

Pathologic Remodeling of Endoneurial Tubules in Human Neuromas

Michael Karsy¹, Cheryl A. Palmer², Mark A. Mahan³

1. Department of Neurosurgery, University of Utah 2. Department of Pathology, University of Utah 3. Neurological Surgery, University of Utah

✉ **Corresponding author:** Mark A. Mahan, mark.mahan@hsc.utah.edu

Disclosures can be found in Additional Information at the end of the article

Abstract

Background: Laminins are extracellular matrix proteins that participate in endoneurial tubule formation and are important in the regeneration of nerves after injury. They act as scaffolds to guide nerves to distal targets and play a key role in neurite outgrowth. Because there is evidence that laminin architecture affects nerve regeneration, we evaluated endoneurial tubules by examining the laminin structure in clinical samples from patients with nerve injuries.

Methods: In a retrospective review of eight nerve injury cases, we evaluated nerve histology in relation to clinical history and injury type. The immunohistochemical delineation of the laminin structure in relationship with the neuroma type was performed.

Results: Five cases of upper-trunk stretch injuries—four from childbirth injury and one from a motorcycle accident—and three cases of nerve laceration leading to neuroma formation were examined. In the upper-trunk stretch injuries, avulsed nerves demonstrated no neuroma formation with a linear laminin architecture and a regular Schwann cell arrangement, but increased fibrous tissue deposition. For neuromas-in-continuity after a stretch injury, laminin immunohistochemistry demonstrated a double-lumen laminin tubule, with encapsulation of the Schwann cells and axonal processes. Nerve laceration leading to stump neuroma formation had a similar double-lumen laminin tubule, but less severe fibrosis.

Conclusions: In nerve injuries with regenerative capacity, endoneurial tubules become pathologically disorganized. A double-lumen endoneurial tubule of unclear significance develops. The consistency of this pattern potentially suggests a reproducible pathophysiologic process. Further exploration of this pathophysiologic healing may provide insight into the failure of programmed peripheral nerve regeneration after injury.

Received 12/11/2017

Review began 12/19/2017

Review ended 01/02/2018

Published 01/18/2018

© Copyright 2018

Karsy et al. This is an open access article distributed under the terms of the Creative Commons Attribution License CC-BY 3.0., which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Categories: Pathology, Neurosurgery

Keywords: endoneurial tubule, extracellular matrix, laminin, peripheral nerve injury, schwann cell

Introduction

Traumatic peripheral nerve injury (PNI) occurs in a wide array of situations, including laceration, concussion, stretch, rupture, or avulsion of peripheral nerves. PNI occurs in 2.8% of all trauma patients, with an incidence of 13–23:100,000 persons/year in developed countries, which represents a notably higher incidence than spinal cord injury [1-2]. Various mechanistic patterns of injury, levels of injury severity, and patient-specific factors (e.g., age, associated secondary injuries) can make PNI a heterogeneous disease process. A hallmark of severe injury

How to cite this article

Karsy M, Palmer C A, Mahan M A (January 18, 2018) Pathologic Remodeling of Endoneurial Tubules in Human Neuromas. Cureus 10(1): e2087. DOI 10.7759/cureus.2087

is the development of a neuroma, which occurs with ineffective nerve fiber regeneration to its target tissue.

The term “neuroma” was first coined by Odier of Geneva in 1811 to describe deep lesions of nerves but did not distinguish nerve tumors from lesions with other mechanisms [3]. Current thought suggests that neuromas are the result of sprouting axons that exit from disrupted perineurium to form a fibrous, disorganized mass of fibroblasts and macrophages [4]. A related aspect, the neuroma-in-continuity, as described by Sunderland [5], describes a partial nerve injury, thickened tissue, Wallerian degeneration of fascicles, and the growth of nerve tissue out of endoneurial growth tubes but within the nerve epineurium. Extracellular matrix (ECM) scaffold proteins are postulated to play a role in nerve regeneration by organizing endoneurial tubules, macrophages, and Schwann cells [6].

We suspected that the pathophysiology of a neuroma-in-continuity is more than fibrous tissue that prevents neurite outgrowth because we recognize that there is fibrosis from surgical manipulation after every nerve grafting surgery. To evaluate our hypothesis that the endoneurial tubule may reveal aspects of the failure of regeneration, we assayed clinical samples from various neuroma-in-continuity and stump neuroma pathologic specimens. One of the important ECM proteins involved in the promotion of neurite outgrowth is laminin. Thus, we suspected that laminin may have the most to reveal about neuromas.

