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Exercise for osteoporosis: A literature review of pathology and mechanism

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Osteoporosis (OP) is a disease that weakens bones and has a high morbidity rate worldwide, which is prevalent among the elderly, particularly, women of postmenopausal age. The dynamic balance between bone formation and resorption is necessary for normal bone metabolism. Many factors, including aging, estrogen deficiency, and prolonged immobilization, disrupt normal apoptosis, autophagy, and inflammation, leading to abnormal activation of osteoclasts, which gradually overwhelm bone formation by bone resorption. Moderate exercise as an effective non-drug treatment helps increase bone formation and helps relieve OP. The possible mechanisms are that exercise affects apoptosis and autophagy through the release of exercise-stimulated myohormone and the secretion of anti-inflammatory cytokines via mechanical force. In addition, exercise may also have an impact on the epigenetic processes involved in bone metabolism. Mechanical stimulation promotes bone marrow mesenchymal stem cells (BMSCs) to osteogenic differentiation by altering the expression of non-coding RNAs. Besides, by reducing DNA methylation, the mechanical stimulus can also alter the epigenetic status of osteogenic genes and show associated increased expression. In this review, we reviewed the possible pathological mechanisms of OP and summarized the effects of exercise on bone metabolism, and the mechanisms by which exercise alleviates the progression of OP, to provide a reference for the prevention and treatment of OP.

KEYWORDS

osteoporosis, exercise, inflammatory reaction, cytokines, mechanical stress

Introduction

Osteoporosis (OP) refers to a skeletal disease characterized by low bone density and microarchitectural deterioration of bone tissue consequently increasing bone fragility and susceptibility to fracture as defined by the World Health Organization. OP causes more than 8.9 million fractures annually worldwide, resulting in an osteoporotic fracture

every 3 seconds (1). One in three women and one in five men of age over 50 years will experience osteoporotic fractures (2).

OP can occur due to many factors, including senility, reduction in mechanical stimulation, bone and hormone metabolism disorders, and varying properties of stress associated with transcriptional changes in osteogenic genes (3). These factors can lead to a disturbance in the dynamic balance between bone formation dominated by osteoblasts (OBs) and bone resorption dominated by osteoclasts (OCs). In clinical practice, drug therapy such as calcium, vitamin D, hormone replacement therapy, bisphosphonates, etc., are commonly used to treat OP. However, drug therapy has disadvantages such as a long treatment cycle, various drug side effects, high treatment cost, and low compliance (4). Therefore, exercise therapy has attracted increasing attention because of its few adverse reactions, high safety, practicality, and simple operation.

Rahimi et al. (5) conducted a meta-analysis, including 16 randomized controlled trials (RCTs), that summarized the curative efficacy of different training patterns on bone mineral density (BMD) in postmenopausal women. There was no crucial change in BMD of the lumbar spine or femoral neck after exercise training. However, subgroup analysis by exercise training type showed that lumbar BMD was significantly higher when whole-body vibration (WBV) was used as an intervention than with randomized controlled trials with aerobic, resistance, and combined training. This suggests that WBV is an effective method for improving lumbar BMD among elderly postmenopausal patients. In addition, another metaanalysis of 97 RCTs showed that in patients with OP or osteopenia, mind-body exercises (such as Tai Chi, yoga, and dancing) could improve BMD in the lumbar vertebra and femoral neck. Meanwhile, resistance training was more likely to improve BMD in the total hip (6).

However, there was no review on the effect of exercise on the pathological mechanism of OP. Therefore, this review mainly summarizes the possible pathological mechanisms of OP and the mechanisms of exercise alleviating the progression of OP, to provide a reference for the prevention and treatment of OP.

Pathological changes mechanism of OP

When osteoclastogenesis exceeds osteoblastogenesis, the resulting condition is an OP (3). Bone metabolism is in a state of dynamic balance, which is jointly maintained by bone formation led by OBs and bone resorption led by OCs (coupling between OBs and OCs). However, when osteoclast-dominated bone resorption activity is enhanced and if there is no corresponding bone formation activity, it leads to BMD loss and even OP. Estrogen levels in postmenopausal women drop because of ovarian failure. Estrogen deficiency increases bone

turnover and imbalances in bone metabolism by affecting levels of T lymphocyte, B lymphocyte, monocytes, and cytokines (7). This leads to increased BMD loss, destruction of trabecular microstructure, and increased risk of fracture. The pathological mechanisms of OP mainly include the abnormal activation of OCs resulting from changes in apoptosis, inflammatory reaction, and autophagy. Besides, epigenetic changes, such as changes in non-coding RNA and DNA methylation, can also reduce the expression of osteogenic genes (Figure 1).

Apoptosis

Apoptosis is a normal physiological process. It refers to the process of cell death caused by internal and external factors triggering the preexisting intracellular death process. However, excessive or defects of apoptosis can lead to bone-related diseases (8). Some studies have shown that the maintenance of BMD depends not only on the absorption function of OCs and the osteogenic function of OBs but also on the changes in the lifespan of the two kinds of cells by apoptosis (9). Osteocytes, derived from OBs, are the major cells in mature bone tissue, and they can survive for decades (9).

