

RESEARCH ARTICLE

Open Access

Protective role of heparin in the injury of the liver and kidney on the experimental model of ischemia/reperfusion

Ali Ümit Yener^{1,7*}, Mustafa Cüneyt Çiçek², Serhat Bahadır Genç¹, Turgut Özkan⁷, Emre Doğan³,
Bülent Çağlar Bilgin⁴, Tezcan Akın⁴, Havva Erdem⁵ and Handan Ankaralı⁶

Abstract

Background: Surgery of thoracoabdominal aortic aneurysms (TAAA) is associated with high incidence of serious complications. Ischemia/reperfusion (I/R) injury may be responsible for these complications. We investigated the effect of degree of anticoagulation on remote organ I/R injuries and whether heparin is protective against I/R injury in addition to its anticoagulant properties.

Methods: Sprague Dawley rats were used to determine both liver and kidney concentrations of HSP-70, IL-6, MPO in four groups: ischemic control (operation with cross-clamping and intraperitoneal administration of 0.9% saline, n = 7), sham (operation without cross-clamping, n = 7), heparin (ACT level about 200), and high dose heparin (ACT level up to 600). Histological analyses of the organs were performed.

Results: Histopathological evaluation of kidney presented significant differences between groups with regards to the cytoplasmic vacuole formation, hemorrhage, tubular cell degeneration and tubular dilatation while heparinized group had best results. The kidney MPO and HSP-70 levels significantly decreased ($p < 0.05$), but IL-6 level was not significant ($p > 0.05$) in heparinized group when compared to ischemic control group. No statistically significant intergroup differences were detected in the tissue samples of liver. Immunohistochemical markers of the liver were compared and no statistically significant difference was found among the groups.

Conclusion: Heparin is an important anticoagulation agent in TAAA surgical procedures but the use of higher levels of heparin in the present study revealed no beneficial effects. Bleeding complications is much less when heparin is used in the real-world clinical practice as ACT levels of 200.

Keywords: Heparin, Ischemia, Reperfusion, Lung, Kidney

Background

Surgical repair of thoracoabdominal aortic aneurysms (TAAAs) are highly complex and challenging interventions. It is not surprising that this operation is associated with a high incidence of serious complications, often leading to respiratory failure, renal failure, neurological deficits or even death.

Multiorgan dysfunction, presumably related to activation of inflammatory pathways and over production of

cytokines and other inflammatory mediators, is a major cause of death. The systemic inflammatory response syndrome (SIRS), ischemia-reperfusion injury (I/Ri), and inflammatory pathway activation are considered precursors of morbidity and mortality after open TAAA repair [1-3].

Cytokines such as IL-6 are important mediators of the inflammatory response in ischemia. Plasma IL-6 concentration increased gradually, reached its peak 24 hours after surgery, and correlated with aortic cross-clamp time and morbidity. The observed correlation between plasma IL-6 and aortic cross-clamp time supports the hypothesis that the IL-6 response mainly reflects the impact of ischemia and reperfusion rather than the impact of surgical trauma in elective TAAA surgery. The strong

* Correspondence: yener@comu.edu.tr

¹Department of Cardiovascular Surgery, Ankara Numune Education and Research Hospital, 06330 Ankara, Turkey

⁷Department of Cardiovascular Surgery, Çanakkale Onsekiz Mart University, Kepez, Çanakkale, Turkey

Full list of author information is available at the end of the article

correlation between plasma IL-6 and morbidity in previous reports suggesting that plasma IL-6 is a reliable predictor of outcome in elective TAAA repair [4-6].

Heat shock proteins (HSPs) are cellular stress proteins which have been shown to have an important role for the survival of cells under stress conditions [7]. Zhang et al. [8] pointed out that, HSP70 could respond to a wide variety of stress conditions such as ischemia, and inflammation. It can prevent the irreversible denaturation of proteins [9]. Elevated expression of HSP70 can prevent cell death processes [10]. It has been showed that overexpression of HSP70 attenuate the release of inflammatory factors and interferes with the process of apoptotic cell death [10-12].

Myeloperoxidase (MPO) is one of the obvious indicators for the tissue infiltration of neutrophilic granulocytes. MPO activity increases in response to the I/R injury [13].

Heparin continues to dominate anticoagulation therapy for surgical repair of TAAA and other cardiovascular surgical procedures. Heparin is best recognized for its ability to prevent blood coagulation by catalytically accelerating the interaction of antithrombin III (AT III) with thrombin, as well as with factors XIIa, IXa, VIIa, and Xa, thereby inhibiting the proteases necessary for completion of the coagulation cascade [14].

