

## RESEARCH ARTICLE

## RE-PERG in early-onset Alzheimer's disease: A double-blind, electrophysiological pilot study

Alberto Mavilio<sup>1\*</sup>, Dario Sisto<sup>2</sup>, Florenza Prete<sup>3</sup>, Viviana Guadalupi<sup>3</sup>, Rosanna Dammacco<sup>2</sup>, Giovanni Alessio<sup>2</sup>

**1** Social Health District, Glaucoma Center, Azienda Sanitaria Locale—Brindisi, Brindisi, Italy, **2** Department of Neurosciences, Institute of Ophthalmology, University of Bari, Bari, Italy, **3** Social Health District, Alzheimer Evaluation Units, Azienda Sanitaria Locale—Brindisi, Brindisi, Italy

\* [a.mavilio@gmail.com](mailto:a.mavilio@gmail.com)

## OPEN ACCESS

**Citation:** Mavilio A, Sisto D, Prete F, Guadalupi V, Dammacco R, Alessio G (2020) RE-PERG in early-onset Alzheimer's disease: A double-blind, electrophysiological pilot study. PLoS ONE 15(8): e0236568. <https://doi.org/10.1371/journal.pone.0236568>

**Editor:** Stephen D. Ginsberg, Nathan S Kline Institute, UNITED STATES

**Received:** November 18, 2019

**Accepted:** July 8, 2020

**Published:** August 13, 2020

**Peer Review History:** PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: <https://doi.org/10.1371/journal.pone.0236568>

**Copyright:** © 2020 Mavilio et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the manuscript.

**Funding:** The Authors received no specific funding for this work.

## Abstract

## Purpose

To evaluate the ability of re-test pattern electroretinogram (RE-PERG), a non-invasive and fast steady-state PERG, to detect inner retinal bioelectric function anomalies in patients with early-onset Alzheimer's disease (AD).

## Methods

The study population consisted of 17 patients with AD-related mild cognitive impairment (MCI), 16 patients with vascular dementia (VD)-related MCI, both assessed using the neuropsychological Mini-Mental State Examination (MMSE) and by structural magnetic resonance imaging, and 19 healthy, age-matched normal controls (NC). All participants were visually asymptomatic, had normal or near-normal general cognitive functioning and no or minimal impairments in daily life activities. Visual field (VF) test, optical coherence tomography (OCT) and RE-PERG, sampled in five consecutive blocks of 130 events, were performed.

## Results

There was no statistically significant difference among the three groups with respect to age, VF parameters (mean and pattern standard deviations) and OCT parameters (ganglion cell complex thickness and retinal nerve fiber layer thickness). The mean amplitude in the RE-PERG was significantly lower, but only weakly in the AD group than in NC ( $p = 0.1$ ) whereas the intrinsic variability of the 2nd harmonic phase was significantly higher in the AD group than in either the VD or NC group ( $p < 0.001$ ).

## Conclusions

RE-PERG is altered in early-stage AD, showing a reduced amplitude with high intrinsic phase variability. It also allows the discrimination of AD from VD. A high intrinsic variability in the PERG signal, determined using RE-PERG, may thus be a new promising test for neurodegenerative diseases.

**Competing interests:** The Authors have declared that no competing interest exist.

## Introduction

Alzheimer's disease (AD), the most common type of dementia, is characterized by the extracellular accumulation of amyloid- $\beta$  protein (A $\beta$ ) plaques and intraneuronal aggregates of hyperphosphorylated tau that form neurofibrillary tangles in the brain. AD develops in ~5% of individuals over 65 years of age and in about 20% of those over 85 years of age. Currently, AD affects 26 million people around the world, and by 2050 over 100 million are expected to be affected. [1] A rare, early-onset familial AD has also been reported. [2] A non-specific cognitive decline, referred to as mild cognitive impairment (MCI), may precede AD and is frequent in the elderly population. [3,4] In addition, there are several recognized risk factors for AD, including diabetes, obesity and hypercholesterolemia. [5] The diagnosis of AD is currently made using a series of tests, beginning with questionnaires, such as the Mini-Mental State Examination, designed to assess the intellectual, emotional and functional status of the patient. [6] Second-level tests include positron emission tomography (PET), single photon emission computed tomography (SPECT), cerebrospinal fluid A $\beta$ 42 level measurement, and assessment of medial temporal lobe atrophy via brain magnetic resonance imaging (MRI). [7]

Worsening of visual function is a common feature of AD, [8] and the accumulation of A $\beta$  plaques and aggregates of hyperphosphorylated tau in the visual association cortices, [9,10] primary visual cortex, [11,12] lateral geniculate nuclei, [13,14] and the retina [15–17] has been reported. The visual disturbances in AD were long considered to be due to damage in the primary and associative visual cortex, but a primary involvement of retinal ganglion cells (RGCs) and their axons has also been proposed. [15,18–19]

Furthermore, AD patients may suffer deficits in contrast sensitivity. [20–22]

The visual pathway is composed of two different systems. The magnocellular (M) system recognizes achromatic stimuli. It originates from large RGCs and projects first to the magnocellular layers of the lateral geniculate nucleus and then to lamina 4C- $\alpha$  of the visual cortex. The second system is specific for color discrimination. It originates from small RGCs and projects first to the parvocellular layers of the lateral geniculate nucleus and then to lamina 4C- $\beta$  of the visual cortex. This system can be further divided in two different color-based pathways: the red-green parvocellular (P) and the blue-yellow koniocellular (K) subsystems. The M system responds to achromatic stimuli, and the P subsystem to chromatic stimuli but also to achromatic contrast stimuli of high spatial frequency. However, within the same range of spatial frequencies, M-cells are more sensitive to achromatic stimuli, especially at higher temporal frequencies. [23] Whether AD specifically affects one or the other sub/system is unclear. Pathologies in both the M system and the P subsystems have been described in the lateral geniculate nucleus and in the retina, [23–25] but other evidence suggests a specific M pathway involvement. [23,25–28] VD is the second most common type of dementia worldwide, and its prevalence in individuals age 65 and older is expected to double every 5 years. [29] It leads to several cognitive disorders as well as behavioral and locomotor abnormalities. The most important cause of VD is cerebral small-vessel disease; other causes are cardiac and carotid atherosclerosis, cardioembolism, hypertensive vasculopathy, aneurysm, vascular malformations, amyloid angiopathies, monogenic disorders involving stroke as well as metabolic, hematological and vasospastic disorders. Although, like AD, a diagnosis of VD can be made with certainty only post-mortem, strong clinical suspicion is based on history, timing of the event, cardiovascular and hematological assessment, psychometric evaluation and neuroimaging features. [29]

In the evaluation of AD, neuroimaging techniques include structural MRI and PET (tracing amyloid, fluorodeoxyglucose, tau). The typical MRI features of AD are a reduction of gray-matter volume, cortical atrophy and a reduced hippocampal volume. Amyloid PET is

recommended especially in patients with otherwise unexplained cognitive impairment or an atypical clinical presentation. Other types of PET are mainly used in clinical research. [30]

In VD, typical imaging features are white matter lesions, cortical and subcortical infarctions and intracerebral microhemorrhage. Extensive parenchymal infarctions are due to large-artery disease, and small infarctions especially to small-vessel disease. [29] In the eye, the primary involvement in AD patients is the RGCs [15–19] whereas the visual disturbances found in VD are often due to cerebral infarctions involving the optic pathway, leading to typical visual field alterations according to the affected site; retrogeniculate alterations do not determine subsequent optic atrophy [31] Primary involvement of the retina has also been documented in patients with cerebral autosomal arteriopathy with subcortical infarcts and leukoencephalopathy, [32] hereditary endotheliopathy with retinopathy, nephropathy and stroke, cerebroretinal vasculopathy and hereditary vascular retinopathy, which are interpreted as different phenotypes of the same disease, i.e., autosomal dominant retinal vasculopathy with cerebral leukodystrophy. [33–35] In all of these diseases, retinal damage is due to vascular retinopathy, not to primary neurodegeneration. Unlike other parts of the central nervous system (CNS), RGCs are relatively accessible and can be studied both anatomically and functionally to obtain information related to the state of neurons, including in patients with AD. [36] The properties of RGCs are similar in many ways to those of brain neurons such that anomalies in these cells can be related to brain dysfunction. In patients with AD both optic nerve degeneration and a loss of ganglion cells have been demonstrated. [37–39]

The pattern electroretinogram (PERG) is an electrophysiological test used to assess RGCs function. [40,41] Although developed for the early diagnosis of glaucoma, its utility in neurological diseases, including multiple sclerosis, [42] AD [23, 43–45] and Parkinson's disease, [46] all of which are characterized by inflammation, neurotransmission anomalies, and neurodegeneration, has also been demonstrated. PERG can provide useful diagnostic, prognostic and follow-up information on these diseases.

A specific form of PERG is steady-state PERG (SS-PERG), in which a fast (steady-state) stimulus generates a sinusoidal response that can be analyzed by Fourier transform. This allows the isolation of a second harmonic whose amplitude and phase delay can be evaluated. The PERG amplitude is related to the number of surviving neurons, and the PERG phase delay to synaptic dysfunctions of living neurons. [47] Synaptic damage and remodeling of the RGCs dendritic tree have also been histologically demonstrated in mouse models of glaucoma. [48,49] However, while a reduced amplitude is observed in patients with glaucoma and in those with ocular hypertension, [50–52] it is also a feature of conditions not related to glaucoma, such as cataract and myopia. [53–55] To overcome the limits of SS-PERG, a new test, the re-test PERG (RE-PERG), was recently introduced for the more accurate diagnosis of glaucoma. It is based on five consecutive SS-PERG stimulations without pause and evaluates the individual-intrinsic within-test phase variability of the second harmonic, rather than strictly the amplitude. Phase variability was shown to be very low in healthy controls but the standard deviation of the phase is higher in glaucoma patients. [56] Moreover, unlike the amplitude, phase variability is not influenced by optical media opacities and myopia. [57,58] Second-level imaging-based tests for the diagnosis of AD and VD are often expensive and not always available, especially in rural hospitals, such that diagnostic tools based on biomarkers able to distinguish among the various types of dementia are needed. RE-PERG uses high temporal frequency stimuli able to evoke a response of the M system. A higher phase variability is related to RGCs dysfunction, which precedes ganglion cells loss. Thus, the current research evaluated the ability of RE-PERG to detect anomalies in the primary inner retinal bioelectric function of M-cells in patients with early-onset AD compared to those with VD and in NC.

