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Kidins220/ARMS is expressed in neuroblastoma tumors and stabilizes neurotrophic signaling in a human neuroblastoma cell line

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Abstract

Background—Neurotrophic signaling is an important factor in the survival of developing neurons, and the expression of neurotrophic receptors correlates with prognosis in neuroblastoma. Kinase D-interacting substrate of 220kDa (Kidins220) associates with neurotrophic receptors and stabilizes signaling them, but the expression and function of Kidins220 in neuroblastoma are unknown.

Methods—We study Kidins220 expression in human neuroblastoma cell lines and tumor samples by western blot and microarray analyses. We determine the functional consequences of downregulation of Kidins220 for response of cell lines to oxidative stress, chemotherapeutic treatment, and neurotrophins using small interfering RNA (siRNA) silencing and measuring cell survival, signaling, and migration.

Results—Kidins220 is expressed in all neuroblastoma tumors and cell lines studied. Downregulation of Kidins220 leads to attenuation of nerve growth factor- (NGF-) induced, but not brain-derived neurotrophic factor- (BDNF-) induced MAPK signaling. However, downregulation of Kidins220 does not alter the response to chemotherapeutic drugs or oxidative stress or affect cellular motility.

Conclusions—Kidins220 is expressed in neuroblastoma tumors and stabilizes NGF-induced, but not BDNF-induced survival signaling in neuroblastoma cell lines.

INTRODUCTION

Neurotrophic signaling is critical for the normal development and maintenance of the sympathetic nervous system, which arises from neural crest cells early in embryogenesis. Two ligand-receptor pairs, brain-derived growth factor-tropomyosin receptor kinase B (BDNF-TrkB) and nerve growth factor-tropomyosin receptor kinase A (NGF-TrkA),

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mediate the survival of early neuroblasts and the differentiation into mature neurons. Dysfunction of neurotrophic signaling contributes to the pathogenesis of neuroblastoma, a pediatric tumor of sympathetic precursor cells. The clinical characteristics and current treatment of neuroblastoma were recently reviewed (1). Among other factors, clinical prognosis can be predicted by the presence of TrkA and TrkB receptors in the tumor, which also alter the biology of neuroblastoma cell lines *in vitro* (2). Although many of the signaling pathways of these receptors are known, there are no treatments for neuroblastoma that target this system.

The Kinase D-interacting substrate of 220kDa (Kidins220), is a recently-discovered protein highly expressed in the developing nervous system and initially shown to be a target for receptor tyrosine kinases in neurons (3). The large multidomain structure of Kidins220 has subsequently been shown to be involved in neurotrophic and other cytokine signaling (4, 5), vesicle formation and transport (6-10), and cytoskeletal regulation during migration (11, 12), differentiation (13, 14), and plasticity (15). The specificity of Kidins220 for immature neural cells and its involvement in neurotrophic signaling suggest that Kidins220 may have a role in the biology of neuroblastoma. In this regard, Kidins220 acts as a scaffold for MAPK signaling cascades which can be initiated by neurotrophins binding to TrkA and TrkB. Neuroblastoma prognosis is positively related to TrkA expression and negatively related to TrkB expression (2).

In addition, neuroblastic tumors have genomic instability which leads to increased copy number of whole or partial chromosomes. The short arm of chromosome 2 (2p) often has increased copy number in neuroblastoma, and two genes in this region, *Nmyc* and *Alk*, have been correlated with poor prognosis (16). The *Kidins220* locus on chromosome 2p is near the locations of *Nmyc* and *Alk*, so there is an increased copy number of this gene, as well, in many neuroblastomas (17).

There is also evidence that Kidins220 is expressed and has important functions in normal neural crest cells. Kidins220 expression in mammalian embryos colocalizes both spatially and temporally with sympathetic nervous system development (3). Kidins220 knockout causes late fetal death because the neural crest-derived cardiac outflow tracts do not form properly (18).

Kidins220 expression is increased in melanoma, another neural crest tumor, compared to benign skin lesions, and silencing of Kidins220 causes increased apoptosis in tumor xenografts and melanoma cell lines exposed to oxidative stress. The survival effects of Kidins220 are blocked by MAPK inhibitors, and Kidins220-silenced cells are rescued by establishment of constitutive MAPK activity (19).

In the present study, we establish that Kidins220 is expressed in neuroblastoma cell lines and tumors and examine the effects of Kidins220 downregulation on signaling, migratory behavior, and cell survival in a neuroblastoma cell line.

RESULTS

Kidins220 is expressed in neuroblastoma cell lines and tumor samples

Although there are no published reports for neuroblastoma, Kidins220 is expressed in many different tissues and cell lines. Most notably in relation to other neural crest tumors, a rat pheochromocytoma cell line (PC12 cells) has been shown to express Kidins220 and its function in these cells in relation to neurotrophic signaling has been studied extensively (3, 8, 20). Pheochromocytoma is a neuroendocrine tumor that, like neuroblastoma, has its origin in cells of the sympathetic nervous system lineage.

To qualitatively determine if Kidins220 is expressed in neuroblastoma, we tested a group of six human neuroblastoma cell lines for Kidins220 expression by western blot. These six lines were compared to PC12 cells, which have been shown previously to express Kidins220 (3). All six neuroblastoma cell lines express the protein with very little variation in the level of expression (Figure 1A). All six lines also express more Kidins220 qualitatively than the PC12 line with respect to actin.

