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Review article

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Improving cell reinfusion to enhance the efficacy of chimeric antigen receptor T-cell therapy and alleviate complications

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ABSTRACT

Adoptive cell therapy (ACT) is a rapidly expanding area within the realm of transfusion medicine, focusing on the delivery of lymphocytes to trigger responses against tumors, viruses, or inflammation. This area has quickly evolved from its initial promise in immuno-oncology during preclinical trials to commercial approval of chimeric antigen receptor (CAR) T-cell therapies for leukemia and lymphoma (Jun and et al., 2018) [1]. CAR T-cell therapy has demonstrated success in treating hematological malignancies, particularly relapsed/refractory B-cell acute lymphoblastic leukemia and non-Hodgkin's lymphoma (Qi and et al., 2022) [2]. However, its success in treating solid tumors faces challenges due to the short-lived presence of CAR-T cells in the body and diminished T cell functionality (Majzner and Mackall, 2019) [3]. CAR T-cell therapy functions by activating immune effector cells, yet significant side effects and short response durations remain considerable obstacles to its advancement. A prior study demonstrated that the therapeutic regimen can induce systemic inflammatory reactions, such as cytokine release syndrome (TLS), off-target effects, and other severe complications. This study aims to explore current research frontiers in this area.

1. Introduction

CARs are designed to equip T cells with the ability to target and damage cancer cells. They combine an external domain, which identifies cancer-specific antigens using a single-chain variable fragment (scFv) from antibody segments, with an internal domain that activates T cells through elements of the T cell receptor (TCR) and costimulatory signals. This structure ensures precise targeting and effective T cell activation against tumors [1,2]. CAR T-cell therapy is a powerful treatment option for hematologic malignancies [3–5]. In this approach, the patient's own T cells are collected and engineered to recognize target antigens. These cells are then expanded in vitro before being reintroduced into patients for *in vivo* treatment [6]. Subsequently, CAR T cells engage and eliminate targeted tumor cells they encounter. Briefly, the progress of CAR T-cell approach has been described as follows (Fig. 1). CAR T-cell therapy exhibits high effectiveness against certain challenging cancers, yet it can also lead to serious or even life-threatening side effects. This method activates CAR T cells when they identify particular antigens on cancer cells inside the body. As CAR T cells proliferate, they can release significant amounts of cytokines into the bloodstream, leading to heightened immune activity. This release may result in severe side effects such as high fever, chills, respiratory difficulties, intense nausea, vomiting, diarrhea, dizziness, headaches, rapid heartbeat, extreme fatigue, and muscle or joint pain [7–9]. Clinical strategies have been employed to enhance the efficacy of CAR T-cell therapy

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while mitigating potential risks. Studies have indicated that improving the CAR T cells reinfusion scheme can enhance the therapeutic impact and lower complication rates. Similarly, this study aims to summarize the research scope and application status of improving the CAR T cells reinfusion scheme to enhance its therapeutic effect and avoid complications.

2. Concise overview concerning CAR T-cell therapy

Artificial transmembrane receptors known as CAR T-cell receptors are composed of a single-chain extracellular antibody fragment and intracellular signaling domains taken from the T-cell receptor and costimulatory molecules. These receptors are designed for specific recognition and activation within T cells [10]. As engineered proteins, CAR-T cells can control cell activation and guide cytotoxic lymphocytes to specific antigens [11]. The antigen-recognition domain of CAR T cells primarily relies on sequences from monoclonal antibodies, which interact with tumor epitopes without the requirement for MHC presentation. Currently, CAR T-cell technology shows great potential in addressing hematologic malignancies and other medical conditions [12], suggesting its potential as a broadly applicable cancer treatment modality [13].

3. Pretreatment of CAR T cells reinfusion

3.1. Mechanism of pretreatment to improve therapeutic effect

Pretreatment preceding CAR T-cell immunotherapy involves lymphocyte depletion primarily through chemotherapy or radiotherapy [14]. This pretreatment regimen aims to decrease lymphocyte numbers while maintaining stable cytokine levels, thereby facilitating enhanced contact between CAR T cells and cytokines to support their stable proliferation [15,16]. Some studies have shown that administering cyclophosphamide and fludarabine before CAR T cells reinfusion can inhibit the expression of indoleamine 2,



Fig. 1. Chimeric antigen receptor T-cell therapy.

3-dioxygenase in mouse B-cell malignant tumors, thereby reducing tryptophan metabolism and increasing CAR T-cell proliferation and activity [17]. Clinical evidence suggests that pretreatment can enhance the efficacy of CAR T-cell immunotherapy [18,19]. Brentjens studied 10 patients with acute lymphocytic leukemia and chronic lymphoblastic leukemia to compare the efficacy of cyclophosphamide pretreatment and non-pretreatment. The study showed that patients receiving cyclophosphamide pretreatment had stronger CAR T cells persistence and clinical activity than those with non-pretreatment [18]. Zhang performed a meta-analysis of the clinical data of 131 patients with recurrent/refractory B-cell malignancies and compared the 6 months progression free survival rate of patients with or without pretreatment before CAR T cells reinfusion [20]. The study showed that the progression free survival rate of patients with pretreatment before CAR T cells reinfusion was 94.6%, which was higher than the 54.5% of the with-pretreatment group (P < 0.001) [21]. A study by Caimi proved that administering tocilizumab prophylactically prior to CD19-CAR T cell reinfusion significantly mitigates CRS severity [22]. Therefore, the pretreatment scheme is of great importance for CAR T-cell immunotherapy.

