

Ovarian teratoma displaying a wide variety of tissue components in a broiler chicken (*Gallus Domesticus*): morphological heterogeneity of pluripotential germ cell during tumorigenesis

S. Ohfuji*

Department of Histopathology, Diagnostic Animal Pathology Office, Hokkaido, Japan

Abstract

Spontaneous ovarian teratoma was found in a seven-week-old female Chunky broiler chicken that was slaughtered for food. On *post-mortem* inspection, a spherical tumor mass attaching to a juvenile ovary was found in the abdominal cavity. Histopathologically, the tumor was comprised of immature mesenchymal stroma and a variety of mature tissue elements of mesodermal and ectodermal origin. In addition, there were multiple indistinguishable tissue elements, which showed no malignant cytological features but were unidentifiable as to corresponding embryological layer of origin. These heterogeneous teratoma tissues consisted of a variety of glandular, cystic, duct-like, and tubular structures, some of which exhibited a lining by a mixture of both keratinizing/non-keratinizing stratified squamous epithelial cells and cuboidal/columnar epithelial cells. The ovarian teratoma was considered a benign and congenital one. The highly diverse differentiation of the teratoma might have manifested a morphological aspect of intrinsic character of the pluripotential germ cells during tumorigenesis.

Keywords: Chicken, Heterogeneous differentiation, Ovary, Teratoma.

Introduction

Although uncommon, ovarian teratomas that are classified into a category of the germ cell tumors have been previously described in human beings (Kraus, 1977; Outwater *et al.*, 2001; Kuno *et al.*, 2004; Schanmughapriya *et al.*, 2011), nonhuman primates (Baskin *et al.*, 1982), experimental rats (Tsubota *et al.*, 2004), and domestic animals, including horses, pigs, dogs, and cattle (Dehner *et al.*, 1970; Gruys *et al.*, 1976; Rodriguez *et al.*, 1994; Basaraba *et al.*, 1998; MacLachlan and Kennedy, 2002; Sato *et al.*, 2003; Lefebvre *et al.*, 2005; Schlafer and Miller, 2007; Gamba *et al.*, 2014). Spontaneous ovarian or intra-abdominal teratomas have been reported in the avian species, including chickens (Campbell and Appleby, 1966; Helmboldt *et al.*, 1974; Gupta, 1976; Mohamed *et al.*, 2006), ducks (Cullen *et al.*, 1991; Bolte and Burkhardt, 2000), a goose (Reece and Lister, 1993), and a bald eagle (Ford *et al.*, 2006). It appears that ovarian teratomas are rare in broiler chickens possibly due to their young age; a short life span.

Most ovarian teratomas are benign in domestic animals, with the exception of their malignant counterparts that have rarely been reported in the bitch and mare (Gruys *et al.*, 1976; McEntee, 1990). Unlike most of other tumor types, ovarian teratomas certainly differentiate to produce a wide variety of tissue components which are composed of well-differentiated mature tissues, and this phenomenon is explained by the fact that the tumors arise from pluripotential germ cells that

have undergone somatic differentiation (Dahl *et al.*, 1990; MacLachlan and Kennedy, 2002; Schlafer and Miller, 2007). Although ovarian teratomas commonly show variable tissue components, the occurrence of heterogeneous components which are difficult to give appropriate anatomical designation may be uncommon in the tumor. Therefore, the purpose of the present study was to describe the histopathological features of a spontaneously occurring benign ovarian teratoma in a broiler chicken, which presented with not only well-differentiated mature tissues of mesodermal and ectodermal origin, but also remarkably diverse differentiation of teratoma components unidentifiable as to corresponding embryological layer of origin.