Evidence obtained from studies conducted in mice illustrates that increased osteocyte apoptosis is partly responsible for osteoporosis caused by sex steroid deficiency, overuse of glucocorticoids (GCs), or senescence (10). The upregulated expressions of receptor activator of nuclear factor K-B ligand (RANKL) and vascular endothelial growth factor (VEGF) in aged mice may inhibit osteoblast maturation or induce osteocyte death (11). By increasing the signal of Ras/Rac1/Erk and PI3 kinase/mTOR/S6K, cytokines such as macrophage colonystimulating factor (M-CSF), RANKL, tumor necrosis factor (TNF), and VEGF can slow down apoptosis and improve the survival rate of OCs, which is linked to the rise in bone resorption (12). Weinstein and Manolagas (9) found that osteoblast apoptosis increased 10-fold, meanwhile, osteoblast apoptosis increased fourfold in the vertebrae of ovariectomized mice as compared with the control group. This indicated that the acceleration of bone loss after the lack of estrogen is caused by an earlier reduction in the working time of OBs. Hughes et al. (13) reported that 17\beta-estradiol increased the apoptosis rate of rat OCs from 0.5% to 2.7%, and estrogen deficiency prolonged the survival period of OCs, increasing the number of cells by 2-3 times, and leading to enlargement of the bone absorption cavity. In addition, OBs, as the main target of GCs, play an important role in the pathogenesis of GC-induced OP. GCs can increase the expression of BH3-only protein Bim and downregulate tissue inhibitor of metalloproteinase-1 (TIMP-1) to promote osteoblast apoptosis, thereby leading to the occurrence of OP (14, 15). The apoptosis of osteocytes and OBs in GC-induced OP is caused by the loss of extracellular matrix attachment mediated by the



internal and external signaling pathways of Pyk2/JNK kinase (16). GCs, increase reactive oxygen species (ROS) production *in vivo* bone and *in vitro* OBs (17), which will promote the activation of a PKC β /p66(shc)/JNK signaling cascade.

In addition, mechanical unloading, such as bed rest, reduced activity during senescence, or space travel, frequently causes BMD loss. Aguirre et al. (18) found that increased apoptosis of osteocytes was directly affected by mechanical unloading through tail suspension mice models. Although reducing osteoblast apoptosis can raise BMD, p53 mostly controls how apoptosis affects bone formation. Preventing osteoblast proliferation and apoptosis and encouraging osteoblast differentiation via the Akt-FoxOS pathway, has the opposite effect on the development of OCs (11). Connexin43 (Cx43) expression in the bone can decline with age, and Cx43 knockout increases osteocyte apoptosis, the number of OCs, and bone resorption on the cortical bone surface (19). In addition, Davis et al. identified a brand-new Cx43/miR21/HMGB1/RANKL pathway that controls osteoclast production and recruitment, is decreased with aging, and is important in preventing osteocyte apoptosis.

Inflammatory reaction

A higher incidence of various age-related illnesses, including OP and fragile fractures, is linked to chronic inflammation (20). Osteoclast-mediated bone resorption is accelerated by proinflammatory signals acting on mesenchymal stem cells and osteoclast precursors. (21). Age-related BMD loss and increased bone resorption are predicted by higher serum inflammatory marker levels in older adults (22). According to research by Cauley et al. (20), high serum levels of inflammatory markers in older adults who were in good health indicated a greater risk of fractures during a 5.8-year follow-up period.

The immune system's role in the development of senile OP arises mainly through the effects of estrogen deficiency and secondary hyperparathyroidism. In addition to directly affecting bone cells, the significant postmenopausal estrogen deficiency among postmenopausal women, indirectly causes them to exhibit chronic low-grade inflammatory phenotypes, namely altered cytokine expression and altered immune cell morphology (23). According to studies, a lack of estrogen led to a large rise in the pro-inflammatory cytokines, which include interleukin-1(IL-1), interleukin-6(IL-6), TNF-a, M-CSF, and prostaglandin E2(PGE2) (24-26). Most of these cytokines, directly and indirectly, act on OBs and OCs, improve the differentiation of mononuclear cells to mature OCs, and regulate osteoclast function. TNF- α can increase osteoblast apoptosis and the expression of RANKL. This indirectly leads to the increase of osteoclast differentiation and activity and inhibits osteoclast apoptosis (27), thus leading to BMD loss in the postmenopausal OP. Zha et al. (28) found that postmenopausal women with OP (PMOP) had higher levels of TNF- α than those without the condition. By stimulating the NFκB and PI3K/Akt pathway in vitro, TNF-α synergistically

enhances RANKL-induced osteoclast formation. Some of these cytokines, like IL-1 and TNF- α , may interact with one another in a way that promotes increased production of the other, which would further stimulate bone resorption. While PGE2 raises RANKL and lowers OPG, IL-1 and TNF- α enhance RANKL, OPG, and M-CSF (25, 29).