3.2. Pretreatment scheme selection

To explore the best pretreatment scheme of CAR T-cell therapy, a series of clinical studies focused on the necessity of pretreatment scheme, the choice of drugs and dosage, and the treatment of subsequent adverse reactions [19]. In the selection of chemotherapy treatments, the combination of fludarabine and cyclophosphamide currently stands as the most commonly adopted regimen for lymphodepletion [23,24]. A clinical trial on 30 patients with B-cell acute lymphoblastic leukemia was carried out in the Fred Hutchinson Cancer Research Center [25,26]. Studies have found that the combination of fludarabine and cyclophosphamide is linked to improved disease-free survival and enhanced persistence of CAR T cells, in comparison to the cyclophosphamide-etoposide combination or cyclophosphamide alone. In addition to this scheme, various other schemes containing bendamustine, pentostatin, and polychemotherapy, although possible, are seldom used [27,28].

In addition to the pretreatment of chemotherapy drugs, total body irradiation (TBI) has also been applied to the pretreatment of adoptive cellular transfer immunotherapy, represented by CAR T cells reinfusion [29,30]. Researchers conducted a clinical trial on patients with metastatic melanoma. Patients were categorized into two cohorts: the first received pretreatment with fludarabine and cyclophosphamide, while the second underwent a regimen of chemotherapy in conjunction with total body irradiation (TBI). The results showed that the response rate of chemotherapy pretreatment alone was 49%, and that of the other group had a maximum of 72% [31]. Comparable studies have demonstrated that mice receiving radiotherapy before the infusion of CD19-CAR T cells exhibited extended survival times compared to those that did not undergo radiotherapy [32]. Therefore, chemotherapy combined with radio-therapy can improve the therapeutic effect, but confirming the efficacy and safety of this method requires further study.

Except of the above pretreatment schemes, some researcher established evidence-based approaches of prophylactic immunoglobulin. They made some suggestions that patients before CAR T cells reinfusion for B-cell malignancies with IgG \leq 400 mg/dL shuold consider prophylactic IgG. Additionally, they recommend monitoring serum IgG levels both before and during the first three months following CAR T-cell approach for B-cell malignancies. Historically, corticosteroids have been omitted from the CAR T-cell therapy protocol because of their ability to harm lymphocytes [27]. However, some clinical trials have proved that prophylactic use of dexamethasone can effectively decrease severity of CRS [33]. Furthermore, research has shown that administering antihistamines and antipyretics prior to CAR T-cell reinfusion can aid in mitigating adverse reactions. The efficacy of this pretreatment approach, however, requires further empirical validation [34].

3.3. Pretreatment dose selection

Regarding the dosage of pretreatment, studies have shown that appropriately reducing the pretreatment dose can achieve similar clinical effects and reduce toxicity [35,36]. However, without any pretreatment, the effective remission rate of the disease is reduced [37]. The National Cancer Institute has conducted two studies at different times. In the first study, patients with B-lymphomas were administered high doses of cyclophosphamide and fludarabine as a preparatory regimen prior to receiving CAR T-cell infusion. The second study utilized low doses of cyclophosphamide and fludarabine for patients with non-Hodgkin lymphoma prior to CAR T-cell reinfusion. The outcomes indicated that 87% of patients in the first study experienced acute adverse reactions following CAR T-cell reinfusion. In contrast, the second study reported a significantly lower incidence of such reactions following CAR T-cell reinfusion [35]. The reasons for the different results may be due to different types of diseases or different dosag of chemotherapy drugs. However, these findings also highlight a valuable research direction, emphasizing that choosing the correct pretreatment dosage is crucial for augmenting treatment effectiveness and diminishing the occurrence of complications.

4. Management in CAR T cells reinfusion process

4.1. Observation of the Patient's condition

Observation of the patient's condition is an important work before CAR T cells reinfusion. Patients should be hospitalized to monitor oxygen saturation, vital signs, urinary output and other indicators. Meanwhile, during reinfusion emergency materials should be placed beside the patients [27,28]. Studies have suggested that the possibility of CRS and adverse nervous system reactions after cell reinfusion is higher and more serious in patients with neoplastic fever before cell reinfusion [38]. Therefore, it is essential to promptly address any discomfort experienced by patients before reinfusion. In the process of reinfusion, the patients may have vomiting, nausea, fever, abdominal pain and tremors. Respiratory depression, neurological symptoms or cardiac arrhythmias also may be observed [39].

In such instances, healthcare professionals, including nurses and doctors, should consider slowing down or temporarily stopping the reinfusion process, initiating emergency interventions, and verifying the product being used [40].