Materials and Methods

A flock of seven-week-old Chunky chickens (*Gallus domesticus*) were slaughtered at an abattoir. These chickens had been managed for meat production by a commercial company where there were no epidemiological or other serious problems. The flock was composed of approximately 150 chickens derived from a similar genetic background. Physical examination prior to slaughter revealed no significant clinical signs in all these chickens. At *post-mortem* examination, a female broiler chicken exhibited a tumor mass in the abdominal cavity. The remaining chickens from the same flock, which were processed on the same day, were reported by the veterinary meat inspectors to be unaffected with neoplastic lesions in the ovary or other visceral organs. For

*Corresponding Author: Susumu Ohfuji. Department of Histopathology, Diagnostic Animal Pathology Office, Hokkaido, Japan. Tel/Fax: +81 11 671 7118. E-mail: tksohfuji@juno.ocn.ne.jp

histopathological diagnosis of the ovarian tumor, tissue samples were taken immediately after slaughter from the tumor. Additional samples were collected from major visceral organs, including the liver, spleen, kidney, heart, lung, and intestine. These tissue samples were fixed in 10% formalin. After fixation, a whole ovarian tumor was cut into many portions that were prepared for histopathology. All tissue blocks were embedded in paraffin wax, processed routinely, sectioned at 4 μ m, and stained with hematoxylin and eosin (HE).

Results

On *post-mortem* examination, the broiler chicken under study exhibited a spherical tumor mass, approximately 9.0 cm in diameter (Fig. 1a). The tumor mass was loosely attached to a juvenile ovary. It was soft as a whole, except for some areas gritty to feel on sectioning, and its cut surface showed a mottled gray and tan appearance with occasional cystic areas. Other organs, such as the heart, lung, kidney, spleen, liver, and bowel, were unremarkable.

On histopathology, the tumor was covered by single layer of cuboidal epithelial cells. The tumor comprised a haphazard arrangement of immature mesenchymal stroma and a variety of disorganized, different, somatic tissue elements, enabling a diagnosis of teratoma to be made. Immature mesenchymal cells exhibited round or ellipsoidal pale nuclei with small nucleoli, a small amount of basophilic cytoplasm, and rare mitotic figures (<1 per 200 x field). Many teratoma elements were mature, including extensive areas of the central nervous gray matter, which either contained polygonal neurons and oligodendroglia (Fig. 1b) or had rarely cuboidal-lined small rosettes; large cavities reminiscent of the cerebral ventricles

which were lined by a single layer of spherical cells; nerve fiber bundles of the peripheral nervous system (Fig. 1c); varying-sized groups of striated myofibers (Fig. 1d); bone trabeculae accompanied by osteoblasts, multinucleated osteoclasts, and hematopoietic (erythropoietic and leukopoietic) bone marrow (Fig. 1e); scattered islands of hyaline cartilage enclosed with a membrane of many collagen fibers (Fig. 1f); scattered areas of adipose tissue; and a few encapsulated lymphoid follicles.

Other teratoma elements consisted of a variety of aberrant tissue types, which exhibited no characteristics to a sufficiently distinctive degree to allow identification of intrinsic somatic organs or tissues. Apparent malignant features, such as anisokaryosis, anisocytosis, substantial mitotic figures indicative of rapid growth, and metastatic or invasive behavior, were not recognized in these tissue types. Such aberrant teratoma elements included secretory glands with or without duct-like lumens (Figs. 2a and 2b); glandular structures lined by a layer of cuboidal or columnar epithelial cells (Fig. 2c); duct-like structures lined by a single to multilayered pale cuboidal or columnar epithelial cells (Fig. 2d); cystic structures lined by a single layer of squamous epithelial cells, which were surrounded by immature mesenchymal cells and collagen fibers (Fig. 2e); and variably-sized cystic structures similar to epidermoid cysts or feather follicles devoid of feathers, which were lined by keratinizing stratified squamous epithelial cells (Fig. 2f). In addition, the most unique teratoma elements were comprised of a triad of epithelial-lined cystic, tubular, and duct-like structures. First, cystic structures were composed of a partial lining by multilayered cuboidal epithelial cells, the other