We next wanted to test the expression in human tumor samples. We used a set of publicly available microarray data (21, 22) to look at expression of Kidins220 in neuroblastoma tumor samples. Table 1 shows the demographics of the patient samples with regard to patient age, tumor stage, and MYCN amplification status. The array analysis (Figure 1B) shows high expression of Kidins220 in all samples tested. When compared with other genes commonly expressed in neuroblastoma, only GAPDH shows consistent expression with the intensity of Kidins220 across all samples. It is notable that the neighboring genes on chromosome 2p, ALK and MYCN, which have been shown to be important in neuroblastoma, are not expressed at high levels in all tumor samples.

MAPK signaling after NGF treatment is attenuated after knockdown of Kidins220

Kidins220 acts as a scaffold to prolong neurotrophic signaling through MAPK signaling cascades in both PC12 cells and neuronal cultures. To check the effects of Kidins220 knockdown on neurotrophic signaling in neuroblastoma cells, we first tested the response of SH-SY5Y neuroblastoma cells to both NGF and BDNF treatment. SH-SY5Y cells respond to both ligands by increasing phosphorylation of ERK, but have differential response kinetics based on the ligand (Figure 2A). BDNF causes an early increase in pERK after addition to the medium which returns to baseline by 30 minutes after treatment. In contrast, NGF causes ERK phosphorylation that persists for up to 30 minutes which is shown by the significant difference in P-ERK/ERK ratio with an average difference in 3 experiments of 0.266 ± 0.042 , $p=0.044$. This prolonged signaling after NGF binding has been ascribed to Kidins220 in other cell types (4). It is interesting to speculate that the absence of prolonged ERK phosphorylation after BDNF treatment may reflect differential affinity of Kidins220 for TrkA and TrkB receptors in these cells.

We next examined NGF-induced phosphorylation of ERK in Kidins220-siRNA- and NS-siRNA-transfected cells (Figure 2B). Kidins220-siRNA cells show peak ERK phosphorylation at 1 minute, with a decrement in phosphorylation thereafter. NS-siRNA cells also peak at 1 minute, but have sustained ERK phosphorylation even until 30 minutes

after treatment (Figure 2C). Significance testing of the quantification of 3 independent experiments shows that the average difference in P-ERK/ERK ratio is 0.326 ± 0.012 , $p=0.024$ in NS-siRNA cells at 30 minutes after treatment as compared to untreated cells; whereas, there is no difference in Kidins220-siRNA cells between untreated cells and cells harvested 30 minutes after treatment (Figure 2D). In contrast and as one might predict from the already attenuated MAPK signaling in BDNF treated SH-SY5Y cells, BDNF signaling is not altered by Kidins220-siRNA transfection (Data not shown.).

Cell culture growth rate under native, NGF-enriched, and BDNF-enriched conditions is unchanged by downregulation of Kidins220

We next examined viability of Kidins220-siRNA- and NS-siRNA-transfected SH-SY5Y cells in culture with and without growth factors. After transfection, the cells were starved of growth factors and then treated with growth factor enriched media for 48 hours. Neither NGF nor BDNF treatment altered viability in Kidins220-siRNA- or NS-siRNA-transfected SH-SY5Y cells relative to untransfected controls (Figure 2E).

Kidins220 downregulation does not affect the susceptibility of neuroblastoma cell lines to chemotherapeutic treatment *in vitro*

The current chemotherapeutic treatment for neuroblastoma consists of high-dose anti-mitotic agents including doxorubicin, etoposide, and vincristine. These cause apoptosis during cell division by intercalating DNA, blocking topoisomerase, or disrupting microtubules respectively. We investigated whether knockdown of Kidins220 expression would enhance the effects of these drugs in the SH-SY5Y neuroblastoma cell line. Twenty-four hours after transfection with siRNA, cells were treated with one of the chemotherapeutic agents in increasing concentrations. All three drugs caused cell death in a dose-dependent manner; however, Kidins220-siRNA cells exhibited responses that were statistically indistinguishable from NS-siRNA cells (Figure 3).

We also performed viability assays after treatment with the chemotherapeutic agents etoposide, doxorubicin, and vincristine in cells maintained in starvation medium and then supplemented with vehicle or NGF and saw no statistically significant difference between Kidins220-siRNA- and NS-siRNA-transfected cells (Supplemental Figure S1, online).

Kidins220 does not alter the response of neuroblastoma cell lines to oxidative injury

In studies of melanoma, another neural crest lineage cancer, Kidins220 depletion was shown to increase apoptosis after oxidative stress caused by UV exposure or H_2O_2 (19). It has also been shown that neurons of the entorhinal cortex and frontal cortex in heterozygous Kidins220 knockout mice undergo pyknotic degeneration in a pattern that resembles oxidative damage (23). Our lab and others have shown that neurotrophic signaling pathways are involved in oxidative stress responses in neuroblastoma (24, 25), and we considered Kidins220 as a potential mediator of this response. After transfecting SH-SY5Y cells with Kidins220 specific siRNA or control siRNA, we treated the cells with $150 \mu M H_2O_2$. We looked at cell survival 24 hours after treatment, and found no significant difference between the Kidins220-siRNA and control groups (Figure 4A).

