



## Draft Genome Sequence of Pseudoalteromonas sp. Strain JC3

Margaret E. Rosario, a Jacqueline Camm, b Damian Cavanagh, b David C. Rowley, a D David R. Nelson b

<sup>a</sup>Department of Biomedical and Pharmaceutical Sciences, University of Rhode Island, Kingston, Rhode Island, USA <sup>b</sup>Department of Cell and Molecular Biology, University of Rhode Island, Kingston, Rhode Island, USA

**ABSTRACT** We report the draft genome sequence for *Pseudoalteromonas* sp. strain JC3, an isolate obtained from an aquaculture facility for whiteleg shrimp (*Litopenaeus vannamei*). The JC3 genome suggests multiple mechanisms for microbial interactions, including a type VI secretion system and potential for antibiotic production.

P seudoalteromonas is a genus of marine gammaproteobacteria with substantial capability for antibiotic production (1, 2). Recently, *Pseudoalteromonas* isolates have been suggested as potential probiotic bacteria to help combat acute hepatopancreatic necrosis disease (AHPND), an emerging and severe shrimp disease affecting aquaculture systems (3–5). Here, we provide details of the genome of a *Pseudoalteromonas* strain derived from a shrimp aquaculture facility to aid future investigations into the microbial and host interactions involving *Pseudoalteromonas* strains.

*Pseudoalteromonas* sp. strain JC3 was isolated from seawater of a whiteleg shrimp (*Litopenaeus vannamei*) culture purchased from Miami Aquaculture (Boynton Beach, FL). Seawater (100  $\mu$ l) was spread onto YP30 agar plates (0.1% yeast, 0.5% peptone from meat, 3% Instant Ocean) and incubated at 25°C for 48 h. A yellow pigmented colony designated JC3 was iteratively inoculated onto YP30 agar and reisolated to ensure a pure culture.

Pseudoalteromonas sp. strain JC3 was grown in mLB30 media (Luria broth with 3% Instant Ocean) for 24 h at 25°C and 100 rpm. Genomic DNA was extracted using the Bio Basic molecular biology kit according to the manufacturer's protocol. Genomic DNA was quantified using a Qubit fluorometer (Invitrogen) and sheared using a Covaris ultrasonicator. Libraries were prepared on an Apollo next-generation sequencing (NGS) library prep system using the PrepX DNA library kit (TaKaRa Bio) and run on an Agilent BioAnalyzer DNA high-sensitivity (HS) chip. Quantification was performed on all samples using quantitative PCR (qPCR) in a Roche LightCycler480 as described by the Roche KAPA Library Quantification Kit for Illumina Platforms technical data sheet (https://rochesequencingstore.com/wp-content/uploads/2017/ 10/KAPA-Lib-Quant-ILMN 9.17-IfU 1.pdf). Sequencing was performed at the Rhode Island Genomics and Sequencing Center using 2  $\times$  250-bp paired-end sequencing on an Illumina MiSeq instrument. The total number of reads was 5,655,047 bp. Sequence trimming and quality control were completed using FastQC v1.0.0. Reads shorter than 64 bp were discarded. De novo assembly was performed using CLC Genomics Workbench v12.0.2, and the resulting contigs were processed using the CLC Microbial Genome Finishing module. The final draft assembly was estimated as 100% complete with 1.03% contamination using CheckM v1.0.18 (6) in KBase (7). The completed draft genome sequence is composed of 112 contigs ( $N_{50}$  contig length, 196,838 bp) with a total sequence length of 5,572,526 bp and an average G+C content of 43.1%. Rapid Annotations using Subsystems Technology (RAST) was utilized for gene annotation, which resulted in 5,021 coding sequences and 100 RNAs (8). The SEED viewer identified 354 subsystems containing 28% of the coding sequences (9). Default settings were used unless otherwise specified.

Assessment of the *Pseudoalteromonas* sp. strain JC3 draft genome sequence suggests multiple avenues for interacting with the surrounding microbial community and environment. BLASTp v12.11.0 (10) was used to search the nonredundant protein database at NCBI to **Citation** Rosario ME, Camm J, Cavanagh D, Rowley DC, Nelson DR. 2021. Draft genome sequence of *Pseudoalteromonas* sp. strain JC3. Microbiol Resour Announc 10:e00212-21. https://doi.org/10.1128/MRA.00212-21.

**Editor** Kenneth M. Stedman, Portland State University

**Copyright** © 2021 Rosario et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to David R. Nelson, dnelson@uri.edu.

Received 9 March 2021 Accepted 17 August 2021 Published 9 September 2021 identify genes for type II, IV, and VI secretion systems and a range of putative proteinand carbohydrate-degrading enzymes. Homologs of the *lodAB* (query coverage, 100%; E value, 0.0; amino acid identity, >99.5%) and *goxAB* (query coverage, 100%; E value, 0.0; amino acid identity, >99%) gene clusters found in *Pseudoalteromonas flavipulchra* JG1 and numerous bacterial groups (11) encoding marinocine (12) and glycine oxidase products (13), respectively, were also present. Analysis using the Antibiotics and Secondary Metabolite Analysis Shell (antiSMASH) (14) revealed 19 putative biosynthetic gene clusters, including one for the production of alterochromide/bromoalterochromide antibiotics (1).