## Materials and methods

From September 1st to December 15th 2018, 52 consecutive patients (33 with MCI and 19 age-matched, healthy controls) were finally enrolled in this study. All patients were recruited at the Alzheimer Evaluation Units of the Brindisi Social Health District, Brindisi, Italy. Neurologic exclusion criteria were: neurological/psychiatric conditions other than mild AD and VD, antidepressant-antipsychotic medication, history of malignancy, head trauma or stroke, drug abuse or addiction and metabolic or endocrine anomalies.

Ophthalmic exclusion criteria were: diabetes even in the absence of retinopathy, [59] ocular hypertension and glaucoma as diagnosed by the EGS guidelines, [60] congenital optic nerve head anomalies, retinopathy or any other ocular or general condition or therapy that might influence visual function, a best corrected visual acuity  $<20/40$  (Snellen acuity), spherical refraction  $>\pm 5.0$  D, cylinder correction  $>\pm 2.0$  D and optic media opacities. The healthy control (HC) group consisted of age- and sex-matched healthy individuals with no evidence of dementia as reported by the participant or his/her family.

## Assessment of cognitive function

In the neuropsychological evaluation, cognitive function was assessed using MMSE, a simple screening test that measures global cognitive function [61] by assessing orientation, memory, concentration, language, and design capacity. The same experienced examiner administered the test. The MMSE total score ranges between 0 and 30, with lower scores indicating a poorer cognitive ability. [62] Scores  $\geq 28$  points indicate normal cognition and  $<28$  points mild (24–27 points), moderate (10–23 points) or severe ( $\leq 9$  points) cognitive impairment. A score of  $\leq 9$  points is considered to be almost diagnostic of dementia. [63]

All patients underwent structural MRI. AD and VD were diagnosed according to international consensus criteria. [64]

## Ophthalmic examination

Each participant underwent a comprehensive ophthalmic evaluation, including a review of medical history, best-corrected visual acuity testing, IOP measurement by Goldmann applanation tonometry, ultrasound pachymetry (Pachmate GH55 DGH Technology, Inc. Exton PA, USA), slit-lamp biomicroscopy, gonioscopy, and dilated fundus examination with a 78 lens. The criteria for the clinical and instrumental ophthalmic evaluation were the same as used in previous studies. [56–58]

## Standard Automated Perimetry (SAP)

The visual field was assessed using a Humphrey field analyzer, model 745i II (Carl Zeiss Meditec, Germany) and the 24–2 SITA standard strategy. Near addition was added to the refractive correction value. If fixation losses were  $>20\%$  and false-positive or false negative results  $>15\%$ , the test was repeated. At least two SAPs were performed to ensure reliable results and minimize the effect of learning. [65]

## Spectral-domain Optical Coherence Tomography (OCT)

Peripapillary retinal nerve fiber layer (RNFL) and ganglion cell complex (GCC) thicknesses were assessed using a Zeiss Cirrus HD OCT-500 (software version 7.0.1.290, Carl Zeiss Meditec, Dublin, CA). The protocol's  $200 \times 200$  optic disc cube was used to perform a circular scan 3.46 mm in diameter. The scan was automatically targeted around the optic disc to provide the RNFL thickness of the four quadrants at positions corresponding to each of the 12 hours of the clock. The protocol's  $512 \times 128$  macular cube was used to measure macular thickness.

The same experienced technician performed all the OCTs. Only images with a quality score of at least 7/10 were used. Three consecutive scans of the optic disc and macular region were acquired and analyzed for each eye. The results of the RNFL and GCC measurements were averaged using the data from each of the three scans.

### Pattern electroretinogram

RE-PERG was recorded using a commercial instrument (RETIMAX Advanced ver. 4.3 CSO Florence, Italy) and a method similar to that employed in the PERGLA paradigm, [66] with a few minor changes made by our laboratories. Specifically, we used a stimulus of horizontal bars with a spatial frequency of 1.7 cycles/degree—based on the results of previous studies showing the high sensitivity of this method in detecting RGCs dysfunction in early glaucoma [67,68]—and modulated in counter phase at 15 reversals/s. The stimulus was electronically generated on a high-resolution ionized-gas electrically charged plasma display (contrast: 90% luminance: 80 cd/m<sup>2</sup>; field size: 24° [width] × 24° [height]).

The pupils of the patients or NC were 3–4 mm, undilated, and an appropriate correction was made for the working distance (57 cm). The signals were recorded from a 9-mm Ag/AgCl skin electrode placed on the lower eyelid. A similar electrode placed on the lid of the non-stimulated eye was used as a reference, as described in other studies. The impedance was maintained below 5 K. The responses were amplified (gain of 100,000), filtered (bandwidth: 130 Hz) and sampled with a resolution of 12 bits. The analysis time was equal to the period of the stimulus (133 ms).

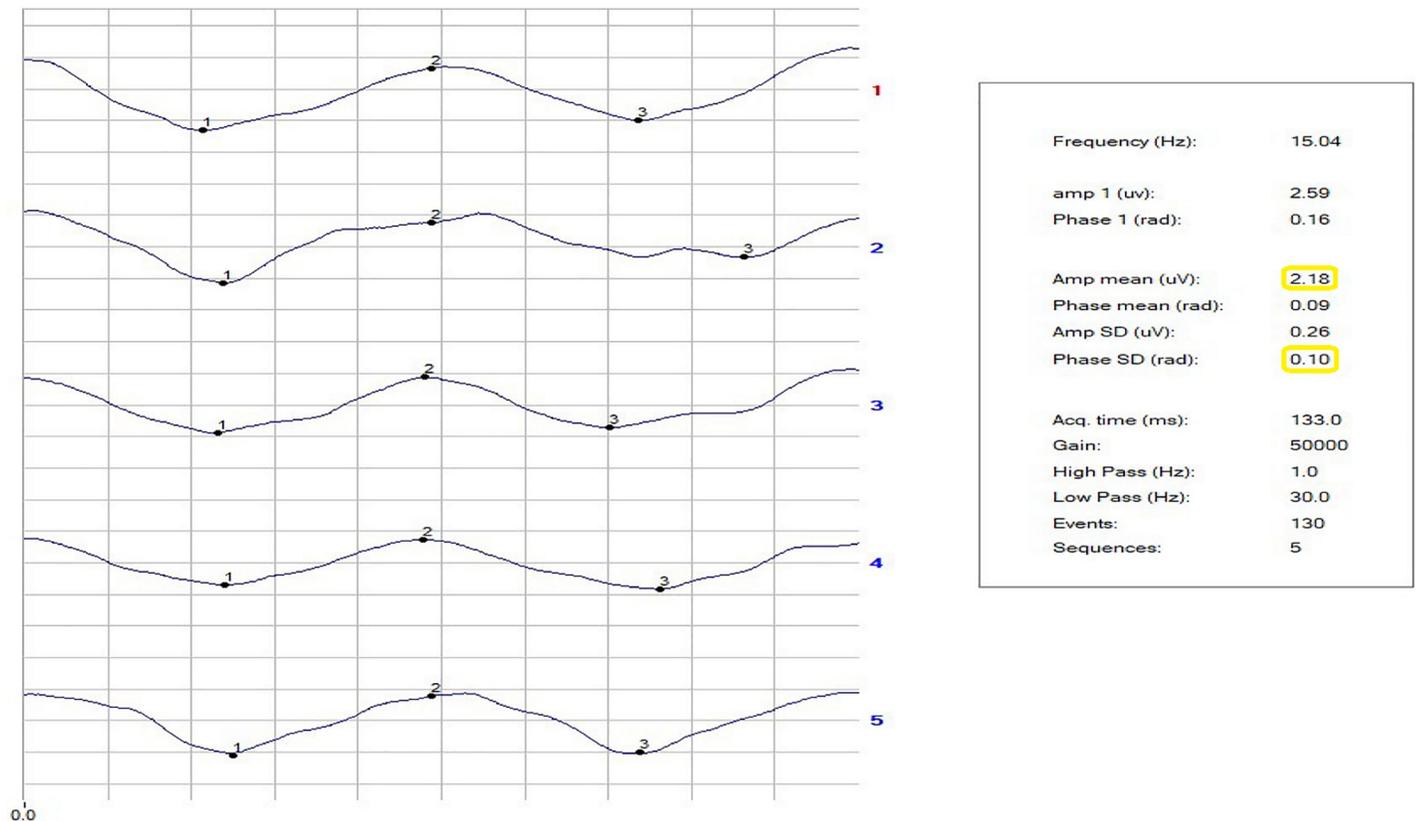
An average of 650 PERG events (5 consecutive blocks of 130 events) for RE-PERG was calculated, with the automatic rejection of artifacts. The data were then exported to a text file and the mean amplitude ( $\mu\text{V}$ ) and phase ( $\pi\text{rad}$ ) of the 2nd harmonic were analyzed by Fourier transform.

The repeatability of the phase of the second harmonic was calculated as the standard deviation of the phase (SDPh). The repeatability of the amplitude (Amp) was not considered, because of a habituation effect. [69] The noise level arising from recording a response to an occluded stimulus was  $\leq 0.087 \pm 0.03 \mu\text{V}$  in both NC and patients. Figs 1–3 show examples of a block of five events in NC and in VD and AD patients. The PERG Amp and PERG SDph values are highlighted. In our laboratory, a PERG Amp value  $< 1.5 \mu\text{V}$  and PERG SDph values  $> 0.15 \text{ SD}$  are considered to indicate pathology. The study was double blind in its design and all RE-PERGs were conducted by the same operator (A. Mavilio).

Statistical analyses were performed using Medcalc® 18.11.3. Because of the high correlation of the responses of the two eyes of the same person, only the data from one randomly chosen eye was included in the analysis. [70]

The distribution of the data was tested for normality using the Shapiro–Wilk test, and a t test was used to determine the differences between two independent groups. Comparisons of more than two independent groups were performed using a one-way ANOVA with post-hoc analyses based on the Scheffé method. The relationships between the electrophysiological values and the SAP, peripapillary RNFL thickness and GCC thickness values were calculated using Pearson's correlation tests. A chi-squared test was used to compare the groups with respect to the categorical variables (sex). A p value  $< 0.05$  was considered to indicate statistical significance.

The Ethics Committee of the Brindisi Social Health District approved the study, and the study protocol adhered to the tenets of the Declaration of Helsinki. Written informed consent was obtained from each participant after administration of the University of California, San Diego Brief Assessment of Capacity to Consent (UBACC). [71]



**Fig 1.**

<https://doi.org/10.1371/journal.pone.0236568.g001>

## Results

The study population consisted of Italians with an education level equal to that of the 8th grade in the USA. All participants lived in Apulia at the time of their enrollment in the study, between 2017 and 2018. For some patients, the family doctor had requested a neuropsychological evaluation for suspected deterioration or dementia, based on cognitive-memory loss reported by the patients; for others, a neurological examination was requested by a neurologist for various reasons, including suspicion of dementia.

