



# Draft Genome Sequence of *Stenotrophomonas maltophilia* MDMC339, Isolated from Soil of Merzouga Desert in Morocco

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**ABSTRACT** Here, we report the draft genome sequence of *Stenotrophomonas maltophilia* MDMC339, a strain able to survive in the difficult conditions imposed by the Merzouga desert. The analyzed genome contains 4,788,525 bp with 4,262 genes coding for proteins, including several genes related to stress.

Merzouga is a small Saharan village in southeastern Morocco known for its arid climate and low rainfall. The region is characterized by a very hot climate with significant variations in temperatures between day and night. In such situations, living organisms have to cope with extreme temperatures, low humidity, and low nutrient availability. However, some bacteria implement several strategies to counter stress (1–3).

*Stenotrophomonas maltophilia* is a ubiquitous, Gram-negative, aerobic, rod-shaped, and mobile bacterium with some polar flagella (4, 5). It is also an opportunistic pathogen that has low sensitivity to many antibiotics (4, 6). Nevertheless, several strains have been described as producing plant growth-promoting substances or enzymes involved in bioremediation, thus bringing out its environmental benefit (7).

In this study, we report the draft genome sequence of *Stenotrophomonas maltophilia* MDMC339, isolated from nonvegetated dunes of the Merzouga region.

The sample was collected (at the coordinates –3.97852083325, 31.1093333325) in three replicates.

Bacterial isolation was done using 1 g of sand, which was suspended in 9 ml of physiological water and then shaken vigorously for 30 min. Serial dilutions (from 10<sup>–3</sup> to 10<sup>–7</sup> ml/ml) were made. Then, 100 μl of each dilution was spread on the surface of a petri dish containing nutrient agar medium supplemented with 50 μg/ml of amphotericin B. The dishes were then incubated at 28°C for 2 days. Once purified, the isolate was seeded on liquid nutrient broth and incubated overnight. Subsequently, the isolate was designated MDMC339 (Moroccan Desert Microorganisms Collection) and stored in 40% (vol/vol) glycerol solution at –80°C.

From the frozen stock, the culture process was carried out in 5 ml of nutrient broth at 28°C for 48 h. After centrifugation, the pellet was used for DNA extraction using the phenol-chloroform method as described by Chen and Kuo (8) and quantified using the Quantus fluorometer (Promega). Isolated bacterial genomic DNA was used both for bacterial identification using the 16S rRNA gene and for the whole-genome sequencing (WGS).

MDMC339 was identified as a *Stenotrophomonas maltophilia* isolate after 16S rRNA gene amplification using the universal primers FD1 (5'-CCGAATTCGTCGACAACAGAGT TTGATCCTGGCTCAG-3') and RS16 (5'-TACGGCTACCTTGTACGACTT-3') followed by Sanger sequencing of amplicons (Genoscreen, Lille, France).

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Using BLAST (9), the obtained 16S rRNA gene sequences showed more than 99% similarity to the 16S rRNA gene sequences of other *S. maltophilia* strains. More information about the identity of the bacterium was revealed after the whole-genome sequencing.

The sequencing library was prepared using the Nextera XT DNA sample preparation kit (Illumina, Inc., San Diego, CA, USA). Sequencing was done in paired-end mode with  $2 \times 300$ -bp reads via the MiSeq sequencer using the MiSeq reagent (600-cycle) kit version 3 (Illumina).

A total of 4,005,852 raw reads were generated and then assembled directly with no filtering according to quality using SPAdes version 3.11.1 (10) with k values of 21, 33, 55, 77, 99, and 127. The quality of the assembly was evaluated using QUAST version 5.0.0 (11), and genome annotation was accomplished using Prokka version 1.12 (12) and the RAST server (13).

Genome assembly yielded 185 contigs with a total length of 4,788,525 bp, an  $N_{50}$  value of 57,069 bp, and a G+C content of 66.14%. The average depth of coverage was 209.0 $\times$ . The annotation of the contigs revealed that the strain possessed 4,262 genes encoding proteins and 76 RNA genes (70 tRNAs and 6 rRNAs). From a total of 4,262 identified proteins, 2,967 were classified as nonhypothetical proteins, while 1,929 sequences were assigned to SEED subsystems.

Using the JSpeciesWS online service (14), average nucleotide identity (ANI) values for the draft genome sequence of *S. maltophilia* MDMC339 compared to other *Stenotrophomonas* spp. were determined. The highest ANI value was 93.52% with *S. maltophilia* strain K279a (GenBank accession number [NC\\_010943](https://ncbi.nlm.nih.gov/nuccore/NC_010943)).

All software used during this study was run with default parameters.

