

Systematic Analysis of the Physiological Importance of Deubiquitinating Enzymes

Wei-Ling Tsou^{1,2}, Michael J. Sheedlo³*, Marie E. Morrow³*, Jessica R. Blount^{1,2}, Kelly M. McGregor¹, Chittaranjan Das³, Sokol V. Todi^{1,2*}

1 Department of Pharmacology, Wayne State University School of Medicine, Detroit, Michigan, United States of America, **2** Department of Neurology, Wayne State University School of Medicine, Detroit, Michigan, United States of America, **3** Department of Chemistry, Purdue University, West Lafayette, Indiana, United States of America

Abstract

Deubiquitinating enzymes (DUBs) are proteases that control the post-translational modification of proteins by ubiquitin and in turn regulate diverse cellular pathways. Despite a growing understanding of DUB biology at the structural and molecular level, little is known about the physiological importance of most DUBs. Here, we systematically identify DUBs encoded by the genome of *Drosophila melanogaster* and examine their physiological importance *in vivo*. Through domain analyses we uncovered 41 *Drosophila* DUBs, most of which have human orthologues. Systematic knockdown of the vast majority of DUBs throughout the fly or in specific cell types had dramatic consequences for *Drosophila* development, adult motility or longevity. Specific DUB subclasses proved to be particularly necessary during development, while others were important in adults. Several DUBs were indispensable in neurons or glial cells during developmental stages; knockdown of others perturbed the homeostasis of ubiquitinated proteins in adult flies, or had adverse effects on wing positioning as a result of neuronal requirements. We demonstrate the physiological significance of the DUB family of enzymes in intact animals, find that there is little functional redundancy among members of this family of proteases, and provide insight for future investigations to understand DUB biology at the molecular, cellular and organismal levels.

Citation: Tsou W-L, Sheedlo MJ, Morrow ME, Blount JR, McGregor KM, et al. (2012) Systematic Analysis of the Physiological Importance of Deubiquitinating Enzymes. PLoS ONE 7(8): e43112. doi:10.1371/journal.pone.0043112

Editor: Brian D. McCabe, Columbia University, United States of America

Received: June 6, 2012; **Accepted:** July 17, 2012; **Published:** August 24, 2012

Copyright: © 2012 Tsou et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by National Institutes of Health grant R00NS064097 to SVT, 1R01RR026273 to CD and a Young Investigator Award to SVT from the National Ataxia Foundation. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: stodi@med.wayne.edu

† These authors contributed equally to this work.

Introduction

Deubiquitinating enzymes (DUBs, also known as deubiquitinases) comprise a large family of proteases with approximately 90 members encoded by the human genome [1] (Figure 1A). DUBs regulate the post-translational modification of proteins by the small protein modifier ubiquitin. Protein ubiquitination controls many different cellular pathways and processes, from gene transcription to protein degradation, from cell division to cell death. Perturbations in ubiquitin-dependent pathways due to mutations in select DUBs are linked to malignancies and to neurological diseases (reviewed in [2–6]). Significant work has been conducted on the structure of several DUBs, their *in vitro* properties, molecular interactions and cell biology [2–6]. However, for the majority of this large family of proteases, it is unclear whether its members are physiologically necessary *in vivo*. It is also uncertain how much functional redundancy exists among a class of proteins with nearly 90 members in humans.

Protein ubiquitination occurs through the coordinated action of three types of enzymes: E1 (ubiquitin activating enzyme), E2 (ubiquitin conjugating enzyme) and E3 (ubiquitin ligase) [7]. Because ubiquitin molecules can be attached to other ubiquitins through any one of seven lysines to form chains, a target protein can be modified by a single ubiquitin or by a poly-ubiquitin chain

[8]. Different types of ubiquitination have divergent effects on the fate of a protein, such as differences in localization (mono-ubiquitin) or its degradation (some types of poly-ubiquitin chains). DUBs mostly act by cleaving bonds between ubiquitin moieties or ubiquitin and another protein, thus editing ubiquitin chains, reversing protein ubiquitination, and recycling ubiquitin. However, DUBs with non-enzymatic functions have also been reported [9–11].

Studies in the fruit fly *Drosophila melanogaster* were among the first to investigate DUBs *in vivo*, including the DUBs Fat facets (faf) [12,13] and Ovarian tumor (otu) [14–16]. While *Drosophila* is a genetically versatile organism with numerous tools that are publicly available, it has not been used extensively to elucidate DUB biology. In part, this could be due to lack of a comprehensive catalog of fly DUBs. Here we identified *Drosophila* DUBs and addressed the following: which DUBs are physiologically necessary in *Drosophila* and to what extent are they functionally redundant?

We conducted targeted RNA-interference (RNAi) screens in which individual DUBs were systematically knocked down in specific cell types or across all cells and tissues of intact flies. Most fly DUBs proved physiologically necessary for fly development, motility or longevity. There was little functional redundancy among these enzymes. To the best of our knowledge, our work comprises the first comprehensive analysis of *Drosophila* DUBs and

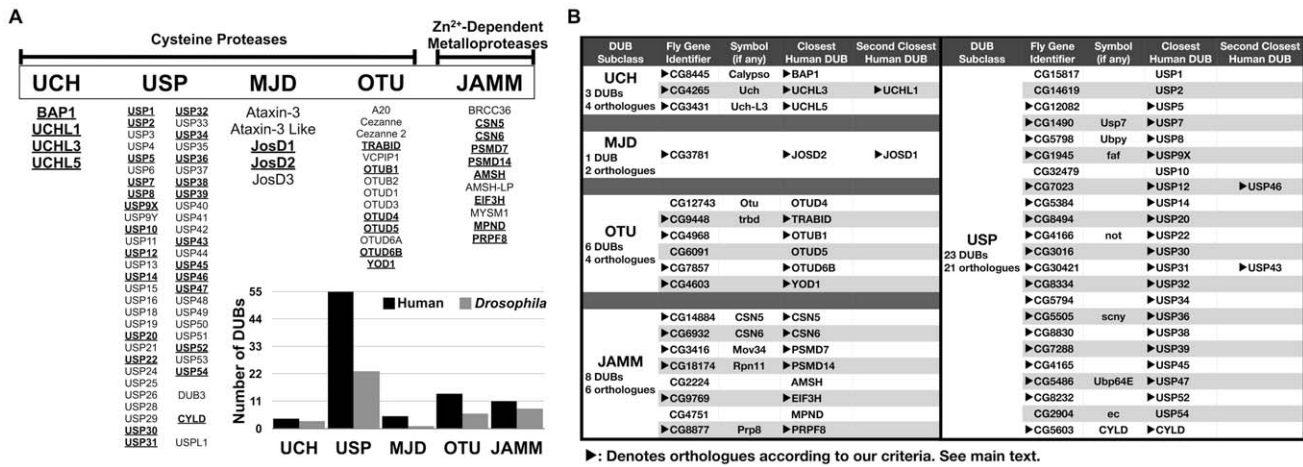


Figure 1. Human and *Drosophila* DUBs. A) Shown are human DUBs categorized into five subclasses based on homology at the catalytic domain [1,3]. UCH: Ubiquitin C-terminal Hydrolases, USP: Ubiquitin-Specific Proteases, MJD: Machado-Joseph Disease Proteases, OTU: Otubain proteases, JAMM: JAB1/MPN/Mov34 Metalloenzymes. All DUB subclasses are cysteine proteases, except JAMM domain DUBs, which are Zn²⁺-dependent metalloproteases [1,3]. Underlined and bolded: human DUBs that aligned with *Drosophila* DUBs (also see File S2). Inset: histograms show the total numbers of human DUBs (black) and fly DUBs (gray) that we identified for each subclass. B) Listed are all the *Drosophila* DUBs that we identified. Arrowheads highlight orthologues based on coverage and domain analyses (complete analysis in File S2). Not all fly DUB genes have symbols designated to them, indicating that they have not been previously characterized. Symbols were obtained from www.flybase.org. doi:10.1371/journal.pone.0043112.g001

provides clear evidence that this family of proteases is indispensable for animal physiology.