In addition, estrogen affects the proliferation, differentiation, activation, and homing of various immune cells (23). Various immune cells directly or indirectly affect bone cells through other mediators secreted by immune cells such as OPG/RANKL, and inflammatory cytokines such as IL-6 and TNF- α . The signaling pathways for NF-KB and c-src/PI 3-kinase/Akt are started when these immune factors activate TNF-related factors (TRAFs). Activating downstream molecules like c-Fos and Fos B causes the nuclear factor of activated T-cells cytoplasmic 1 (NFATc1) to be produced, which in turn causes the maturation and activation of osteoclasts (30). An immunoclinical study shows that CD19+ B lymphocytes are less abundant in postmenopausal women, however, they secrete more granulocyte-macrophage-colony-stimulating factor (GM-CSF) (31). Another study shows that T cells are more likely to express TNF- α in postmenopausal OP patients with bone fractures (32). Two recent studies have shown an increase in circulating T cells and monocytes in postmenopausal women (33) and a higher neutrophil-to-lymphocyte ratio in their peripheral blood (34).

Autophagy

Autophagy is a highly conserved cellular behavior, in which cells enfold intracytoplasmic substrates, such as senescence proteins, organelles, misfolded proteins, and damaged mitochondria, into autophagy by forming a double-layer membrane structure, and transport the substrates to the lysosome for degradation and then release them back to the cytoplasm (35). Autophagy can maintain cell activities and play a protective role through material recycling under the conditions of cell starvation, hypoxia, lack of growth factors, and some pathological conditions (36). The autophagy-associated gene (ATG) family of proteins regulates the formation of autophagosomes, which is the first step at the beginning of autophagy (37). A genome-wide association study confirmed the correlation between genetic variation of several ATG proteins and wrist BMD. It suggested that autophagy alteration may lead to OP phenotype (38).

Age-related autophagy suppression or decreased efficiency might result in BMD loss among the elderly population (39, 40). In addition, an important risk factor for OP associated with aging is vitamin D (VD) insufficiency. VD3 raises cytoplasmic calcium levels, which promotes autophagy. As a result, the CaMKK- β -AMPK signaling pathway is activated, which inhibits the activity of mTORC1 (41). Increased GC levels are another factor that leads to aging and excessive GCs can induce bone marrow mesenchymal stem cells (BMSCs) apoptosis. By inducing autophagy, GC-induced apoptosis in BMSCs may be prevented (37).

Reduced levels of osteoblast autophagy have been reported in rat models of OP (42). Mice with osteoblast-specific deletion of FIP200, a key element of mammalian autophagy, develop osteopenia as a result of OBs' defective terminal differentiation and a reduction in bone production (43). Onal et al. (44) conditionally knocked out the osteocyte's autophagy gene Atg7 in 6-month-old mice. In addition to a decline in the quantity of OCs and OBs, a decline in bone production rates, and an increase in oxidative stress, they discovered a drop in bone mass. These changes are also characteristics of aging mice. In addition, autophagy inhibits osteoblast apoptosis mediated by oxidative stress (45). Liu et al. (46) identified the autophagy receptor Optineurin (OPTN) as a key molecule to determine the cell fate of BMSCs, and its expression was decreased in aged mice. Aged mice and optn ^{-/-} mice showed an osteoporotic bone loss, increased aging of BMSCs, decreased osteogenesis, and increased adipogenesis. On the contrary, the inactivation of mouse monocyte autophagy-associated gene 7 (Atg7) prevented osteoclast differentiation while mitigating bone loss in mice treated with glucocorticoid or ovariectomized (47). Xiu et al. (48) discovered that RelB, a member of the NF-KB family, can cause TRAF3 degradation by controlling the transcription of BECN1, an early autophagy protein, in relation to the impact of autophagy on osteoclasts. The NF-KB signaling pathway is activated by TRAF3 degradation, which encourages RANKLinduced osteoclast development.

Changes in non-coding RNA

Non-coding RNA, a group of functional RNA not encoding proteins, plays a role in the occurrence and progression of several diseases, including OP. According to one study, postmenopausal OP patients (OP group) had differing levels of expression from healthy controls for 13 microRNAs (miRs), 70 long non-coding RNAs (lncRNAs), and 260 circular RNAs (circRNAs) (49).

LncRNAs with lengths longer than 200 nucleotides are transcripts that are not translated into protein (50). Although lncRNA does not encode proteins, it helps in regulating gene transcription in two ways: trans and cis (49). OP patients and ovariectomized mice had higher levels of lncRNA MEG3 in their BMSCs, which suppresses osteogenic development by downregulating miR-133a-3p (51). In response to mechanical stimulation, Liu et al. (52) demonstrated that the lncRNA Neat1 regulates osteoblast activity and bone formation through paraspeckle-dependent E3 ubiquitin ligase Smurf1 mRNA nuclear retention. Following disruption of paraspeckles, Neat1 was deficient, which resulted in lower osteoblast activity and decreased BMD. The alterations in bone formation in response to mechanical loading and unloading were reduced in mice models with Neat1 depletion.

Because their many target mRNA transcripts are involved in cell proliferation, differentiation, and death, miRs play a role in determining the determination of a cell's fate. Chen et al. (53) found that the reduced expression of miR-503 in CD14 + peripheral blood mononuclear cells (PMBCs) may be one of the pathogenesis of postmenopausal OP patients because miR-503 can specifically bind to RANKL, which can suppress the formation of OCs. Sugatani et al. (54) demonstrated that miR-21 high expression can stimulate osteoclast generation by downregulating levels of programmed cell death 4 (PDCD4), a protein that acts as a suppressive regulator of osteoclast production.