Materials And Methods

After receiving institutional review board approval with a waiver of informed consent, we performed a retrospective chart and pathological review to identify patients of the senior author (MM) who underwent surgical neurolysis and resection of neuromas as part of surgical repair. The pathological review was performed to ensure tissue was available. Histology was performed at the discretion of the senior author and neuropathologist (CP); it included hematoxylin and eosin (H&E), trichrome, and laminin immunohistochemistry (IHC) stains. Five-micrometer, formalin-fixed, paraffin-embedded sections of neuroma resections from all patients were cut at regular intervals and mounted on glass slides. H&E and trichrome stains were prepared on all specimens except for Cases Two and Four. Immunohistochemical staining was performed using laminin antibodies (Leica Biosystems, Wetzlar, Germany) at a dilution of 1:100. Staining was performed using the avidin-biotinylated peroxidase complex (ABC) method on a Ventana Staining system (Ventana Medical Systems, Inc., Arizona, United States) and counterstained with hematoxylin.

The means (ranges) of patient ages and follow-up, along with the mechanism of injury and descriptive histopathological findings, were analyzed. Statistical analysis was not performed with this limited sample.

Results

The summary of all eight patients is shown in Table 1. Case descriptions are presented in the Supplemental Material. Five patients, four infants, and one young adult, with a mean age of 5.8 ± 12.1 years (95% confidence interval (CI) 0.3, 27.4 years; median 0.5 years) had stretch-related/avulsion injuries; and three patients with a mean age of 40.1 ± 14.4 years (CI 26.9, 55.5 years) had transection injuries. Stretch-related injuries occurred in two males and three females, mostly infants. Transection injuries occurred in two males and one female, all adults. At surgery, there were five neuroma-in-continuity injuries, four from birth injuries and one stab wound; two stump neuromas, both iatrogenic; and one spinal nerve avulsion injury, with discontinuity of the peripheral nerve from the spinal cord.

Case	Age (yr)	Sex	Mechanism	Injury pattern classification	Neuroma pattern	Staining		
						H&E	Trichrome	Laminin
1	0.5	M	Birth brachial plexopathy	Stretch-related	Neuroma-in-continuity	Whorl-like perineurium	Fibrotic interneuron areas	Aberrant double-lumen
2	0.5	F	Birth brachial plexopathy	Stretch-related	Neuroma-in-continuity	Whorl-like perineurium		Aberrant double-lumen
3	0.3	F	Birth brachial plexopathy	Stretch-related	Neuroma-in-continuity	Whorl-like perineurium	Fibrotic interneuron areas	Aberrant double-lumen
4	0.5	F	Birth brachial plexopathy	Stretch-related	Neuroma-in-continuity	Whorl-like perineurium		Aberrant double-lumen
5	27.4	M	Trauma to brachial plexus from motorcycle	Avulsion	No neuroma	Normal pattern, wavy	Fibrotic perineurial scar	Wavy linear fibers
6	26.9	F	Orthopedic resection of common peroneal nerve	Transection	Stump neuroma	Whorl-like perineurium	Fibrotic perineurial scar	Aberrant double-lumen
7	37.9	M	Iatrogenic femoral nerve injury from inguinal herniorrhaphy	Transection	Stump neuroma	Granular-like perineurium	Fibrotic perineurial scar	Aberrant double-lumen
8	55.5	M	Work-related median nerve laceration	Transection	Neuroma-in-continuity	Granular-like perineurium	Fibrotic perineurial scar	Aberrant double-lumen

TABLE 1: Summary of peripheral nerve injury cases and staining patterns

The four infants with stretch-related injuries demonstrated classic neuroma formation on histology, as did the three patients with laceration/iatrogenic injury. The single patient with the nerve root avulsion injury did not demonstrate neuroma formation.

Stretch neuroma-in-continuity

On H&E stains, neuromas from infants with stretch-related injuries showed a monotonous cellularity and whorl-like perineurium along extensive interfascicular fibrotic scars (Figure 1, A, D, F, I). Trichrome stain showed perineurial fibrotic scar and disorganized nonlamellar architecture (Figure 1, B, G). Laminin IHC showed aberrant ECM formation, with uneven and abnormal double-lumen endoneurial tubules (Figure 1, C, E, H, J).