4.2. CAR T cells thawing

A 10% DMSO solution is widely recognized as the standard freezing medium for CAR T cells [41]. Recently, studies have lowered the concentration of DMSO from 10% to either 7.5% or 5% [42,43]. Furthermore, investigations are underway into a cryopreservation solution comprised of 50/55% DMSO mixed with 5% dextrose as an innovative substitute for the DMSO-dextran combination. This alternative approach is targeted at preventing the aggregation of DNA and proteins from lysed cells, as well as mitigating cell swelling during the thawing phase, potentially improving the overall viability and functional integrity of the cells post-thaw [44]. The thawing process is crucial for ensuring the quality of infused cells and the clinical effectiveness of the product [45]. When thawing the CAR T cell bag, either on-site at the pharmacy or at a subcontracted cell processing facility, double-wrap it in protective plastic and place it in a dedicated 37 ± 2 °C water bath in a clean room, until the product is completely thawed [46]. Unsurprisingly, the optimal thawing rate is influenced by the rate of cooling employed [47]. It is advisable to double-wrap the CAR T cell bag in a waterproof plastic covering during thawing. This measure serves to safeguard the integrity of the bag and enables nurses to detect any unnoticed solution leakage during the transfer process [46]. Recently, researchers have introduced a thawing technique that eliminates the need for a water bath, thereby reducing the risk of contamination during thawing [41]. This alternative approach usually entails applying mechanical heat, such as utilizing a heated metallic plate or sealed warm liquids that do not come into direct contact with the thawed sample [48,49]. However, adoption of this technology remains limited.

4.3. CAR T cells reinfusion

Before reinfusing CAR T cells, patients' clinical status must be thoroughly reassessed, encompassing vital signs (body temperature, blood pressure, blood oxygen saturation, heart rate), presence of active infections, and organ function. Active infection and hypotension requiring vasopressor therapy are contraindications to CAR T cells reinfusion, and CAR T cells reinfusion needs to be delayed until infection or hypotension is completely treated or controlled [27]. Some studies suggest that patients with tumor fever before reinfusion are more likely to have CRS and nervous system adverse reactions after cell reinfusion, and are more serious [50]. In addition, corticosteroids should not be used before infusion unless in life-threatening situations, preventing the destruction of CAR T cells expansion and survival. Acetaminophen derivatives and antihistamines, like chlorphenamine maleate or diphenhydramine, are commonly given 30-60 min prior to CAR T cell reinfusion to decrease the likelihood of infusion reactions [27]. The widespread use of steroids is prohibited during and after transfusion (unless used to treat adverse reactions related to cells reinfusion, such as CRS or neurological adverse reactions) [27]. Mononuclear macrophages are significantly involved in the initiation and progression of adverse reactions associated with CAR T-cell therapy [51,52]. Granulocyte-macrophage colony-stimulating factor (GM-CSF) can stimulate macrophages, aggravating cell reinfusion related adverse reactions. Although some studies have shown that the application of GM-CSF neutralizing antibodies may be able to eliminate CRS and neurological adverse reactions and enhance CAR T cells function, but this mechanism needs further research to confirm [53,54]. In addition, the vital signs of the patients should be closely observed during the CAR T cells reinfusion process. The process should be carried out with reference to the product instructions, and the emergency equipment should be in a standby state [55,56].

5. Observation of complications after CAR T cells reinfusion

5.1. CRS

Cytokine storm, known as CRS, is a severe systemic inflammatory response syndrome that can be initiated by infection or the use of specific medications [57,58]. CRS is accompanied by a series of biochemical changes. Davila argued that IFN- γ , IL-6, IL-5, IL-10, Flt-3L, GM-CSF, and fractalkine are closely related to severe CRS [59]. Different researchers have different conclusions in this respect. A number of cytokines peak after the occurrence of CRS, including ferritin, CRP, AST, ALT, and BUN, thus they are not helpful in

Table 1	
ASTAC CRS consensus grading ^a	[64].

CRS Parameter	Grade 1	Grade 2	Grade 3	Grade 4
Fever ^b	Temperature ≥38.5 °C	Temperature \geq 38.5 °C	Temperature \geq 38.5 °C	Temperature \geq 38.5 °C
Hypotension ^b	None	Not necessitating vasopressors	Necessitating the use of a vasopressor, with or without vasopressin	Necessitating multiple vasopressors (excluding vasopressin)
Hypoxia ^b	None	Necessitating the use of low-flow nasal cannula (≤6 L/minute)	Necessitating high-flow nasal cannula (>6 L/minute) or facemask	Necessitating positive pressure

^a The CRS grade is determined based on the most severe event experienced by the patient: For instance, a patient with a temperature of 39.0 °C, hypotension requiring vasopressors, and hypoxia necessitating positive pressure is classified as grade 4 CRS.