In melanoma (19), changes in the MAPK signaling cascade were observed after oxidative stress and this pathway was altered in Kidins220 depleted cells. Specifically, these authors reported that phosphorylation of ERK after UV exposure was decreased in Kidins220 knockdown cells. We tested the MAPK response to treatment with 150 μ M H₂O₂ in SH-SY5Y cells by looking at phosphorylation of ERK in a time-course after treatment (Figure 4B). This study was also performed in the SK-N-SH and CHP-212 human neuroblastoma cell lines (Supplemental Figure S2, online). Although ERK phosphorylation increases after treatment of neuroblastoma lines with H₂O₂, the duration and magnitude of the increase is equivalent in Kidins220-siRNA and NS-siRNA cells. This is shown by the quantification of 3 independent experiments with no significant difference in the ERK/P-ERK ratio at each time point (Figure 4C). Most notably, the phosphorylation of ERK is not attenuated at the 30 and 60 minute time points in Kidins220-siRNA cells compared to NS-siRNA cells, suggesting that Kidins220 is not involved the MAPK/ERK signaling pathway after oxidative stress in these neuroblastoma cells.

Knockdown of Kidins220 does not change migration behavior of neuroblastoma cells

Cell motility is an important factor in the aggressiveness of neuroblastoma because it determines the capacity of the tumor to form metastases. Kidins220 is involved in regulating motility in T lymphocytes (12) and dendritic cells *in vitro* (11) and might control neural progenitor migration as shown by defects in Kidins220 knockout mice (26). We tested neuroblastoma cell motility *in vitro* using a simple scratch assay where a section of cells is scraped off of the culture dish and then monitored to see if new cells will migrate into the cleared area (Figure 5A). With serum present in the media, the cells cover the cleared area within 24 hours through migration and proliferation. To reduce proliferation and also test whether neurotrophins influence the migration, we performed the same assay using starvation medium with only 0.5% serum, or starvation medium plus NGF. In these conditions there is less proliferation, so the number of cells that migrate into the cleared area more likely reflects migration than proliferation. Quantification of the number of migrating cells per field shows that equal numbers migrate into the cleared area in both Kidins220-siRNA and NS-siRNA cells (Figure 5B). There is no difference in the migration ability of SH-SY5Y cells after Kidins220 knockdown.

DISCUSSION

In this report, we have shown that Kidins220 is expressed in neuroblastoma patient tumor samples and cell lines. We studied the effects of Kidins220 in the SH-SY5Y cell line and showed that Kidins220 stabilizes the NGF/MAPK/ERK signaling pathway in these cells, but that depleting Kidins220 expression does not alter viability or migration of the cells, the compensatory signaling response or cell viability after treatment with hydrogen peroxide, or cell viability after treatment with anti-mitotic chemotherapeutic drugs.

We chose to use transient transfection with siRNA to decrease Kidins220 expression. We did this to (a) mimic the effects of a drug with a short half-life that would block Kidins220 activity and could be given in conjunction with other chemotherapeutics and (b) minimize compensatory transcriptional changes that would occur as a result of the knockdown.

Because of this choice, our results represent only what changes may occur in the 36 hours after blocking Kidins220 signaling and are not useful in predicting the outcome of neuroblastic tumors that do not express Kidins220 in their native state.

The efficacy of Kidins220 knockdown as a potential adjunctive strategy for neuroblastoma treatment hinges on the capability of Kidins220 to promote cell survival by stabilizing MAPK signaling after cellular stress. Liao et al. demonstrated in melanoma that cellular stress caused upregulation of MAPK signaling which was impaired after Kidins220 silencing (19). They proposed that Kidins220 is a functional part of the main survival pathway. Duffy et al. also showed that a decrease in Kidins220 expression results in cellular damage in the developing mouse brain (23). This damage was not proven to be due to oxidative stress or attributed to a particular pathway, but the damage was phenotypically similar to oxidative damage seen in other studies. Here we have shown that Kidins220 does not affect MAPK signaling induced after hydrogen peroxide even though it does affect MAPK signaling after NGF treatment. Thus, either a separate MAPK signaling pathway is induced by oxidative stress in these cells that does not involve Kidins220, or the MAPK pathway is not the dominant survival pathway in these cells. We have shown previously that p75NTR mediated Akt phosphorylation increases survival after hydrogen peroxide treatment in PC12 cells (24). Even though Kidins220 can form complexes that include p75NTR (20), Kidins220 knockdown does not affect Akt signaling in PC12 cells (4). Akt signaling has shown to be influenced, however, by the effects of Kidins220 on ephrin signaling. While our work focused on the effects of Kidins220 on the neurotrophic signaling pathways, Kidins220 associates with ephrin receptors and can modulate ephrin signaling (3, 5, 27). Ephrins and ephrin receptors have been shown to have increased expression low grade tumors that also have high TrkA expression (28, 29). In hippocampal neurons from Kidins220 knockout mouse, decreased ephrin receptor-modulated PI3K signaling was shown to be associated with, a lack of dendrite outgrowth (5). In native SH-SY5Y cells, overexpression of ephrin receptors is associated with growth suppression (29). In our system, we not only expect ephrin receptor expression to be low, but we also assume that knockdown of Kidins220 would interfere with ephrin signaling. The net result is a decrease in ephrin signaling which would not change the growth potential in SH-SY5Y cells. This is supported by our results which showed no difference in survival between Kidins220-siRNA and NS-siRNA cells.