Initially, 58 patients were enrolled. However, because of unreliable visual field examinations or poor-quality OCT images, 6 were excluded (4 from the AD group and 2 from the VD group), leaving 52 patients in the study.

The 17 patients in the AD group (5 males and 12 females) ranged in age between 58 and 81 years. Most were retired and came to the visit with a caregiver (usually a family member). Some had active interests, but others did not.

The 16 age-matched patients in the VD group (9 males and 7 females) had not been diagnosed with AD.

The 19 members of the HC group (12 males and 7 females) were also age-matched with the patients. Demographic and other data of the study participants are summarized in [Table 1](#). The results of the statistical analyses are reported in [Table 2](#). There was no difference between groups with respect to age, mean deviation, pattern standard deviation (PSD), RNFL and GCC, as determined in an ANOVA. AD patients had a slight significant reduction in the PERG Amp ( $1.33 \pm 0.28$  vs  $1.67 \pm 0.16$ ,  $p = 0.01$ ) value compared to NC whereas the difference

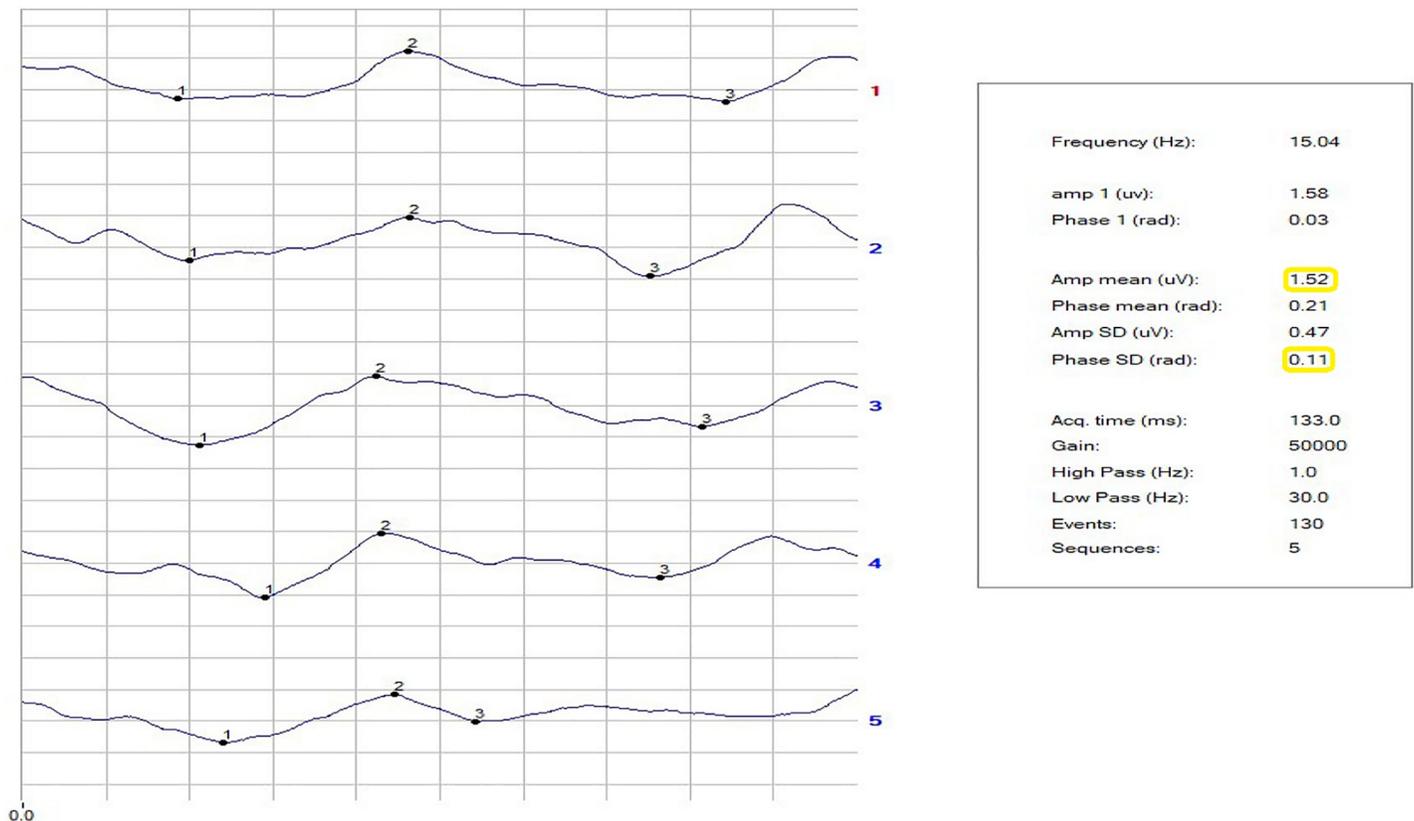


Fig 2.

<https://doi.org/10.1371/journal.pone.0236568.g002>

in the PERG SDph was highly significant between AD and VD patients ( $0.32 \pm 0.91$  vs  $0.12 \pm 0.04$ ,  $p < 0.001$ ) and between AD patients and NC ( $0.32 \pm 0.91$  vs  $0.12 \pm 0.03$ ,  $p < 0.001$ ) (Figs 4 and 5).

The MMSE score was significantly lower in both AD and VD patients than in NC ( $P = 0.02$ ;  $P = 0.01$  respectively).

The results of the correlation analysis are reported in Table 3.

There was a negative correlation between PERG Amp and age and between MMSE and PSD. Positive correlations were determined for PERG Amp and increasing PERG SDPh, for RNFL thinning and GCC thinning and for a reduction in PERG Amp and RNFL thinning.

## Discussion

The two most frequent causes of dementia worldwide are AD and VD, and their prevalence is expected to increase as populations age. Both diseases may be preceded by MCI, which is common in the elderly population but not necessarily associated with subsequent dementia. AD is associated especially with amnesic MCI, and VD with executive dysfunction and psychomotor slowness, [72] but psychometric evaluation findings alone cannot be used to discriminate VD from AD. Both AD and VD are accompanied by visual disturbances, due primarily to retinal degeneration and retrograde degeneration, respectively. The early diagnosis of AD may allow better disease management, including a delay of symptom occurrence. However, the most accurate tests for the diagnosis of AD are expensive or invasive. Consequently, there is a growing need for the detection of new, less-invasive and more cost-effective diagnostic testing.

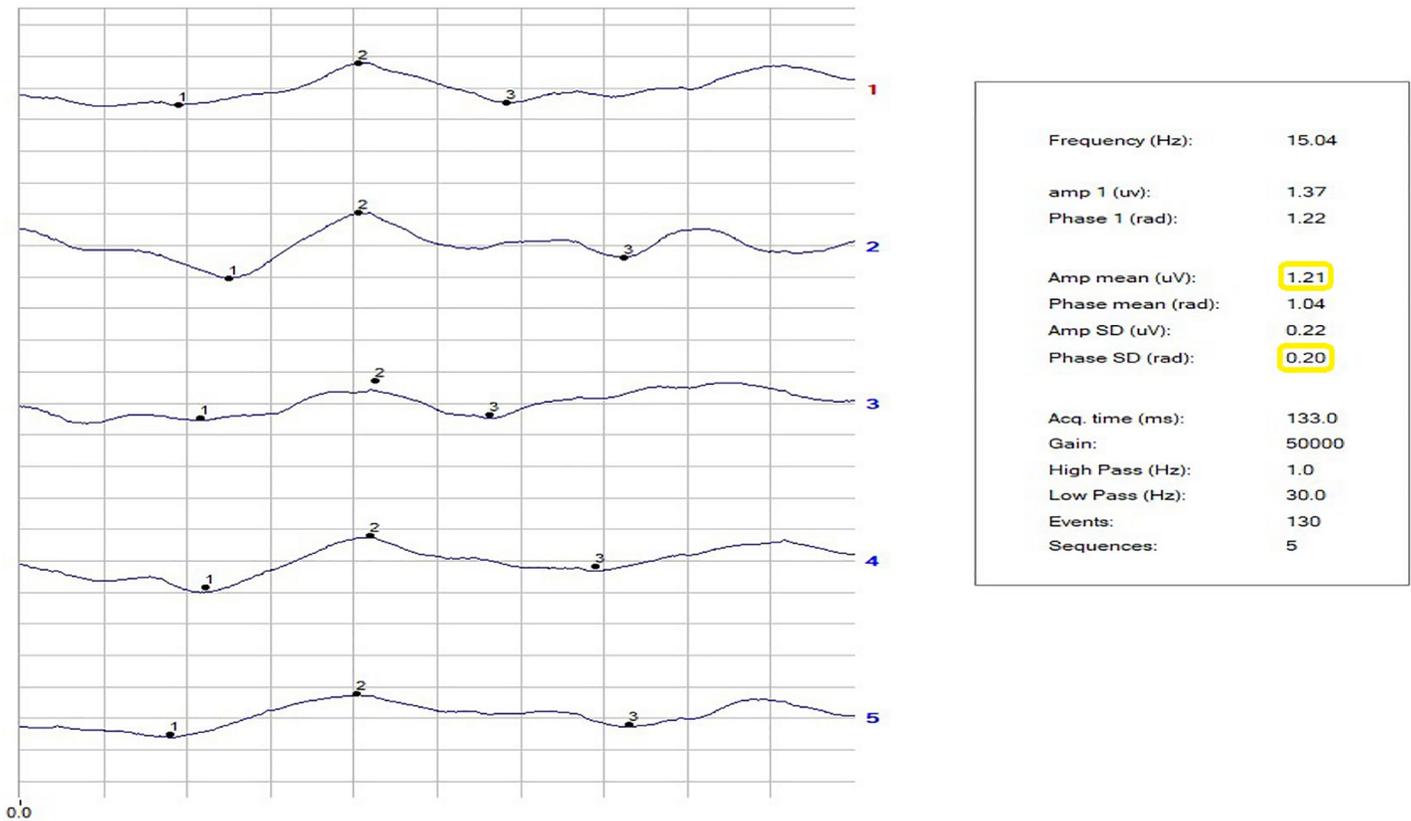


Fig 3.