⁹ Fever hypotension and hypoxia are not attributable to any other cause.

predicting CRS [8,60]. Besides observing the biochemical changes, it is also necessary to observe the symptoms of patients. The symptoms of CRS may manifest progressively, with primary clinical presentations including fever, low blood pressure, hypoxia, and end-organ dysfunction. However, these symptoms are not exclusive to CRS [61–63]. Furthermore, there clinical manifestations must be a reasonable time relationship between diagnosis of CRS and cell therapy. Experts from the American Society for Transplantation and Cellular Therapy (ASTCT) have developed a grading system to accurately evaluate the severity of CRS. This system is currently the predominant standard utilized in the field [64] (Table 1).

5.2. ICANS

Research has shown that patients undergoing CD19-CAR T-cell therapy may develop ICANS at an incidence varying between 21% and 64%. Similarly, BCMA-CAR T-cell therapy has been associated with ICANS in 18%–21% of patients. The severity of ICANS, classified as grade \geq 3, has been reported in 10%–30% of CD19-CAR T-cell cases and in 3%–9% of BCMA-CAR T-cell cases [65–70]. The onset typically occurs around one week after reinfusion [71–73]. Clinical manifestations of ICANS vary widely, spanning from tremors, confusion, and dysphasia/aphasia to deteriorating handwriting or seizures [64]. Schoeberl et al. discovered link between elevated preinfusion serum levels of neurofilament light chain, an indicator of neuroaxonal damage, and the intensity of ICANS [74]. Gust et al. suggest that pre-existing neurological comorbidities (e.g., peripheral neuropathy and headache disorder) are risk factors for ICANS [60]. Nurses should carefully observe the clinical manifestations of patients after CAR T cells reinfusion and strive to identify ICANS in advance to reduce its impact on patients.

5.3. TLS

TLS refers to the rapid release of cell contents into the blood due to the massive dissolution and destruction of tumor cells, where a series of complications are caused by the accumulation of metabolites due to exceeding the liver metabolism and renal excretion capacity. TLS mainly manifests as hyperuricemia, hyperkalemia, hyperphosphatemia, hypocalcemia, and metabolic acidosis [75]. TLS is not a specific complication of CAR T-cell therapy and can occur in patients undergoing chemotherapy. Therefore, it is very important to accurately identify the causes of TLS and early medication is beneficial to prevent the occurrence of TLS [76]. Allopurinol is recommended as the primary preventive medication for patients classified as low or moderate risk of TLS [77,78]. Conversely, rasburicase is the preferred choice for both prevention and treatment of TLS in patients at high risk [79]. To detect TLS, nurses must closely monitor the patient's vital signs and renal function, while accurately documenting their 24-h intake and output. Nurses should adhere to the physician's guidance and provide appropriate care accordingly.

5.4. Off-target effect

Tumor antigens include tumor-specific antigens (TSA) and tumor-associated antigens (TAA). The TAS on the tumor surface are relatively lacking. Consequently, CAR T cells may identify and eradicate tumor cells, potentially causing unintended damage to normal tissues with low expression of target antigens, leading to off-target effects [80]. Therefore, CAR T cells can attack normal tissues and cause damage to them and organs while clearing tumors [81]. Beatty's study pointed out that the off-target effect can cause toxic reactions such as cardiac arrest, respiratory failure, intestinal obstruction, and abdominal pain [82]. To minimize off-target effects, it is recommended to regulate the reinfusion dose and route of CAR T cells. Typically, toxicity levels are positively associated with the dose of CAR T cells, underscoring the importance of carefully selecting the initial dose for the first treatment [83]. It is recommended to use a stepwise dose reinfusion scheme, that is, slowly increasing the dose of reinfused CAR T cells. Change the reinfusion route of CAR T cells can reduce the occurrence of off-target reaction, such as intrapleural injection of mesothelin-targeted CAR T cells for the treatment of malignant pleural tumors, intracranial injection for the treatment of glioblastoma multiforme. Intratumoral injection makes most CAR T cells accumulate in the range of lesions, enhances the therapeutic effect, and limits its targeted toxicity to normal tissues [84,85].

Table 2	
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Other complications	and	related	factors.
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Complications	Symptoms	Related Factors
Infections of CAR T cells reinfusion	Bloodstream infections, respiratory tract infections, hypogammaglobulinemia	Nosocomial infections, unreasonable use of glucocorticoids, B-cell dysplasia and hypogammaglobulinemia, high-grade CRS
Hemophagocytic	Fever, hepatosplenomegaly, abnormal liver function, decreased	A large number of inflammatory cytokines release due to lysis of
lymphohistiocytosis	blood cell count, elevated triglycerides, increased serum	tumor cells and pro-inflammatory cytokines, released cytokines of
	ferritin, decreased fibrinogen levels	activated CAR T cells, high levels of IFN- γ , increased serum ferritin, viral infections
Coagulation disorders	Increased D-dimer, prolonged prothrombin time, increased	Liver damage, increased levels of cytokines, damage to endothelial
	fibrinogen degradation products, decreased fibrinogen, thrombocytopenia	cells, hypercoagulable blood of malignant tumor patients
Cytopenias	Neutropenia, thrombocytopenia, anemia	Higher-intensity chemotherapy, history of hematopoietic stem cell transplantation, high-grade CRS

5.5. Other complications and related factors

CRS, ICANS, TLS, and off-target effect are major complications. In addition, other complications also affect the patient's health. Some of the symptoms and related factors of complications are listed as follows (Table 2).