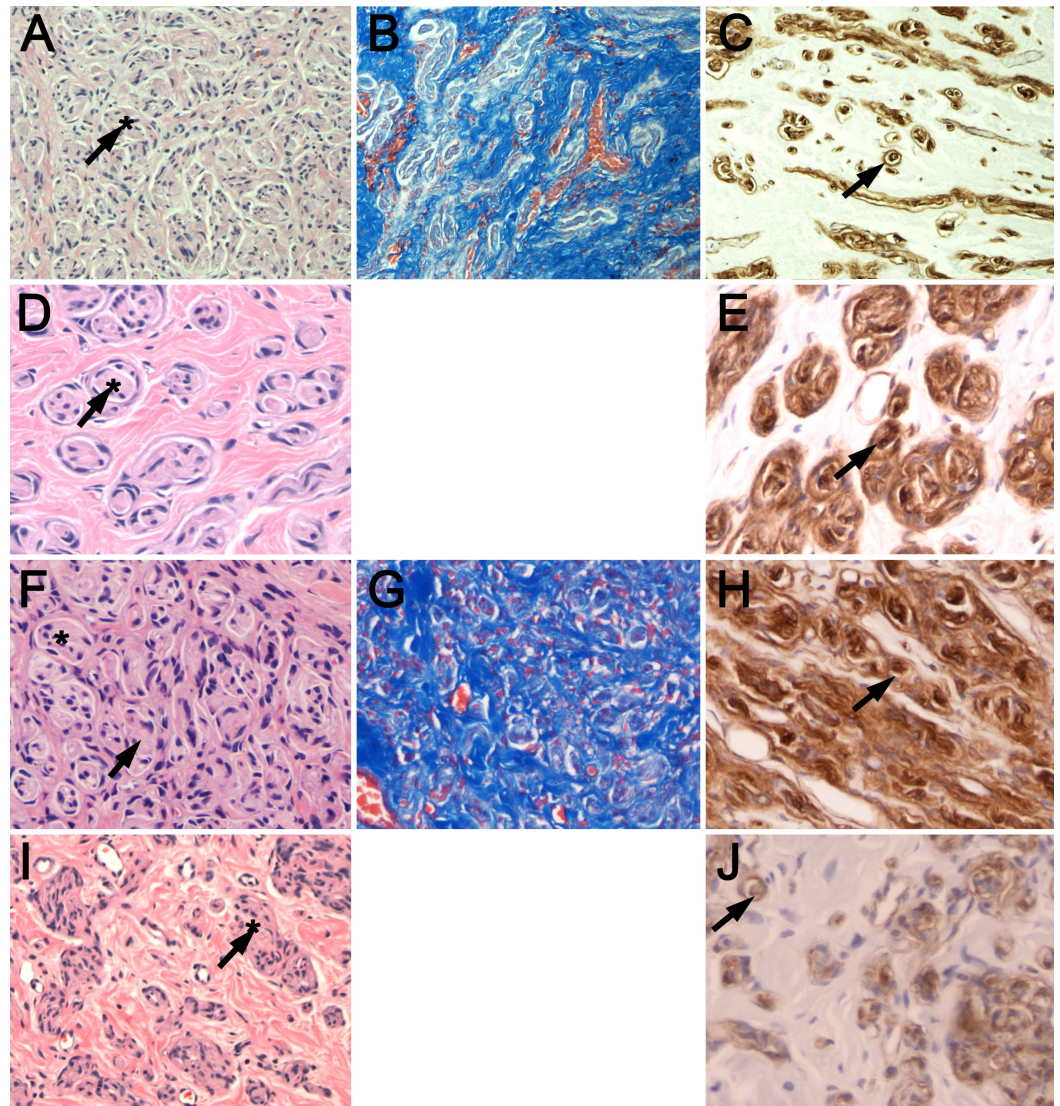


FIGURE 1: Histology of pediatric brachial plexus stretch neuromas-in-continuity

(A, D, F, I) H&E, (B, G) trichrome, and (C, E, H, J) laminin staining of resected neuromas-in-continuity from infants with birth stretch-related injury is shown. Monotonous cellularity and whorl-like endoneurium (asterisk) and extensive intraneural fibrotic scar are seen on H&E and trichrome stains. Laminin staining showing uneven, abnormal double-lumen endoneurial tubules (arrows) reflective of severely aberrant regeneration. Case One = A, B, C, $\times 100$; Case Two = D, E, $\times 200$; Case Three = F, G, H, $\times 200$; Case Four = I, J, $\times 100$.

Stretch avulsion injury

Nerve tissue from the trunks of a brachial plexus after avulsion from the spinal cord showed parallel, wavy fibers with minimal fibrotic scar on H&E (Figure 2, A-C). Nerve fibers showed evidence of stretch, with relative straightening of the nerve fibers and wide fiber spacing. Laminin stain reflected the H&E showing laminar, parallel extracellular protein deposition with minimal disruption (Figure 2A).