Results and Discussion

Identification of *Drosophila* DUBs

We used published lists of human DUBs [1,3] to identify *Drosophila* counterparts through two sets of alignments. First, the amino acid sequence of each human DUB was aligned against the *Drosophila* proteome to uncover similarities. Next, through reverse alignments, the amino acid sequence of fly proteins identified by the first set of comparisons was used to examine domain composition and to uncover potential human orthologues (Figure 1A, File S1 and S2). For this study, we defined orthologues as protein pairs that are conserved at most domains, with at least 50% coverage (File S2). Exceptions to our criteria exist, as in the case of orthologues that have been reported previously (e.g. CYLD [17], USP36 [18]). According to our analyses, about 40% of human DUBs have *Drosophila* orthologues at the amino acid level (Figure 1B).

We found 41 fly DUBs, of which only 18 had been previously reported (Figure 1B). Out of the 41, 33 *Drosophila* DUBs fulfill our criteria as orthologues. Based on the number of members in each subdivision, the UCH and JAMM subclasses appear to be most highly conserved evolutionarily (Figure 1A). Nearly all fly DUBs aligned best with a single human DUB, except four that aligned with two human counterparts: CG4265, CG3781, CG7023, and CG30421 (File S2). These examples could be due to evolution-related specialization of certain biological processes that require specific human DUBs to perform divergent functions in the same cell, or the same functions in different cell types. Functional studies will determine whether the fly protein serves as an orthologue for both human DUBs.

The Physiological Significance of *Drosophila* DUBs

We next examined the physiological importance of *Drosophila* DUBs by using the Gal4-UAS system [19] and RNAi to systematically knock down individual DUBs either in all cells

and tissues or only in neurons (Figure S1 summarizes the Gal4-UAS system). RNAi-based approaches are powerful because they enable selective knockdown to explore the significance of individual genes *in vivo* and they provide the capacity to expediently test the physiological importance of genes either everywhere or only in specific tissues. Additionally, through different Gal4 drivers, UAS-RNAi enables gene dosage studies to uncover subtle effects that might be missed by null mutations or near-complete knockdown. RNAi-based screens were implemented successfully in the past to identify DUBs in specific cellular processes in mammalian cells (e.g. [20–23]).

We used the following Gal4 drivers for our systematic genetic screens (Figure 2): 1) sqh-Gal4, which drives the expression of UAS-constructs everywhere throughout development [24,25]; 2) a strong elav-Gal4 that drives the expression of UAS-constructs pan-neuronally throughout development; and 3) a weak elav-Gal4 that also drives the expression of UAS-constructs pan-neuronally. As shown in Figure 2, the strong elav-Gal4 driver expresses robustly, particularly when considering that neuronal cells do not constitute a majority of the total fly volume.

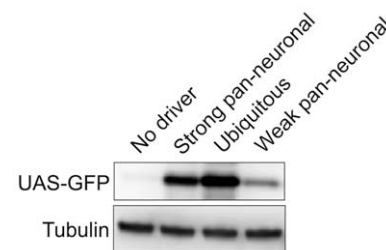


Figure 2. Gal4 drivers. Flies carrying UAS-GFP were crossed to flies carrying the Gal4 driver specified in the figure. Newly-emerged adults heterozygous for UAS-GFP and the Gal4 driver were homogenized in SDS lysis buffer and loaded into SDS/PAGE gels. Western blots show how strongly each Gal4 driver expresses UAS-GFP. Blots were probed with anti-GFP and anti-tubulin antibodies. Ubiquitous driver: sqh-Gal4. Pan-neuronal drivers: elav-Gal4. doi:10.1371/journal.pone.0043112.g002

We examined the extent of knockdown for each fly DUB through quantitative RT-PCR (see Materials and Methods). The phenotypes that we monitored included fly development, adult motility (geotaxis response) and adult longevity (Figure S2). To address non-target specificity, we used RNAi lines with a specificity score between 0.99–1 (see Materials and Methods). Also, for 20 out of a total of 33 fly DUBs, we tested two or more different RNAi lines (Figure S3). Controls were offspring from crosses of Gal4 drivers to the isogenic backgrounds of RNAi lines.

As shown in Figure 3 and Figure S3, we achieved ~80%–100% knockdown for 14 out of 33 fly DUBs tested when using the ubiquitous *sqh*-Gal4 driver. For most of the tested DUBs, however, knockdown was less than 80%. Nonetheless, ubiquitous knockdown of most fly DUBs had dramatic consequences for fruit fly development, adult motility or longevity, establishing a broad requirement for DUB functions *in vivo* (Figure 3). Out of 33 DUBs tested, 9 did not show a discernible phenotype when they were knocked down everywhere, and for three out of these 9 we did not observe any knockdown. Therefore, ubiquitous knockdown of 80% of tested fly DUBs had physiological consequences (Figure 3). This proportion is actually higher when considering DUBs whose pan-neuronal knockdown caused a phenotype even though ubiquitous knockdown did not, as discussed further below.

Even relatively low knockdown had physiological consequences for some DUBs. This includes the fly orthologues of USP20, USP30 and USP45, where moderate knockdown throughout the fly caused developmental lethality or earlier adult death (Figure 3). Reducing the mRNA of several DUBs by half, nearing hemizygosity, had physiological effects on the fly orthologues of BAP1, USP8, USP36 and TRABID. In other instances (e.g. CG4265) near-complete knockdown led to no discernible phenotype. Even with the latter exception, the DUB family of proteases is essential for *Drosophila* physiology.

Functional Redundancy. Individual knockdown of the vast majority of DUBs throughout the fly had dramatic outcomes for *Drosophila* physiology (Figure 3, Figure S3), indicating that there is little functional redundancy among fly DUBs. Based on our findings, redundancy is likely limited among human DUBs as well. This is an important notion because nearly 90 DUBs are encoded by the human genome [1] and one could assume functional overlap. Instead, according to our data, most DUBs might have evolved to fulfill distinct physiological functions by being expressed only in certain cells or tissues, by being produced during specific physiological states, by interacting with specific pools of partners, or by acting on specific substrates. This is not to say that there is no functional overlap at any level. We propose that many, or indeed most, DUBs perform at least some functions that are uniquely theirs.

Our *in vivo* analyses provide strong physiological support for studies conducted *in vitro* and in mammalian cell culture, where it was found that human DUBs have divergent structures [3], distinct substrates [2,4,26] and largely separate pools of interacting partners [27], indicating little functional redundancy.