<https://doi.org/10.1371/journal.pone.0236568.g003>

In most AD patients, the visual association cortices are altered whereas the primary visual cortex is spared. [73] Involvement of other areas of the visual pathway is controversial: as in some patients alterations of stereopsis and contrast sensitivity have been reported even in those without evidence of plaques and neurofibrillary tangles. [74] Other studies have demonstrated an involvement of the magnocellular pathway (a visual pathway extending from the inner layers of the retina to the primary visual cortex) in the form of the deposition of a specific type of plaque in the lateral geniculate nucleus as well as in RGCs and their axons. [75] Based on these observations, an evaluation of the macular RGC layer may provide useful diagnostic information for patients with suspected AD. The RGC layer can be studied by imaging and electrophysiological tests, PERG and visual evoked potentials. [23,76,77] For example, ssPERG tests conducted in a mouse model of AD showed alterations in the amplitude of the second wave. [78] However, in ssPERG testing the studied parameter is usually the amplitude, but it can be influenced by causes not related to neurodegeneration, such as optic media opacities and myopia, whereas the phase is not. Thus, we developed a new test, RE-PERG, in which the variability of the phase is studied based on five consecutive ssPERG stimulations. In previous studies we showed that phase variability is higher in glaucoma patients and that it is not influenced by cataract or myopia. [57,58] Since the neurodegeneration of RGCs shows similar features in glaucoma and AD, we examined the ability of RE-PERG to identify early-stage AD and to discriminate AD from VD on the basis of the different mechanisms of neurodegeneration. The results showed a slightly significant reduction in the PERG Amp value in AD patients vs. NC, but no difference between VD patients and NCs. However, the difference in the PERG SDPh in AD vs. VD patients and in AD versus NC patients was highly significant; therefore,

Table 1. Demographics and specific data.

No	Type	gender	age, years	PERG SDPh	PERG Amp (μV)	MD (dB)	PSD (dB)	RNFL (μm)	GCC (μm)	MMSE
1	VD	m	68	0.07	1.09	-1.69	1.54	88	76	25
2	VD	m	57	0.12	1.89	-0.5	1.3	85	90	24
3	VD	m	75	0.1	1.55	-1	1	94	81	25
4	VD	m	72	0.15	1.47	1.46	1.4	90	77	27
5	VD	m	64	0.09	1.44	1.44	0.8	85	72	28
6	VD	f	66	0.15	1.6	-0.97	1	93	67	28
7	VD	f	76	0.1	1.72	0.63	1.2	77	73	27
8	VD	m	68	0.15	1.77	-0.97	1	93	67	27
9	VD	f	80	0.11	1.42	1.01	1.4	95	92	26
10	VD	f	62	0.06	1.55	0.77	0.74	96	82	28
11	VD	f	67	0.23	1.53	0.85	1.02	92	80	27
12	VD	m	84	0.17	1.42	0.93	1.24	81	79	26
13	VD	m	75	0.12	1.55	0.47	0.94	90	77	26
14	VD	m	79	0.14	1.75	1.01	0.87	94	80	18
15	VD	f	72	0.14	1.7	0.88	1.5	94	90	22
16	VD	f	67	0.05	2.1	-0.04	2.2	99	81	18
17	AD	f	69	0.2	1.21	0.04	1.17	93	77	19
18	AD	m	81	0.61	1	-0.43	1.47	87	72	27
19	AD	f	74	0.15	1.34	0.89	1.34	84	77	24
20	AD	f	60	0.07	1.5	-1.29	2.05	98	84	23
21	AD	f	81	0.14	1.28	-0.78	1.28	81	77	28
22	AD	m	66	0.48	1.16	-0.68	1.9	76	74	24
23	AD	f	70	0.22	1.67	0.47	1.37	97	82	24
24	AD	f	58	0.18	1.82	0.71	1.39	106	92	24
25	AD	f	78	0.39	0.93	1.51	1.43	88	80	24
26	AD	m	81	0.47	1.57	1.51	1.43	93	72	23
27	AD	f	67	0.11	1.57	0.23	1.8	81	77	21
28	AD	m	72	0.25	1.32	0.5	1.8	70	55	22
29	AD	m	77	0.66	0.84	-0.75	1.22	82	76	25
30	AD	f	79	0.58	1.11	1.34	0.97	80	65	28
31	AD	f	60	0.3	1.1	0.8	1.12	77	75	29
32	AD	f	76	0.14	1.52	-0.33	1.12	80	81	28
33	AD	f	70	0.46	1.66	1.15	1.1	87	80	28
34	NC	m	74	0.09	1.62	-0.4	0.8	88	72	28
35	NC	m	74	0.13	1.78	-0.5	1.3	74	64	27
36	NC	m	78	0.1	1.46	-1.01	1.2	88	70	29
37	NC	m	68	0.12	1.65	2	1.5	104	91	27
38	NC	m	70	0.11	1.58	1.3	1.55	101	91	26
39	NC	f	74	0.12	1.79	1.1	0.88	87	71	27
40	NC	m	70	0.1	1.71	-0.97	1	95	76	28
41	NC	m	65	0.1	1.5	0.63	1.2	96	83	27
42	NC	f	60	0.1	1.81	1.01	0.87	80	80	25
43	NC	f	64	0.08	1.59	0.85	1.02	93	87	26
44	NC	m	65	0.1	2.14	-0.23	1.1	79	70	28
45	NC	m	60	0.11	1.51	-2.1	1.53	99	90	28
46	NC	m	68	0.16	1.53	0.93	1.46	84	80	29
47	NC	m	66	0.1	1.69	1.01	1.4	88	74	30
48	NC	m	77	0.16	1.75	1.81	1.41	82	80	28

(Continued)

Table 1. (Continued)

No	Type	gender	age, years	PERG SDPh	PERG Amp ( $\mu$ V)	MD (dB)	PSD (dB)	RNFL ( $\mu$ m)	GCC ( $\mu$ m)	MMSE
49	NC	f	86	0.1	1.65	-0.5	1.3	92	86	25
50	NC	f	62	0.15	1.5	-0.45	1.34	70	87	25
51	NC	m	66	0.2	1.75	1	1.4	100	90	27
52	NC	m	66	0.17	1.76	0.4	1.5	99	90	25

Mean Deviation (MD) Pattern Standard Deviation (PSD), Retinal Nerve Fiber Layer Thickness (RNFL), ganglion cell complex (GCC), steady-state intrinsic phase variability (PERG SDph) steady-state PERG amplitude (PERG Amp), Mini-Mental State Examination (MMSE) in Early Alzheimer disease (AD), Vascular Dementia-related MCI (VD) and Normal Controls (NC)

<https://doi.org/10.1371/journal.pone.0236568.t001>

PERG SDPh may be of value not only in detecting inner retinal dysfunction in AD, but also in distinguishing between AD and VD.

Correlation studies showed a negative correlation between PERG Amp and age, as expected due to the physiological loss of RGCs. The negative correlation between MMSE and PSD, that is, a worsening of the visual field related to a reduction in the MMSE score, may reflect the neurodegeneration occurring both in the retina and in the brain. The positive correlation between PERG Amp reduction and an increased PERG SDPh can be explained by a worsening of all parameters with disease progression, and that between RNFL and RGC thinning by the parallel degeneration of neuronal cell bodies and axons (Table 3). The positive correlation between PERG Amp reduction and RNFL thinning indicates that the amplitude is related to the number of surviving RGCs. The findings of our study suggest that PERG SDph is a suitable parameter to detect early damage to magnocellular RGCs in AD patients. While the M system has been shown to respond to stimuli of high temporal frequency, a response by the P system cannot be excluded, also because the K and P visual streams were not specifically tested. However, there are fewer P cells and they tend to be scattered, such that the increased PERG SDph could be predominantly attributed to M dysfunction. Our finding is in agreement with other studies in which involvement of the M pathway was reported. [23]

As noted above, the phase variation is related to the synaptic loss and dendritic degeneration that may precede ganglion cell loss. [47] Such alterations have been described in early AD, but also in Parkinson's and Huntington's diseases. [79] Normal neuronal activity is

Table 2. Demographic and relevant ocular characteristic of study participants.

	AD (17)		VD (16)		NC (19)		P-value <sup>®</sup>		
	Mean	SD	Mean	SD	Mean	SD	AD vs VD	VD vs NC	AD vs NC
age	71.71	7.63	70.75	7.17	69.10	6.67	P = 0.72	P = 0.5	P = 0.44
PERG Amp ( $\mu$ v)	1.33	0.28	1.59	0.23	1.67	0.16	P = 0.2	P = 0.26	P = 0.01
PERG SDph	0.32	0.19	0.12	0.04	0.12	0.03	P < 0.001	P = 0.95	P < 0.001
MD (db)	0.29	0.88	0.27	0.99	0.31	1.07	P = 0.5	P = 0.90	P = 0.31
PSD (db)	1.41	0.31	1.20	0.37	1.25	0.24	P = 0.6	P = 0.24	P = 0.52
GCC ( $\mu$ m)	76.23	7.96	79.00	7.39	80.63	8.60	P = 0.6	P = 0.24	P = 0.23
RNFL ( $\mu$ m)	85.89	9.18	90.38	5.85	89.42	9.56	P = 0.1	P = 0.73	P = 0.77
MMSE	24.76	2.84	25.12	3.20	27.10	1.49	P = 0.73	P = 0.02	P = 0.01

Mean Deviation (MD) Pattern Standard Deviation (PSD), Retinal Nerve Fiber Layer Thickness (RNFL), ganglion cell complex (GCC), steady-state intrinsic phase variability (PERG SDph) steady-state PERG amplitude (PERG Amp), Mini-Mental State Examination (MMSE) in Early Alzheimer disease (AD), Vascular Dementia-related MCI (VD) and Normal Controls (NC)