CircRNAs are another non-coding RNA mainly found in the cytoplasm in mammalian cells. They function as both indicators for various illnesses and essential components in tissue growth. Huang et al. (55) discovered that during osteoblast generation induced by recombinant NELL-1, the expression of circRFWD2 and circINO8 increased and was inversely linked with hsa-miR-6817-5p. In BMSCs from nonunion patients, Ouyang et al. (56) examined the differential expression of circRNAs and discovered that hsa_circ_0074834 had a much lower expression level. By controlling the expression of f Zeb1 and VEGF through miR-942-5p, has_circ_0074834 encourages BMSCs to differentiate into osteogenic tissue.

DNA methylation

The process of DNA methylation, one of the most durable epigenetic changes of gene transcription, is carried out by the DNA methyltransferase family (DNMT), which adds a methyl group to the cytosine base's five carbon position (57, 58). Increased levels of DNA methylation can suppress gene expression, possibly through transcriptional repression by recruiting proteins that prevent transcription factors from binding to DNA (59). Recent research has demonstrated that aberrant epigenetic alteration, which may result from both individual genetic variables and environmental triggers, is connected with a homeostatic imbalance between bone creation and resorption (60, 61). Reppe et al. (62) revealed an association between DNA methylation and BMD and fracture risk in postmenopausal women.

Raje et al. (63) compared CpG methylation levels of BMP2 promoters in OP patients and healthy people. They found that transcriptional activity and gene expression of BMP2 promoters in OP patients were reduced, which could downregulate the osteoblast markers involved in bone formation. According to Chen et al. (3), abnormal DNA methylation changes resulted in diminished femoral nuclear factor E2-related factor 2 (Nrf2), which is directly related to the development of OP. The simultaneous increase of DNA methyltransferase (Dnmt)1/ Dnmt3a/Dnmt3b and hypermethylation of the Nrf2 promoter dramatically suppressed femur Nrf2 in OP patients and ovariectomized mice. In addition, Dnmt3a also has a significant impact on the abnormal activation of OCs. Nishikawa et al. (64) found that, through epigenetic inhibitions of the IRF8 (a negative regulator of osteoclastinduced differentiation) gene, RANKL-induced activation of Dnmt3a can mediate DNA methylation via Sadenosylmethionine (SAM), this promotes osteoclast formation. Moller et al. (65) induced the differentiation of blood CD14+ monocytes into OCs from women of different ages and menopausal states and found that older women also had higher bone resorption activity of differentiated OCs in vitro, which may be related to the decreased DNA methylation of the TM7SF4 promoter, a key OCs gene.

Mechanisms of exercise in alleviating OP

Regular exercise helps keep the bone healthy and improve bone health by stimulating bone formation and strength through mechanical loading, even if it does not significantly improve BMD (66). Skeletal unloading occurs to some extent as people age due to decreases in physical activity and increases in sedentary time (67). From a mechanically-centric point of view, activities that generate higher intensity or quicker loads (such as resistance training and leaping) are excellent for promoting bone health because they stimulate existing bone cells in a significant way. Consequently, bone-healthy exercise encourages mesenchymal stem cells to differentiate into osteoblast lineages, thereby producing more healthy bone cells (68). As described by Robling et al. (69), mechanical forces applied to bone tissue induce the movement of interstitial fluid along tubules and bone cell pores, thereby causing cell-level shear stress and deformation of bone cell plasma membrane. These changes lead to the beginning of the bone remodeling process and stimulate bone resorption and formation cycles (70). Studies have shown that weight-bearing exercise, resistance training, or WBV training help maintain or improve bone mass, and improve the BMD of postmenopausal women, thereby promoting health and improved quality of life (71-73).

In addition, independent of mechanical loads, aerobic exercise can enhance osteocyte survival through altered macronutrient transport, the release of exercise-stimulated myohormones, and preservation of cellular or mitochondrial repair (68). The likely reason is that muscle factors (such as myostatin and irisin) secreted by muscle and bone factors (such as osteocalcin, TGF- β , and PGE2) secreted by osteocytes can interact with each other, and their secretion is regulated by

mechanical load (74). Other studies have shown that by enhancing aerobic glycolysis, irisin can promote osteoblast proliferation (75). Several studies have shown that regular endurance exercise can slow the age-related degeneration of mitochondrial number and capacity and dysfunction (76, 77).

In addition, proper exercise helps improve balance and postural stability (e.g., Tai Chi and Yoga) and reduces the frequency of falls, thus effectively reducing the occurrence of OP fractures thereby protecting the bone from trauma (Turner et al., 2003). A meta-analysis has shown that Tai Chi practice can reduce the risk of falls and fall-related injuries by approximately 43% and 50% in the short term (< 12 months) in at-risk adults and older adults, respectively (78). Moreover, a 12-week Iyengar yoga program can significantly improve standing balance by conducting sit-to-stand tests and a 4-minute walk for elderly community residents as compared to the control group (79).