Developmental roles for DUBs. Many *Drosophila* DUBs are clearly required during development. Ubiquitous knockdown of over 40% of the tested DUBs caused lethality in larval, pupal or pharate adult stages; others were required for both adult and developmental stages or only in adults (Figure 4A, Figure S3). Based on proportionality, ubiquitous knockdown of JAMM DUBs had a greater impact during development than other subclasses (Figures 3 and 4A), highlighting the particular importance of this subclass of DUBs for developmental physiology. Perhaps JAMM DUBs are functionally indispensable for most cell types. For example, the orthologue of PSMD14, which is involved with

ubiquitin recycling and is an integral component of the 19S proteasome [3], could be required more generally to assist with ubiquitin homeostasis. This orthologue can be compared to that of USP20, which may function only in specific tissues, cells, or stages. Unlike the JAMM subclass, USPs appear equally important during developmental stages and in adults (Figure 4A).

Cell Type Specificity. The importance of DUBs in neurons is becoming increasingly recognized, partly because mutations in a few DUBs are linked to neurological disease or neurodegeneration in mammals [5,28]. The neuronal significance for DUBs in general, however, is unclear and much remains to be discovered about ubiquitin-dependent pathways and deubiquitination in the nervous system. Consequently, we investigated the importance of individual DUBs for the nervous system by systematically knocking down fly DUBs in neurons.

Pan-neuronal knockdown of most fly DUBs caused defects during development or in adults (Figure 5). Out of the 33 DUBs tested, only 8 did not show discernible phenotypes when knocked down pan-neuronally. The distribution of phenotypic stages with pan-neuronal knockdown differed markedly from that observed with ubiquitous knockdown (Figure 4B). Pan-neuronal knockdown of ~24% of DUBs proved lethal during developmental stages, compared to ~42% of DUBs with ubiquitous knockdown (Figures 4B and 5).

For some DUBs, including the fly orthologues of USP7, USP8, USP34, USP47, USP54 and PSMD14, phenotypes from pan-neuronal and ubiquitous knockdowns were generally comparable, demonstrating the importance of these enzymes to overall neuronal development or function (Figure 5). For other DUBs, whose ubiquitous knockdown had dramatic developmental consequences, pan-neuronal knockdown led to considerably milder phenotypes, including the orthologues of USP39 and USP45. DUBs whose pan-neuronal knockdown caused milder phenotypes than their ubiquitous knockdown might be required by only a subset of neurons or select neuronal networks.

In a few other cases, including the USP14 and USP31/USP43 orthologues, ubiquitous knockdown did not lead to discernible problems, but pan-neuronal knockdown resulted in motility phenotype (Figure 5). This difference probably stems from stronger knockdown achieved in neurons by the pan-neuronal *elav*-Gal4 than by the ubiquitous *sqh*-Gal4 driver. However, this finding indicates the importance of USP31/USP43, USP14 and other similar DUBs, to neuronal cells. USP14 appears to function in part by maintaining ubiquitin homeostasis at synapses and its mutations cause ataxia in mice [29–32]. The functions of USP31 and USP43 in mammals are unclear. It will be of great interest to identify which types of neuronal cells, or networks, rely most heavily on which DUBs for ubiquitin-dependent processes.

Since pan-neuronal knockdown of the orthologues of USP39 and a few other DUBs led to a considerably milder phenotype than their ubiquitous knockdown (Figure 5), we tested whether they are required for non-neural cells. Using *repo*-Gal4 [33], we knocked down the mRNA of select DUBs only in glial cells. As shown in Figure 6, knockdown of the fly orthologue of USP39 only in glia caused lethality during development, indicating that it is overall more important to glial cells than neurons. In other cases (e.g. the EIF3H orthologue), glial or neuronal knockdown led to generally comparable phenotypes. However, in the case of the USP47 orthologue, we did not observe an overt phenotype when this DUB was knocked down in glia, demonstrating the requirement of this DUB more specifically by neuronal cells (Figure 6; see also drooping wings phenotype discussed below).

One of the advantages in using RNAi with the Gal4-UAS system is the capacity to examine gene dosage effects. For select

	Gene Identifier	Symbol	Closest Human DUB	Knockdown	Strongest Phenotype with Ubiquitous Knockdown								
					Larval Death	Pupal Death	Pharate Adult Death	Dead soon after Eclosion	Dead by 20 Days	Dead by 40 Days	Adults slower than controls	Other Remarks	
UCH	◆CG8445	Calypso	BAP1	55%								✓	
	CG4265	Uch	UCHL3	94%									No phenotype observed
	◆CG3431	Uch-L3	UCHL5	100%		✓	✓						
USP	CG15817		USP1	52%									No phenotype observed
	◆CG14619		USP2	84%						✓		✓	
	◆CG12082		USP5	84%		✓							
	◆CG1490	Usp7	USP7	99%			✓						
	◆CG5798	Ubp7	USP8	50%	✓								
	CG7023		USP12	42%									No phenotype observed
	CG5384		USP14	80%									No phenotype observed
	◆CG8494		USP20	33%						✓			
	◆CG3016		USP30	36%				✓	✓			✓	Majority died soon after eclosion
	CG30421		USP31	68%									No phenotype observed
	CG8334		USP32	39%									No phenotype observed
	◆CG5794		USP34	88%			✓	✓					Majority died as pharate adults
	◆CG5505	scny	USP36	60%	✓								
	◆CG8830		USP38	84%								✓	Late, mild phenotype
	◆CG7288		USP39	90%	✓								
	◆CG4165		USP45	23%		✓	✓	✓					Majority died in pupal and pharate stages
	◆CG5486	Ubp64	USP47	89%				✓	✓			✓	Most died soon after eclosion
	◆CG2904	ec	USP54	90%			✓	✓	✓				Majority died as pharate adults
◆CG5603	CYLD	CYLD	58%								✓	Late, mild phenotype	
MJD	◆CG3781		JOSD2	67%								✓	Late, mild phenotype
OTU	◆CG9448	trbd	TRABID	53%			✓	✓		✓		✓	Majority died as pharates or young adults
	CG4968		OTUB1	None									No phenotype observed
	CG6091		OTUD5	None									No phenotype observed
	CG7857		OTUD6B	None									No phenotype observed
	◆CG4603		YOD1	36%								✓	Late, mild phenotype
JAMM	◆CG6932	CSN6	CSN6	100%		✓							
	◆CG3416	Mov34	PSMD7	36%	✓	✓							
	◆CG18174	Rpn11	PSMD14	89%	✓								
	◆CG2224		AMSH	73%								✓	Late, mild phenotype
	◆CG9769		EIF3H	94%	✓								

Figure 3. List of phenotypes when individual DUBs are knocked down throughout the fly. Listed are the strongest phenotypes associated with ubiquitous knockdown of individual fly DUBs. Diamonds highlight DUBs whose knockdown led to observable phenotypes. Experimental groups consisted of flies heterozygous for sqh-Gal4 and UAS-RNAi. Controls were heterozygous for sqh-Gal4 on the isogenic background of RNAi lines. "Dead soon after eclosion" category denotes flies that eclosed successfully from the pupal case but fell on food and died within a few hours. Late motility phenotype means it was first observed 20 or more days after eclosing from the pupal case. doi:10.1371/journal.pone.0043112.g003

