Aging Cell* 17, e12745 https://doi.org/10.1111/acel.12745
- 46 Li, M.Y., Guo, H.Z., Carey, M. and Huang, C.Y. (2024) Transcriptional and epigenetic dysregulation impairs generation of proliferative neural stem and progenitor cells during brain aging. *Nat. Aging* **4**, 62–79 https://doi.org/10.1038/s43587-023-00549-0
- 47 Ma, C.Y., Yao, M.J., Zhai, Q.W., Jiao, J.W., Yuan, X.B. and Poo, M.M. (2014) SIRT1 suppresses self-renewal of adult hippocampal neural stem cells. Development 141, 4697–4709 https://doi.org/10.1242/dev.117937
- 48 Wani, G.A., Sprenger, H.G., Ndoci, K., Chandragiri, S., Acton, R.J., Schatton, D. et al. (2022) Metabolic control of adult neural stem cell self-renewal by the mitochondrial protease YME1L. *Cell Rep.* **38**, 110370 https://doi.org/10.1016/j.celrep.2022.110370
- 49 Khacho, M., Clark, A., Svoboda, D.S., Azzi, J., MacLaurin, J.G., Meghaizel, C. et al. (2016) Mitochondrial dynamics impacts stem cell identity and fate decisions by regulating a nuclear transcriptional program. *Cell Stem Cell* 19, 232–247 https://doi.org/10.1016/j.stem.2016.04.015
- 50 Beckervordersandforth, R., Ebert, B., Schäffner, I., Moss, J., Fiebig, C., Shin, J. et al. (2017) Role of mitochondrial metabolism in the control of early lineage progression and aging phenotypes in adult hippocampal neurogenesis (vol 93, pg 560, 2017). *Neuron* **93**, 1518–1518 https://doi.org/10.1016/ineuron.2017.03.008
- 51 Wang, W., Osenbroch, P., Skinnes, R., Esbensen, Y., Bjorås, M. and Eide, L. (2010) Mitochondrial DNA integrity is essential for mitochondrial maturation during differentiation of neural stem cells. *Stem Cells* **28**, 2195–2204 https://doi.org/10.1002/stem.542
- 52 Hämäläinen, R.H., Ahlqvist, K.J., Ellonen, P., Lepistö, M., Logan, A., Otonkoski, T. et al. (2015) mtDNA mutagenesis disrupts pluripotent stem cell function by altering redox signaling. *Cell Rep.* **11**, 1614–1624 https://doi.org/10.1016/j.celrep.2015.05.009
- 53 Ali, A.A.H., Schwarz-Herzke, B., Stahr, A., Prozorovski, T., Aktas, O. and von Gall, C. (2015) Premature aging of the hippocampal neurogenic niche in adult Bmal1-deficient mice. *Aging* **7**, 435–449 https://doi.org/10.18632/aging.100764
- 54 Prozorovski, T., Schulze-Topphoff, U., Glumm, R., Baumgart, J., Schröter, F., Ninnemann, O. et al. (2008) Sirt1 contributes critically to the redox-dependent fate of neural progenitors. Nat. Cell Biol. 10, 385–U335 https://doi.org/10.1038/ncb1700
- 55 Burda, J.E. and Sofroniew, M.V. (2014) Reactive gliosis and the multicellular response to CNS damage and disease. *Neuron* **81**, 229–248 https://doi.org/10.1016/j.neuron.2013.12.034
- 56 Escartin, C., Galea, E., Lakatos, A., O'Callaghan, J.P., Petzold, G.C., Serrano-Pozo, A. et al. (2021) Reactive astrocyte nomenclature, definitions, and future directions. *Nat. Neurosci.* **24**, 312–325 https://doi.org/10.1038/s41593-020-00783-4
- 57 Alliot, F., Godin, I. and Pessac, B. (1999) Microglia derive from progenitors, originating from the yolk sac, and which proliferate in the brain. *Dev. Brain Res.* **117**, 145–152 https://doi.org/10.1016/S0165-3806(99)00113-3
- 58 Tan, Y.L., Yuan, Y. and Tian, L. (2020) Microglial regional heterogeneity and its role in the brain. *Mol. Psychiatry* **25**, 351–367 https://doi.org/10.1038/s41380-019-0609-8
- 59 Park, D.S., Kozaki, T., Tiwari, S.K., Moreira, M., Khalilnezhad, A., Torta, F. et al. (2023) iPS-cell-derived microglia promote brain organoid maturation via cholesterol transfer. *Nature* **623**, 397–405 https://doi.org/10.1038/s41586-023-06713-1