This review summarizes the pathogenic mechanisms of exercise in the therapy of OP in rodents by summarizing the related literature (Table 1), and different types of exercise can improve human OP (Table 2). Mechanistically, exercise reduces the harmful OP alterations *via* affecting apoptosis, inflammatory response, and autophagy, and exercise may affect the epigenetic mechanisms of bone metabolism by regulating non-coding RNAs and DNA methylation (Figure 2).

The role of exercise in apoptosis

One of the processes causing the pathogenic alterations in OP is apoptosis. Aveline et al. (80) measured osteocyte apoptosis

as a percentage of caspase-3 immunostaining. They found that ovariectomized rats improved BMD and bone mineral content (BMC), trabecular parameters, cross-sectional area, a moment of inertia and OCN levels, and reduced osteocytes apoptosis and lipid content after 8 weeks of jumping training. In a rat model of secondary OP brought on by excessive alcohol intake, Maurel et al. (81) trained the experimental group on the treadmill. After 14 weeks of training, trabecular and cortical thickness, cortical pores, and osteocyte apoptosis of the rat model were improved.

Wen et al. (82) identified that OBs from old rats all showed senescence changes, among which osteocytes manifest the most evident senescence changes. After low magnitude vibration (LMV) treatment, the expression of anti-aging protein SIRT1 was significantly upregulated, whereas the expressions of p53 and P21 were significantly downregulated. LMV can promote bone formation in aged rats, upregulate the expression of antiaging protein Sirt1 in osteoblasts, and downregulate the expression of p53 and P21. Thus, LMV can partially inhibit the aging of osteoblasts through Sirt1/p53/P21 axis.

High-intensity exercise raises irisin levels in the blood (100). Irisin is an exercise-related myokine that is crucial for bone remodeling. A study revealed that exercise-mimetic myokine irisin can increase osteocyte functions and exhibits antiapoptotic effects (101). Irisin's activation of MAP kinases Erk1 and Erk2 and subsequent upregulation of the transcription factor Atf4 through an Erk-dependent pathway in osteocytes was the underlying mechanism. Moreover, according to Xu et al. (102), irisin reduced the prevalence of postmenopausal OP by upregulating Nrf2, blocking inflammatory bodies containing Pyrin domain protein 3 (NLRP3), and reducing the content of

| TABLE 1 | Mechanism of | exercise in | the treatment of | f osteoporosis (| OP) in rodents. |
|---------|--------------|-------------|------------------|------------------|-----------------|
|---------|--------------|-------------|------------------|------------------|-----------------|

| Authors | Model | Exercise types | Related gene/cytokines/ protein | Involved in pathways | Improved organization | Functions | Change |
|--------------------------------------|---------------------------|-------------------------------|---|----------------------|---|------------------------|----------------------|
| Chen et al. (3) | OP mice model | Running exercise | Nrf2, Dnmt1/3a/3b, SOD | Keap1-Nrf2 | Femur bone mass and trabecular microstructure | DNA Methylation | Ļ |
| Aveline et al. (80) | OP rat model | Jumping exercise | Caspase-3 | - | Whole body BMC and BMD, femur trabecular bone, and cortical microarchitecture | Osteocyte Apoptosis | Ļ |
| Maurel et al. (<mark>81</mark>) | OP rat model | Treadmill training | Caspase-3 | - | Femur trabecular microstructure | Osteocyte Apoptosis | Ļ |
| Wen et al. (82) | Aged rat model | Low magnitude vibration | P53, P21 | Sirt1/p53/p21 | Femur BMD and trabecular microstructure | Apoptosis | Ļ |
| Li et al. (83) | OP rat model | Running | IL-1b, IL-6, Cox-2 | - | Tibias trabecular microstructure | Inflammation | Ļ |
| Gao et al. (84) | Female rat model | Treadmill training | serum corticosterone, cortisol, pregnenolone, and estradiol | BDNF/AKT | - | Inflammation | Ļ |
| Lee et al. (85) | Middle-aged mice model | Treadmill training | - | - | Femur and tibias BMD and trabecular microstructure and skeletal nerve fiber density | miRNA | $\downarrow\uparrow$ |
| Liu et al. (52) | OP mice model | Treadmill training | OCN, Col1α1, PINP | - | Femur bone mass and thickness of cortical bone and trabecular microstructure | lncRNA Neat1 | Ť |
| Zuo et al. (<mark>86</mark>) | OP cell model | Mechanical stretch | Runx2 | - | Osteoblast differentiation and bone formation | miR-103a | Ļ |

↑: increase; ↓: decrease; ↑↓: The expression of some miRNAs was increased, and the expression of some miRNAs was decreased.