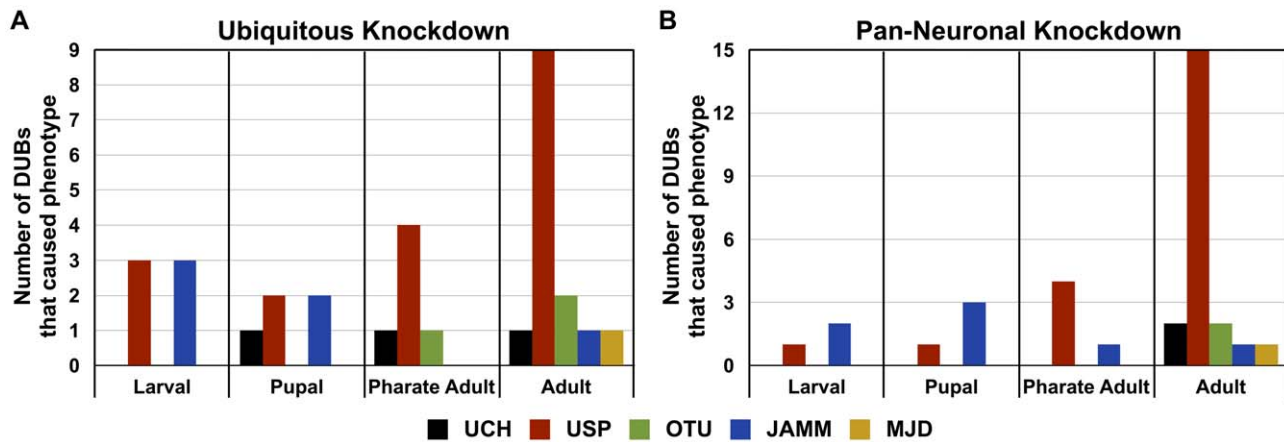


Figure 4. Phenotypic distribution by developmental stage. Histograms show the number of DUBs whose knockdown led to phenotype at each stage. A) *sqh-Gal4* (ubiquitous) driver; details in Figure 3. B) strong *elav-Gal4* (pan-neuronal) driver; details in Figure 5. In cases where two different RNAi lines targeting the same DUB led to phenotype in different stages, only the earlier stage was counted. Instances where knockdown of a specific DUB by one RNAi line led to defects in two stages were counted for each stage (e.g. knockdown of CG3416 led to death in both larval and pupal stages).

doi:10.1371/journal.pone.0043112.g004

DUBs, we used a weaker pan-neuronal driver to compare severity of phenotype to that observed with the stronger pan-neuronal driver (Figure 2). When we expressed hairpins through the weaker driver, we noticed less severe or lack of phenotype across all DUBs tested (Figure 7). The weaker driver presumably leads to a lower degree of knockdown. Thus, DUB requirement in neurons and other cell types need not be binary; different DUB levels might be required by specific cell types, populations or developmental stages.

A few points about the work presented thus far merit further discussion. Null mutations might uncover a physiological significance for DUBs whose knockdown did not lead to observable defects. Also, larger knockdown or null mutations might cause a more severe phenotype and a different distribution of phenotypes among developmental stages than what we observed. For example, null mutation of the fly orthologue of USP8 (CG5798) is embryonic lethal [34]. In our hands, ubiquitous knockdown of CG5798 did not completely eliminate its mRNA and instead led to larval or pupal death (details in Figure S3). Lower, or developmentally timed, knockdown could lead to milder phenotype. This example highlights the strength the Gal4-UAS-RNAi approach, which allows for the examination of pathways that are controlled by specific DUBs after bypassing earlier lethality and without the need to generate hypomorphic mutations.

Similar to null mutations, RNAi-based approaches cannot discriminate between catalytic and non-catalytic roles of DUBs. According to recent evidence, DUBs can play roles independent of their catalytic activity, in some cases by regulating protein complex formation [10,11]. For some DUBs we can conclude that phenotypes stem directly from lack of protease activity; knockdown of proteasome-associated DUBs PSMD14 or UCHL5 would hamper ubiquitin recycling at the proteasome. For others, the generation of domain mutations will likely be required to define their functions.

Finally, for most cases where we used two different RNAi lines to target the same DUB, we observed either similar phenotypes or phenotype severity correlated with efficiency of knockdown (Figure S3). However, this was not as clear for the orthologues of USP36 and UCHL5. For CG5505, the USP36 orthologue, we observed a more severe phenotype with lower knockdown. For CG3431, the UCHL5 orthologue, we achieved comparable knockdown with

two different RNAi lines, but phenotype severity was not identical. Perhaps there is a confounding effect from the site of insertion of UAS-RNAi. Nonetheless, because we observed clear phenotypes with all RNAi lines targeting CG5505 or CG3431, we conclude that these genes are essential for *Drosophila* development.

Insight into Molecular Mechanisms of DUBs in *Drosophila*

To examine cellular effects from the ubiquitous knockdown of select DUBs, we analyzed the distribution and levels of ubiquitinated proteins through western blotting from whole fly lysates. Western blotting has been used previously to demonstrate that knockdown, knockout, or mutations in a single DUB can be associated with differences in the levels or distribution of ubiquitinated species in mammalian cell culture and tissue [29,35–37].

As shown in Figure 8, knockdown of several DUBs throughout the fly led to markedly different levels and distribution of ubiquitinated species compared to controls. Differences in ubiquitin smears involved multiple species (Figure 8). Generally, ubiquitous knockdown led to higher levels of ubiquitinated species, as in the case of CG8494 (USP20 orthologue), CG14619 (closest to USP2) and CG5794 (USP34 orthologue). Ubiquitous knockdown of these DUBs caused early lethality (Figure 3). USP2 and USP20 have been linked to TNF pathways in mammalian cells [38–40], while USP34 has been implicated in Wnt/ β -catenin signaling [41]. Other cellular functions are also likely for these DUBs, but previous reports will inform future endeavors to identify substrates and cellular pathways through quantitative mass spectrometry using intact animals.

Ubiquitous knockdown of the USP14 orthologue did not result in discernible phenotype (Figure 3), although we noticed differences in intensity of ubiquitinated species in western blots (Figure 8). Sensitized tests, stronger ubiquitous knockdown or null mutations might uncover phenotypes that our assays did not. In support of this notion, pan-neuronal knockdown of the USP14 orthologue caused locomotion defects in adults (Figure 5). Western blotting does not indicate whether knockdown of a specific DUB leads to overall differences in ubiquitinated species because of multiple substrates or a single one. However, this analysis provides valuable information on global effects that DUBs can have on ubiquitin-dependent pathways.