Beside heparin is known to possess effects independent of its anticoagulant activity, it plays a role in inhibition of leukocyte-mediated damage [15,16]. Other important and often unrecognized actions of heparin and other glycosaminoglycans are their ability to inhibit complement activation [17] and to exhibit anti-inflammatory effects [18-20].

In this study, we investigated the effect of the degree of anticoagulation on remote organ I/R injury and whether heparin is protective against I/R injury beside its anticoagulant properties.

Methods

Animal and surgical procedure

This study was approved by the Animal Experiment Committee of Düzce University Graduate School of Medicine and all animal care and use were in accordance with the European Convention for Animal Care. 28 male Sprague–Dawley rats weighing 250 to 350 g were used for the experiment. Rats were fed standard laboratory diets and were maintained in a temperature- and photoperiod-controlled (12 hr/day) room. The rats were followed for 15 days before the procedure. None of the animals had any abnormality before operation.

Study groups

The animals were randomly divided into four groups (7 rats each). 1) Sham-operation group. The operation was performed in the same form, but without aortic occlusion and heparin administration. 2) Non-heparinized control group rats' abdominal aortas were clamped for

45 minutes. 3) Low-dose heparin treated group animals' aortas were cross-clamped for 45 minutes and 400 IU/kg of heparin was administered. ACT was kept around 200 during the procedure. 4) High-dose heparin treated group. In this group rats' aortas were cross-clamped for 45 minutes and 800 IU/kg of heparin was administered. ACT was kept around 600 during the procedure.

Operative procedure and technique

Premedication was intraperitoneally applied with ketamine (50 mg/kg) and xylazine (5 mg/kg) to rats. Anesthesia was continued by intermittent injections of ketamine, without endotracheal intubation and mechanical ventilation. Temperature probe was placed in the rectum.

Rats were placed in supine position. After sterile preparation of the surgical area, a standard midline laparotomy incision was made, and after the intestines were retracted the abdominal aorta was explored transperitoneally. The abdominal aorta at the below renal arteries and above the iliac bifurcation levels was cross-clamped during surgical procedures. The duration of the ischemic insult was 45 minutes. Temperature was maintained between 36.5 and 37.5°C during the procedure. Following removal of the aortic clamps, abdominal wall was repaired by using 5/0 polypropylene suture. Pentobarbital (20 mmg/kg) was used as anesthetic at the 48th hour and all the animals were sacrificed. The liver and kidney was dissected totally and fixed in buffered formalin for 7 days.

Histopathological assessment

The liver and kidney were removed and fixed in 10% formalin for histopathologic examination. About 4 µm thick paraffin sections were cut and collected on glass slides. 5 µm thick sections were located on polylysine-coated slides and were stained with hematoxylin and eosin (H & E). The glass slides were examined by light microscopy (Olympus BX51; Olympus Corp. Tokyo, Japan) at 400× magnification.

Immunohistochemical assessment

Blood samples were obtained directly from the cardiac of groups of four rats at the end of 48 h immediately before the sacrifice. Blood samples were obtained with sterile 10 ml syringe after the chest wall was cleansed with chlorohexidine in spirit. Blood samples for cytokine assay were collected into heparinized (20 unit/ml blood) sterile tubes and immediately transferred on ice to be centrifuged at 2000 rpm (at 4°C) for 10 minutes and stored (-70°C) until the time of assay for IL-6, MPO and HSP-70. These marker levels were obtained from blood samples together with kidney and liver tissue.

Analysis of HSP

4 µm thick paraffin sections were prepared. Deparaffinization and hydration of tissue sections was performed in

xylens and graded alcohol. The sections were incubated with primary anti-HSP70 (clone BRM.22, dilution 1/80, Biogenex, San Ramon, California) diluted in buffer. Negative control was PBS.

Analysis of IL-6

Immunohistochemical detection of IL-6 receptor was performed with polyclonal anti-human IL-6 receptor antibody C-20 (Santa Cruz Biotechnologies, Santa Cruz, CA, USA). The antibody was diluted 1: 20. Streptavidin-biotin-peroxidase protocol was applied in IL-6 receptor immunostaining. The secondary anti-rabbit antibody was diluted 1: 500. After the first antibody was omitted negative controls were conducted.