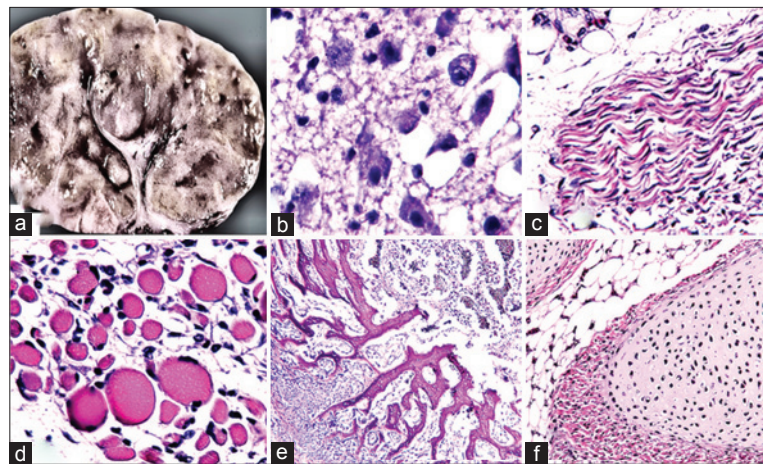


Fig. 1. (a) Teratoma after fixation shows a cavitory appearance on cut section. (b) Polygonal neurons in the neuropil-like area of teratoma (HE x 200). (c) Nerve bundle similar to peripheral nerve (HE x 100). (d) A group of varying sized striated myofibers (HE x 100). (e) Bone trabeculae accompanied by hematopoietic bone marrow (HE x 100). (f) Insular hyaline cartilage surrounded by a membrane of collagen fibers (HE x 100).