In neuroblastoma, blocking Kidins220 might actually increase cell survival. In PC12 cells, short-lived signaling through the MAPK/ERK signaling cascade leads to proliferation; whereas NGF treatment caused prolonged ERK activation leading to differentiation (30). Our study (Figure 2A) shows that BDNF treatment causes a short-term phosphorylation of ERK in SH-SY5Y cells, and Kidins220 silencing can block the prolonged signaling after NGF treatment, making it analogous to BDNF treatment (Figure 2C). Thus, blocking Kidins220 may cause NGF/TrkA signaling to stimulate proliferation, rather than differentiation, in neuroblastoma cells. This could explain why Kidins220 does not make these cells more susceptible to cell death.

Furthermore, Kidins220 may differentially associate with TrkA and TrkB receptors, and thereby mediate the differential response of neuroblastoma cells to NGF and BDNF,

respectively. This is of clinical interest because TrkA and TrkB expression influence neuroblastoma prognosis in opposite directions (2). A full understanding of the role of Kidins220 in neuroblastoma clinical behavior must await further studies.

METHODS

Cell Culture

The following human neuroblastoma cell lines were obtained from American Type Culture Collection (ATCC, Manassas, VA): SK-N-SH, SH-SY5Y, SK-N-AS, SK-N-MC, CHP-212, and IMR-32. PC12 cells were also from ATCC. All cells were grown at 37°C in 5% CO₂. SH-SY5Y cells were cultured in a 1:1 mixture of Dulbecco's Modified Eagle's Medium (DMEM) and Ham's F12 medium (Gibco Life Technologies, Grand Island, NY) supplemented with 10% v/v fetal bovine serum (FBS) (Innovative, Novi, MI), and 1% v/v penicillin/streptomycin (Gibco). PC12 cells were grown in low glucose DMEM (Gibco) supplemented with 10% v/v horse serum (Innovative), 5% v/v FBS, and 1% v/v penicillin/streptomycin (Gibco). All other cell lines were cultured in Eagle's Minimum Essential Medium (MEM) (Gibco) supplemented with 1% v/v 100mM sodium pyruvate, 1% v/v non-essential amino acids, 10% v/v FBS, and 1% v/v penicillin/streptomycin. Before treatment with neurotrophins, SH-SY5Y cells were cultured in starvation medium (DMEM:F12 media supplemented with 0.5% v/v FBS and 1% v/v penicillin/streptomycin).

Western Blotting

Native neuroblastoma cell lines or transfected and treated cells were washed with ice-cold PBS and lysed in RIPA buffer (PBS plus 1% NP40, 0.5% sodium deoxycholate, 0.1% SDS) with proteinase and phosphatase inhibitors. Lysates were then sonicated, and incubated on ice for 30 min, vortexing every 10 min. After centrifugation, the supernatant representing the soluble fraction was isolated. Protein content in the supernatant was analyzed using the Bio-Rad Protein Assay (Bio-Rad, Hercules, CA). Protein loading was normalized based on the quantification. After SDS-PAGE, proteins were transferred to a nitrocellulose membrane. Membranes were blocked for non-specific binding using 5% w/v non-fat dry milk in PBS for 1 hour at room temperature. Primary antibody staining was performed in 5% milk in PBS overnight at 4°C. The following antibodies and concentrations were used: ARMS/Kidins220- R-300 antibody recognizes C-terminus of Kidins220 (Santa Cruz Biotechnology, Santa Cruz, CA), 1:1000 dilution. This antibody was characterized by comparison to the Kidins220 antibody generated by Maggiwar lab (31). In our experiments, it can detect rat, mouse, and human Kidins220; P-ERK- E-4 antibody recognizes phospho-Tyr 204 in ERK1 and ERK2 (Santa Cruz); ERK- K-23 antibody recognizes ERK1 and ERK2 (Santa Cruz); β -Actin, 1:5000 (Sigma-Aldrich, St. Louis, MO). Membranes were washed 3 times for 5 minutes in PBS with 0.05% Tween-20 (Fisher Scientific, Fair Lawn, NJ). Secondary staining was done with the following horse radish peroxidase (HRP) conjugated antibodies in 5% milk in PBS for 1 hour at room temperature: goat anti-rabbit IgG-HRP, 1:5000 (Santa Cruz); goat anti-mouse IgG-HRP, 1:5000 (Santa Cruz). Membranes were washed and developed using ECL (Santa Cruz).

siRNA transfection

Neuroblastoma cell lines were transfected with either a mixture of siRNA oligonucleotides specific for Kidins220 (referred to as Kidins220-siRNA) or non-silencing scrambled siRNA (referred to as NS-siRNA). Both siRNA cocktails were obtained from Santa Cruz Biotechnology. Transfections were performed using Lipofectamine 2000 (Invitrogen, Carlsbad, CA). Cells were plated in appropriate sized plates and allowed to grow overnight. The siRNA oligos were mixed with Lipofectamine reagent in Opti-MEM I medium to form complexes and then added to the cells in media without antibiotics. The final concentration of siRNA in the cultures was 20nM. The cells were then incubated for 8 hours, after which the media was changed and treatments could begin.