Analysis of MPO

Immuno-histochemical evaluation of MPO activities of the liver and kidney tissues was performed with an anti-MPO kit according to the manufacturer's protocol. Deparaffinization and hydration of samples on polylysine-coated slides was performed. Then, the microwave antigen retrieval procedure was performed, and the samples were incubated in a 3% H₂O₂ solution to inhibit endogenous peroxidase. The sections were incubated with a blocking solution for blocking nonspecific background staining. Then incubation of the sections with primary anti-MPO antibody and with biotinylated goat anti-mouse antibody followed. After incubation with chromogenic substrate (DAB), the sections were counterstained with hematoxylin and eosin (H & E). The slides were examined under a light microscope and two pathologists who did all analyses were blinded to group assignments. The staining of cytoplasmic MPO in the neutrophils was evaluated, and the results were expressed as the percentage of neutrophils cytoplasmically stained positive for MPO. Tissues with no evidence of staining, or only rare scattered positive cells, less than 3%, were recorded as negative. The immunohistochemical results were evaluated for intensity and frequency of staining. The intensity of staining was graded as 0 (negative), 1 (weak), 2 (moderate), and 3 (strong). The frequency was graded from 0 to 4 by the percentage of positive cells as follows: grade 0, <3%; grade 1, 3-25%; grade 2, 25-50%; grade 3, 50-75%; grade 4, more than 75%. The index score was the product of multiplication of the intensity and frequency grades, which was then classified into a 4 point scale: index score 0 = product of 0, index score 1 = products 1 and 2, index score 2 = products 3 and 4, index score 3 = products 6 through 12.

Statistical analysis

Statistical analysis and calculations were performed with SPSS 15 for Windows (Chicago, IL). Results were expressed as the mean (standard error mean). Intergroup differences are detected by Kruskal-Wallis analysis of variance and

Mann-Whitney U test was used for statistical comparisons. While p value of < .01 was accepted as statistically significant p value of < .05 was accepted as statistically less significant.

Results

There was no significant difference with regards to body temperature, mean arterial blood pressure and heart rate among the groups.

Histopathological evaluation of the tissue samples taken from the kidney showed significant intergroup differences in terms of the cytoplasmic vacuole formation, hemorrhage, tubular cell degeneration and tubular dilatation ($P = 0.031$, $P = 0.003$, $P = 0.007$, $P = 0.011$ respectively).

There was no cytoplasmic vacuole formation in the kidney specimens of high dose-heparinized group, while cytoplasmic vacuole formation was evident in the other animals' kidney; as shown in Table 1.

As depicted in Table 2, heparinized group of the examination compared with other groups showed no hemorrhage according to histopathologic evaluation. Hemorrhage was worst in the ischemic control group followed by sham and high dose-heparinized group. Hemorrhage in the kidney specimens of high dose-heparinized group may be due to high levels of ACT (Table 1).

Examination of the kidney tissue revealed that ischemic control group showed most tubular cell degeneration according to histopathologic evaluation ($P = 0.007$); whereas there was no degeneration in the high dose-heparinized group.

There was no tubular dilatation in the kidney specimens of sham and heparinized groups while tubular dilatation was worst in the ischemic control group ($P = 0.011$).

Immunohistochemical markers as, MPO, HSP-70 and IL-6 parameters were studied from the kidney tissue samples. While there was no intergroup difference with regards to IL-6 stain ($p = 0.205$), there were statistical significant differences among the groups with regards to MPO and HSP-70 stains (p value 0.01 and 0.004, respectively) (Table 2). Grade 1, 2 and 3 staining with MPO was strongest in the ischemic control group and high dose-heparinized group whereas there was no staining in the heparinized group ($P = 0.01$). The lowest HSP-70 level was measured in the sham group, after heparinized groups while the highest HSP-70 level was measured in the high dose-heparinized and ischemic control group. IL6 staining results are given in Table 2. There was no significant difference found among the groups ($P = 0.205$).

Histopathological evaluation of the tissue samples taken from the liver showed no statistically significant intergroup differences in terms of the hepatocyte degeneration, hepatocyte steatosis, single cell necrosis, inflammatory cell, sinusoidal dilatation and periportal bridging necrosis ($P = 0.587$, $P = 0.277$, and $P = 0.196$ respectively) (Table 3).

cellular protection. We could not find a significant difference between the groups in terms of the degree of inflammatory response, degree of IL6 kidney tissue.

Cytoplasmic vacuole formation, hemorrhage, tubular cell degeneration and tubular dilatation in the kidney tissue after I/R were decreased after heparin treatment and the histopathologic protective effect of heparin was shown in the present study.