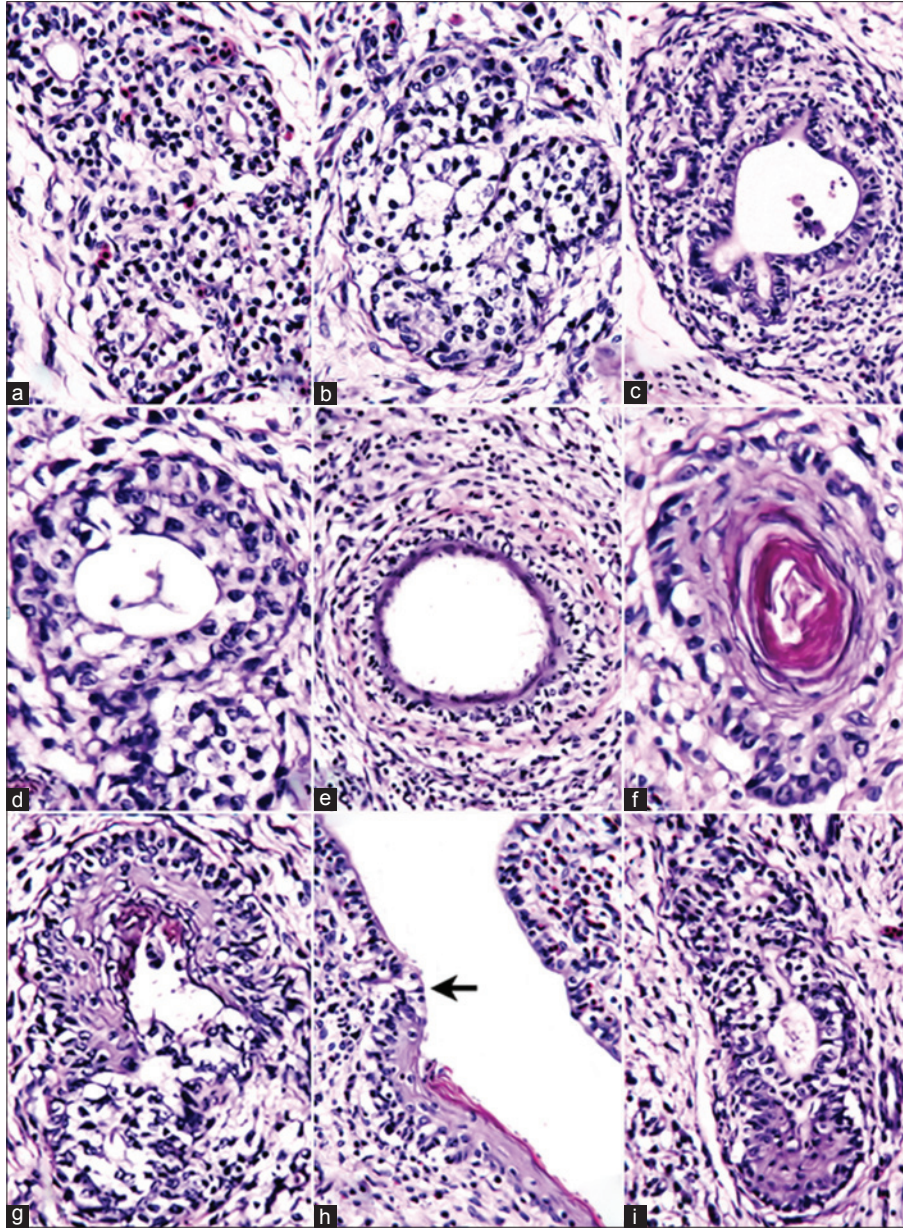


Fig. 2. (a) Teratoma represents secretory gland composed of cuboidal-lined acinar structure which has duct-like lumens (HE x 100). (b) Secretory gland composed of cuboidal-lined acinar structure without ducts (HE x 100). (c) Glandular structure lined by a layer of cuboidal or columnar epithelial cells (HE x 100). (d) Tubular structure lined by a single to multilayered, pale, cuboidal or columnar epithelial cells (HE x 200). (e) Cystic structure lined by a single layer of squamous epithelial cells, which is surrounded by immature mesenchymal cells and collagen fibers (HE x 100). (f) Cystic structure similar to epidermoid cyst or feather follicle lacking feathers which is lined by keratinizing stratified squamous epithelial cells (HE x 200). (g) Cystic structure composed of a partial lining of both multilayered cuboidal epithelial cells and keratinizing stratified squamous epithelial cells (HE x 100). (h) Tubular structure is lined by both columnar epithelial cells and keratinizing stratified squamous epithelial cells, with sharp transition boundary (arrow) between the two types of epithelial cells (HE x 200). (i) Duct-like structure lined by cuboidal epithelial cells which are connected with stratified squamous epithelial cells (HE x 200).

half being lined by keratinizing stratified squamous epithelial cells (Fig. 2g). Second, tubular structures were lined by both a layer of columnar epithelial cells and keratinizing stratified squamous epithelial cells (Fig. 2h). Third, duct-like structures, which consisted

of pale cuboidal epithelial cells constituting duct-like lumens of secretory glands, were connected with non-keratinizing stratified squamous epithelial cells (Fig. 2i). Other than various teratoma components described above, relatively poor capillary networks

were present throughout the tumor. Tissues that can determine or suggest the origin of the tumor were not recognized elsewhere in the tumor. Other visceral organs did not exhibit metastatic tumor lesions or any other significant lesions.

Discussion

Teratoma of the avian species has been recognized in the ovary, testes, kidney, adrenal gland, cerebrum, spinal cord, pineal body, and eye (Hooper, 2008; Reece, 2008; Paździor *et al.*, 2012). In the current broiler chicken, the normal appearance of every other abdominal visceral organ, and the anatomical location of the tumor, (which was closely associated with intact ovary and covered by a single layer of cuboidal epithelial cells, as seen in normal ovaries), indicated that this teratoma was most likely a primary tumor originating from the ovary. Low mitotic rate of mesenchymal stromal cells indicating slow growth of the tumor, which was recognized in a seven-week-old chicken, and the absence of metastasis and invasion suggested this ovarian teratoma to be a benign and congenital one.