Neurotrophin treatment

For experiments that required neurotrophin treatment, transfected cells were transitioned to starvation media 8 hours after transfection. Cells were starved overnight, and then treated with the desired growth factors, transitioned to standard media with 10% FBS, or left in starvation media. All treatments with recombinant human nerve growth factor (NGF, Peprotech, Rocky Hill, NJ) were done at a concentration of 100 ng/ml. All treatments with recombinant human brain derived neurotrophic factor (BDNF, Peprotech) were done at 50 ng/ml.

Chemotherapeutic drug treatment

SH-SY5Y cells were plated in 96 well tissue culture dishes at a density of 10,000 cells/well. After transfection with Kidins220-siRNA or NS-siRNA, cells were maintained in standard media, or starved and treated with growth factors as needed. Stock solutions of etoposide (Sigma), doxorubicin (Toronto Research Chemicals, Toronto, Canada), and vincristine (Sigma) were diluted in standard media or starvation media to the appropriate concentrations. Cells were treated with the drugs, incubated with for a further 48 hours, and then assayed for viability. Cell viability after drug treatments was performed with the Alamar Blue viability assay (Invitrogen). At the specified time after drug treatment, 10X Alamar blue reagent was added directly to Kidins220-siRNA or NS-siRNA cells in the 96 well plate to get the proper final concentration. Cells were then incubated for 1-2 hours at 37° C. In a Spectromax M5 plate reader (Molecular Devices, Sunnyvale, CA), the fluorescent signal of the reduced dye was excited using 570 nm wavelength light and the emission was read at 585 nm. Relative viability of the cells was determined by comparing fluorescence of drug treated cells to untreated controls.

Hydrogen peroxide treatment

Neuroblastoma cells were cultured in standard media in 24 well tissue culture dishes at a density of 40,000 cells/well. After transfection with siRNA constructs, the cells were treated with a stock solution of 3% H₂O₂ (Sigma) diluted in media to a final concentration of 150 μM. This solution was immediately added to the cells after making it. For western blots, cells were harvested minutes to hours after treatment. For viability, cells were incubated for 24 or 48 hours after treatment. Because the Alamar blue assay is based on a reduction reaction which can be influenced by H₂O₂, we used Trypan blue (Hyclone, Logan, UT) to

quantify survival in the H₂O₂ treated cells. At the indicated timepoint after treatment, cells were trypsinized and collected along with the supernatant to make sure that non-adherent cells were included in the counts. Suspended cells were mixed with Trypan blue dye at a 9:1 ratio meaning that the final mixture was 0.04% Trypan blue. This mixture was loaded on a hemacytometer and counted to determine the percent survival.

Migration Assay

After 24 hours, cells were starved of growth factors overnight. A plastic pipette tip was used to scrape away the cells in the middle of the plate. The plates were rinsed and then flooded with media plus 0.5% FBS, media plus 100ng/ml NGF, or media plus 10% FBS. Images were taken of the cells immediately after scraping and the locations of the images on the plate were marked. After 1 and 2 days, cells were re-imaged to determine the extent of cell migration into the scraped area. Quantification was done by tracing the borders of the scraped area in the initial image, and then overlaying the border onto the later images and counting the number of cells within the outline. Cell counts were then normalized based on the area of the outlined section.

Microarray Analysis

The microarray data used was originally published in by Lavarino and colleagues (21, 22) and included a total of 62 unique neuroblastoma samples which were all isolated from adrenal tumors. Data was accessed in the caArray database which is part of the caBIG project (National Cancer Institute, Bethesda, MD). Normalization of the dataset was done through the Automated Microarray Pipeline using Robust Multiarray Averaging (RMA) (Dana-Farber Cancer Institute, Boston, MA). The open source MeV experiment viewer (Dana-Farber Cancer Institute) was used to analyze and output the data graphically.

Imaging

Brightfield images of migrating SH-SY5Y cells were taken on a Nikon Eclipse TE2000-U inverted microscope (Nikon Instruments Inc., Melville, NY) with a SPOT RT Monochrome digital camera and SPOT software (Spot Imaging Solutions, Sterling Heights, MI).