TABLE 2 Different exercise types on human osteoporosis (OP).

| Authors | Participants | Duration | Intervention group | Control group | Outcome | Exercise effect |
|---------------------------------------|---|-----------|--|--|--|--|
| FILIPOVIĆ et al. (87) | Postmenopausal OP women | 12 weeks | N = 47 Resistance training, balance exercise, and aerobic exercise | N = 49 No train | TUG, STS, and OLST | Improved the TUG, STS, and OLST. |
| Hettchen et al. (88) | Postmenopausal OP women | 13 months | N = 27 High impact weight-bearing/ high-intensity/ velocity resistance training | N = 27 Low-intensity exercise | Lumbar spine BMD and total hip BMD | Improved the lumbar spine BMD. |
| Kistler Fischbacher et al. (89) | Postmenopausal women with low bone mass | 8 months | N = 15 HiRIT-med, N = 14 BB-med | N = 42 HiRIT N = 44 BB | Lumbar spine BMD and total hip BMD | HiRIT improved the lumbar spine BMD more than BB. Antiresorptive medication may enhance exercise efficacy at the proximal femur and lumbar spine. |
| Stanghelle et al. (90) | 65+ years old women diagnosed with OP and vertebral fracture | 12 weeks | N = 76 Multicomponent resistance and balance exercise programme | N = 73 No train | Habitual walking speed, physical fitness, health-related quality of life, and fear of falling | Improved muscle strength, and balance and reduce fear of falling. |
| Stanghelle et al. (91) | 65+ years old women diagnosed with OP and vertebral fracture | 12 weeks | N = 76 Multicomponent resistance and balance exercise programme | N = 73 No train | Habitual walking speed, physical fitness, health-related quality of life, and fear of falling | Improved muscle strength, balance, and mobility and reduces fear of falling 3 months post- intervention. |
| Kemmler et al. (92) | Sedentary community- dwelling older men with osteopenia/OP and SMI-based sarcopenia | 12 months | N = 21 low-volume/HIT- DRT, with whey protein, VD, and calcium | N = 22 No train, with whey protein, VD, and calcium | Lumbar spine BMD, SMI, total hip BMD, maximum isokinetic hip-/leg- extensor strength | Improved lumbar spine BMD, SMI, and maximum isokinetic hip-/leg-extensor strength. |
| Harding et al. (93) | Older men with low hip and/or lumbar spine BMD | 8 months | N = 34 HiRIT. N = 33 Machine-based IAC | N = 26 No train | Lumbar spine and hip BMD, calcaneal ultrasound parameters, anthropometry, body composition, function (TUG, FTSTS, BES, LES) | Compared with CON, HiRIT improved trochanteric BMD, lumbar spine BMD, BUA, stiffness index, lean mass, TUG, FTSTS, BES, and LES. Compared with CON, IAC improved lean mass and FTSTS. Compared with IAC, HiRIT improved lumbar spine BMD, stiffness index, and FTSTS. |
| Filipović et al. (94) | Postmenopausal osteoporotic patients | 12 weeks | N = 37 Aerobic exercise, resistance training, and balance exercise, with alendronate therapy | N = 31 No train, with alendronate therapy | Activities of serum MMP-9 and TIMP-1 | Exercises decreased the activity of serum MMP-9 and increased the activity of TIMP-1. |
| Harding et al. (95) | Men with low lumbar spine and/ or proximal femur BMD | 8 months | N = 34 HiRIT N = 33 Machine-based IAC | N = 26 No train | Femoral neck and total hip BMC, volume, and vBMD for total, trabecular, and cortical bone compartments, total femoral neck cortical thickness, geometric and bone structural strength indices | Compared with IAC, HiRIT improved medial femoral neck cortical thickness. Both HiRIT and IAC preserve bone strength at the distal tibia and distal radius. |
| ElDeeb et al. (96) | Postmenopausal women with low BMD | 24 weeks | N = 25 WBV, with VD and calcium | N = 24 No train, with VD and calcium | BMD of the lumbar vertebrae and femur, and hip/knee/ankle muscle work during gait | Improved the leg muscle work and lumbar and femoral BMD. |

(Continued)

TABLE 2 Continued

| Authors | Participants | Duration | Intervention group | Control group | Outcome | Exercise effect |
|--------------------------------|--|----------|---|---|--|--|
| | | | supplementations once daily | supplementations once daily | | |
| Sen et al. (97) | Postmenopausal women | 6 months | N = 19 WBV N = 19 High impact training, with 1500 mg of calcium and 880 IU of VD per day | N=20 No train, with 1500 mg of calcium and 880 IU of VD per day | BMD of the lumbar spine and femur, serum markers, functional mobility, fall index, HRQoL, and depressive symptoms | WBV can prevent bone loss, and WBV and high impact training can improve functional mobility, HRQoL, and depressive symptoms. |
| Pérez- Gómez et al. (98) | Postmenopausal women | 12 weeks | N = 13 WBV | - | SMP30, body composition, (fat mass, lean mass, and bone mass) physical fitness (balance, TUG, and 6MWT) | Increased SMP30 in plasma, and 6MWT, reduced SMP30 in platelets, TUG, and total body fat mass. |
| Pasqualini et al. (99) | Postmenopausal women with a T- score at the lumbar spine or femoral neck between - 1 and - 2.5 SD | 3 months | N = 33 weight-bearing and resistance exercise | - | Anthropometric and fitness parameters, bone-remodeling markers, OCs, and QoL | Increased the markers of bone formation and the commitment of immature OCs, and improved the score of QoL with pain, physical function, and mental function. |