Ovarian teratoma, which has been reported previously in domestic animal species, such as the bitch, cat, sow, mare, and cow, may generally contain mature rather than undifferentiated somatic tissues (Basaraba *et al.*, 1998; MacLachlan and Kennedy, 2002; Lefebvre *et al.*, 2005; Schlafer and Miller, 2007). Likewise, tissue types of ovarian teratoma reported previously in the avian species may usually display well-differentiated adult morphology, even if they vary from teratoma to teratoma. In this species, teratoma tissues consist of keratinized epithelium-lined sac containing feathers, epithelial pearls formed by squamous epithelium, bone, cartilage, smooth muscle, nerve, fat, trachea, or melanocytes (Reece and Lister, 1993; Bolte and Burkhardt, 2000; Reece, 2008), and individual teratoma component shows a built-in mature anatomical feature characteristic of each somatic tissue, which can be easily identified as such. Ovarian teratoma described in the current chicken was composed of not only a variety of well-differentiated mature tissue components of mesodermal and ectodermal origin, but also multiple aberrant and heterogeneous components, particularly a variety of glandular, cystic, duct-like, and tubular structures. These heterogeneous teratoma components were unidentifiable as to corresponding embryological layer of origin. Some of these unusual structures exhibited a lining by a mixture of both keratinizing/non-keratinizing stratified squamous epithelial cells and cuboidal/columnar epithelial cells, showing simultaneous differentiation of tumor cells into two different types of epithelial cells within a single teratoma component. Although ectodermal origin was suggested by the presence of partial lining by squamous

epithelium (Berman, 2009), this notion could not be established because of the participation of cuboidal/columnar epithelial lining. All such epithelial-lined structures did not exhibit cytologically malignant features, namely anisokaryosis, anisocytosis, and frequent mitotic figures, and therefore were regarded as mature rather than anaplastic teratoma components. In the present ovarian teratoma, histological features of those aberrant teratoma components, which were lined by a combination of two different types of epithelial cells (keratinizing/non-keratinizing stratified squamous epithelial cells and cuboidal/columnar epithelial cells) were reminiscent of those in transdifferentiation, dedifferentiation, or simple squamous metaplasia. The phenomenon of transdifferentiation is suggested to have played a role in sacrococcygeal teratoma in human beings (Jurić-Lekić *et al.*, 1993). Identification of dedifferentiation is documented in an ovarian teratoma (Yasunaga *et al.*, 2011) and other teratomas involving internal visceral organs or tissues in human beings (Game *et al.*, 2001; Kim *et al.*, 2012). Although ovarian carcinoma associated with squamous metaplasia is described in the bovine species (McEntee, 1990) and human beings (Kay, 1961), and even if the stratified squamous epithelium is commonly seen in ovarian teratoma, there are no reports of ovarian teratoma displaying features of ongoing squamous metaplasia in either the medical or veterinary literature. In this chicken's ovarian teratoma, the epithelia-lined structures did not show any changes associated with metaplasia. Therefore, it was not possible to accurately determine which event (transdifferentiation, dedifferentiation, or squamous metaplasia) was implicated in the development of the aberrant teratoma components during tumorigenesis.

Even if the pluripotential cell lineage can differentiate to produce a variety of somatic tissues, it seems unusual that ovarian teratoma of the broiler chicken described in this report exhibited so many morphologically diverse tissue elements. In conclusion, it may be thought that such a highly heterogeneous variant of ovarian teratoma might have indicated a curious morphological aspect of the pluripotential germ cells during tumorigenesis. This morphological expression would be largely attributable to intrinsic character of the pluripotential germ cell lineage. The present study was limited to the histopathological investigation. Thus, further cytogenetic, ultrastructural, and immunohistochemical investigations appear warranted to better understand the highly heterogeneous differentiation of tissue components in the ovarian teratoma such as that described in this report.

Conflict of interest

Author declares that he has no conflict of interest.

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