Statistical Analysis

Significance of quantitative results was determined using an unpaired, two-tailed t-test to compare groups. In experiments comparing the response after NGF or BDNF treatment, significance was determined by comparing treated cells at a certain timepoint to untreated cells. In viability experiments comparing Kidins220-siRNA to NS-siRNA cells, comparisons were made between groups at the same time point or treatment dose. Statistics were computed using SPSS 16.0.2 software.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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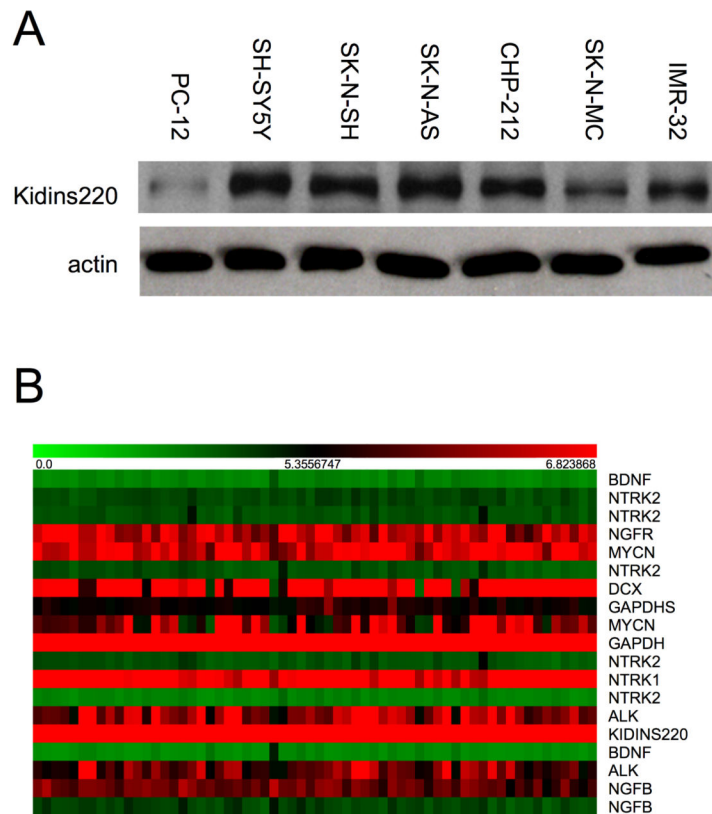
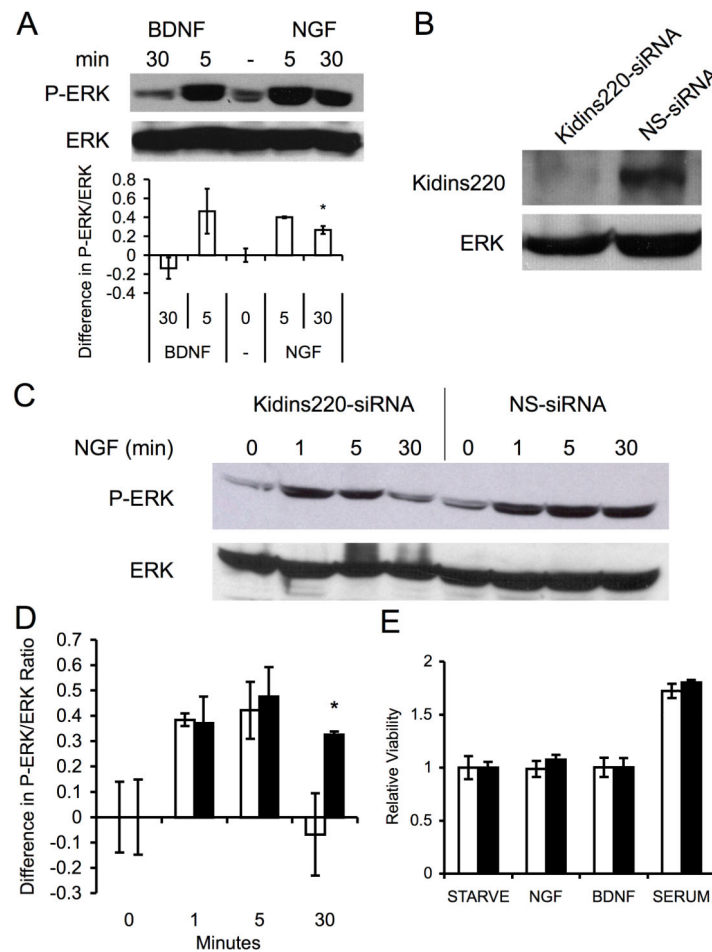


Figure 1.

Kidins220 is expressed in neuroblastoma cell lines and tumor samples. (A) Western blot from six different neuroblastoma cell lines as compared to the PC12 rat pheochromocytoma cell line. Expression is seen in all the cell lines tested. (B) Microarray analysis of neuroblastoma tumor samples assayed for mRNA expression using the Affymetrix U95Av2 whole genome expression array (21, 22). This analysis of 62 samples shows high expression of Kidins220 across all samples in comparison to other genes which are important in neuroblastoma prognosis. Gene expression is represented on a color scale with green boxes representing low, black boxes are intermediate, and red boxes are high expression. Data was normalized with GAPDH representing high expression across all cell lines. Genes are presented in the order they appear on the array. Genes listed more than once are represented by multiple elements on the array.