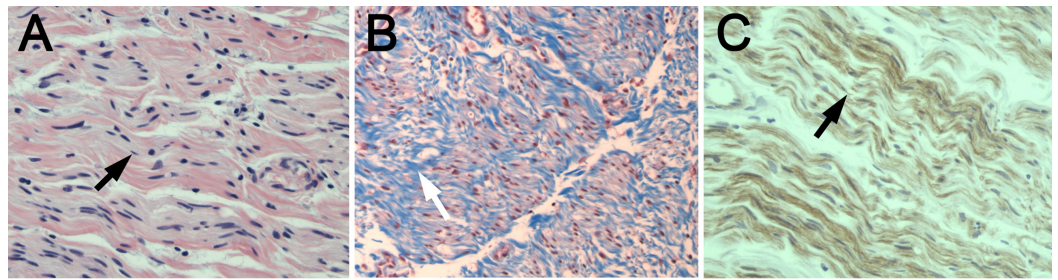


FIGURE 2: Histology of brachial plexus avulsion injury

(A) H&E, (B) trichrome, and (C) laminin staining (all $\times 200$) of the proximal end of the lower trunk from an adult with avulsion is shown (Case Five). The lower trunk was trimmed prior to transfer to the C7 spinal nerve. Parallel, wavy fibers (black arrow, A) with minimal fibrotic scar (white arrow, B) and laminar laminin (black arrow, C) with minimal disruption is seen. Minimal regeneration is seen along with limited fibrosis.

Laceration neuromas

Neuromas from the transection injuries were similar in microscopic appearance to the stretch-related injuries. H&E stains showed well-encapsulated perineurial tissue, heterogeneous nuclei, with reduced interneural fibrotic scars (Figure 3, A, D, G). Trichrome reflected the presence of fibrotic intraneural tissue (Figure 3, B, E, H). Laminin IHC showed separated groupings of double-lumen endoneurial tubules in all three cases (Figure 3, C, F, I).

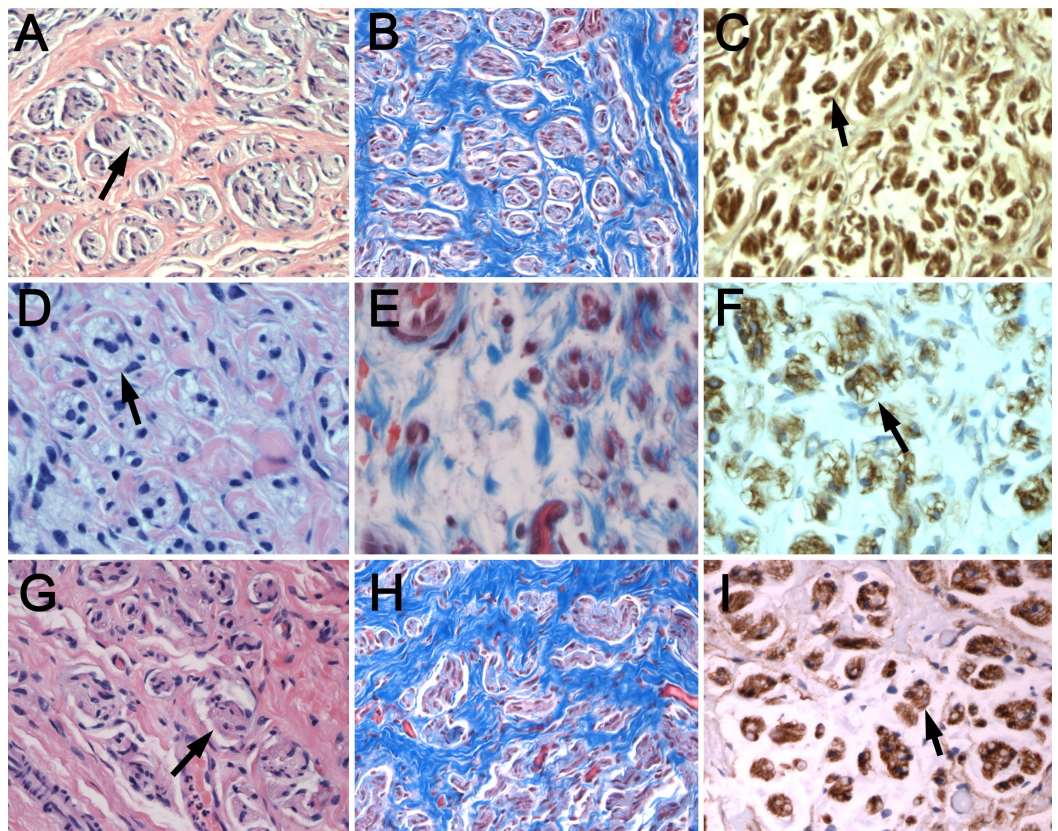


FIGURE 3: Histology of nerve laceration injuries

(A, D, G) H&E, (B, E, H) trichrome, and (C, F, I) laminin staining of neuromas from adult patients with transections is shown. Well-encapsulated endoneurial tissue, heterogeneous nuclei, with