\*—One Way Analysis of Variance (Bonferroni corrected); \*\*—Chi-Square

<https://doi.org/10.1371/journal.pone.0236568.t002>

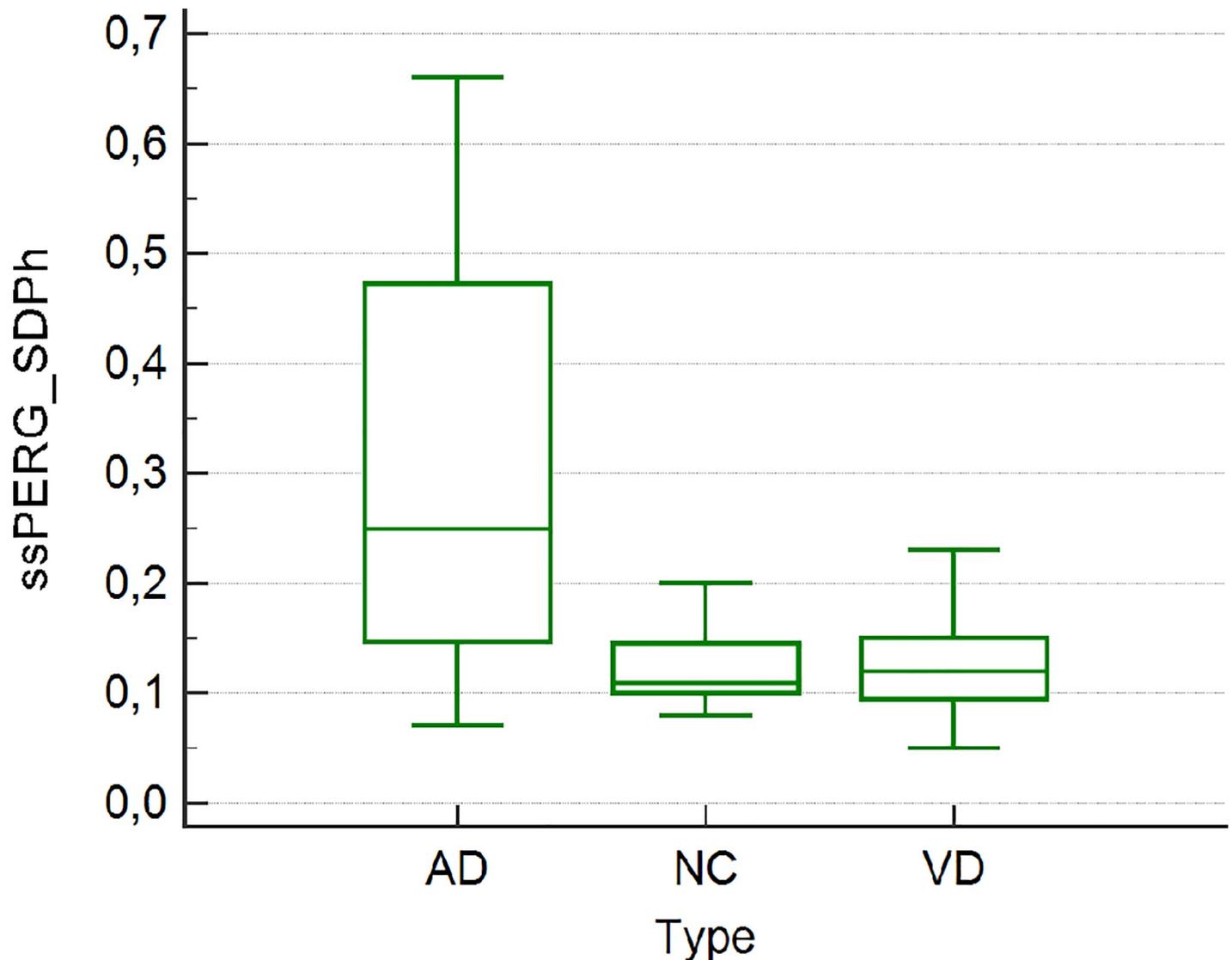


Fig 4.

<https://doi.org/10.1371/journal.pone.0236568.g004>

accompanied by a high energy demand; [80] such that RE-PERG serves as a metabolic stress test able to show early damage to RGCs.

In glaucoma patients, functional and anatomical changes may be present in RGCs before any damage of the optic nerve is detectable. [52] Thus, in CNS diseases that share some features of the degeneration seen in glaucoma, the same may be true.

Our study may have been biased by several factors. First, the diagnosis of MCI was based exclusively on the MMSE, which cannot replace a full psychometric evaluation. Tests specific for AD include the Alzheimer's Disease Assessment Scale (ADAS-Cog), the Clinical Dementia Rating (CDR) score, and the Repeatable Battery for the Assessment of Neuropsychological Status (RBANS). In the diagnosis of VD, the Montreal Cognitive Assessment (MoCA) has a higher sensitivity and specificity than the MMSE. [81] However, our study was performed in a National Health Service setting, and MMSE is the only psychometric test available. Thus, it cannot be ruled out that a more specific evaluation would have led to a different definition of mental status and influenced our results.

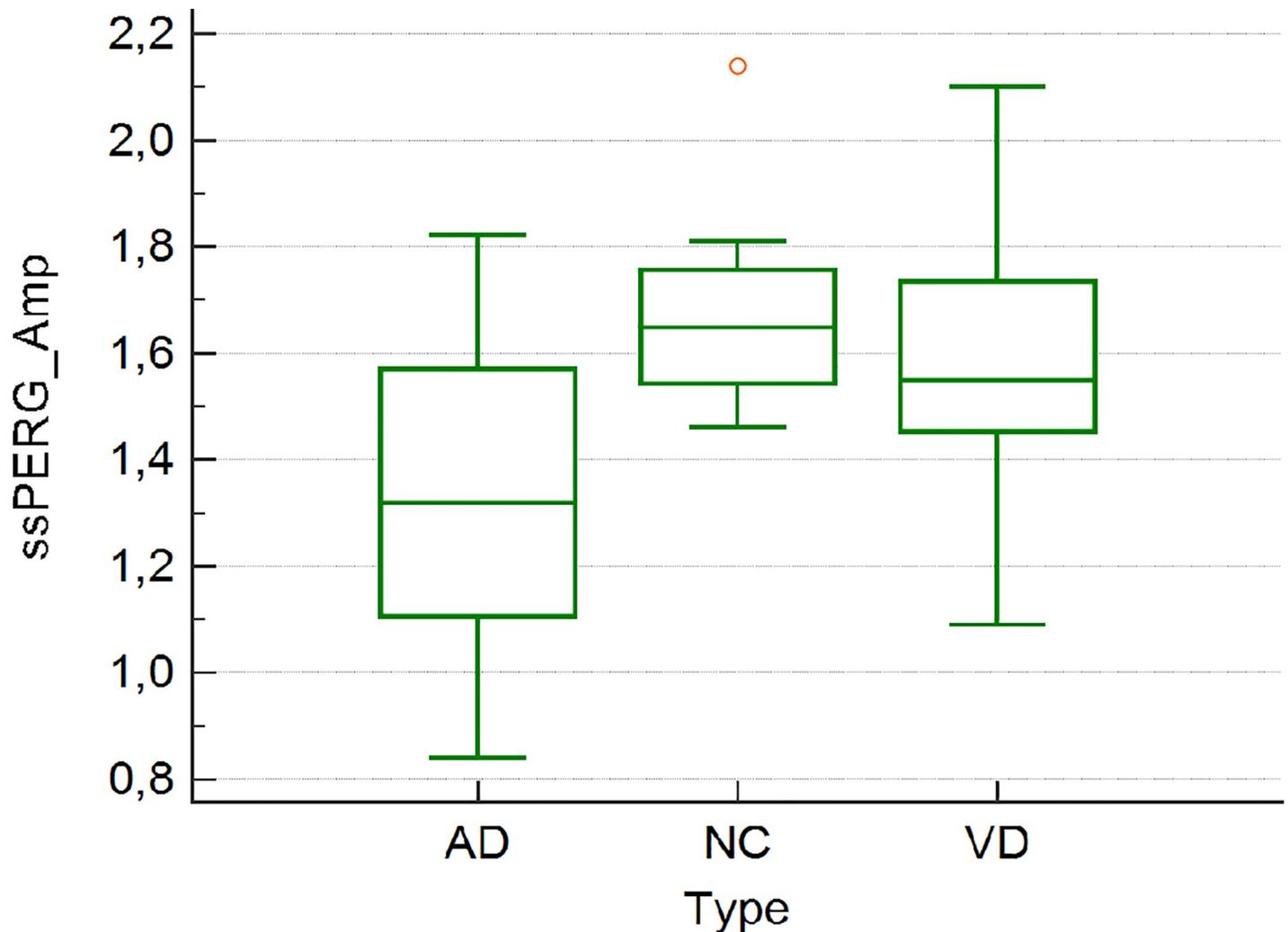


Fig 5.

<https://doi.org/10.1371/journal.pone.0236568.g005>

Another possible source of bias was the small number of enrolled patients. Further studies with a larger cohort of patients are required to confirm our preliminary results.

Two issues emerge from this study. The first is the question whether the alteration in PERG SDPh is a sign of primary RGCs degeneration or related to transsynaptic degeneration in the visual cortex. In our opinion, the first hypothesis is more likely, as RGC thinning has been found both in prodromal and in preclinical AD as well as in patients without other signs of visual cortex involvement. [82,83]

The second is the shared finding of an altered PERG SDPh in both glaucoma and AD. AD and glaucoma have several common features. Epidemiological studies have shown that the prevalence of glaucoma in AD patients is about 25% vs. 5–6% in the non-AD population. [84,85] Abnormal folded amyloid beta ( $A\beta$ ) and tau protein, typical findings in AD, have been demonstrated both in mouse models of glaucoma and in humans with the disease. [86,87] In addition, several studies have shown OCT alterations typical of glaucoma, such as RNFL and RGC thinning, in patients with early and even preclinical AD, [88,89] and visual field alterations detected in glaucoma, including arcuate defects, also occur in AD. [90] Finally, an

Table 3. Correlation table.

		age	MD (dB)	PSD (dB)	GCC ( $\mu\text{m}$ )	RNFL ( $\mu\text{m}$ )	PERG Amp ( $\mu\text{V}$ )	PERG SDPh	MMSE
age	CC		0.12	-0.09	-0.27	-0.17	-0.29	0.3	-0.02
	SL-P		0.4	0.5	0.05	0.22	0.03	0.03	0.90
MD (db)	CC	0.12		-0.13	0.13	0.06	0.09	0.11	0.002
	SL-P	0.4		0.36	0.35	0.67	0.53	0.43	0.99
PSD (db)	CC	-0.09	-0.13		0.17	0.06	-0.07	0.07	-0.42
	SL-P	0.54	0.36		0.22	0.69	0.64	0.64	0.0018
GCC ( $\mu\text{m}$ )	CC	-0.27	0.13	0.17		0.59	0.21	-0.27	-0.14
	SL-P	0.051	0.35	0.22		<0.0001	0.13	0.054	0.31
RNFL ( $\mu\text{m}$ )	CC	-0.17	0.06	0.06	0.59		0.28	-0.24	-0.15
	SL-P	0.22	0.67	0.69	<0.0001		0.04	0.08	0.29
PERG Amp ( $\mu\text{V}$ )	CC	-0.29	0.09	-0.07	0.21	0.28		-0.6	-0.05
	SL-P	0.038	0.52	0.63	0.13	0.05		<0.0001	0.71
PERG SDPh	CC	0.3	0.11	0.07	-0.27	-0.24	-0.6		-0.006
	SL-P	0.03	0.43	0.64	0.054	0.08	<0.0001		0.97
MMSE	CC	-0.02	0.002	-0.42	-0.14	-0.15	-0.05	-0.006	
	SL-P	0.91	0.99	0.0018	0.31	0.29	0.71	0.97	

Pearson Correlation Coefficient (CC) and Significance Level P (SL-P) between Mean Deviation (MD) Pattern Standard Deviation (PSD), Retinal Nerve Fiber Layer Thickness (RNFL), ganglion cell complex (GCC), steady-state intrinsic phase variability (PERG SDph) steady-state PERG amplitude (PERG Amp) and Mini-Mental State Examination (MMSE) in all participants

<https://doi.org/10.1371/journal.pone.0236568.t003>

enlarged cup-to-disc ratio of the optic nerve, the most typical feature of glaucoma, has also been detected in some, [91–93] but not all [94,95] AD patients.