	Gene Identifier	Symbol	Closest Human DUB	Phenotype with Ubiquitous Knockdown	Strongest Phenotype with Pan-Neuronal Knockdown								
					Larval Death	Pupal Death	Pharate Adult Death	Dead soon after Eclosion	Dead by 20 Days	Dead by 40 Days	Adults slower than controls	Other Remarks	
UCH	◆CG8445	Calypso	BAP1	Slower adults								✓	Late, mild phenotype
	CG4265	Uch	UCHL3	No phenotype									No phenotype observed
	◆CG3431	Uch-L3	UCHL5	Pupal, pharate death				✓					
USP	◆CG15817		USP1	No phenotype								✓	Late, mild phenotype
	◆CG14619		USP2	Slower adults, earlier death						✓		✓	
	◆CG12082		USP5	Pupal death			✓				✓	✓	Many died as pharates
	◆CG1490	Usp7	USP7	Pharate adult death			✓	✓					
	◆CG5798	Ubp7	USP8	Larval death	✓	✓							
	CG7023		USP12	No phenotype									No phenotype observed
	◆CG5384		USP14	No phenotype						✓		✓	
	◆CG8494		USP20	Earlier adult death								✓	Late, mild phenotype
	◆CG3016		USP30	Early adult death, Slower adults								✓	Late, mild phenotype
	◆CG30421		USP31	No phenotype								✓	Late, mild phenotype
	CG8334		USP32	No phenotype									No phenotype observed
	◆CG5794		USP34	Pharate, young adult death			✓	✓					
	◆CG5505	scny	USP36	Larval death							✓	✓	Mild motility phenotype
	◆CG8830		USP38	Slower adults								✓	Late, mild phenotype
	◆CG7288		USP39	Larval death								✓	
	◆CG4165		USP45	Pupal, pharate, young adult death								✓	Late, mild phenotype
	◆CG5486	Ubp64	USP47	Slower adults, early death				✓			✓	✓	Most died soon after eclosion
	◆CG2904	ec	USP54	Pharate, young adult death			✓	✓					
CG5603	CYLD	CYLD	Slower adults										No phenotype observed
MJD	◆CG3781		JOSD2	Slower adults								✓	Late, mild phenotype
OTU	◆CG9448	trbd	TRABID	Pharate, young adult death						✓		✓	Mild motility phenotype
	CG4968		OTUB1	No phenotype									No phenotype observed
	CG6091		OTUD5	No phenotype									No phenotype observed
	CG7857		OTUD6B	No phenotype									No phenotype observed
	◆CG4603		YOD1	Slower adults								✓	Late, mild phenotype
JAMM	◆CG6932	CSN6	CSN6	Pupal death				✓	✓			✓	Most died soon after eclosion
	◆CG3416	Mov34	PSMD7	Larval, pupal death		✓	✓						
	◆CG18174	Rpn11	PSMD14	Larval death	✓	✓							
	CG2224		AMSH	Slower adults									No phenotype observed
	◆CG9769		EIF3H	Larval death	✓	✓							

Figure 5. List of phenotypes when individual DUBs are knocked down pan-neuronally. Listed are the strongest phenotypes associated with pan-neuronal knockdown of individual fly DUBs. Diamonds highlight DUBs whose knockdown led to phenotypes. Experimental groups consisted of flies heterozygous for the strong elav-Gal4 driver and UAS-RNAi. Controls were heterozygous for the strong elav-Gal4 driver on the isogenic background of RNAi lines. “Dead soon after eclosion” category denotes flies that eclosed successfully from the pupal case but fell on food and died within a few hours. Late motility phenotype means it was first observed 20 or more days after eclosing from the pupal case. doi:10.1371/journal.pone.0043112.g005

Gene Identifier	Closest Human DUB	PHENOTYPE		
		Ubiquitous Knockdown	Pan-Neuronal Knockdown	Glial Knockdown
CG12082	USP5	Pupal death	Pharate adult and earlier adult death	Pharate adult and young adult death
CG7288	USP39	Larval death	Slower adults	Pupal and pharate adult death
CG5486	USP47	Young adult death	Young adult death	No phenotype observed
CG9769	EIF3H	Larval death	Larval and pupal death	Larval, pupal and pharate adult death

Figure 6. Phenotype when select DUBs are knocked down in glial cells. List compares phenotypes associated with knockdown of select fly DUBs in all cells and tissues, pan-neuronally or only in glial cells.
doi:10.1371/journal.pone.0043112.g006

Further insight into cell type specificity and function of some DUBs can also be gained from a drooping wings phenotype that we observed with the knockdown of select proteases (Figure 9). Drooping wings can result from diseased, malfunctioning or degenerating flight muscles [42,43]. Two DUBs whose ubiquitous knockdown led to drooping wings were CG8445 (the orthologue of BAP1) and CG5486 (the orthologue of USP47). In both cases, this was a progressive phenotype that corresponded with decreased motility. Interestingly, drooping wings were also observed when select DUBs were knocked down only pan-neuronally (Figure 9). Abnormal wing posture as a result of neuronal knockdown implicates DUBs in neurons supplying flight muscles. Importantly, this phenotype identifies cell types that should be investigated first to elucidate the function of these proteases *in vivo*. Future work will determine the role of these DUBs in neurons.

Conclusions

We have presented a comprehensive list of *Drosophila* DUBs and examined their physiological significance through systematic knockdown screens in intact flies. We found 41 fly DUBs, most of which were not characterized previously (Figure 10). Thirty-three of the DUBs identified fulfill our criteria for human orthologues. The vast majority of fly DUBs proved indispensable for *Drosophila* development, adult motility, or longevity (Figures 3, 5, 10). Some DUBs were necessary for neuronal cells, while others were required by cells other than neurons (e.g. glia; Figure 6), providing critical insight into cell type specificity for these proteases. Our data demonstrate that the DUB family of enzymes is necessary *in vivo*. Our genetic screens also lead us to conclude that there is little functional redundancy among DUBs. Future studies will extend our work to provide mechanistic and molecular

details into the biology of this large class of proteases that controls organismal development, function and disease.

Materials and Methods

Fly stocks, crosses and observations

Flies were reared and maintained in standard cornmeal/molasses/yeast medium in diurnally-controlled, 25°C environment. *Drosophila* RNAi stocks were purchased from the Vienna *Drosophila* RNAi Center (VDRC) and Bloomington *Drosophila* Stock Center (BDSC). We thank Dr. Norbert Perrimon (Harvard Medical School) for providing the *Drosophila* RNAi stock TRiP-CG5798. Gal4 stocks have been previously described [24,25,44]. Figure S3 lists the stock number for each *Drosophila* RNAi line and the extent of knockdown we achieved through the ubiquitous sqh-Gal4 driver, as assayed by quantitative RT-PCR (see below). To address non-target specificity, we only used VDRC lines with a specificity (s19) score of 0.99 or higher. The specificity score is determined as follows: $s19 = \frac{\sum \text{ON-target matches}}{(\sum \text{ON-target matches} + \sum \text{OFF-target matches})}$, and is reported at www.vdrc.at. Similar scores were not available for non-VDRC stocks, but the lines we utilized do not target other genes (as reported by BDSC; flystocks.bio.indiana.edu).

Four or more separate sets of crosses were set up for each RNAi line with each Gal4 driver, yielding similar results. At least 200 flies per genotype were examined. For experimental groups, parents from RNAi lines were crossed to parents from Gal4 drivers and heterozygous offspring containing both Gal4 driver and UAS-RNAi were collected and monitored. Controls were heterozygous offspring from isogenic non-RNAi lines crossed to Gal4 drivers, and in cases of balanced RNAi stocks also from balanced sibling controls containing Gal4 driver but lacking the UAS-RNAi construct. Qualitative examination of adult motility was per-

Gene Identifier	Closest Human DUB	Weak Neuronal Driver	Strong Neuronal Driver
CG3431	UCHL5	No phenotype observed	Adults dead soon after eclosion
CG12082	USP5	Mildly slower adults	Largely pharate adult death. Surviving adults progressively slower; earlier death
CG1490	USP7	No phenotype observed	Pharate adult and young adult death
CG18174	PSMD14	Slower adults	Larval and pupal death
CG3416	PSMD7	Adults dead soon after eclosion	Pupal and pharate adult death
CG9769	EIF3H	Pharate adult death	Larval and pupal death