reduced interneural fibrotic scars (black arrows, A, D, G) as compared with samples from neonatal stretch injuries. Double-lumen endoneurial tubules are seen (black arrows, C, F, I) are similar to those from birth stretch injuries. Case Six = A, B, C, $\times 200$; Case Seven = D, E, F, $\times 400$; Case Eight = G, H, I, $\times 200$.

Discussion

The ECM, Schwann cells, and signal transduction from trophic factors play an important role in axon regeneration and functional recovery [7]. These three factors have been suggested to work in concert in promoting successful regeneration; however, the tissue architecture mediating this process is not clearly understood. Upon losing contact with an axon because of Wallerian degeneration, Schwann cells revert to an immature, proregenerative state [8]. When transformed to this state, they proliferate and signal macrophage infiltration to clear debris. Schwann cells reorganize into columns along the ECM, termed the Bands of Büngner, to guide neuronal axon growth [9].

In the case of a neuroma-in-continuity, a distinct pathophysiology likely occurs to prevent the choreography of neurite, Schwann cell, and ECM from successful regeneration. We hypothesized that the endoneurial tubule may play a role in the failure of regeneration, and we selected laminin as a potential marker of the particular failure within the endoneurial tubule.

Laminins play an important role in axonal guidance. Laminin has been used as a substrate for nerve conduits to guide axonal growth in a variety of settings [10-11] and has been used during tissue engineering approaches [12-13]. Laminin specifically induces signaling pathways, including PI-3-kinase [14]. Within Schwann cells, an early pro-myelinating pathway is driven by neuregulin 1 while a later anti-myelinating pathway is driven by laminin [14]. Schwann cells lacking laminin induction were shown to express decreased schwannomin (ser518) phosphorylation, as well as CDC42 and Rac1 activation [15]. Furthermore, decreased levels of these proteins reduced Schwann cell-dependent myelination. Laminins were able to enhance the phosphorylation of I κ B and p65 NF- κ B signaling proteins in schwannoma cells [16]. A study by Chen and Strickland [17] demonstrated the importance of laminins using a Cre-loxP system to disrupt laminin γ 1 in Schwann cells. This model showed the motor deficits of laminin resulted in hind leg paralysis, tremor, and Schwann cell inability to differentiate and synthesize myelin proteins as well as Schwann cell apoptosis. In addition, after a sciatic nerve crush, axons showed significantly impaired regeneration. These results suggest that laminin plays a key role in organizing neurite outgrowth as well as proper signaling to regulate regeneration.

Our pathologic specimens demonstrated a consistent double lumen of a laminin-encasing axon and Schwann cells (Figure 4) in all forms of neuromas. The origins and consequences of this histopathologic observation are unknown. One possible cause may be the pathophysiologic remodeling of damaged ECM by regenerating axons and Schwann cells. Alternatively, the invasion of inflammatory cells after trauma may also participate in the pathology of extracellular proteins.