Recently, optical coherence tomography angiography (OCT-A) has been used to study AD. Bulut et al. reported a lower retinal vascular density (VD) and choroidal thickness [96] together with an enlargement of the foveal avascular zone (FAZ) in AD patients compared to NC. In a comparison of AD and primary open-angle glaucoma (POAG) patients, Zabel et al. found a larger FAZ and a reduced vascular density in the deep vascular plexus in the AD group [97] whereas in POAG patients reductions in the vascular density of the superficial vascular plexus and in radial peripapillary capillaries were detected. However, a reduced VD and FAZ enlargement have also been reported in normal-tension glaucoma (NTG). [98] In addition, an even larger FAZ occurs in progressed glaucoma (both NTG and POAG) [99] and in POAG patients with central visual field defects. [100] The FAZ is also variably influenced by glaucoma surgery. [101]

Finally, the reduced VD of the deep macular plexus, such as reported by Zabel et al. in AD patients, is also a feature of progressed NTG. [99] Thus, whether OCT-A findings comprise a specific biomarker of AD remains to be determined in further studies. Moreover, these studies also demonstrate that all of the tools used to diagnose glaucoma may be biased by the presence of AD. In the absence of an elevated intraocular pressure, i.e. in a patient with NTG, the differential diagnosis can be particularly challenging and AD has to be carefully ruled out.

Other causes of inner retinal dysfunction, detectable by electrophysiological tests, as stated before, are Multiple Sclerosis and Parkinson's disease; we didn't test RE-PERG in these diseases, but its alteration cannot be excluded. At the same way, it is also known that age-related visual conditions such as age itself, presbyopia and cataract can influence PERG. As for cataract, we showed reduced amplitude with small intrinsic variability of the phase in a RE-PERG pilot study,[57] but further studies are required also in the other above-mentioned conditions. Our results suggest that RE-PERG is a quick, easy to perform and non-invasive test able to

detect RGC dysfunction in AD, but despite its promise its utility must be confirmed in other laboratories and in larger cohorts of patients.

## Author Contributions

**Conceptualization:** Alberto Mavilio.

**Data curation:** Alberto Mavilio.

**Funding acquisition:** Alberto Mavilio.

**Investigation:** Alberto Mavilio, Florenza Prete, Viviana Guadalupi, Rosanna Dammacco.

**Methodology:** Alberto Mavilio.

**Software:** Alberto Mavilio.

**Validation:** Alberto Mavilio.

**Visualization:** Giovanni Alessio.

**Writing – review & editing:** Dario Sisto.

## References

1. Prince M., Wimo A., Guerchet M. et al. An analysis of prevalence, incidence, cost & trends. Alzheimer's Disease International; London: 2015. World Alzheimer Report 2015.
2. Hickman RA, Faustin A, Wisniewski T. Alzheimer disease and its growing epidemic: risk factors, biomarkers, and the urgent need for therapeutics. *Neurol Clin* 2016; 34:941–53. <https://doi.org/10.1016/j.ncl.2016.06.009> PMID: 27720002
3. Petersen RC, Smith GE, Waring SC, et al. Mild Cognitive Impairment. *Arch. Neurol.* 1999; 56(3): 303. <https://doi.org/10.1001/archneur.56.3.303> PMID: 10190820;
4. Petersen RC. Mild cognitive impairment as a diagnostic entity. *J. Intern. Med.* 2004; 256(3):183–194. <https://doi.org/10.1111/j.1365-2796.2004.01388.x> PMID: 15324362
5. Bates KA, Sohrabi HR, Rodrigues M et al. Association of cardiovascular factors and Alzheimer's disease plasma amyloid-beta protein in subjective memory complainers. *J Alzheimers Dis* 2009; 17(2): 305–318 <https://doi.org/10.3233/JAD-2009-1050> PMID: 19363264
6. Folstein MF, Folstein SE, McHugh PR "Mini-mental state". A practical method for grading the cognitive state of patients for the clinician. *J Psychiatr Res* 1875; 12(3): 189–198
7. Jack CR Jr Alzheimer disease: new concepts on its neurobiology and the clinical role imaging will play. *Radiology* 2012; 263(2): 344–361 <https://doi.org/10.1148/radiol.12110433> PMID: 22517954
8. Cerquera-Jaramillo MA, Nava-Mesa MO, González-Reyes RE et al. Visual Features in Alzheimer's Disease: From Basic Mechanisms to Clinical Overview *Neural Plast.* 2018 Oct 14; 2018:2941783. <https://doi.org/10.1155/2018/2941783> eCollection 2018 PMID: 30405709
9. Whitehouse PJ, Price DL, Clark AW, Coyle JT, DeLong MR. Alzheimer disease: evidence for selective loss of cholinergic neurons in the nucleus basalis. *Ann Neurol.* 1981; 10:122–6. <https://doi.org/10.1002/ana.410100203> PMID: 7283399
10. van de Nes JAP, Nafe R, Schlote W. Non-tau based neuronal degeneration in Alzheimer's disease—an immunocytochemical and quantitative study in the supragranular layers of the middle temporal neocortex. *Brain Res.* 2008; 1213:152–65. <https://doi.org/10.1016/j.brainres.2008.03.043> PMID: 18455153
11. Leuba G, Kraftsik R. Visual cortex in Alzheimer's disease: occurrence of neuronal death and glial proliferation, and correlation with pathological hallmarks. *Neurobiol Aging.* 1994; 15:29–43. [https://doi.org/10.1016/0197-4580\(94\)90142-2](https://doi.org/10.1016/0197-4580(94)90142-2) PMID: 8159261
12. Armstrong RA. Is there a spatial association between senile plaques and neurofibrillary tangles in Alzheimer's disease? *Folia Neuropathol.* 2005; 43:133–8. PMID: 16245206
13. Leuba G, Saini K. Pathology of subcortical visual centres in relation to cortical degeneration in Alzheimer's disease. *Neuropathol Appl Neurobiol.* 1995; 21:410–22 <https://doi.org/10.1111/j.1365-2990.1995.tb01078.x> PMID: 8632836
14. Dugger B, Tu M, Murray ME, Dickson DW. Disease specificity and pathologic progression of tau pathology in brainstem nuclei of Alzheimer's disease and progressive supranuclear palsy. *Neurosci Lett.* 2011; 491:122–6 <https://doi.org/10.1016/j.neulet.2011.01.020> PMID: 21236314

15. Hinton DR, Sadun AA, Blanks JC, Miller CA. Optic-nerve degeneration in Alzheimer's disease. *N Engl J Med*. 1986; 315:485–7. <https://doi.org/10.1056/NEJM198608213150804> PMID: 3736630
16. Löffler K, Edward DP, Tso MO. Immunoreactivity against tau, amyloid precursor protein, and beta-amyloid in the human retina. *Invest Ophthalmol Vis Sci*. 1995; 36:24–31. PMID: 7822152
17. La Morgia C, Ross-Cisneros FN, Koronyo Y, et al. Melanopsin retinal ganglion cell loss in Alzheimer disease. *Ann Neurol*. 2016; 79:90–109. <https://doi.org/10.1002/ana.24548> PMID: 26505992
18. Sadun A, Bassi CJ. Optic nerve damage in Alzheimer's disease. *Ophthalmology*. 1990; 97:9–17. [https://doi.org/10.1016/s0161-6420\(90\)32621-0](https://doi.org/10.1016/s0161-6420(90)32621-0) PMID: 2314849
19. Liu S, Ong Y-T, Hilal S, Loke YM, Wong TY, Chen CL-H, Cheung CY, Zhou J. The association between retinal neuronal layer and brain structure is disrupted in patients with cognitive impairment and Alzheimer's disease. *J Alzheimers Dis*. 2016; 54:585–95 <https://doi.org/10.3233/JAD-160067> PMID: 27567815
20. Hutton J, Morris JL, Elias JW, Poston JN. Contrast sensitivity dysfunction in Alzheimer's disease. *Neurology*. 1993; 43:2328–30. <https://doi.org/10.1212/wnl.43.11.2328> PMID: 8232951
21. Gilmore G, Whitehouse PJ. Contrast sensitivity in Alzheimer's disease: a 1-year longitudinal analysis. *Optom Vis Sci*. 1995; 72:83–91. <https://doi.org/10.1097/00006324-199502000-00007> PMID: 7753532
22. Crow R, Levin LB, LaBree L, Rubin R, Feldon SE. Sweep visual evoked potential evaluation of contrast sensitivity in Alzheimer's dementia. *Invest Ophthalmol Vis Sci*. 2003; 44:875–8 <https://doi.org/10.1167/iovs.01-1101> PMID: 12556424
23. Sartucci F, Borghetti D, Bocci T, Murri L, Orsini P, Porciatti V, Origlia N, Domenici L. Dysfunction of the magnocellular stream in Alzheimer's disease evaluated by pattern electroretinograms and visual evoked potentials. *Brain Res Bull* 2010; 82(3):169–176
24. Hof PR, Vogt BA, Bouras C, Morrison JH. Atypical form of Alzheimer's disease with prominent posterior cortical atrophy: a review of lesion distribution and circuit disconnection in cortical visual pathways. *Vision Res*. 1997; 37:3609–3625. [https://doi.org/10.1016/S0042-6989\(96\)00240-4](https://doi.org/10.1016/S0042-6989(96)00240-4) PMID: 9425534
25. Lennie P, Krauskopf J, Sclar G. Chromatic mechanisms in striate cortex of macaque. *J Neurosci*. 1990; 10:649–669. <https://doi.org/10.1523/JNEUROSCI.10-02-00649.1990> PMID: 2303866
26. Levy JA, Chelune GJ. Cognitive-behavioral profiles of neurodegenerative dementias: beyond Alzheimer's disease. *J Geriatr Psychiatry Neurol*. 2007; 20:227–238. <https://doi.org/10.1177/0891988707308806> PMID: 18004009
27. Sadun AA, Borchert M, DeVita E, Hinton DR, Bassi CJ. Assessment of visual impairment in patients with Alzheimer's disease. *Am J Ophthalmol*. 1987; 104:113–120. [https://doi.org/10.1016/0002-9394\(87\)90001-8](https://doi.org/10.1016/0002-9394(87)90001-8) PMID: 3618708
28. Kalaria R. N. (2012). Cerebrovascular disease and mechanisms of cognitive impairment: evidence from clinicopathological studies in humans. *Stroke*, 43(9), 2526–2534. <https://doi.org/10.1161/STROKEAHA.112.655803> PMID: 22879100
29. Femminella G. D., Thayanandan T., Calsolaro V., Komici K., Rengo G., Corbi G., & Ferrara N. (2018). Imaging and molecular mechanisms of Alzheimer's disease: A review. *International journal of molecular sciences*, 19(12), 3702
30. Lachenmayr, Bernhard J., and Patrick MO Vivell. *Perimetry and its clinical correlations*. Georg Thieme Verlag, 1993.
31. Parisi Vincenzo, et al. "Visual electrophysiological responses in subjects with cerebral autosomal arteriopathy with subcortical infarcts and leukoencephalopathy (CADASIL)." *Clinical neurophysiology* 111.9 (2000): 1582–1588. [https://doi.org/10.1016/s1388-2457\(00\)00366-7](https://doi.org/10.1016/s1388-2457(00)00366-7) PMID: 10964068
32. Jen J., et al. "Hereditary endotheliopathy with retinopathy, nephropathy, and stroke (HERNS)." *Neurology* 49.5 (1997): 1322–1330. <https://doi.org/10.1212/wnl.49.5.1322> PMID: 9371916
33. Ophoff Roel A., et al. "Hereditary vascular retinopathy, cerebroretinal vasculopathy, and hereditary endotheliopathy with retinopathy, nephropathy, and stroke map to a single locus on chromosome 3p21. 1-p21. 3." *The American Journal of Human Genetics* 69.2 (2001): 447–453. <https://doi.org/10.1086/321975> PMID: 11438888
34. Terwindt GM1, et al. "Clinical and genetic analysis of a large Dutch family with autosomal dominant vascular retinopathy, migraine and Raynaud's phenomenon." *Brain: a journal of neurology* 121.2 (1998): 303–316.
35. Chiquita S, Rodrigues-Neves AC, Baptista FI, Carecho R, Moreira PI, CasteloBranco M, Ambrosio AF. The Retina as a Window or Mirror of the Brain Changes Detected in Alzheimer's Disease: Critical Aspects to Unravel. *Mol Neurobiol* 2019; 56, 5416–5435 <https://doi.org/10.1007/s12035-018-1461-6> PMID: 30612332
36. Hinton DR, Sadun AA, Blanks JC, Miller CA. Optic nerve degeneration in Alzheimer's disease. *New Engl J Med* 1986; 15:485–487.