6. Conclusion

Many investigations have confirmed that the improved CAR T cells reinfusion scheme can improve its therapeutic effect and avoid or reduce the occurrence of complications. Administering cyclophosphamide and fludarabine prior to CAR T cell reinfusion has been demonstrated to improve the proliferation and efficacy of CAR T cells. Additionally, tocilizumab is frequently utilized as a pretreatment drug [17,18,22]. In terms of the choice of pretreatment dose some studies have shown that appropriately reducing the dose can achieve similar clinical effects [35,36]. In addition, a series of clinical trials have confirmed that TBI is also used as a pretreatment before CAR T cells reinfusion [27]. Further investigations are needed to confirm the efficacy and safety of chemotherapy combined with radiotherapy in improving the effectiveness of pretreatment. Monitoring the vital signs of patients undergoing CAR T cells reinfusion is crucial. In addition, future research directions may include exploring optimal concentrations for frozen solutions of CAR T cells. CRS, ICANS, TLS and off-target effect are common complications after CAR T cells reinfusion. Many cytokines have important predictive effects on them [8,59,60]. However, how to accurately and early identify the risk factors of complications and take appropriate treatment measures is an important issue for current researchers. In addition, some complications have different stages of development, and how to prevent the progression of complications is also an urgent problem for researchers.

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Data availability statement

There is no additional data available for this study.

Ethics declarations

Review and approval by an ethics committee were not necessary for this study, and informed consent was not required as the study did not involve animal experiments or human behavioral studies. Instead, we conducted a review of published research and provided appropriate references.

CRediT authorship contribution statement

Zhihao Han: Writing – original draft, Conceptualization. Xiaoqin Ma: Writing – review & editing, Methodology. Guiyue Ma: Writing – review & editing.

Declaration of competing interest

The authors declare no competing financial interests or commercial relationships that could be perceived as influencing the outcome of this research.

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References

- [1] J. Chen, et al., Tuning charge density of chimeric antigen receptor optimizes tonic signaling and CAR-T cell fitness, Cell Res. 33 (2023) 341–354.
- [2] P. Ramesh, et al., Chimeric antigen receptor T-cells: Properties, production, and quality control, Int. J. Lit. Humanit. 45 (2023) 425–435.
- [3] J.N. Kochenderfer, et al., Eradication of B-lineage cells and regression of lymphoma in a patient treated with autologous T cells genetically engineered to recognize CD19, Blood 116 (2010) 4099–4102.

[4] Y. Zhao, et al., Multiple injections of electroporated autologous T cells expressing a chimeric antigen receptor mediate regression of human disseminated tumor, Cancer Res. 70 (2010) 9053–9061.

- [5] M. Kalos, et al., T cells with chimeric antigen receptors have potent antitumor effects and can establish memory in patients with advanced leukemia, Sci. Transl. Med. 3 (2011), 95-73.
- [6] C.H. Jun, et al., CAR T cell immunotherapy for human cancer, Science (New York, N.Y.) 359 (2018) 1361–1365.
- [7] D.W. Lee, et al., Current concepts in the diagnosis and management of cytokine release syndrome, Blood 124 (2014) 188–195.
- [8] K.A. Hay, et al., Kinetics and biomarkers of severe cytokine release syndrome after CD19 chimeric antigen receptor-modified T-cell therapy, Blood 130 (2017) 2295–2306.
- [9] A. Shimabukuro-Vornhagen, et al., Cytokine release syndrome, Journal for immunotherapy of cancer 6 (2018) 56.
- [10] J.N. Kochenderfer, et al., Construction and preclinical evaluation of an anti-CD19 chimeric antigen receptor, J. Immunother. 32 (2009) 689–702.