The genome of *Stenotrophomonas maltophilia* MDMC339 contains genes that code for proteins induced by heat shock, notably *dnaK*, *dnaJ*, and *grpE*, and transcription repressor *hrcA* (15). In addition to heat shock proteins, the genome also contains genes involved in the response to cold shock (*cspA* and *cspD*) (16).

Genome analysis also reveals the presence of the *betA* and *betB* genes involved in the enzymatic conversion of choline and glycine betaine aldehyde to glycine betaine. The import of choline is done using high-affinity choline uptake gene *betT*, the sequence of which is present in four copies (17). Finally, the genome encodes proteins of the type VI secretion system, which is known for its role in interbacterial competition (18) and which could be useful in competition for environmental resources.

**Data availability.** This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number [RXLZ00000000](https://ncbi.nlm.nih.gov/nuccore/RXLZ00000000). The version described in this paper is the first version, [RXLZ01000000](https://ncbi.nlm.nih.gov/nuccore/RXLZ01000000). The raw sequencing data are available from the Sequence Read Archive (SRA) under the accession number [SRR11886254](https://ncbi.nlm.nih.gov/sra/SRR11886254). The bacterial 16S rRNA marker gene sequenced during this study is available at the NCBI GenBank database under accession number [KX013440](https://ncbi.nlm.nih.gov/nuccore/KX013440).

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## REFERENCES

1. Aanniz T, Ouadghiri M, Melloul M, Swings J, Elfahime E, Ibjibijen J, Ismaili M, Amar M. 2015. Thermophilic bacteria in Moroccan hot springs, salt marshes and desert soils. *Braz J Microbiol* 46:443–453. <https://doi.org/10.1590/S1517-838246220140219>.
2. Gommeaux M, Barakat M, Montagnac G, Christen R, Guyot F, Heulin T. 2010. Mineral and bacterial diversities of desert sand grains from south-east Morocco. *Geomicrobiol J* 27:76–92. <https://doi.org/10.1080/01490450903393066>.
3. Bär M, von Hardenberg J, Meron E, Provenzale A. 2002. Modelling the survival of bacteria in drylands: the advantage of being dormant. *Proc Biol Sci* 269:937–942. <https://doi.org/10.1098/rspb.2002.1958>.
4. Brooke JS. 2012. *Stenotrophomonas maltophilia*: an emerging global opportunistic pathogen. *Clin Microbiol Rev* 25:2–41. <https://doi.org/10.1128/CMR.00019-11>.
5. Carmody LA, Spilker T, LiPuma JJ. 2011. Reassessment of *Stenotrophomonas maltophilia* phenotype. *J Clin Microbiol* 49:1101–1103. <https://doi.org/10.1128/JCM.02204-10>.
6. Sánchez MB. 2015. Antibiotic resistance in the opportunistic pathogen

- Stenotrophomonas maltophilia*. *Front Microbiol* 6:658–658. <https://doi.org/10.3389/fmicb.2015.00658>.
7. Mukherjee P, Roy P. 2016. Genomic potential of *Stenotrophomonas maltophilia* in bioremediation with an assessment of its multifaceted role in our environment. *Front Microbiol* 7:967–967. <https://doi.org/10.3389/fmicb.2016.00967>.
  8. Chen WP, Kuo TT. 1993. A simple and rapid method for the preparation of Gram-negative bacterial genomic DNA. *Nucleic Acids Res* 21:2260–2260. <https://doi.org/10.1093/nar/21.9.2260>.
  9. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
  10. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
  11. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
  12. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
  13. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75–75. <https://doi.org/10.1186/1471-2164-9-75>.
  14. Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>.
  15. Roncarati D, Scarlato V. 2017. Regulation of heat-shock genes in bacteria: from signal sensing to gene expression output. *FEMS Microbiol Rev* 41:549–574. <https://doi.org/10.1093/femsre/fux015>.
  16. Keto-Timonen R, Hietala N, Palonen E, Hakakorpi A, Lindström M, Korkeala H. 2016. Cold shock proteins: a minireview with special emphasis on Csp-family of enteropathogenic *Yersinia*. *Front Microbiol* 7:1151. <https://doi.org/10.3389/fmicb.2016.01151>.
  17. Mandon K, Østerås M, Boncompagni E, Trinchant JC, Spennato G, Poggi MC, Le Rudulier D. 2003. The *Sinorhizobium meliloti* glycine betaine biosynthetic genes (*betICBA*) are induced by choline and highly expressed in bacteroids. *Mol Plant Microbe Interact* 16:709–719. <https://doi.org/10.1094/MPMI.2003.16.8.709>.
  18. Coulthurst S. 2019. The type VI secretion system: a versatile bacterial weapon. *Microbiology* 165:503–515. <https://doi.org/10.1099/mic.0.000789>.