37. Leuba G, Saini K. Pathology of subcortical visual centers in relation to cortical degeneration in Alzheimer's disease. *Neuropathol Appl Neurobiol* 1995; 21:410–422 <https://doi.org/10.1111/j.1365-2990.1995.tb01078.x> PMID: 8632836
38. Berisha F, Feke GT, Trempe CL, McMeel JW, Schepens CL. Retinal abnormalities in early Alzheimer's disease. *Invest Ophthalmol Vis Sci* 2007; 48:2285–2289 <https://doi.org/10.1167/iovs.06-1029> PMID: 17460292
39. Maffei L and Fiorentini L. Electroretinographic responses to alternating gratings before and after section of the optic nerve. *Science*. 1981; 211(4485):953–955. <https://doi.org/10.1126/science.7466369> PMID: 7466369
40. Zrenner E. The physiological basis of the pattern electroretinogram. *Progress in Retinal Research*. 1990; 427–464
41. Holder G.E., Gale R.P., Acheson J.F., Robson A.G. Electrodiagnostic assessment in optic nerve disease. *Curr. Opin. Neurol* 2009; 22, 3–10. <https://doi.org/10.1097/WCO.0b013e328320264c> PMID: 19155758
42. Krasodomska K., Lubiński W., Potemkowski A., Honczarenko K. Pattern electroretinogram (PERG) and pattern visual evoked potential (PVEP) in the early stages of Alzheimer's disease. *Doc. Ophthalmol. Adv. Ophthalmol.* 2010; 121, 111–121. <http://dx.doi.org/10.1007/s10633-010-9238-x>.
43. Katz B, Rimmer S, Iragui V, Katzman R. Abnormal pattern electroretinogram in Alzheimer's disease: evidence for retinal ganglion cell degeneration? *Ann Neurol* 1989; 26(2):221–225. <https://doi.org/10.1002/ana.410260207> PMID: 2774509
44. Trick GL, Barris MC, Bickler-Bluth M. Abnormal pattern electroretinograms in patients with senile dementia of the Alzheimer type. *Ann Neurol* 1989; 26(2):226–231. <https://doi.org/10.1002/ana.410260208> PMID: 2774510
45. Garcia-Martin E., Rodriguez-Mena D., Satue M., Almarcegui C., Dolz I., Alarcia R., Seral M., Polo V., Larrosa J.M., Pablo L.E., Electrophysiology and optical coherence tomography to evaluate Parkinson disease severity. *Invest. Ophthalmol. Vis. Sci.* 2014; 55, 696–705. <https://doi.org/10.1167/iovs.13-13062> PMID: 24425856
46. Porciatti V, Ventura LM. Physiologic significance of steady-state pattern electroretinogram losses in glaucoma: clues from simulation of abnormalities in normal subjects. *J Glaucoma*. 2009; 18(7): 535–542. <https://doi.org/10.1097/IJG.0b013e318193c2e1> PMID: 19745668
47. Della Santina L, Inman DM, Lupien CB, et al. Differential progression of structural and functional alterations in distinct retinal ganglion cell types in a mouse model of glaucoma. *J Neurosci* 2013; 33(44), 17444–57. <https://doi.org/10.1523/JNEUROSCI.5461-12.2013> PMID: 24174678
48. Jacobs TC, Libby RT, Ben Y, et al. Retinal ganglion cell degeneration is topological but not cell type specific in DBA/2J mice. *J Cell Biol* 2005; 17(2), 313–25.
49. Ventura LM, Sorokac N, Los Santos NR et al. The relationship between retinal ganglion cell function and retinal nerve fiber thickness in early glaucoma. *Investigative Ophthalmology and Visual Science*. 2006; 47(9):3904–3911. <https://doi.org/10.1167/iovs.06-0161> PMID: 16936103
50. Bach M and Hoffmann MB. Update on the pattern electroretinogram in glaucoma. *Optometry and Vision Science*. 2008; 85(6):386–395. <https://doi.org/10.1097/OPX.0b013e318177ebf3> PMID: 18521020
51. Pfeiffer N and Bach M. The pattern-electroretinogram in glaucoma and ocular hypertension. A cross-sectional and longitudinal study. *German Journal of Ophthalmology*, 1992; 1(1):35–40. PMID: 1477616
52. Oner A, Gumus K, Arda H, et al. Pattern electroretinographic recordings in eyes with myopia. *Eye Contact Lens* 2009; 35(5): 238–41 <https://doi.org/10.1097/ICL.0b013e3181b343d9> PMID: 19672200
53. Ventura LM, Porciatti V, Ishida K et al. Pattern electroretinogram abnormality and glaucoma. *Ophthalmology*. 2005; 112(1):10–19. <https://doi.org/10.1016/j.ophtha.2004.07.018> PMID: 15629814
54. Ventura LM, Golubev I, Feuer WJ, and Porciatti V. The PERG in diabetic glaucoma suspects with no evidence of retinopathy. *Journal of Glaucoma*. 2010; 19(4):243–247 <https://doi.org/10.1097/IJG.0b013e3181a990ea> PMID: 19528818
55. Mavilio A, Scrimieri F, Errico D. Can variability of pattern ERG signal help to detect retinal ganglion cells dysfunction in glaucomatous eyes? *Biomed Res Int* 2015; 2015:571314. <https://doi.org/10.1155/2015/571314> Epub 2015 Jun 8 PMID: 26167489
56. Mavilio A, Sisto D, Ferreri P, et al. RE-PERG, a new procedure for electrophysiologic diagnosis of glaucoma that may improve PERG specificity. *Clin Ophthalmol* 2017 Jan 23; 11:209–218. <https://doi.org/10.2147/OPHTH.S122706> PMID: 28176965
57. Mavilio A, Sisto D, Ferreri P, et al. RE-PERG, a new paradigm for glaucoma diagnosis, in myopic eyes. *Clin Ophthalmol*. 2019; 13:1315–22 <https://doi.org/10.2147/OPHTH.S211337>