- [11] L.L. Li, et al., A brief review concerning Chimeric Antigen Receptors T cell therapy, J. Cancer 11 (2020) 5424–5431.
- [12] W.A. Lim, C.H. June, The Principles of engineering immune cells to treat cancer, Cell 168 (2017) 724–740.
- [13] S. Gill, C.H. June, Going viral: chimeric antigen receptor T-cell therapy for hematological malignancies, Immunol. Rev. 263 (2015) 68-89.
- [14] L. Hershkovitz, et al., Focus on adoptive T cell transfer trials in melanoma, Clin. Dev. Immunol. 2010 (2010) 260-267.
- [15] L. Gattinoni, et al., Removal of homeostatic cytokine sinks by lymphodepletion enhances the efficacy of adoptively transferred tumor-specific CD8+ T cells, J. Exp. Med. 202 (2005) 907–912.
- [16] L. Bracci, et al., Cyclophosphamide enhances the antitumor efficacy of adoptively transferred immune cells through the induction of cytokine expression, B-cell and T-cell homeostatic proliferation, and specific tumor infiltration, Clin. Cancer Res. : an official journal of the American Association for Cancer Research 13 (2007) 644–653.
- [17] S. Ninomiya, et al., Tumor indoleamine 2,3-dioxygenase (Ido) inhibits CD19-CAR T cells and is downregulated by lymphodepleting drugs, Blood 125 (2015) 3905–3916.
- [18] R.J. Brentjens, et al., Safety and persistence of adoptively transferred autologous CD19-targeted T cells in patients with relapsed or chemotherapy refractory B-cell leukemias, Blood 118 (2011) 4817–4828.
- [19] H.J. Pegram, et al., Tumor-targeted T cells modified to secrete IL-12 eradicate systemic tumors without need for prior conditioning, Blood 119 (2012) 4133-4141
- [20] M. Yu, et al., Efficacy and safety of Dual-targeting chimeric antigen receptor-T therapy for relapsed or refractory B cell lymphoid malignancies: a systematic review and meta-analysis, Hum. Gene Ther. 34 (2023) 192–202.
- [21] T. Zhang, et al., Efficiency of CD19 chimeric antigen receptor-modified T cells for treatment of B cell malignancies in phase I clinical trials: a meta-analysis, Oncotarget 6 (2015) 33961–33971.
- [22] P.F. Caimi, et al., Prophylactic tocilizumab prior to anti-CD19 CAR-T cell therapy for non-hodgkin lymphoma, Front. Immunol. 12 (2021) 745320.
- [23] C.J. Turtle, et al., Durable molecular remissions in chronic lymphocytic leukemia treated with CD19-specific chimeric antigen receptor-modified T cells after failure of ibrutinib, J. Clin. Oncol. : official journal of the American Society of Clinical Oncology 35 (2017) 3010–3020.
- [24] S.S. Neelapu, et al., Chimeric antigen receptor T-cell therapy assessment and management of toxicities, Nat. Rev. Clin. Oncol. 15 (2018) 47-62.
- [25] C.J. Turtle, et al., Immunotherapy of non-Hodgkin's lymphoma with a defined ratio of CD8+ and CD4+ CD19-specific chimeric antigen receptor-modified T cells, Sci. Transl. Med. 8 (2016), 355-116.
- [26] K.A. Hay, et al., Factors associated with durable EFS in adult B-cell ALL patients achieving MRD-negative CR after CD19 CAR T-cell therapy, Blood 133 (2019) 1652–1663.
- [27] I. Yakoub-Agha, et al., Management of adults and children undergoing chimeric antigen receptor T-cell therapy: best practice recommendations of the European Society for Blood and Marrow Transplantation (EBMT) and the Joint Accreditation Committee of ISCT and EBMT (JACIE), Haematologica 105 (2020) 297–316.
 [28] K.M. Mahadeo, et al., Management guidelines for paediatric patients receiving chimeric antigen receptor T cell therapy, Nat. Rev. Clin. Oncol. 16 (2019) 45–63.
- [29] B.R. Shank, et al., Chimeric antigen receptor T cells in hematologic malignancies, Pharmacotherapy 37 (2017) 334-345.
- [30] M.C. Jensen, et al., Antitransgene rejection responses contribute to attenuated persistence of adoptively transferred CD20/CD19-specific chimeric antigen receptor redirected T cells in humans, Biology of blood and marrow transplantation, journal of the American Society for Blood and Marrow Transplantation 16 (2010) 1245–1256.
- [31] M.E. Dudley, et al., Adoptive cell therapy for patients with metastatic melanoma: evaluation of intensive myeloablative chemoradiation preparative regimens, J. Clin. Oncol. : official journal of the American Society of Clinical Oncology 26 (2008) 5233–5239.
- [32] J.N. Kochenderfer, et al., Adoptive transfer of syngeneic T cells transduced with a chimeric antigen receptor that recognizes murine CD19 can eradicate lymphoma and normal B cells, Blood 116 (2010) 3875–3886.
- [33] O.O. Oluwole, et al., Prophylactic corticosteroid use in patients receiving axicabtagene ciloleucel for large B-cell lymphoma, Br. J. Haematol. 194 (2021) 690–700.