Figure 7. Gene dosage effects with pan-neuronal knockdown. List of phenotypes observed when individual fly DUBs were knocked down by pan-neuronal drivers with different expression strengths.
doi:10.1371/journal.pone.0043112.g007

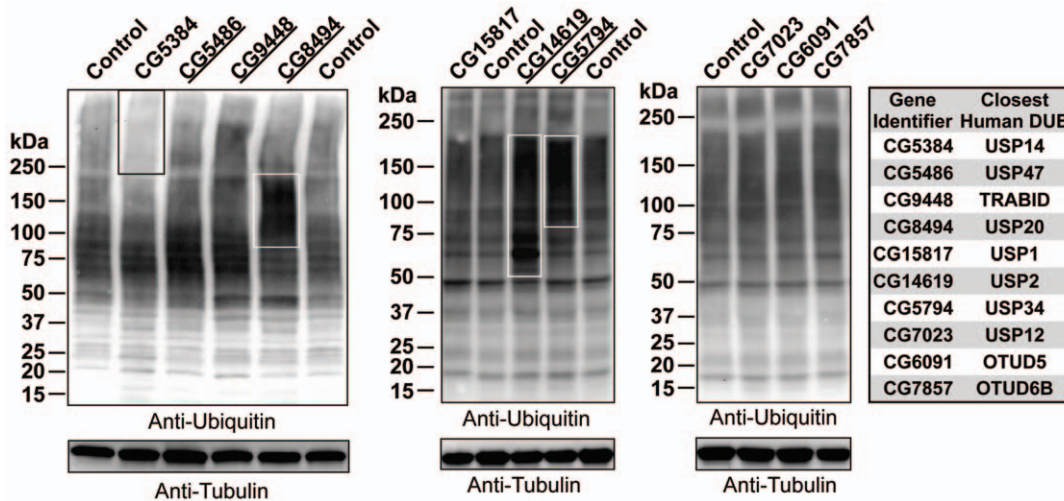


Figure 8. Differences in distribution and levels of ubiquitinated species in whole fly lysates. Shown are western blots of whole, newly-eclosed adult flies homogenized in SDS lysis buffer and electrophoresed in SDS-PAGE gels. Experimental groups were heterozygous for sqh-Gal4 and UAS-RNAi. Controls were heterozygous for sqh-Gal4 on the isogenic background of RNAi lines. Boxes highlight some areas with visible differences in ubiquitinated species. Western blots are representative of at least three independent repeats with similar results. Underlined: ubiquitous knockdown led to phenotype (Figure 3). doi:10.1371/journal.pone.0043112.g008

formed through the geotaxis response: vials containing 20–25 adult flies were tapped down lightly and fly movement up the vial wall was compared to controls. Groups were considered slower if they lagged behind controls approximately 1/5th or more of the total distance travelled in 15 seconds or when the controls reached the top, whichever occurred first. “Mild motility phenotype” denotes flies around this margin. Locomotion was examined every two or three days for a total of 40 days. Because motility decreases normally with age, we monitored same-age cohorts. Fly death was monitored daily. Longevity of control flies and those without a longevity phenotype was 60–70+ days.

Amino acid alignments and domain analyses

The amino acid sequence of each human DUB was obtained from Ensembl, using the longest isoform when multiple were predicted. Amino acid alignments against the *Drosophila* proteome were conducted using BLASTp from NCBI. Amino acid sequences for fly DUBs were retrieved from FlyBase and are listed in File S1. Domain architectures for human and fly DUBs were compiled from UniProt, SMART, PROSITE and InterPro. Most DUB domains could be found on UniProt and PROSITE, MIT domains on InterPro and coiled coil domains on UniProt and SMART. A pairwise local BLAST was obtained using the

ClustalW algorithm to validate identification of fly domains by percent identity and E-values (cutoff of 0.001).

Quantitative RT-PCR

Total RNA was extracted from young adult flies, except cases of larval, pupal or pharate adult lethality, where RNA was extracted from stages preceding death. With the exception of larvae, total RNA was extracted using TRIzol Reagent (Invitrogen). For larvae, RNA was extracted using RNAqueous®-Micro Scale RNA Isolation Kit (Ambion). Extracted RNA was treated with TURBO DNase (Ambion) to eliminate contaminating DNA. Reverse transcription was performed with the High Capacity Kit (ABI) and DUB knockdown was quantified by using the PlusOne real-time quantitative system with fast SYBR green (ABI). Rp49 and actin were used as control primers, with similar results. Primer sequences are listed in File S3. Similar to phenotypic observations, animals heterozygous for sqh-Gal4 and UAS-RNAi were compared to animals heterozygous for sqh-Gal4 on the isogenic background of UAS-RNAi lines of the same stage.

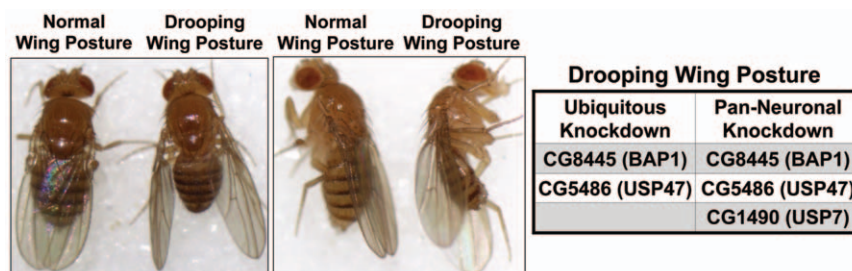


Figure 9. Wing postural defects. Shown are representative cases of flies with normal or drooping wings. Drooping wings was generally an age-dependent phenotype. Box lists DUBs whose knockdown throughout the fly or only in neurons led to this phenotype. Ubiquitous knockdown of CG1490 was lethal before adults eclosed from the pupal case. doi:10.1371/journal.pone.0043112.g009

DUB Subclass	Gene Identifier	Symbol	Closest Human DUB	Our Observations (Neuronal only where noted)	Reported Function in <i>Drosophila</i> (www.flybase.org)
UCH	CG8445	Calypso	BAP1	Slower adults	Segmental identity specification; histone deubiquitination
	CG4265	Uch	UCHL3	No phenotype observed	
	CG3431	Uch-L3	UCHL5	Pupal and pharate adult death	Neurogenesis
USP	CG15817		USP1	(Neuronal) Slower adults	
	CG14619		USP2	Slower adults, early death	
	CG12082		USP5	Pupal death	
	CG1490	Usp7	USP7	Pharate adult death	Chromatin silencing; histone deubiquitination
	CG5798	Ubpy	USP8	Larval death	Regulator of Wnt signaling pathway
	CG1945	faf	USP9X	Not tested	Embryo development; nervous system development
	CG32479		USP10	Not tested	
	CG7023		USP12	No phenotype observed	
	CG5384		USP14	(Neuronal) Slower adults, earlier death	
	CG8494		USP20	Earlier adult death	
	CG4166	not	USP22	Not tested	Transcription regulation; nervous system development
	CG3016		USP30	Slower adults, early death	
	CG30421		USP31	(Neuronal) Slower adults	
	CG8334		USP32	No phenotype observed	
	CG5794		USP34	Pharate adult and young adult death	Regulation of gene silencing
	CG5505	scny	USP36	Larval death	Regulation of apoptotic processes; histone deubiquitination
	CG8830		USP38	Slower adults	
	CG7288		USP39	Larval death	Nuclear mRNA splicing
	CG4165		USP45	Pupal, pharate adult and young adult death	
	CG5486	Ubp64E	USP47	Slower adults, early death	Regulation of chromatin silencing
CG8232		USP52	Not tested	mRNA polyA tail shortening	
CG2904	ec	USP54	Pharate adult and young adult death	Programmed cell death; eye morphogenesis	
CG5603	CYLD	CYLD	Slower adults	Determination of adult longevity; response to stress and pathogens	
MJD	CG3781		JOSD2	Slower adults	
OTU	CG12743	Otu	OTUD4	Not tested	Regulation of DNA endoreduplication
	CG9448	trbd	TRABID	Pharate adult and young adult death	Regulation of Wnt receptor signaling
	CG4968		OTUB1	No knockdown	
	CG6091		OTUD5	No knockdown	Phagocytosis
	CG7857		OTUD6B	No knockdown	
	CG4603		YOD1	Slower adults	
JAMM	CG14884	CNS5	CSN5	Not tested	Neuronal differentiation; sensory organ development; system development; axis specification
	CG6932	CSN6	CSN6	Pupal death	Mitotic progression; lateral inhibition
	CG3416	Mov34	PSMD7	Larval and pupal death	Mitotic organization; neurogenesis
	CG18174	Rpn11	PSMD14	Larval death	Positive regulation of protein turnover
	CG2224		AMSH	Slower adults	
	CG9769		EIF3H	Larval death	Autophagic cell death
	CG4751		MPND	Not tested	
	CG8877	Prp8	PRPF8	Not tested	Nuclear mRNA splicing; neurogenesis