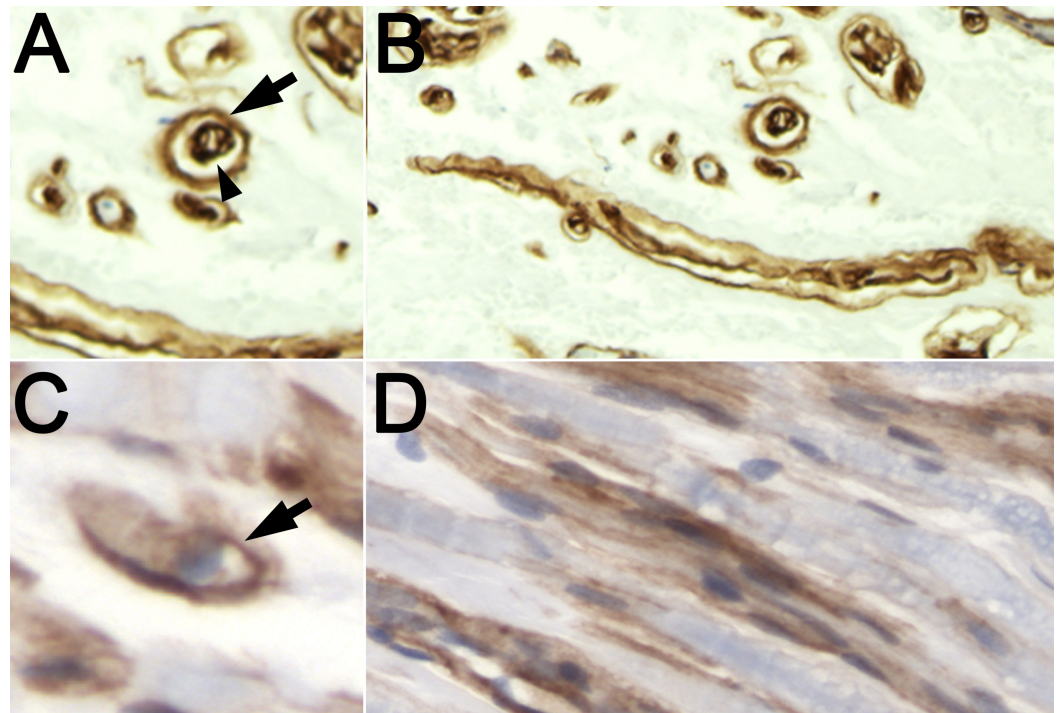


FIGURE 4: Double-lumen laminin pattern compared with typical laminin pattern (anti-laminin IHC and hematoxylin)

(A,B) neuroma-in-continuity specimen presented with cross-sectional (A) and longitudinal (B) slicing of the endoneurial tubule. The outer lumen (arrowhead) typically has thinner staining and an eccentric nucleus outside the laminin. The inner lumen (arrow) laminin is irregular, appears to have multiple channels, and may contain one or more nuclei. (C,D) normal nerve specimen removed for non-pathologic indications presented with cross-sectional (C) and longitudinal (D) slicing of the endoneurial tubule. Laminin staining is around a single channel, no more than one internal nucleus (C) and is thin around Schwann cells (D) (all $\times 400$).

IHC: immunohistochemistry

Two interesting features are notable. First, the pattern of a double-lumen tubule was consistent, regardless of whether the lesion was a neuroma-in-continuity or a stump neuroma and of the age of the patient, suggesting a conserved or consistent response to severe injury. Second, injured or regenerating axons seem to be required. As shown in the histology of the avulsed spinal nerves, where motor neurons are not present and the distal sensory axons are presumably intact, the architecture of the laminin tubules was unremarkable. Because Schwann cells produce the ECM and regenerating axons appear to be essential for the formation of the double-lumen tubule, it would seem to be a shared process. It is possible that loss of endoneurial tubule integrity produces axon-Schwann cell-mediated remodeling, as has been shown when large somatic fibers remodel the smaller endoneurial tubules of autonomic nerves [18-19]. However, much further work is necessary to identify the mechanism of this laminin deposition.

There are several limitations of this study. The sample is currently a small sample of overall nerve injury patterns. There was some heterogeneity of IHC staining among samples. In addition, we have only one sample from an avulsion injury, which served as a key comparison.

Conclusions

Our study used laminin antibodies to assess pathophysiologic regeneration in neuromas. Laminin IHC showed disorganized double-lumen endoneurial tubules in pathologic specimens of neuroma-in-continuity and stump neuromas, whereas avulsion injury maintained good nerve architecture and a relatively normal laminin pattern. This observation of the pathologic remodeling of endoneurial tubules during neuroma formation suggests pair interaction between regenerating axons and Schwann cells in response to injury to the ECM. Better understanding these patterns may help to generate better-directed treatment approaches.

Appendices

Case descriptions

Case One

A 6.5-month-old boy with shoulder dystocia during vaginal delivery due to macrosomia and anoxic injury at time of delivery presented with a flaccid left arm and progressive waiter's tip position and active grasp (Figure 1 A–C). Magnetic resonance imaging (MRI) demonstrated absent ventral rootlets at C8 with a clumping of nerve roots and hyperintensity of the C5–C7 roots suggestive of a neuroma. The patient underwent the grafting of C5 to the suprascapular nerve, C5 to the axillary nerve, and C6 to the lateral cord.

Case Two

A six-month-old girl presented with shoulder dystocia and flail right arm after birth by vaginal delivery (Figure 1 D, E). MRI demonstrated neuromas-in-continuity of the upper trunk along with a pseudomeningocele at C7. She underwent a brachial plexus neurolysis and a cabled graft repair of C5 to the posterior division of the upper trunk and the supraclavicular nerve and of C6 to the anterior division of the upper trunk.