- [34] G. Granroth, et al., Supportive care for patients with lymphoma undergoing CAR-T-cell therapy: the advanced practice provider's perspective, Curr. Oncol. Rep. 24 (2022) 1863–1872.
- [35] J.N. Kochenderfer, et al., Chemotherapy-refractory diffuse large B-cell lymphoma and indolent B-cell malignancies can be effectively treated with autologous T cells expressing an anti-CD19 chimeric antigen receptor, J. Clin. Oncol. : official journal of the American Society of Clinical Oncology 33 (2015) 540–549.
- [36] J.N. Kochenderfer, et al., Lymphoma remissions caused by anti-CD19 chimeric antigen receptor T cells are associated with high serum interleukin-15 levels, J. Clin. Oncol. : official journal of the American Society of Clinical Oncology 35 (2017) 1803–1813.
- [37] J.N. Brudno, et al., Allogeneic T cells that express an anti-CD19 chimeric antigen receptor induce remissions of B-cell malignancies that progress after allogeneic hematopoietic stem-cell transplantation without causing graft-versus-host disease, J. Clin. Oncol. : official journal of the American Society of Clinical Oncology 34 (2016) 1112–1121.
- [38] F.L. Locke, et al., Phase 1 results of ZUMA-1: a multicenter study of KTE-C19 anti-CD19 CAR T cell therapy in refractory aggressive lymphoma, molecular therapy, the journal of the American Society of Gene Therapy 25 (2017) 285–295.
- [39] D.V. Clé, et al., Associação Brasileira de Hematologia, Hemoterapia e Terapia Celular Consensus on genetically modified cells. I: structuring centers for the multidisciplinary clinical administration and management of CAR-T cell therapy patients, Hematology, transfusion and cell therapy 43 (2021) 3–12.
- [40] Z. Shu, S. Heimfeld, D. Gao, Hematopoietic SCT with cryopreserved grafts: adverse reactions after transplantation and cryoprotectant removal before infusion, Bone Marrow Transplant. 49 (2014) 469–476.
- [41] R. Li, , et al.A. Hubel, Preservation of cell-based immunotherapies for clinical trials, Cytotherapy 21 (2019) 943–957.
- [42] A.L. Garfall, et al., Chimeric antigen receptor T cells against CD19 for multiple myeloma, N. Engl. J. Med. 373 (2015) 1040–1047.
- [43] X. Wang, et al., Phase 1 studies of central memory-derived CD19 CAR T-cell therapy following autologous HSCT in patients with B-cell NHL, Blood 127 (2016) 2980–2990.
- [44] D. Raffo, et al., Evaluation of DMSO dextrose as a suitable alternative for DMSO dextran in cord blood cryopreservation, Vox Sang. 114 (2019) 283-289.
- [45] A. Joules, et al., Comparative analysis of cell therapy infusion workflows at clinical sites, Cytotherapy 23 (2021) 285–292.
- [46] C. Rioufol, et al., The EBMT/EHA CAR-T Cell Handbook, The Author(s), Springer Copyright 2022, Cham, 2022, pp. 37-43 (CH).
- [47] P. Mazur, A biologist's view of the relevance of thermodynamics and physical chemistry to cryobiology, Cryobiology 60 (2010) 4-10.
- [48] P. Kilbride, et al., Automated dry thawing of cryopreserved haematopoietic cells is not adversely influenced by cryostorage time, patient age or gender, PLoS One 15 (2020) 0240310.
- [49] C.J. Hunt, Technical considerations in the freezing, low-temperature storage and thawing of stem cells for cellular therapies, Transfus. Med. Hemotherapy : offizielles Organ der Deutschen Gesellschaft fur Transfusionsmedizin und Immunhamatologie 46 (2019) 134–150.
- [50] J. Gauthier, et al., Factors associated with outcomes after a second CD19-targeted CAR T-cell infusion for refractory B-cell malignancies, Blood 137 (2021) 323–335.
- [51] T. Giavridis, et al., CAR T cell-induced cytokine release syndrome is mediated by macrophages and abated by IL-1 blockade, Nat. Med. 24 (2018) 731–738.
- [52] M. Norelli, et al., Monocyte-derived IL-1 and IL-6 are differentially required for cytokine-release syndrome and neurotoxicity due to CAR T cells, Nat. Med. 24 (2018) 739–748.
- [53] R.M. Sterner, et al., GM-CSF inhibition reduces cytokine release syndrome and neuroinflammation but enhances CAR-T cell function in xenografts, Blood 133 (2019) 697–709.
- [54] S. Shang, et al., RNA silencing of GM-CSF in CAR-T cells reduces the secretion of multiple inflammatory cytokines, Invest. N. Drugs 41 (2023) 220–225.
- [55] E.M. Holland, et al., Chimeric antigen receptor T cells as salvage therapy for post-chimeric antigen receptor T cell failure, Transplantation and cellular therapy 29 (2023) 1–10.
- [56] D.P. Turicek, et al., CAR T-cell detection scoping review: an essential biomarker in critical need of standardization, Journal for immunotherapy of cancer 11 (2023).