**Figure 2.**

SH-SY5Y cells have attenuated NGF signaling after Kidins220 knockdown. (A) Representative western blot showing the downstream signaling after treatment of the native SH-SY5Y with neurotrophins. Cells were starved of growth factors overnight and then treated with 50 ng/ml BDNF or 100 ng/ml NGF. Cells were harvested either 5 minutes or 30 minutes after treatment. Accompanying bar graph shows quantification of the difference in P-ERK/ERK ratio at different time points after treatment. Sample size: n=3. Results show that phosphorylation of ERK remains significantly elevated at 30 minutes after treatment with NGF; whereas, BDNF treatment results in only a short duration increase in ERK phosphorylation with a subsequent return to below baseline. (B) Western blot showing the extent of Kidins220 silencing in this experiment. SH-SY5Y cells were transfected with either Kidins220 specific siRNA duplexes or non-silencing scrambled siRNA control duplexes. (C) Transfected cells were then treated with 100ng/ml NGF and harvested after 1, 5, or 30 minutes of treatment. Representative western blot shows that phosphorylation of ERK returned to pretreatment levels after 30 minutes in the Kidins220-siRNA treated cells. NS-siRNA cells showed prolonged phosphorylation of ERK. (D) Quantification of the difference in P-ERK/ERK ratio after NGF treatment by densitometric analysis of 3 independent experiments. P-ERK levels are significantly elevated at 30 minutes after treatment in the NS-siRNA group compared to the 0 minute timepoint, whereas the level of

P-ERK in Kidins220-siRNA group is not significantly different at 30 minutes. White bars, Kidins220-siRNA; black bars, NS-siRNA. (E) SH-SY5Y cells are not dependent on neurotrophins for survival or growth. After starvation from growth factors overnight, cells were grown in starvation media supplemented with either 50 ng/ml BDNF or 100 ng/ml NGF. An Alamar blue viability assay shows that there is no difference in viability after treatment with these growth factors in both the Kidins220-siRNA and NS-siRNA groups. In both cases, addition of serum enhances viable cell number, assumedly because of its positive effect on proliferation. Data represents 3 independent experiments performed in triplicate. White bars, Kidins220-siRNA; black bars NS-siRNA. In all panels, error bars represent \pm SEM. * $p < 0.05$.

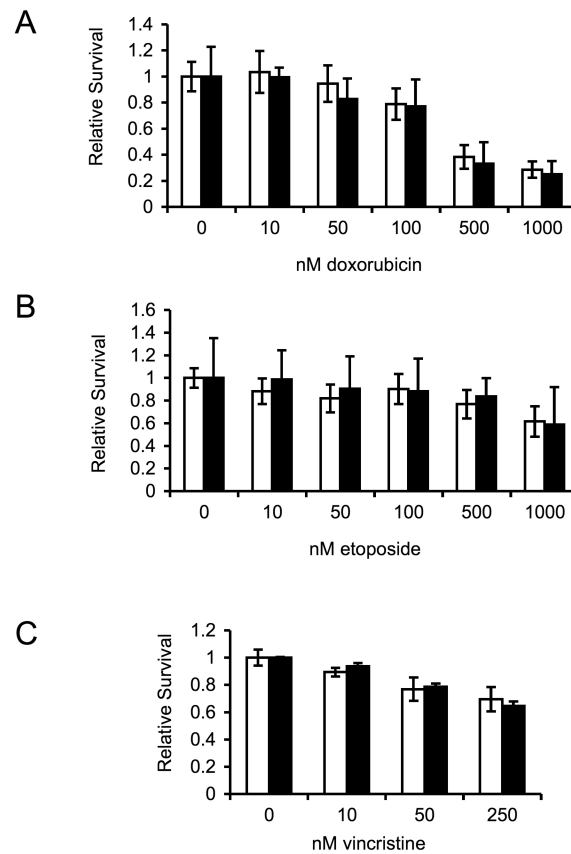
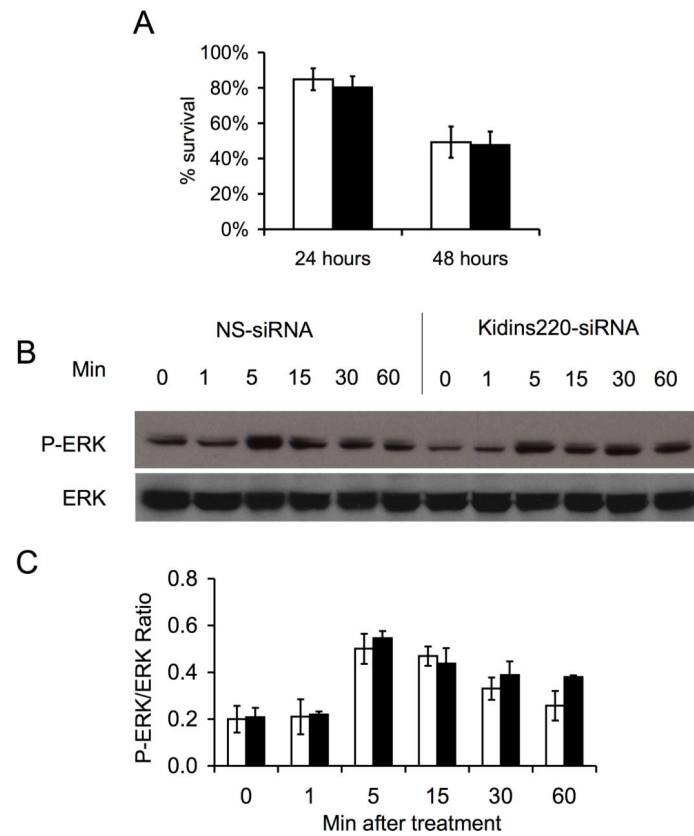


Figure 3.