**Data availability.** This whole-genome shotgun project has been deposited at DDBJ/ ENA/GenBank under the accession number JAFEKQ000000000. The version described in this paper is the first version. The contigs are available at https://www.ncbi.nlm.nih.gov/Traces/ wgs/JAFEKQ01?display=contigs. The raw reads can be found at the SRA under the accession number PRJNA699263.

## **ACKNOWLEDGMENTS**

Funding for this work was provided by the USDA (2019-67016-29868) to D.C.R. and D.R.N. We thank Janet Atoyan at the Rhode Island Genomics and Sequencing Center for assistance with DNA sequencing.

## REFERENCES

- Suria AM, Tan KC, Kerwin AH, Gitzel L, Abini-Agbomson L, Bertenshaw JM, Sewell J, Nyholm SV, Balunas MJ. 2020. Hawaiian bobtail squid symbionts inhibit marine bacteria via production of specialized metabolites, including new bromoalterochromides BAC-D/D'. mSphere 5:e00166-20. https:// doi.org/10.1128/mSphere.00166-20.
- Paulsen SS, Strube ML, Bech PK, Gram L, Sonnenschein EC. 2019. Marine chitinolytic Pseudoalteromonas represents an untapped reservoir of bioactive potential. mSystems 4:e00060-19. https://doi.org/10.1128/mSystems.00060-19.
- Li P, Kinch LN, Ray A, Dalia AB, Cong Q, Nunan LM, Camilli A, Grishin NV, Salomon D, Orth K. 2017. Acute hepatopancreatic necrosis disease-causing *Vibrio parahaemolyticus* strains maintain an antibacterial type VI secretion system with versatile effector repertoires. Appl Environ Microbiol 83:e00737-17. https://doi.org/10.1128/AEM.00737-17.
- Lai H-C, Ng TH, Ando M, Lee C-T, Chen I-T, Chuang J-C, Mavichak R, Chang S-H, Yeh M-D, Chiang Y-A, Takeyama H, Hamaguchi H-O, Lo C-F, Aoki T, Wang H-C. 2015. Pathogenesis of acute hepatopancreatic necrosis disease (AHPND) in shrimp. Fish Shellfish Immunol 47:1006–1014. https:// doi.org/10.1016/j.fsi.2015.11.008.
- Sánchez-Díaz R, Molina-Garza ZJ, Cruz-Suárez LE, Selvin J, Kiran GS, Ibarra-Gámez JC, Gómez-Gil B, Galaviz-Silva L. 2019. Draft genome sequence of *Pseudoalteromonas piscicida* strain 36Y\_RITHPW, a hypersaline seawater isolate from the south coast of Sonora, Mexico. J Glob Antimicrob Resist 16:83–86. https://doi.org/10.1016/j.jgar.2018.09.003.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome Res 25:1043–1055. https:// doi.org/10.1101/gr.186072.114.
- Arkin AP, Cottingham RW, Henry CS, Harris NL, Stevens RL, Maslov S, Dehal P, Ware D, Perez F, Canon S, Sneddon MW, Henderson ML, Riehl WJ, Murphy-Olson D, Chan SY, Kamimura RT, Kumari S, Drake MM, Brettin TS, Glass EM, Chivian D, Gunter D, Weston DJ, Allen BH, Baumohl J, Best AA, Bowen B, Brenner SE, Bun CC, Chandonia J-M, Chia J-M, Colasanti R, Conrad N, Davis JJ, Davison BH, DeJongh M, Devoid S, Dietrich E, Dubchak I, Edirisinghe

JN, Fang G, Faria JP, Frybarger PM, Gerlach W, Gerstein M, Greiner A, Gurtowski J, Haun HL, He F, Jain R, Joachimiak MP, et al. 2018. KBase: the United States Department of Energy Systems Biology Knowledgebase. Nat Biotechnol 36:566–569. https://doi.org/10.1038/nbt.4163.

- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42:D206–D214. https:// doi.org/10.1093/nar/gkt1226.
- Gish W, States DJ. 1993. Identification of protein coding regions by database similarity search. Nat Genet 3:266–272. https://doi.org/10.1038/ ng0393-266.
- Campillo-Brocal JC, Chacón-Verdú MD, Lucas-Elío P, Sánchez-Amat A. 2015. Distribution in microbial genomes of genes similar to *lodA* and *goxA* which encode a novel family of quinoproteins with amino acid oxidase activity. BMC Genomics 16:231. https://doi.org/10.1186/s12864-015-1455-y.
- Lucas-Elío P, Gómez D, Solano F, Sanchez-Amat A. 2006. The antimicrobial activity of marinocine, synthesized by *Marinomonas mediterranea*, is due to hydrogen peroxide generated by its lysine oxidase activity. J Bacteriol 188:2493–2501. https://doi.org/10.1128/JB.188.7.2493-2501.2006.
- Campillo-Brocal JC, Lucas-Elio P, Sanchez-Amat A. 2013. Identification in Marinomonas mediterranea of a novel quinoprotein with glycine oxidase activity. Microbiologyopen 2:684–694. https://doi.org/10.1002/mbo3.107.
- Blin K, Shaw S, Steinke K, Villebro R, Ziemert N, Lee SY, Medema MH, Weber T. 2019. AntiSMASH 5.0: updates to the secondary metabolite genome mining pipeline. Nucleic Acids Res 47:W81–W87. https://doi.org/10 .1093/nar/gkz310.