

Draft Genome Sequences of Two Drug-Resistant Isolates of *Pseudomonas aeruginosa* **Obtained from Keratitis Patients in India**

Ramesh K. Aggarwal,^a Chhavi Dawar,^a Satrupa Das,^a Savitri Sharma^b

Centre for Cellular & Molecular Biology (CSIR-CCMB), Tarnaka, Hyderabad, India^a; Brien Holden Eye Research Centre, Hyderabad Eye Research Foundation, L.V. Prasad Eye Institute (LVPEI), L. V. Prasad Marg, Banjara Hills, Hyderabad, Indiab

We report here the draft genomes of two drug (fluoroquinolone)-resistant clinical isolates of *Pseudomonas aeruginosa* **obtained from the corneal scrapings of keratitis patients from India. The two annotated genomes are 6.31 Mb and 6.41 Mb in size. These genomes are expected to facilitate the identification and understanding of the genes associated with acquired multidrug resistance.**

Received 26 November 2014 **Accepted** 1 December 2014 **Published** 8 January 2015

Citation Aggarwal RK, Dawar C, Das S, Sharma S. 2015. Draft genome sequences of two drug-resistant isolates of *Pseudomonas aeruginosa* obtained from keratitis patients in India. Genome Announc 3(1):e01404-14. doi:10.1128/genomeA.01404-14.

Copyright © 2015 Aggarwal et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Ramesh K. Aggarwal, rameshka@ccmb.res.in.

P*seudomonas aeruginosa* is a Gram-negative bacterial pathogen associated with a variety of infections [\(1\)](#page-1-0). The emergence of multidrug resistance in *P. aeruginosa* isolates has been increasingly documented worldwide. Fluoroquinolones are the most potent agents for the treatment of *P. aeruginosa* infections [\(2\)](#page-1-1). However, in recent years, a number of clinical *P. aeruginosa* isolates have shown reduced susceptibilities or resistances to fluoroquinolones [\(3,](#page-1-2) [4\)](#page-1-3). Here, we report draft genomes of two resistant isolates (P2-L230/95 and P7-L633/96) obtained from the corneal scrapings of keratitis patients seen and treated at the L. V. Prasad Eye Institute, Hyderabad, India.

The availability of whole-genome information of pathogens can help in unraveling the genetics/mechanism(s) of acquired drug resistance. Thus, we performed the whole-genome sequencing (WGS) of the two resistant isolates using the Illumina HiSeq and Roche 454 (FLX Titanium) platforms. The isolate identity was confirmed to the species level by typing the 16S rRNA. The reads from Roche 454 (95,836 reads for P2 and 204,602 reads for P7) and Illumina (1,781,644 reads for P2 and 1,271,248 reads for P7) were assembled initially using the GS *de novo* Assembler (version 2.8) and A5-MiSeq assembly pipeline, respectively [\(5\)](#page-1-4). The resulting contigs were then used to assemble the draft genomes of the two isolates using the Contig Integrator for Sequence Assembly (CISA) of bacterial genomes [\(6\)](#page-1-5). The draft genome assembly of P2-L230/95 is 6.31 Mb, comprising 110 contigs (range, 4,197 to 371,711 bp in size), with an average length (N_{50}) of 95,132 bp and 65.70% G-C content. Similarly, the P7-L633/96 genome has 72 contigs (range, 924 to 671,326 bp), with a genome size of 6.41 Mb, an *N*₅₀ of 108,322 bp, and 65.97% G+C content. Both genomes were submitted to Rapid Annotations using Subsystems Technology (RAST) and the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (http://ncbi.nlm.nih.gov/genomes/static/Pipeline.html) for annotation.

In total, 5,823 protein-coding sequences and 63 RNA-coding genes in P2 and 5,949 protein-coding sequences and 60 RNAcoding genes in P7 were annotated using the NCBI annotation

pipeline. The RAST server gave comparable results covering 541 subsystems in P2 and 537 in P7. Under the virulence, disease, and defense subsystems in the RAST annotation, the subcategories of resistance to antibiotics and toxic compounds had 121 and 135 genes annotated in P2 and P7, respectively. The alterations in the quinolone resistance-determining regions within topoisomerase II (GyrA and GyrB subunits) and topoisomerase IV (ParC and ParE subunits) are the major mechanisms for fluoroquinolone resistance in Gram-negative bacteria, in addition to a decreased accumulation of fluoroquinolones due to the impermeability of the membrane and/or overexpression of the efflux pump system [\(7](#page-1-6)[–](#page-1-7)[10\)](#page-1-8). Thus, comparative studies were done using the *P. aeruginosa* PAO1 genome (GenBank accession no. NC_002516.2) to examine topoisomerase II, topoisomerase IV, and two efflux pump regulatory genes, *mexR* and *nfxB*, to identify putative mutations that might have led to fluoroquinolone resistance. Indeed, we observed several mutations specifically in *gyrA, gyrB*, and *parC* that were reported earlier in drug-resistant strains [\(11\)](#page-1-9). It is expected that the genomes described here will facilitate detailed studies to identify/elucidate mutation(s)/possible mechanism(s) of acquired drug resistance and better drug discovery.