Figure 10. Comprehensive list of fly DUBs and their physiological significance. Listed are all the fly DUBs that we identified, highlighting previously reported functions and our current findings. Not tested: DUBs that we did not examine either because of lack of reagents, too many non-specific targets from existing RNAi lines, or because the function of these DUBs is well characterized in flies. Empty cells: as of this publication, no information had been previously reported for these DUBs.
doi:10.1371/journal.pone.0043112.g010

Western Blotting and antibodies

15–20 flies per genotype were frozen in dry ice and homogenized in boiling 2%SDS/100 mM DTT lysis buffer by mechanical disruption. 50 μ L of homogenizing buffer were used per fly. Homogenates were boiled 10 minutes, sonicated for 20 seconds and centrifuged at top speed for 15 minutes. Supernatant was loaded into SDS/PAGE gels and transferred onto PVDF membrane for western blotting. Western blotting was conducted and imaged as described previously [37,45]. Antibodies used were: mouse monoclonal anti-tubulin (Sigma-Aldrich) at 1:15,000; rabbit anti-GFP (Santa Cruz Biotechnology) at 1:500; rabbit anti-ubiquitin (DAKO) at 1:500. Peroxidase-conjugated secondary antibodies (Jackson ImmunoResearch) were used at 1:10,000.

Supporting Information

Figure S1 Gal4-UAS approach to RNAi. Flies encoding a Gal4 driver are crossed to flies encoding UAS-RNAi targeting a specific DUB gene. In the resulting offspring, the Gal4 driver binds to UAS sequences and drives the expression of inverted repeats in a tissue-specific manner. Hairpin RNAs that result from the expression of the inverted repeat are processed and bind to DUB mRNA, leading to their destruction. Schematic was redrawn from a similar diagram posted by VDRC on www.vdrc.at.
(TIFF)

Figure S2 Phenotypic observations. Diagram depicts the developmental stages of the fruit fly and what was monitored in adults. Developmental time is not drawn to scale.
(TIFF)

Figure S3 RNAi lines used and phenotypic observations. Complete list of the fly DUBs that we targeted with RNAi and phenotypic observations from each line. Also shown are the stock numbers for lines from VDRC and BDSC, as well as extent of knockdown achieved by the ubiquitous driver sqh-Gal4. ND: not determined. Diamonds highlight DUBs whose knockdown led to discernible phenotype. *Drosophila* stock HMS TRIP-CG5798 was a kind gift from the laboratory of Dr. Norbert Perrimon (Harvard Medical School) before the line was deposited into BDSC.
(TIFF)

File S1 Amino acid sequences of fly DUBs.
(PDF)

File S2 Human and fly DUB domain designations and comparisons.
(PDF)

File S3 Quantitative RT-PCR primers.
(PDF)

Author Contributions

Conceived and designed the experiments: WLT SVT. Performed the experiments: WLT MJS MEM JRB KMM SVT. Analyzed the data: WLT MJS MEM CD SVT. Wrote the paper: WLT CD SVT.

References

- Nijman SM, Luna-Vargas MP, Velds A, Brummelkamp TR, Dirac AM, et al. (2005) A Genomic and Functional Inventory of Deubiquitinating Enzymes. *Cell* 123: 773–786.
- Clague MJ, Coulson JM, Urbe S (2012) Cellular Functions of the Dubs. *J Cell Sci* 125: 277–286.
- Komander D, Clague MJ, Urbe S (2009) Breaking the Chains: Structure and Function of the Deubiquitinases. *Nat Rev Mol Cell Biol* 10: 550–563.
- Reyes-Turcu FE, Ventii KH, Wilkinson KD (2009) Regulation and Cellular Roles of Ubiquitin-Specific Deubiquitinating Enzymes. *Annu Rev Biochem* 78: 363–397.
- Todi SV, Paulson HL (2011) Balancing Act: Deubiquitinating Enzymes in the Nervous System. *Trends Neurosci* 34: 370–382.
- Sacco JJ, Coulson JM, Clague MJ, Urbe S (2010) Emerging Roles of Deubiquitinases in Cancer-Associated Pathways. *Iubmb Life* 62: 140–157.
- Pickart CM (2000) Ubiquitin in Chains. *Trends Biochem Sci* 25: 544–548.
- Pickart CM, Fushman D (2004) Polyubiquitin Chains: Polymeric Protein Signals. *Curr Opin Chem Biol* 8: 610–616.
- Luna-Vargas MP, Faesen AC, Van Dijk WJ, Rape M, Fish A, et al. (2011) Ubiquitin-Specific Protease 4 Is Inhibited by Its Ubiquitin-Like Domain. *Embo Rep* 12: 365–372.
- Skaug B, Chen J, Du F, He J, Ma A, et al. (2011) Direct, Noncatalytic Mechanism of Ikk Inhibition by A20. *Mol Cell* 44: 559–571.
- Sun XX, Challagundla KB, Dai MS (2012) Positive Regulation of P53 Stability and Activity by the Deubiquitinating Enzyme Otubain 1. *Embo J* 31: 576–592.
- Fischer JA, Overstreet E (2002) Fat Facets Does a Highwire Act at the Synapse. *Bioessays* 24: 13–16.
- Fischer-Vize JA, Rubin GM, Lehmann R (1992) The Fat Facets Gene Is Required for *Drosophila* Eye and Embryo Development. *Development* 116: 985–1000.
- King RC, Storto PD (1988) The Role of the Otu Gene in *Drosophila* Oogenesis. *Bioessays* 8: 18–24.
- Steinhauer WR, Walsh RC, Kalfayan IJ (1989) Sequence and Structure of the *Drosophila* Melanogaster Ovarian Tumor Gene and Generation of an Antibody Specific for the Ovarian Tumor Protein. *Mol Cell Biol* 9: 5726–5732.
- Goodrich JS, Clouse KN, Schupbach T (2004) Hrb27c, Sqd and Otu Cooperatively Regulate Gurken Rna Localization and Mediate Nurse Cell Chromosome Dispersion in *Drosophila* Oogenesis. *Development* 131: 1949–1958.
- Xue L, Igaki T, Kuranaga E, Kanda H, Miura M, et al. (2007) Tumor Suppressor Cyld Regulates Jnk-Induced Cell Death in *Drosophila*. *Dev Cell* 13: 446–454.
- Thevenon D, Engel E, Avet-Rochex A, Gottar M, Bergeret E, et al. (2009) The *Drosophila* Ubiquitin-Specific Protease Dusp36/Scny Targets Imd to Prevent Constitutive Immune Signaling. *Cell Host Microbe* 6: 309–320.
- Brand AH, Perrimon N (1993) Targeted Gene Expression As a Means of Altering Cell Fates and Generating Dominant Phenotypes. *Development* 118: 401–415.
- Brummelkamp TR, Nijman SM, Dirac AM, Bernards R (2003) Loss of the Cylindromatosis Tumour Suppressor Inhibits Apoptosis by Activating NF-Kappab. *Nature* 424: 797–801.
- Lavorgna A, Harhaj EW (2012) An Rna Interference Screen Identifies the Deubiquitinase Stambp11 as a Critical Regulator of Human T-Cell Leukemia Virus Type 1 Tax Nuclear Export and NF-Kappab Activation. *J Virol* 86: 3357–3369.
- Moffat J, Sabatini DM (2006) Building Mammalian Signalling Pathways with Rnai Screens. *Nat Rev Mol Cell Biol* 7: 177–187.
- Metzig M, Nickles D, Falschlehner C, Lehmann-Koch J, Straub BK, et al. (2011) An Rnai Screen Identifies Usp2 as a Factor Required for Tnf-Alpha-Induced NF-Kappab Signaling. *Int J Cancer* 129: 607–618.
- Franke JD, Montague RA, Kiehart DP (2005) Nonmuscle Myosin II Generates Forces that Transmit Tension and Drive Contraction in Multiple Tissues During Dorsal Closure. *Curr Biol* 15: 2208–2221.
- Todi SV, Franke JD, Kiehart DP, Eberl DF (2005) Myosin ViiA Defects, which Underlie the Usher 1b Syndrome in Humans, Lead to Deafness in *Drosophila*. *Curr Biol* 15: 862–868.
- Ventii KH, Wilkinson KD (2008) Protein Partners of Deubiquitinating Enzymes. *Biochem J* 414: 161–175.
- Sowa ME, Bennett EJ, Gygi SP, Harper JW (2009) Defining the Human Deubiquitinating Enzyme Interaction Landscape. *Cell* 138: 389–403.
- Yi JJ, Ehlers MD (2007) Emerging Roles for Ubiquitin and Protein Degradation in Neuronal Function. *Pharmacol Rev* 59: 14–39.