TUG, timed up-and-go; STS, sit to stand test; OLST, one leg stance test; BMD, bone mineral density; HiRIT, high intensity progressive resistance and impact training; BB, Buff Bones®, SMI, skeletal muscle mass index; HIT-DRT, high intensity dynamic resistance exercise; VD, vitamin D; IAC, isometric axial compression; FTSTS, five-times sit-to-stand; BES, back extensor strength; LES, leg extensor strength; CON, control; BUA, broadband ultrasound attenuation; BES, back extensor strength; BMC, bone mineral content; vBMD, volumetric bone mineral density; WBV, whole-body vibration; IU, international unit; HRQoL, health-related quality of life; SMP30, Regucalcin or senescence marker protein 30; 6MWT, 6-min walk test; OCs, $osteoclasts. \uparrow: increase; \downarrow: decrease; \uparrow \downarrow: The expression of some miRNAs was increased, and the expression of some miRNAs was decreased.$

inflammatory factors. These actions together prevented OBs from apoptosis and inhibited osteoblast death. Furthermore, studies have shown that Senescence marker Protein-30 (SMP-30) can protect against cell apoptosis (103), however, the serum

level of SMP30 decreases with age in elderly women (104). Pérez-Gómez et al. (98) showed that after 12 weeks of three timesweekly WBV exercise, postmenopausal women's circulating SMP30 levels, gait efficiency, and fat mass were all improved.



FIGURE 2

The mechanism of exercise improving osteoporosis. Exercise-induced changes in mechanical stress, hormones, and cytokines can regulate the pathological changes of osteoporosis. Mechanistically, exercise reduces the harmful osteoporosis alterations via affecting apoptosis, inflammatory response, and autophagy, and exercise may affect the epigenetic mechanisms of bone metabolism by regulating non-coding RNAs and DNA methylation.

The role of exercise in inflammatory reaction

Exercise can prevent and reduce OP by reducing proinflammatory cytokines and suppressing inflammation. Exercise increases the secretion of anti-inflammatory cytokines like IL-2, IL-10, IL-12, and interferon (IFN), which are beneficial to bone formation, and decreases the secretion of proinflammatory cytokines such as IL-1, IL-6, and TNF- α to prevent bone resorption (105). IL-10 is a cytokine that limits the host's immune response to pathogens and prevents tissue damage. A systematic review discovered a strong correlation between the duration of exercise and the rise in serum IL-10 (106). Santos et al. (107) reported that after 6 months of moderate exercise, older individuals' quality of life improved with higher serum IL-10 levels and significantly lower IL-6 and TNF- α levels. In addition, according to a meta-analysis, regular exercise reduces IL-6 and C-reactive protein levels among older persons (108). Reducing the expression of monocyte Toll-like receptor (TLR) is one of the potential mechanisms of the antiinflammatory effects of physical activity. Stewart et al. (109) found that 12 weeks of endurance and resistance exercise can effectively reduce the activation of TLR4 and the generation of pro-inflammatory cytokines IL-6 in previously sedentary older adults and younger adults.

Gao et al. (84) showed that mice that started moderateintensity continuous training at 8 months had higher levels of serum corticosterone, cortisol, pregnenolone, and estradiol compared to mice that started training at 18 months, and this suggests that reducing aging-related steroid hormone and antiinflammatory factors is more effectively accomplished by beginning training at 8 months.

The role of exercise in autophagy

Exercise can regulate bone metabolism by regulating the autophagy of bone tissue cells. Dalle et al. (110) found that physical exercise promoted the differentiation of BMSCs. After a half marathon, the expression of genes associated with telomerase, autophagy, and the genes Runx2, Msx1, and Spp1 was also altered. These changes were positively correlated with the differentiation of BMSCs.

The content of LC3-II, which is the surface marker of autophagic vacuoles, can inadvertently indicate the degree of autophagy (111). Zhang et al. (112) found that fluid shear stress (FSS) causes protective autophagy in osteocytes by inducing autophagic vacuoles, and higher levels of the LC3-II isoform, and the degeneration of P62 in osteocyte-like MLO-Y4 cells. In addition, the survival of osteocytes and ATP metabolism are linked to the autophagy induced by mechanical stimulation.

Treadmill exercise increased AMP-activated protein kinase (AMPK) phosphorylation in mice models, which promotes metabolism for glucose and fatty acid utilization (113). It has been hypothesized that human mesenchymal stem cells' ability to differentiate into OBs is controlled by AMPK through late activation of the Akt/mTOR signaling axis and early mTOR inhibition-mediated autophagy (114). In addition, irisin increased during and after exercise, may upregulate autophagy by increasing the Atg12-Atg5-Atg16L complex, thereby promoting osteogenesis of BMSCs and enhancing the Wnt/ β -catenin signaling pathway (115).