Case Three

A four-month-old girl presented with a flail left arm after shoulder dystocia complicated by macrosomia and prolonged labor during birth by vaginal delivery (Figure 1 F–H). Examination at the time of birth showed an inability to move the left arm with a waiter's tip position. MRI showed traction injury to C5–7 with neuroma formation. The patient underwent a brachial plexus neurolysis and a cabled graft repair of C5 to the posterior division and the suprascapular nerve and of C6 to the anterior division.

Case Four

A five-month-old girl presented with obstetrical brachial plexus injury secondary to macrosomia and prolonged descent (Figure 1 I, J). The neurological examination at presentation showed a flail arm in the waiter's tip position without functional recovery. MRI demonstrated pseudomeningoceles at C6, C7, and T1. The patient underwent brachial plexus neurolysis and cabled graft repair of C5 to the posterior division and suprascapular nerve, C6 to the anterior division, and contralateral C7 to the lower trunk.

Case Five

A 27-year-old man presented four months after a motorcycle accident with associated clavicular fracture and pan-brachial plexopathy (Figure 2 A–C). MRI demonstrated absent rootlets at C8–T1 and a clumping of rootlets at C7, with an electromyogram (EMG) notable for modest denervation–reinnervation of the rhomboids. The patient underwent a multistage

nerve transfer, including neurolysis of the brachial plexus, distal spinal accessory-to-suprascapular transfer, C6 transfer to the posterior division of the upper trunk, and ipsilateral C7 transfer to the lower trunk. A second-stage procedure performed two weeks later included transfers of the motor intercostal nerves to musculocutaneous and biceps branches and to a sural nerve graft for later, staged, free functioning muscle transfer, and transfers of sensory intercostal nerves to lateral cord contribution to the median nerve, lateral antebrachial cutaneous nerve, and medial antebrachial cutaneous nerve.

Case Six

A 26-year-old woman presented with significant left lateral leg pain after being struck in the fibular head with a softball 11 years prior (Figure 3 A–C). She had undergone over 29 knee surgeries for pain, including decompression of the common peroneal nerve, and underwent a common peroneal nerve resection with Bridle procedure (posterior tibialis tendon to the insertion of tibialis anterior) at another institution. MRI revealed a large neuroma at the common peroneal nerve resection. She underwent resection of the neuroma and transfer of the common peroneal nerve to the nerve pedicle of the short head of the biceps femoris muscle.

Case Seven

A 38-year-old man presented with continued left leg pain and new leg weakness three months after a revision herniorrhaphy with an intended division of the ilioinguinal and iliohypogastric nerves for pain control at an outside institution (Figure 3 D–F). He had a quadriceps muscle weakness of 1/5 on the Medical Research Council scale and diminished sensation in the femoral and saphenous cutaneous distributions. A pelvis MRI showed hyperintensity along the femoral nerves with a thickening of the femoral nerve suggestive of an iatrogenic neuroma. The patient underwent repair of the lacerated medial femoral nerve branches and definitive neurectomy of the genitofemoral and ilioinguinal nerves.

Case Eight

A 55-year-old man presented five months after accidentally stabbing his left medial forearm, six inches proximal to the distal crease, in a cattle-handling accident (Figure 3 G–I). His symptoms include persistent numbness in the 4th and 5th digits and atrophy of the distal ulnar distribution in the hand. EMG results show no sensory recording of the 5th digit, prolonged latency and decreased conduction velocity of the ulnar motor nerve, and fibrillation with positive sharp waves of the first dorsal interosseous and abductor quinti minimi. MRI suggested partial ulnar nerve laceration. The patient underwent an ulnar nerve internal neurolysis and fascicular repair.

Additional Information

Disclosures

Human subjects: Consent was obtained by all participants in this study. University of Utah Institutional Review Board issued approval #IRB_00095096. Institutional Review Board approval was granted with a waiver of informed consent. **Animal subjects:** All authors have confirmed that this study did not involve animal subjects or tissue. **Conflicts of interest:** In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared

that there are no other relationships or activities that could appear to have influenced the submitted work.

Acknowledgements

We thank Kristin Kraus, MSc, for editorial assistance with this paper.