Nucleotide sequence accession numbers.This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession numbers JMBR00000000 for P2-L230/95 and JMBS00000000 for P7-L633/96. The version described in this paper is the second version, with accession numbers JMBR02000000 and JMBS02000000, respectively.

ACKNOWLEDGMENTS

R.K.A. thanks the Council for Scientific and Industrial Research, India, for the FYP grant Genomics and Informatics Solutions for Integrating Biology (GENESIS)_BSC0121: WP4-Decoding Indian Genome: Genome view of people, pathogen and organisms of agricultural and evolutionary importance. We thank the NCBI PGAP team for genome annotation services, Debabrata Dash for help with Roche 454 run, SciGenom Labs Pvt.

Ltd., Cochin, India, for help with Illumina HiSeq data, and R. Phanindranath and Evangelene for technical help.

REFERENCES

- 1. **Klockgether J, Munder A, Neugebauer J, Davenport CF, Stanke F, Larbig KD, Heeb S, Schöck U, Pohl TM, Wiehlmann L, Tümmler B.** 2010. Genome diversity of *Pseudomonas aeruginosa* PAO1 laboratory strains. J Bacteriol **192:**1113–1121. http://dx.doi.org/10.1128/JB.01515-09.
- 2. **Schmitz FJ, Verhoef J, Fluit AC.** 1999. Comparative activities of six different fluoroquinolones against 9,682 clinical bacterial isolates from 20 European university hospitals participating in the European SENTRY surveillance programme. The SENTRY participants group. Int J Antimicrob Agents **12:**311–317.
- 3. **Thomson CJ.** 1999. The global epidemiology of resistance to ciprofloxacin and the changing nature of antibiotic resistance: a 10 year perspective. J Antimicrob Chemother **43**(Suppl A):31–40. http://dx.doi.org/10.1093/ jac/43.suppl_1.31.
- 4. **Karlowsky JA, Draghi DC, Jones ME, Thornsberry C, Friedland IR, Sahm DF.** 2003. Surveillance for antimicrobial susceptibility among clinical isolates of *Pseudomonas aeruginosa* and *Acinetobacter baumanii* from hospitalized patients in the United States, 1998 –2001. Antimicrob Agents Chermother **47:**1681–1688.
- 5. **Coil D, Jospin G, Darling AE.** 2014. A5-miseq: an updated pipeline to

assemble microbial genomes from Illumina MiSeq data. arXiv: 1401.5130v2. http://arxiv.org/abs/1401.5130.

- 6. **Lin SH, Liao YC.** 2013. CISA: contig integrator for sequence assembly of bacterial genomes. PLoS One **8:**e60843. http://dx.doi.org/10.1371/ journal.pone.0060843.
- 7. **Hancock REW, Burman WJ, Wilson ML.** 1998. Resistance mechanisms in *Pseudomonas aeruginosa* and other nonfermentative Gram-negative bacteria. Clin Infect Dis **27:**93–99. http://dx.doi.org/10.1086/514909.
- 8. **Ruiz J.** 2003. Mechanisms of resistance to quinolones: target alterations, decreased accumulation and DNA gyrase protection. J Antimicrob Chemother **51:**1109 –1117. http://dx.doi.org/10.1093/jac/dkg222.
- 9. **Lindgren PK, Karlsson A, Hughes D.** 2003. Mutation rate and evolution of fluoroquinolone resistance in *Escherichia coli* isolates from patients with urinary tract infections. Antimicrob Agents Chermother **47:**3222–3232. http://dx.doi.org/10.1128/AAC.47.10.3222-3232.2003.
- 10. **Valdezate S, Vindel A, Echeita A, Baquero F, Cantó R.** 2002. Topoisomerase II and IV quinolone resistance-determining regions in *Stenotrophomonas maltophilia* clinical isolates with different levels of quinolone susceptibility. Antimicrob Agents Chemother **46:**665–671. http://dx.doi.org/ 10.1128/AAC.46.3.665-671.2002.
- 11. **Lee JK, Lee YS, Park YK, Kim BS.** 2005. Alterations in the GyrA and GyrB subunits of topoisomerase II and the ParC and ParE subunits of topoisomerase IV in ciprofloxacin-resistant clinical isolates of *Pseudomonas aeruginosa*. Int J Antimicrob Agents **25:**290 –295. http://dx.doi.org/ 10.1016/j.ijantimicag.2004.11.012.