The role of exercise in non-coding RNA

By controlling the expression of non-coding RNA, exercise can lessen the harmful effects of OP. In the study of An et al. (116), miRs, lncRNAs, and circRNAs of Type 2 Diabetes with Depression (DD) patients were analyzed before and after Baduanjin treatment at 12 weeks. It was confirmed that Baduanjin can effectively improve the depressive symptoms and blood glucose level of DD patients by regulating the abnormal expression of lncRNAs, miRs, and circRNAs. Zhu et al. (117) found that appropriate tensile stress can promote BMSCs to osteogenic differentiation, prevent adipocyte differentiation, and induce the production of lncRNA-MEG3. Furthermore, overexpressed lncRNA-MEG3 further stimulated osteogenic differentiation in stressed BMSCs and inhibited the expression of miR-140-5p. According to Liu et al. (52), OP or unloading-induced bone loss can be prevented by increasing the Neat1 levels or stabilizing the paraspeckle structures by exercise and mechanical stress in conjunction with certain small molecules and oligonucleotides (52). In addition, exercise affects the expression of miRs that control inflammation and triggers changes in the gene expression of neutrophils and peripheral blood mononuclear cells (118). In ovariectomized rats, Li et al. (83) showed that 3-month running exercise can effectively reduce miRs, IL-1, IL-6, and Cox-2 expression levels, inhibit bone resorption, and improve bone trabecular formation. In addition, Zuo et al. (86) identified that in vivo and in vitro osteogenesis and bone production may be controlled by mechanical stress. Periodic mechanical stretch downregulates miRNA-103A and its host gene PANK3, and promotes the expression of Runx2 protein (a major osteogenic transcription factor), suggesting that the down-regulation of miRNA-103A may be a crucial factor for mechanically stimulating bone growth.

Few research has been done so far on the impact of circRNAs on bone during exercise, however, to understand their purpose, further research is needed to elaborate on their effects. Lee et al. (85) found that improvements in trabecular bone microarchitecture in middle-aged mice were directly

associated with increased skeletal nerves after aerobic exercise training. And the expression of seven upstream osteogeneses and neuroplasticity non-coding RNAs, including miR-491-3p, miR-470-5p, let-7a-5p, miR-137-3p, miR-130a3p, and miR-29b-3p, were upregulated after 8 weeks of treadmill activity training. Guo et al. (119) conducted a study and confirmed that the expression of circBBS9 in the quadriceps femoris of the aging mice decreased with age compared to the young mice, however, it was reversed after 2 months of treadmill exercise. Fang et al. (120) found that aerobic exercise helps reduce cartilage tissue damage, inflammatory cytokines content, type II collagen, chondrogenic differentiation-related genes, and circUNK expression induced by knee osteoarthritis.

The role of exercise in DNA methylation

DNA methylation is one of the known epigenetic mechanisms modulated by exercise (57). When it comes to aging, cancer, and type II diabetes, appropriate physical activity can help to control DNA methylation, as discovered by recent studies (121). A study was conducted involving 12 young men and 11 young women who underwent 3 months of one-legged knee-extension exercise and found that methylation at 4, 919 sites in the leg genome was altered after endurance training as compared with pre-training (122). From 23 healthy men, Rönn et al. (123) analyzed genome-wide patterns of DNA methylation in human adipose tissue. After a 6-month exercise intervention, including spinning and aerobics, global DNA methylation changed and a total of 17, 975 individual CpG sites of 7, 663 distinct genes showed altered DNA methylation levels. These results suggest that exercise can induce genome-wide alternations in DNA methylation in human adipose tissue and may affect the metabolism of adipocytes. Arnsdorf et al. (124) demonstrated that mechanical stimulation alters the epigenetic status of osteogenic genes by reducing DNA methylation and shows associated increased expression. This promotes the migration of MSCs and increases the potential for osteogenic differentiation.

In addition, Chen et al. (3) further demonstrated that running exercise (RE) ameliorated OP by correcting hypermethylation of the anti-osteoporotic factor Nrf2 promoter in an ovariectomized OP mouse model. The possible mechanism is that RE may reduce Nrf2 inhibition by blocking the abnormally elevated Dnmt, which demethylates the Nrf2 promoter. Nakajima et al. (125) investigated the epigenetic effects of exercise and aging on CpG island methylation in the ASC gene, which is involved in IL-1 β and IL-18 secretion and whose expression increases with age. Age-induced reductions in ASC methylation led to an increase in pro-inflammatory status, however, 6 months of high-intensity intermittent walking reduced age-dependent reductions in ASC methylation and inhibited excess pro-inflammatory cytokines by reducing ASC expression.

Conclusion and outlook

This review describes that the changes in apoptosis, inflammatory reaction, and autophagy are significant contributors to the pathogenesis of OP. Besides, epigenetic changes, such as changes in non-coding RNA and DNA methylation, are also crucial in the pathogenesis of OP. Exercise-induced changes in mechanical stress, hormones, cytokines, epigenetics, and signaling pathways can regulate these pathological changes, thus regulating bone metabolism, thereby promoting bone formation. Different sports items, intensity, duration, and frequency may have varying effects on the body.

At present, there are few studies on the effects of exercise on circRNAs, DNA methylation, and autophagy in osteoporotic people. To investigate the mechanism of exercise to alleviate OP and to establish the foundation for clinical treatment, more clinical research as well as *in vitro* and *in vivo* trials are required.

Author contributions

X-QW and HZ led the conception and design. LZ led the drafting and reviewing the manuscript. Y-LZ, RW were involved in the editing/revision process. All authors approved the final version and were involved in realization of this review.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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