References

1. Noble J, Munro CA, Prasad VS, Midha R: Analysis of upper and lower extremity peripheral nerve injuries in a population of patients with multiple injuries. *J Trauma*. 1998, 45:116–122.
2. Taylor CA, Braza D, Rice JB, Dillingham T: The incidence of peripheral nerve injury in extremity trauma. *Am J Phys Med Rehabil*. 2008, 87:381–385. [10.1097/PHM.0b013e31815e6370](https://doi.org/10.1097/PHM.0b013e31815e6370)
3. Odier L: *Manuel de Medecine Pratique* [Book in French] . Bibliothèque Britannique & J.J. Paschoud, Geneva; 1803.
4. Chen ZL, Yu WM, Strickland S: Peripheral regeneration. *Annu Rev Neurosci*. 2007, 30:209–233. [10.1146/annurev.neuro.30.051606.094337](https://doi.org/10.1146/annurev.neuro.30.051606.094337)
5. Evans GR: Peripheral nerve injury: a review and approach to tissue engineered constructs . *Anat Rec*. 2001, 263:396–404. [10.1002/ar.1120](https://doi.org/10.1002/ar.1120)
6. Hallin RG, Wu G: Fitting pieces in the peripheral nerve puzzle . *Exp Neurol*. 2001, 172:482–492. [10.1006/exnr.2001.7813](https://doi.org/10.1006/exnr.2001.7813)
7. Fex Svennigsen A, Dahlin LB: Repair of the peripheral nerve-remyelination that works . *Brain Sci*. 2013, 3:1182–1197. [10.3390/brainsci3031182](https://doi.org/10.3390/brainsci3031182)
8. Fu SY, Gordon T: The cellular and molecular basis of peripheral nerve regeneration . *Mol Neurobiol*. 1997, 14:67–116. [10.1007/BF02740621](https://doi.org/10.1007/BF02740621)
9. Yang DP, Zhang DP, Mak KS, Bonder DE, Pomeroy SL, Kim HA: Schwann cell proliferation during Wallerian degeneration is not necessary for regeneration and remyelination of the peripheral nerves: axon-dependent removal of newly generated Schwann cells by apoptosis. *Mol Cell Neurosci*. 2008, 38:80–88. [10.1016/j.mcn.2008.01.017](https://doi.org/10.1016/j.mcn.2008.01.017)
10. Gonzalez-Perez F, Udina E, Navarro X: Extracellular matrix components in peripheral nerve regeneration. *Int Rev Neurobiol*. 2013, 108:257–275. [10.1016/B978-0-12-410499-0.00010-1](https://doi.org/10.1016/B978-0-12-410499-0.00010-1)
11. Zhang Q, Yan S, You R, et al.: Multichannel silk protein/laminin grafts for spinal cord injury repair. *J Biomed Mater Res A*. 2016, 104:3045–3057. [10.1002/jbm.a.35851](https://doi.org/10.1002/jbm.a.35851)
12. Fereol S, Fodil R, Barnat M, Georget V, Milbreta U, Nothias F: Micropatterned ECM substrates reveal complementary contribution of low and high affinity ligands to neurite outgrowth. *Cytoskeleton (Hoboken)*. 2011, 68:373–388. [10.1002/cm.20518](https://doi.org/10.1002/cm.20518)
13. Seggio AM, Narayanaswamy A, Roysam B, Thompson DM: Self-aligned Schwann cell monolayers demonstrate an inherent ability to direct neurite outgrowth. *J Neural Eng*. 2010, 7:046001. [10.1088/1741-2560/7/4/046001](https://doi.org/10.1088/1741-2560/7/4/046001)
14. Heller BA, Ghidinelli M, Voelkl J, et al.: Functionally distinct PI 3-kinase pathways regulate myelination in the peripheral nervous system. *J Cell Biol*. 2014, 204:1219–1236. [10.1083/jcb.201307057](https://doi.org/10.1083/jcb.201307057)
15. Yu WM, Chen ZL, North AJ, Strickland S: Laminin is required for Schwann cell morphogenesis. *J Cell Sci*. 2009, 122:929–936. [10.1242/jcs.033928](https://doi.org/10.1242/jcs.033928)
16. Armstrong SJ, Wiberg M, Terenghi G, Kingham PJ: Laminin activates NF-kappaB in Schwann cells to enhance neurite outgrowth. *Neurosci Lett*. 2008, 439:42–46. [10.1016/j.neulet.2008.04.091](https://doi.org/10.1016/j.neulet.2008.04.091)
17. Chen ZL, Strickland S: Laminin gamma1 is critical for Schwann cell differentiation, axon myelination, and regeneration in the peripheral nerve. *J Cell Biol*. 2003, 163:889–899. [10.1083/jcb.200307068](https://doi.org/10.1083/jcb.200307068)
18. Evans DH: Endings produced by somatic nerve fibres growing into the adrenal gland . *J Anat*. 1947, 81:225–232.
19. Simpson SA, Young JZ: Regeneration of fibre diameter after cross-unions of visceral and somatic nerves. *J Anat*. 1945, 79:48–65.