Comparison of immunohistochemical markers and histopathological evaluation of the ischemia-reperfusion injury of the liver tissue in an aortic occlusion model of rats revealed that there was no statistically significant difference. The reason for this outcome for us that; the afferent blood supply to the liver arises from two sources: (i) the hepatic artery, which carries oxygenated blood and accounts for approximately 25% of hepatic blood flow and 50% of total oxygen, and (ii) the portal vein, which drains the splanchnic circulation and accounts for approximately 75% of hepatic blood flow. Because of its protection by a dual blood supply and the capacity for anaerobic metabolism of glycogen stored in the liver, hypoxic damage can occur later in this organ [35].

The half-life of heparin is approximately 30 minutes, 60 minutes and 150 minutes respectively following IV bolus of 25 U/kg, 100 U/kg and 400 U/kg [36]. Our clinical routine in abdominal aortic surgery is to administer bolus of 100 U/kg heparin and to maintain ACT level around 200 seconds. In the present study there was no significant difference between heparin groups (ACT: 200 and ACT: 600) in terms of histopathologic changes or biochemical inflammatory response in the liver and kidney. Also, keeping ACT level around 200 sec during thoracoabdominal surgery seems both to ensure the adequate level of anticoagulation and to avoid the adverse effects such as bleeding.

Conclusions

Many current analysis clearly evidences that the surgical repair of TAAA remains a challenge even in this century. Recent major progress in our understanding of the pathophysiology and operative strategy have decreased the risk of open TAAA repair complications but obviously further investigation is necessary. Heparin is an important anticoagulation agent in thoracoabdominal and abdominal surgical procedures but no beneficial effects were seen with the use of higher levels of heparin in the present study. Taking into account that, bleeding complications will be much less when heparin used in the real-world clinical practice as ACT levels of 200. Furthermore, the results of this investigation indicate that heparin molecule has provide to potential anti-inflammatory property and renal protective action.

Abbreviations

TAAA: Thoracoabdominal aortic aneurysm; IL-6: Interleukin-6; MPO: Myeloperoxidase; HSP-70: Heat shock protein 70; ACT: Activated

clotting time; SIRS: Systemic inflammatory response syndrome; AT III: Antithrombin III; I/R: Ischemia reperfusion; I/Ri: Ischemia reperfusion injury; H & E: Hematoxylin and eosin.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AÜY and MCÇ carried out the design and conduction of the study. SBG, BÇB and ED participated in the design of the study. HE carried out the histopathological examination. HA and TA performed the statistical analysis. TÖ revised and improved the last version of the text. All authors read and approved the final manuscript.

Acknowledgments

We would like to thank Dr. Ömer Faruk Çiçek for his help in revising the manuscript critically for important intellectual content. The authors received no financial support for the research, authorship, and/or publication of this article.

Author details

¹Department of Cardiovascular Surgery, Ankara Numune Education and Research Hospital, 06330 Ankara, Turkey. ²Department of Cardiovascular Surgery, Nevşehir State Hospital, 50200 Nevşehir, Turkey. ³Department of Cardiovascular Surgery, Ahi Evren Education and Research Hospital, 61040 Trabzon, Turkey. ⁴Department of General Surgery, Ankara Numune Education and Research Hospital, 06330 Ankara, Turkey. ⁵Department of Histopathology, Medical Faculty, Duzce University, Konuralp, Duzce, Turkey. ⁶Department of Biostatistics, Medical Faculty, Duzce University, Konuralp, Duzce, Turkey. ⁷Department of Cardiovascular Surgery, Çanakkale Onsekiz Mart University, Kepez, Çanakkale, Turkey.