58. Ventura LM, Golubev I, Feuer WJ, and Porciatti V. The PERG in diabetic glaucoma suspects with no evidence of retinopathy. *Journal of Glaucoma*.2010; 19(4):243–247 <https://doi.org/10.1097/IJG.0b013e3181a990ea> PMID: 19528818
59. Terminology and guidelines for glaucoma (3rd edition) [http://www.eugs.org/eng/EGS\\_guidelines.asp](http://www.eugs.org/eng/EGS_guidelines.asp)
60. Cockrell J. R., & Folstein M. F. (2002). Mini-mental state examination. *Principles and practice of geriatric psychiatry*, 140–141.
61. Benson A. D. et al. Screening for Early Alzheimer's Disease: Is There Still a Role for the Mini-Mental State Examination? *Prim Care Companion J Clin Psychiatry* 7, 62–69 (2005)
62. Mungas D. In-office mental status testing: a practical guide. *Geriatrics*.1991; 46, 54–58;
63. Albert MS, DeKosky ST, Dickson D, et al. The diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers. Dement.* 2011; 7(3):270–279. <https://doi.org/10.1016/j.jalz.2011.03.008> PMID: 21514249 NIA-AA workgroup diagnostic and research biomarker guidelines for MCI.
64. Heijl A, Lindgren G, Olsson J. The effect of perimetric experience in normal subjects. *Arch Ophthalmol* 1989; 107(1): 81–6 <https://doi.org/10.1001/archophth.1989.01070010083032> PMID: 2642703
65. Porciatti V, Ventura LM. Normative data for a user-friendly paradigm for pattern electroretinogram recording. *Ophthalmology* 2004; 111(1): 161–8 <https://doi.org/10.1016/j.ophtha.2003.04.007> PMID: 14711729
66. Porciatti V, Falsini B, Scalia G, et al. The pattern electroretinogram by skin electrodes: effect of spatial frequency and age. *Doc Ophthalmol* 1988; 70(1): 117–22. <https://doi.org/10.1007/BF00154742> PMID: 3229289
67. Falsini B, Marangoni D, Salgarello T, et al. Structure-function relationship in ocular hypertension and glaucoma: interindividual and interocular analysis by OCT an pattern ERG. *Graefes Arch Clin Exp Ophthalmol* 2008; 246(8): 1153–62 <https://doi.org/10.1007/s00417-008-0808-5> PMID: 18386035
68. Porciatti V, Sorokoc N, Buchser W. Habituation of retinal ganglion cell activity in response to steady state pattern visual stimuli in normal subjects. *Invest Ophthalmol Vis Sci.* 2005; 46: 1296–1302
69. Bunce C, Patel KV, Xing W, Freemantle N, Dore´ CJ (2014) Ophthalmic statistics note 1: unit of analysis. *Br J Ophthalmol* 98(3):408–412 <https://doi.org/10.1136/bjophthalmol-2013-304587> PMID: 24357496
70. Jeste DV, Palmer BW, Appelbaum PS, et al. A New Brief Instrument for Assessing Decisional Capacity for Clinical Research. *Arch Gen Psych* 2007; 64(8): 966–74
71. Gorelick Philip B., et al. "Vascular contributions to cognitive impairment and dementia: a statement for healthcare professionals from the American Heart Association/American Stroke Association." *Stroke* 42.9 (2011): 2672–2713 <https://doi.org/10.1161/STR.0b013e3182299496> PMID: 21778438
72. Gorelick PB, Scuteri A, Black SE, et al. Vascular contributions to cognitive impairment and dementia: a statement for healthcare professionals from the American heart association/America stroke association. *Stroke* 2011; 42: 2672–713 <https://doi.org/10.1161/STR.0b013e3182299496> PMID: 21778438
73. Holroyd S, Shepherd ML. Alzheimer's disease: a review for the ophthalmologist. *Surv Ophthalmol* 2001; 45:516–524. [https://doi.org/10.1016/s0039-6257\(01\)00193-x](https://doi.org/10.1016/s0039-6257(01)00193-x) PMID: 11425357
74. Lewis DA, Campbell MJ, Terry RD, Morrison JH. Laminar and regional distributions of neurofibrillary tangles and neuritic plaques in Alzheimer's disease. A quantitative study of visual and auditory cortices. *J Neurosci* 1987; 7: 1799–1808. <https://doi.org/10.1523/JNEUROSCI.07-06-01799.1987> PMID: 2439665
75. Parisi V, Restuccia R, Fattapposta F, et al. Morphological and functional retinal impairment in Alzheimer's disease patients. *Clin Neurophysiol.* 2001 Oct; 112(10):1860–7 [https://doi.org/10.1016/s1388-2457\(01\)00620-4](https://doi.org/10.1016/s1388-2457(01)00620-4) PMID: 11595144
76. Krasodomska K, Lubinski W, Potemowski A. et al. Pattern electroretinogram (PERG) and pattern visual evoked potential (PVEP) in the early stages of Alzheimer's disease *Doc Ophthalmol.* 2010 Oct; 121(2): 111–121. <https://doi.org/10.1007/s10633-010-9238-x> PMID: 20549299
77. Criscuolo C, Cerri E, Fabiani C et al. The retina as a window to early dysfunctions of Alzheimer's disease following studies with a 5xFAD mouse model *Neurobiology of Aging*, 2018; vol. 67, pp. 181–188. <https://doi.org/10.1016/j.neurobiolaging.2018.03.017> PMID: 29735432
78. Di Prospero NA, Chen E-Y, Charles V, Plomann M, Kordower JH, Tagle DA. Early changes in Huntington's disease patient brains involve alterations in cytoskeletal and synaptic elements. *J Neurocytol* 2004; 33: 517–33 <https://doi.org/10.1007/s11068-004-0514-8> PMID: 15906159
79. Ames A III. CNS energy metabolism as related to function. *Brain Res Brain Res Rev.* 2000; 34:42–68. [https://doi.org/10.1016/s0165-0173\(00\)00038-2](https://doi.org/10.1016/s0165-0173(00)00038-2) PMID: 11086186

80. Freitas S, Simoes MR, Alves L, et al. Montreal Cognitive Assessment (MoCA): validation study for vascular dementia. *J Int Neuropsychol Soc* 2012; 18: 1031–40 <https://doi.org/10.1017/S135561771200077X> PMID: 22676901
81. Santos CY, Johnson LN, Sinoff SE, et al. Change in retinal structure anatomy during the preclinical stage of Alzheimer's disease. *Alzheimer Dement (Amst)* 2018; 10: 196–209
82. Lopez-de-Eguileta A, Lage C, et al. Ganglion cell layer thinning in prodromal Alzheimer's disease defined by amyloid-PET. *Alzheimer Dement (NY)* 2019; 5: 570–8
83. Bayer AU, Keller ON, Ferrari F, et al. Association of glaucoma with neurodegenerative diseases with apoptotic cell death: Alzheimer's disease and parkinson's disease. *Am J Ophthalmol* 2002; 133(1): 135–7. [https://doi.org/10.1016/s0002-9394\(01\)01196-5](https://doi.org/10.1016/s0002-9394(01)01196-5) PMID: 11755850
84. Bayer AU, Ferrari F, Erb C. High occurrence rate of glaucoma among patients with Alzheimer's disease. *Eur Neurol* 2002; 47(3): 165–8 <https://doi.org/10.1159/000047976> PMID: 11914555
85. Guo L, Salt TE, Luong V, et al. Targeting amyloid- $\beta$  in glaucoma treatment. *Proc Natl Acad Sci USA* 2007; 104(33): 13444–9 <https://doi.org/10.1073/pnas.0703707104> PMID: 17684098
86. Gasparini L, Crowther RA, Martin KR, et al. Tau inclusions in retinal ganglion cells of human P301S tau transgenic mice: effects on axonal viability. *Neurobiol Aging* 2011; 32(3): 419–33
87. Salobarra-Garcia E, de Hoz R, Ramirez AI, et al. Changes in visual function and retinal structure in the progression of Alzheimer's disease. *PloS One* 2019; 14(8):e0220535 <https://doi.org/10.1371/journal.pone.0220535> PMID: 31415594
88. Mutku U, Colijn JM, Ikram MA, et al. Association of retinal neurodegeneration on optical coherence tomography with dementia. *JAMA Neurol* 2018; 75(10): 1256–63 <https://doi.org/10.1001/jamaneurol.2018.1563> PMID: 29946702
89. Trick GL, Trick LR, Morris P, et al. Visual field loss in senile dementia of the Alzheimer's type. *Neurology* 1995; 54(1) 68–74
90. Lu Y, Li Z, Zhang X, Ming B, Jia J, Wang R, Ma D. Retinal nerve fiber layer structure abnormalities in early Alzheimer's disease: evidence in optical coherence tomography. *Neurosci Lett*. 2010; 480:69–72. <https://doi.org/10.1016/j.neulet.2010.06.006> PMID: 20609426
91. Danesh-Meyer H, Birch H, Ku JY, Carroll S, Gamble G. Reduction of optic nerve fibers in patients with Alzheimer disease identified by laser imaging. *Neurology*. 2006; 67:1852–4 <https://doi.org/10.1212/01.wnl.0000244490.07925.8b> PMID: 17130422
92. Berisha F, Fekete GT, Trempe CL, McMeel JW, Schepens CL. Retinal abnormalities in early Alzheimer's disease. *Invest Ophthalmol Vis Sci*. 2007; 48:2285–2289. <https://doi.org/10.1167/iovs.06-1029> PMID: 17460292
93. Kergoat H, Kergoat MJ, Justino L, Chertkow H, Robillard A, Bergman H. An evaluation of the retinal nerve fiber layer thickness by scanning laser polarimetry in individuals with dementia of the Alzheimer type. *Acta Ophthalmol Scand*. 2001; 79:187–91. <https://doi.org/10.1034/j.1600-0420.2001.079002187.x> PMID: 11284761
94. Kurna SA, Akar G, Altun A, Agirman Y, Gozke E, Sengor T. Confocal scanning laser tomography of the optic nerve head on the patients with Alzheimer's disease compared to glaucoma and control. *Int Ophthalmol*. 2014; 34:1203–11 <https://doi.org/10.1007/s10792-014-0004-z> PMID: 25284015
95. Bulut M, Kurtulu? F, G?zkaya O, et al. Evaluation of optical coherence tomography angiographic findings in Alzheimer's type dementia. *Br J Ophthalmol* 2018; 102: 233–237
96. Zabel P, Kaluzny JJ, Wilcosc-Debczynska M, et al. Comparison of retinal microvasculature in patients with Alzheimer's disease and primary open-angle glaucoma by optical coherence tomography angiography. *Invest Ophthalmol Vis Sci* 2019; 60: 3447–3455 <https://doi.org/10.1167/iovs.19-27028> PMID: 31408108
97. Zivkovic M, Dayanir V, Kocaturk T, et al. Foveal avascular zone in normal tension glaucoma measured by optical coherence tomography angiography. *Biomed Res Int* 2017; 2017:3079141 <https://doi.org/10.1155/2017/3079141> PMID: 29392131
98. Lee CY, Liu CH, Chen HC, et al. Correlation between basal macular circulation and following glaucomatous damage in progressed high-tension and normal-tension glaucoma. *Ophthalmic Res* 2019; 62(1): 46–54 <https://doi.org/10.1159/000499695> PMID: 31104053
99. Kwon J, Choi J, Shin JVV, et al. Alterations of the foveal avascular zone measured by optical coherence tomography angiography in glaucoma patients with central visual field defects. *Invest ophthalmol Vis Sci* 2017; 58: 1637–45. <https://doi.org/10.1167/iovs.16-21079> PMID: 28297029
100. Ch'ng TW, Gillmann K, Hoskens K, et al. Effect of surgical intraocular pressure lowering on retinal structures—nerve fibre layer, foveal avascular zone, peripapillary and macular vessel density: 1 year results. *Eye (Lond)* 2019 <https://doi.org/10.1038/s41433-019-0560-6> PMID: 31409906

101. Hof PR, Morrison JH. Quantitative analysis of a vulnerable subset of pyramidal neurons in Alzheimer's disease: II. Primary and secondary visual cortex. *J Comp Neurol*. 1990; 301:55–64. <https://doi.org/10.1002/cne.903010106> PMID: [1706358](https://pubmed.ncbi.nlm.nih.gov/1706358/)