- 60 Huang, Y., Liu, B., Sinha, S.C., Amin, S. and Gan, L. (2023) Mechanism and therapeutic potential of targeting cGAS-STING signaling in neurological disorders. Mol. Neurodegener. 18, 79 https://doi.org/10.1186/s13024-023-00672-x
- 61 Damani, M.R., Zhao, L.A., Fontainhas, A.M., Amaral, J., Fariss, R.N. and Wong, W.T. (2011) Age-related alterations in the dynamic behavior of microglia. *Aging Cell* **10**, 263–276 https://doi.org/10.1111/j.1474-9726.2010.00660.x
- 62 Yanguas-Casás, N., Crespo-Castrillo, A., Arevalo, M.A. and Garcia-Segura, L.M. (2020) Aging and sex: impact on microglia phagocytosis. *Aging Cell* 19, e13182 https://doi.org/10.1111/acel.13182
- 63 Purkayastha, S. and Cai, D.S. (2013) Disruption of neurogenesis by hypothalamic inflammation in obesity or aging. *Rev. Endocr. Metab. Disorders* **14**, 351–356 https://doi.org/10.1007/s11154-013-9279-z
- 64 Morsch, M., Radford, R., Lee, A., Don, E.K., Bedrock, A.P., Hall, T.E. et al. (2015) In vivo characterization of microglial engulfment of dying neurons in the zebrafish spinal cord. *Front. Cell. Neurosci.* **9**, 321 https://doi.org/10.3389/fncel.2015.00321
- 65 Montagne, A., Barnes, S.R., Sweeney, M.D., Halliday, M.R., Sagare, A.P., Zhao, Z. et al. (2015) Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* **85**, 296–302 https://doi.org/10.1016/j.neuron.2014.12.032
- 66 Dulken, B., Buckley, M.T., Negredo, P.N., Saligrama, N., Cayrol, R., Leeman, D.S. et al. (2019) Single-cell analysis reveals T cell infiltration in old neurogenic niches. *Nature* 571, 205 https://doi.org/10.1038/s41586-019-1362-5
- 67 Derecki, N.C., Cardani, A.N., Yang, C.H., Quinnies, K.M., Crihfield, A., Lynch, K.R. et al. (2010) Regulation of learning and memory by meningeal immunity: a key role for IL-4. *J. Exp. Med.* **207**, 1067–1080 https://doi.org/10.1084/jem.20091419



- Wolf, S.A., Steiner, B., Akpinarli, A., Kammertoens, T., Nassenstein, C., Braun, A. et al. (2009) CD4-positive t lymphocytes provide a neuroimmunological link in the control of adult hippocampal neurogenesis. *J. Immunol.* **182**, 3979–3984 https://doi.org/10.4049/jimmunol.0801218
- 69 Radjavi, A., Smirnov, I. and Kipnis, J. (2014) Brain antigen-reactive CD4+ T cells are sufficient to support learning behavior in mice with limited T cell repertoire. *Brain Behav. Immun.* **35**, 58–63 https://doi.org/10.1016/j.bbi.2013.08.013
- 70 Wang, H., Xu, Z.Y., Xia, Z.Y., Rallo, M., Duffy, A. and Matise, M.P. (2021) Inactivation of Hedgehog signal transduction in adult astrocytes results in region-specific blood-brain barrier defects. *Proc. Natl Acad. Sci. U.S.A.* 118, e2017779118 https://doi.org/10.1073/pnas.2017779118
- Hasel, P., Dando, O., Jiwaji, Z., Baxter, P., Todd, A.C., Heron, S. et al. (2018) Neurons and neuronal activity control gene expression in astrocytes to regulate their development and metabolism (vol 8. 15132, 2017). *Nat. Commun.* **9.** 16176 https://doi.org/10.1038/ncomms16176
- 72 Clarke, L.E., Liddelow, S.A., Chakraborty, C., Münch, A.E., Heiman, M. and Barres, B. (2018) Normal aging induces A1-like astrocyte reactivity. *Proc. Natl Acad. Sci. U.S.A.* **115**, E1896–E1905 https://doi.org/10.1073/pnas.1800165115
- 73 Sierra, A., Gottfried-Blackmore, A.C., McEwen, B.S. and Bulloch, K. (2007) Microglia derived from aging mice exhibit an altered inflammatory profile. Glia 55, 412–424 https://doi.org/10.1002/glia.20468
- 74 Herrmann, J.E., Imura, T., Song, B.B., Qi, J.W., Ao, Y., Nguyen, T.K. et al. (2008) STAT3 is a critical regulator of astrogliosis and scar formation after spinal cord injury. *J. Neurosci.* 28, 7231–7243 https://doi.org/10.1523/JNEUROSCI.1709-08.2008