Received: 1 October 2013 Accepted: 3 February 2014

Published: 17 February 2014

References

1. Vasdekis SN, Argentou M, Kakisis JD, Bossios A, Gourgiotis D, Karanikolas M, Karatzas G: A global assessment of the inflammatory response elicited upon open abdominal aortic aneurysm repair. *Vasc Endovasc Surg* 2008, **42**(1):47-53.
2. Walker PM: Ischemia/reperfusion injury in skeletal muscle. *Ann Vasc Surg* 1991, **5**(4):399-402.
3. Bown MJ, Nicholson M, Bell PR, Sayers RD: Cytokines and inflammatory pathways in the pathogenesis of multiple organ failure following abdominal aortic aneurysm repair. *Eur J Vasc Endovasc Surg* 2001, **22**(6):485-495.
4. Roumen RM, Hendriks T, Van Der Ven-Jongekrijg J, et al: Cytokine patterns in patients after major vascular surgery, hemorrhagic shock, and severe blunt trauma. Relation with subsequent adult respiratory distress syndrome and multiple organ failure. *Ann Surg* 1993, **218**:769-776.
5. Adembri G, Kastamoniti E, Bertolozzi I, et al: Pulmonary injury follows systemic inflammatory reaction in infrarenal aortic surgery. *Crit Care Med* 2004, **32**:1170-1177.
6. Rajmakers PG, Groeneveld AB, Rauwerda JA, Teule GJ, Hack CE: Acute lung injury after aortic surgery: the relation between lung and Leg microvascular permeability to indium-labelled transferrin and circulating mediators. *Thorax* 1997, **52**:866-871.
7. Bao XQ, Liu GT: Bicyclol: a novel antihepatitis drug with hepatic heat shock protein 27/70-inducing activity and cytoprotective effects in mice. *Cell Stress Chaperones* 2008, **13**(3):347-355.
8. Zhang K, Zhao T, Huang X, Liu ZH, Xiong L, Li MM, Wu L, Zhao Y, Zhu L, Fan M: Preinduction of Hsp70 promotes hypoxic tolerance and facilitates acclimatization to acute hypobaric hypoxia in mouse brain. *Cell Stress Chaperones* 2009, **14**:407-415.
9. Latchman DS: Heat shock proteins and cardiac protection. *Cardiovasc Res* 2001, **51**:637-646.
10. Mosser DD, Caron AW, Bourget L, Denis-Larose C, Massie B: Role of the human heat shock Protein hsp70 in protection against stress-induced apoptosis. *Mol Cell Biol* 1997, **17**:5317-5327.
11. Fujibayashi T, Hashimoto N, Jijiwa M, Hasegawa Y, Kojima T, Ishiguro N: Protective effect of geranylgeranylacetone, an inducer of heat shock