Survival of cultured neuroblastoma cell lines after chemotherapeutic treatment is unaffected by Kidins220 silencing. SH-SY5Y cells were transfected with Kidins220-siRNA or NS-siRNA. Cells were treated with doxorubicin (A), etoposide (B), or vincristine (C) at increasing concentrations. Alamar blue assay was used 48 hours after drug treatment to assess cell viability. Although these cells are sensitive to all three drugs in a dose-dependent manner, no significant differences were seen in survival between the NS-siRNA and Kidins220-siRNA cells. All panels: White bars, NS-siRNA; black bars, Kidins220-siRNA. Data represents two independent experiments performed in quadruplicate samples. Error bars represent \pm SEM.

**Figure 4.**

Response of neuroblastoma cells to oxidative stress after Kidins220 depletion. SH-SY5Y cells were transfected with Kidins220-siRNA or NS-siRNA. After Kidins220 silencing, SH-SY5Y cells were treated with 150 μ M hydrogen peroxide. (A) 24 and 48 hours after hydrogen peroxide treatment, cells were stained with Trypan blue and counted. The survival percentage was calculated as the number of cells that are not stained by the dye divided by the total number of cells. $n=3$. Error bars represent \pm SEM. White bars, NS-siRNA; black bars, Kidins220-siRNA. (B) MAPK signaling in response to hydrogen peroxide treatment is unchanged in Kidins220-siRNA cells. Kidins220-siRNA and NS-siRNA cells were treated with 150 μ M hydrogen peroxide. Cells were harvested after 1, 5, 15, 30, or 60 minutes of treatment. Western blotting shows the survival response of the cell by measuring the phosphorylation of ERK. (C) The quantification of the treatment time course shows that ERK phosphorylation increases 5 minutes after hydrogen peroxide treatment in both the Kidins220-siRNA and NS-siRNA cells. At each time point, there is no significant difference between Kidins220-siRNA and NS-siRNA cells suggesting Kidins220 is not involved in ERK signaling in neuroblastoma cells after oxidative stress. Sample size: $n=3$. White bars, NS-siRNA; black bars, Kidins220-siRNA.

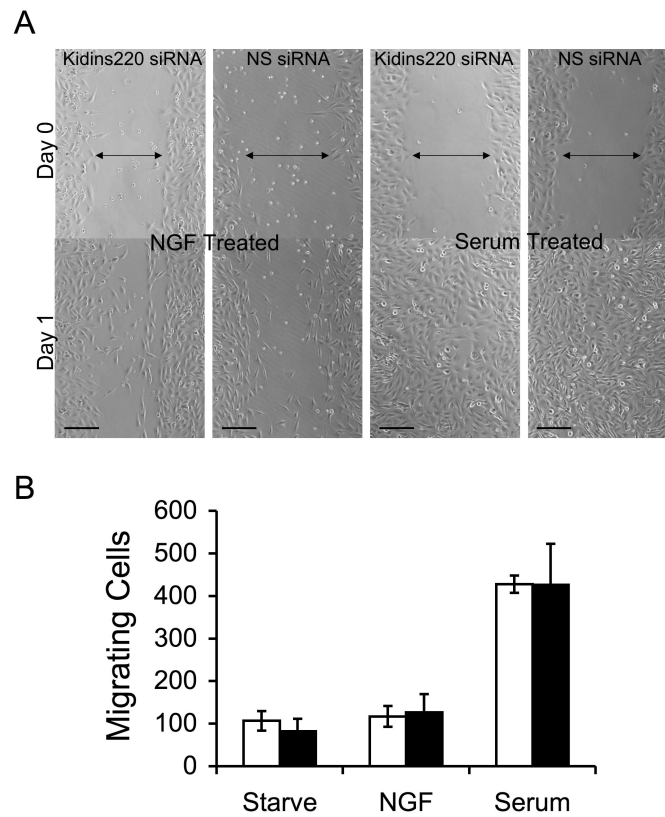


Figure 5.

Cell migration is unchanged by Kidins220 silencing. SH-SY5Y cells transfected with either Kidins220-siRNA or NS-siRNA were assayed for migration using a scratch assay. (A) Representative brightfield images show plates immediately and 24 hours after scratching. Double-headed arrows designate the area that was cleared by scratching. NGF cells treated show modest migration into the area on Day 1. Serum treated cells cover the area through migration and proliferation. (B) Quantification of cells that migrated into the scratched area shows no significant difference between the NS-siRNA groups and the Kidins220-siRNA groups. $n=4$. Error bars represent \pm SEM. White bars, Kidins220-siRNA; black bars, NS-siRNA. Scale bars represent 100 μ m.

Table 1

Demographics for tumor samples used in microarray analysis.

	# of Tumors	% of Tumors
Patient Age		
18m	44	71.0%
>18m	18	29.0%
INSS Stage		
1	4	6.5%
2	10	16.1%
3	9	14.5%
4	30	48.4%
4S	9	14.5%
MYCN status		
Amplified	15	24.2%
not amplified	47	75.8%
% Tumor in sample		
100%	4	6.5%
90%	14	22.6%
80%	23	37.1%
70%	21	33.9%
Total Samples	62	

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