- 75 Segel, M., Neumann, B., Hill, M.F.E., Weber, I.P., Viscomi, C., Zhao, C. et al. (2019) Niche stiffness underlies the ageing of central nervous system progenitor cells. *Nature* **573**, 130–134 https://doi.org/10.1038/s41586-019-1484-9
- 76 Wernig, M., Zhao, J.P., Pruszak, J., Hedlund, E., Fu, D.D., Soldner, F. et al. (2008) Neurons derived from reprogrammed fibroblasts functionally integrate into the fetal brain and improve symptoms of rats with Parkinson's disease. *Proc. Natl Acad. Sci. U.S.A.* **105**, 5856–5861 https://doi.org/10.1073/pnas. 0801677105
- 77 Wijeyekoon, R. and Barker, R.A. (2009) Cell replacement therapy for Parkinson's disease. Biochim. Biophys. Acta 1792, 688–702 https://doi.org/10.1016/j.bbadis.2008.10.007
- 78 Katsimpardi, L., Litterman, N.K., Schein, P.A., Miller, C.M., Loffredo, F.S., Wojtkiewicz, G.R. et al. (2014) Vascular and neurogenic rejuvenation of the aging mouse brain by young systemic factors. *Science* **344**, 630–634 https://doi.org/10.1126/science.1251141
- 79 Schepici, G., Silvestro, S., Bramanti, P. and Mazzon, E. (2020) Traumatic brain injury and stem cells: an overview of clinical trials, the current treatments and future therapeutic approaches. *Medicina* **56**, 137 https://doi.org/10.3390/medicina56030137
- Kirkeby, A., Nelander, J., Hoban, D., Rogelius, N., Bjartmarz, H., Storm, P. et al. (2023) Preclinical quality, safety, and efficacy of a human embryonic stem cell-derived product for the treatment of Parkinson's disease, STEM-PD. *Cell Stem Cell* 30, 1299–1314.e9 https://doi.org/10.1016/j.stem.2023.
- 81 Eguchi, R., Hamano, M., Iwata, M., Nakamura, T., Oki, S. and Yamanishi, Y. (2022) TRANSDIRE: data-driven direct reprogramming by a pioneer factor-guided trans-omics approach. *Bioinformatics* **38**, 2839–2846 https://doi.org/10.1093/bioinformatics/btac209
- 82 Guo, Z.Y., Zhang, L., Wu, Z., Chen, Y.C., Wang, F. and Chen, G. (2014) In vivo direct reprogramming of reactive glial cells into functional neurons after brain injury and in an Alzheimer's disease model. *Cell Stem Cell* 14, 188–202 https://doi.org/10.1016/j.stem.2013.12.001
- 83 Rao, Z.P., Wang, R., Sanlan, L., Shi, Y.H., Licun, M., Su'e, H. et al. (2021) Molecular mechanisms underlying Ascl1-mediated astrocyte-to-neuron conversion. Stem Cell Rep. 16, 534–547 https://doi.org/10.1016/j.stemcr.2021.01.006
- 84 Liu, F., Zhang, Y.J., Chen, F.L., Yuan, J.C., Li, S.L., Han, S. et al. (2021) Neurog2 directly converts astrocytes into functional neurons in midbrain and spinal cord. Cell Death Dis. 12, 225 https://doi.org/10.1038/s41419-021-03498-x
- 85 Berninger, B., Costa, M.R., Koch, U., Schroeder, T., Sutor, B., Grothe, B. et al. (2007) Functional properties of neurons derived from in vitro reprogrammed postnatal astroglia. *J. Neurosci.* **27**, 8654–8664 https://doi.org/10.1523/JNEUROSCI.1615-07.2007
- 86 Todd, L., Finkbeiner, C., Wong, C.K., Hooper, M.J. and Reh, T.A. (2020) Microglia suppress Ascl1-induced retinal regeneration in mice. *Cell Rep.* 33, 108507 https://doi.org/10.1016/j.celrep.2020.108507
- 87 Sun, D., Bullock, M.R., Altememi, N., Zhou, Z.W., Hagood, S., Rolfe, A. et al. (2010) The effect of epidermal growth factor in the injured brain after trauma in rats. *J. Neurotrauma* 27, 923–938 https://doi.org/10.1089/neu.2009.1209
- 88 Sun, D., Bullock, M.R., McGinn, M.J., Zhou, Z., Altememi, N., Hagood, S. et al. (2009) Basic fibroblast growth factor-enhanced neurogenesis contributes to cognitive recovery in rats following traumatic brain injury. *Exp. Neurol.* **216**, 56–65 https://doi.org/10.1016/j.expneurol.2008.11.011