29. Chen PC, Qin LN, Li XM, Walters BJ, Wilson JA, et al. (2009) The Proteasome-Associated Deubiquitinating Enzyme Usp14 Is Essential for the Maintenance of Synaptic Ubiquitin Levels and the Development of Neuromuscular Junctions. *J Neurosci* 29: 10909–10919.
30. Crimmins S, Jin Y, Wheeler C, Huffman AK, Chapman C, et al. (2006) Transgenic Rescue of Ataxia Mice with Neuronal-Specific Expression of Ubiquitin-Specific Protease 14. *J Neurosci* 26: 11423–11431.
31. Lappe-Siefke C, Loeblich S, Hevers W, Waidmann OB, Schweizer M, et al. (2009) The Ataxia (Axj) Mutation Causes Abnormal Gabaa Receptor Turnover in Mice. *Plos Genet* 5: E1000631.
32. Wilson SM, Bhattacharyya B, Rachel RA, Coppola V, Tessarollo L, et al. (2002) Synaptic Defects in Ataxia Mice Result from a Mutation in Usp14, Encoding A Ubiquitin-Specific Protease. *Nat Genet* 32: 420–425.
33. Sepp KJ, Schulte J, Auld VJ (2001) Peripheral Glia Direct Axon Guidance across the Cns/Pns Transition Zone. *Dev Biol* 238: 47–63.
34. Mukai A, Yamamoto-Hino M, Awano W, Watanabe W, Komada M, et al. (2010) Balanced Ubiquitylation and Deubiquitylation of Frizzled Regulate Cellular Responsiveness to Wg/Wnt. *Embo J* 29: 2114–2125.
35. Scaglione KM, Zavodszky E, Todi SV, Patury S, Xu P, et al. (2011) Ube2w and Ataxin-3 Coordinately Regulate the Ubiquitin Ligase Chip. *Mol Cell* 43: 599–612.
36. Schmitt I, Linden M, Khazneh H, Evert BO, Breuer P, et al. (2007) Inactivation of the Mouse Atxn3 (Ataxin-3) Gene Increases Protein Ubiquitination. *Biochem Biophys Res Commun* 362: 734–739.
37. Winborn BJ, Travis SM, Todi SV, Scaglione KM, Xu P, et al. (2008) The Deubiquitinating Enzyme Ataxin-3, A Polyglutamine Disease Protein, Edits K63-Linkages in Mixed Linkage Ubiquitin Chains. *J Biol Chem* 283: 26436–26443.
38. Krzystanek K, Rasmussen HB, Grunnet M, Staub O, Olesen SP, et al. (2012) Deubiquitylating Enzyme Usp2 Counteracts Nedd4-2-Mediated Downregulation of Kcnq1 Potassium Channels. *Heart Rhythm* 9: 440–448.
39. Mahul-Mellier AL, Pazarentzos E, Datler C, Iwasawa R, Abuali G, et al. (2012) De-Ubiquitinating Protease Usp2a Targets Rip1 and Traf2 to Mediate Cell Death by Tnf. *Cell Death Differ* 19: 891–899.
40. Yasunaga J, Lin FC, Lu X, Jeang KT (2011) Ubiquitin-Specific Peptidase 20 Targets Traf6 and Human T Cell Leukemia Virus Type 1 Tax to Negatively Regulate Nf-Kappab Signaling. *J Virol* 85: 6212–6219.
41. Lui TT, Lacroix C, Ahmed SM, Goldenberg SJ, Leach CA, et al. (2011) The Ubiquitin-Specific Protease Usp34 Regulates Axin Stability and Wnt/Beta-Catenin Signaling. *Mol Cell Biol* 31: 2053–2065.
42. Nongthomba U, Ramachandra NB (1999) A Direct Screen Identifies New Flight Muscle Mutants on the Drosophila Second Chromosome. *Genetics* 153: 261–274.
43. Liu S, Lu B (2010) Reduction of Protein Translation and Activation of Autophagy Protect Against Pink1 Pathogenesis in Drosophila Melanogaster. *Plos Genet* 6: E1001237.
44. Franke JD, Montague RA, Kiehart DP (2010) Nonmuscle Myosin Ii Is Required for Cell Proliferation, Cell Sheet Adhesion and Wing Hair Morphology during Wing Morphogenesis. *Dev Biol* 345: 117–132.
45. Todi SV, Winborn BJ, Scaglione KM, Blount JR, Travis SM, et al. (2009) Ubiquitination Directly Enhances Activity of the Deubiquitinating Enzyme Ataxin-3. *Embo J* 28: 372–382.