- protein 70, against drug-induced lung injury/fibrosis in an animal model. *BMC Pulm Med* 2009, **9**:45.
12. Choudhury S, Bae S, Ke Q, Lee JY, Kim J, Kang PM: Mitochondria to nucleus translocation of Aif in mice lacking Hsp70 during ischemia/reperfusion. *Basic Res Cardiol* 2011, **106**:397–407.
 13. Qian M, Eaton JW, Wolff SP: Cyanate-mediated inhibition of neutrophil myeloperoxidase activity. *Biochem J* 1997, **326**(1):159–166.
 14. Maillet F, Petitou M, Choay J, Kazatchkine MD: Structure function relationships in the inhibitory effect of heparin on complement activation: independency of the anticoagulant and anticomplement sites on the heparin molecule. *Mol Immunol* 1988, **25**:917–923.
 15. Redini F, Tixier J-M, Petitou M, Choay J, Robert L, Hornbeck S: Inhibition of leukocyte elastase by heparin and its derivatives. *Biochem J* 1988, **252**:515–519.
 16. Pasini FL, Pasqui AL, Ceccatelli L, Capecci PL, Orrico A, Perri TD: Heparin inhibition of polymorphonuclear leukocyte activation in vitro: a possible pharmacological approach to granulocytemediated vascular damage. *Thromb Res* 1984, **35**:527–537.
 17. Ecker EE, Gross P: Anticomplementary power of heparin. *J Infect Dis* 1929, **44**:250–253.
 18. Brestel EP, McClain EJ: A mechanism for inhibition of luminol dependent neutrophil chemiluminescence by polyanions. *J Immunol* 1983, **131**:2515–2519.
 19. Ekre HP, Naparstek Y, Lider O, Hyden P, Hagermark O, Nilsson T, Vlodavsky I, Cohn I: Anti-inflammatory effects of heparin and its derivatives: inhibition of complement and lymphocyte migration. *Adv Exp Med Biol* 1992, **313**:329–340.
 20. Nelson RM, Cecconi O, Roberts WG, Aruffo A, Linhardt RJ, Bevilacqua MP: Heparin oligosaccharides bind L- and P-selectin and inhibit acute inflammation. *Blood* 1993, **82**:3253–3258.
 21. Katz DJ, Stanley JC, Zelenoek GB: Operative mortality rates for intact and ruptured abdominal aortic aneurysms in Michigan: an eleven-year statewide experience. *J Vasc Surg* 1994, **19**:804–815.
 22. Lafci G, Selcuk Gedik H, Korkmaz K, Erdem H, Cicek OF, Nacar OA, Yildirim L, Kaya E, Ankarali H: Efficacy of iloprost and montelukast combination on spinal cord ischemia/reperfusion injury in a rat model. *J Cardiothorac Surg* 2013, **8**:64.
 23. Korkmaz K, Gedik HS, Budak AB, Erdem H, Lafci G, Karakilic E, Nacar OA, Yildirim L, Ankarali H: Effect of heparin on neuroprotection against spinal cord ischemia and reperfusion in rats. *Eur Rev Med Pharmacol Sci* 2013, **17**:522–530.
 24. Schneiderman J, Walden R, Bass A, Broieris S, Segal E, Adar R: Surgery for thoraco-abdominal aortic aneurysm. *Harefuah* 1991, **120**(4):179–181.
 25. Głowiczki P: Surgical repair of thoracoabdominal aneurysms: patient selection, techniques and results. *Cardiovasc Surg* 2002, **10**(4):434–441.
 26. Gedik HS, Korkmaz K, Erdem H, Karakilic E, Lafci G, Ankarali H: Protective effect of heparin in the End organ ischemia/reperfusion injury of the lungs and heart. *J Cardiothorac Surg* 2012, **7**:123.
 27. Inokawa H, Date H, Okazaki M, Okutani D, Aokage K, Nagahiro I, Aoe M, Sano Y, Shimizu N: Effects of postmortem heparinization in canine lung transplantation with Non-heart-beating donors. *J Thorac Cardiovasc Surg* 2005, **129**(2):429–434.
 28. Etz CD, Di Luozzo G, Bello R, Luehr M, Khan MZ, Bodian CA, Griep RB, Plestis KA: Pulmonary complications after descending thoracic and thoracoabdominal aortic aneurysm repair: predictors, prevention, an treatment. *Ann Thorac Surg* 2007, **83**(2):870–876. Discussion 890–872.
 29. Marumoto Y, Kaibara M, Murata T: Hemorheological studies on platelet counts and size in normal pregnancy and pregnancies with preeclampsia and intrauterine growth retardation. *Nihon Sanka Fujinka Gakkai Zasshi* 1989, **41**(9):1380–1386.
 30. Mauney MC, Blackbourne LH, Langenburg SE, Buchanan SA, Il K, Tribble CG: Prevention of spinal cord injury after repair of the thoracic or thoracoabdominal aorta. *Ann Thorac Surg* 1995, **59**:245–252.
 31. Sahin MA, Onan B, Guler A, Oztas E, Uysal B, Arslan S, Demirkilic U, Tatar H: Cilostazol, A type III phosphodiesterase inhibitor, reduces ischemia/reperfusion-induced spinal cord injury. *Heart Surg Forum* 2011, **14**:E171–E177.
 32. Budak B, Seren M, Turan NN, Sakaogullari Z, Ulus AT: The protective effects of resveratrol and L-name on visceral organs following aortic clamping. *Ann Vasc Surg* 2009, **23**:675–685.
 33. Severin IC, Soares A, Hantson J, Teixeira M, Sachs D, Valognes D, Scheer A, Schwarz MK, Wells TN, Proudfoot AE, Shaw J: Glycosaminoglycan analogs as a novel anti-inflammatory strategy. *Front Immunol* 2012, **3**:293.
 34. Norwood MG, Bown MJ, Sutton AJ, Nicholson ML, Sayers RD: Interleukin 6 production during abdominal aortic aneurysm repair arises from the gastrointestinal tract and not the legs. *Br J Surg* 2004, **91**(9):1153–1156.
 35. Teoh NC, Farrell GC: Hepatic ischemia reperfusion injury: pathogenic mechanisms and basis for hepatoprotection. *J Gastroenterol Hepatol* 2003, **18**(8):891–902.
 36. Hirsh J, Warkentin TE, Shaughnessy SG, Anand SS, Halperin JL, Raschke R, Granger C, Ohman EM, Dalen JE: Heparin and Low-molecular-weight heparin: mechanisms of action, pharmacokinetics, dosing, monitoring, efficacy, and safety. *Chest* 2001, **119**(1 Suppl):64–94.

doi:10.1186/1749-8090-9-35

Cite this article as: Yener et al.: Protective role of heparin in the injury of the liver and kidney on the experimental model of ischemia/reperfusion. *Journal of Cardiothoracic Surgery* 2014 **9**:35.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