Z. Han et al.

- [57] F. Korell, et al., EASIX and severe endothelial complications after CD19-directed CAR-T cell therapy-A cohort study, Front. Immunol. 13 (2022) 877477.
- [58] M.L. Schubert, et al., Side-effect management of chimeric antigen receptor (CAR) T-cell therapy, Ann. Oncol. : official journal of the European Society for Medical Oncology 32 (2021) 34–48.
- [59] M.L. Davila, et al., Efficacy and toxicity management of 19-28z CAR T cell therapy in B cell acute lymphoblastic leukemia, Sci. Transl. Med. 6 (2014) 224–225.
- [60] J. Gust, et al., Endothelial activation and blood-brain barrier disruption in neurotoxicity after adoptive immunotherapy with CD19 CAR-T cells, Cancer Discov. 7 (2017) 1404–1419.
- [61] K.L. Chohan, E.L. Siegler, S.S. Kenderian, CAR-T cell therapy: the efficacy and toxicity balance, Current hematologic malignancy reports 18 (2023) 9-18.
- [62] N. Gazeau, et al., Anakinra for refractory cytokine release syndrome or immune effector cell-associated neurotoxicity syndrome after chimeric antigen receptor T cell therapy, Transplantation and cellular therapy 29 (2023) 430–437.
- [63] M.D. Jain, M. Smith, N.N. Shah, How I treat refractory CRS and ICANS after CAR T-cell therapy, Blood 141 (2023) 2430-2442.
- [64] D.W. Lee, et al., ASTCT Consensus grading for cytokine release syndrome and neurologic toxicity associated with immune effector cells, biology of blood and marrow transplantation, journal of the American Society for Blood and Marrow Transplantation 25 (2019) 625–638.
- [65] S.L. Maude, et al., Tisagenlecleucel in children and young adults with B-cell lymphoblastic leukemia, N. Engl. J. Med. 378 (2018) 439-448.
- [66] S.J. Schuster, et al., Tisagenlecleucel in adult relapsed or refractory diffuse large B-cell lymphoma, N. Engl. J. Med. 380 (2019) 45–56.
- [67] J.S. Abramson, et al., Lisocabtagene maraleucel for patients with relapsed or refractory large B-cell lymphomas (TRANSCEND NHL 001): a multicentre seamless design study, Lancet (London, England) 396 (2020) 839–852.
- [68] M. Wang, et al., KTE-X19 CAR T-cell therapy in relapsed or refractory mantle-cell lymphoma, N. Engl. J. Med. 382 (2020) 1331–1342.
- [69] N.C. Munshi, et al., Idecabtagene vicleucel in relapsed and refractory multiple myeloma, N. Engl. J. Med. 384 (2021) 705-716.
- [70] J.G. Berdeja, et al., Ciltacabtagene autoleucel, a B-cell maturation antigen-directed chimeric antigen receptor T-cell therapy in patients with relapsed or refractory multiple myeloma (CARTITUDE-1): a phase 1b/2 open-label study, Lancet (London, England) 398 (2021) 314–324.
- [71] D.B. Rubin, et al., Clinical predictors of neurotoxicity after chimeric antigen receptor T-cell therapy, JAMA Neurol. 77 (2020) 1536–1542.
- [72] P. Strati, et al., Clinical and radiologic correlates of neurotoxicity after axicabtagene ciloleucel in large B-cell lymphoma, Blood advances 4 (2020) 3943–3951.
 [73] C. Belin, et al., Description of neurotoxicity in a series of patients treated with CAR T-cell therapy, Sci. Rep. 10 (2020) 18997.
- [74] F. Schoeberl, et al., Neurofilament light chain serum levels correlate with the severity of neurotoxicity after CAR T-cell treatment, Blood advances 6 (2022) 3022–3026.
- [75] L. Baeksgaard, J.B. Sørensen, Acute tumor lysis syndrome in solid tumors–a case report and review of the literature, Cancer Chemother. Pharmacol. 51 (2003) 187–192.
- [76] M. Takai, et al., Controlling serum uric acid using febuxostat in cancer patients at risk of tumor lysis syndrome, Oncol. Lett. 8 (2014) 1523–1527.
- [77] T. Barbar, I. Jaffer Sathick, Tumor lysis syndrome, Adv. Chron. Kidney Dis. 28 (2021) 438–446.
- [78] L. Zafrani, E. Canet, M. Darmon, Understanding tumor lysis syndrome, Intensive Care Med. 45 (2019) 1608–1611.
- [79] G.L. Jones, et al., Guidelines for the management of tumour lysis syndrome in adults and children with haematological malignancies on behalf of the British Committee for Standards in Haematology, Br. J. Haematol. 169 (2015) 661–671.
- [80] R.A. Morgan, et al., Case report of a serious adverse event following the administration of T cells transduced with a chimeric antigen receptor recognizing ERBB2, Mol. Ther. : the journal of the American Society of Gene Therapy 18 (2010) 843–851.
- [81] L. Miao, et al., Reactions related to CAR-T cell therapy, Front. Immunol. 12 (2021) 663201.
- [82] G.L. Beatty, et al., Mesothelin-specific chimeric antigen receptor mRNA-engineered T cells induce anti-tumor activity in solid malignancies, Cancer Immunol. Res. 2 (2014) 112–120.
- [83] M. Frigault, et al., Dose fractionation of CAR-T cells. A systematic review of clinical outcomes, J. Exp. Clin. Cancer Res. : CR 42 (2023) 11.
- [84] P.S. Adusumilli, et al., Regional delivery of mesothelin-targeted CAR T cell therapy generates potent and long-lasting CD4-dependent tumor immunity, Sci. Transl. Med. 6 (2014) 151–261.
- [85] C.E. Brown, et al., Regression of glioblastoma after chimeric antigen receptor T-cell therapy, N. Engl. J. Med. 375 (2016) 2561–2569.