

RESEARCH

Open Access



The genetic diversity and geographical separation study of *Oncomelania hupensis* populations in mainland China using microsatellite loci

Wei Guan^{1,2}, Shi-Zhu Li^{1*}, Eniola Michael Abe³, Bonnie L. Webster^{4*}, David Rollinson⁴ and Xiao-Nong Zhou^{1,2}

Abstract

Background: *Oncomelania hupensis* is the unique intermediate host of *Schistosoma japonicum*, which plays a crucial role in the transmission of schistosomiasis. The endemic area of *S. japonicum* is strictly consistent with the geographical distribution of *O. hupensis*.

Methods: A total of 24 populations of *O. hupensis* from four ecological landscapes were selected for analysis of genetic diversity by screening eight microsatellite DNA polymorphic loci.

Results: The number of alleles per locus ranged from 29 to 70 with an average of 45.625 and that of effective alleles were 18.5 to 45.8 with an average of 27.4. The observed (H_o) and expected (H_e) heterozygosities varied from 0.331 to 0.57 and from 0.888 to 0.974, respectively. The mean of polymorphism information content (PIC) for all populations was 0.940, appearing polymorphic for all loci. For the fixation index of F -Statistics, Fit and Fst were 54.95 and 37.62 %, respectively. Variation of *O. hupensis* chiefly exists among individuals, accounting for 60.58 % of the total variation determined by Analysis of Molecular Variation (AMOVA). Variation among individuals within populations, among populations within groups and among groups only accounted for 26.60, 8.04 and 4.78 %, respectively. This distribution of variation suggests that genetic differences principally originate from within-populations rather than among-populations. Moreover, UPGMA cluster analysis showed that the populations spreading within middle and lower reaches of the Yangtze River (HBWH, JSYZ, JXNC, HNHS, JXJJ, AHWW, HBJL, JXDC, HNNX, JSYZJZ, ZJJH, AHNG and AHWJ) clustered together first, then gathered with the populations in the high mountains (SCMS, SCYA, SCPJ, YNEY, SCLS, YNWS and SCXC), coastal hills (FJFQ and FJFZ) and Karst landform (GXBS and GXYZ) successively.

Conclusion: This study provides novel insight into the theoretical source of genetic differentiation of *Oncomelania hupensis* in mainland China, which is critical for the epidemiological investigation and surveillance of *S. japonicum*.

Keywords: *Oncomelania hupensis*, *Schistosoma japonicum*, Microsatellites DNA, Polymorphism, Genetic differentiation

* Correspondence: Lisz@chinacdc.cn; bonw@nhm.ac.uk

¹National Institute of Parasitic Diseases, Chinese Center for Disease Control and Prevention, Shanghai 200025, People's Republic of China

⁴Wolfson Wellcome Biomedical Laboratories, Department of Zoology, Natural History Museum, Cromwell Road, London SW7 5BD, UK

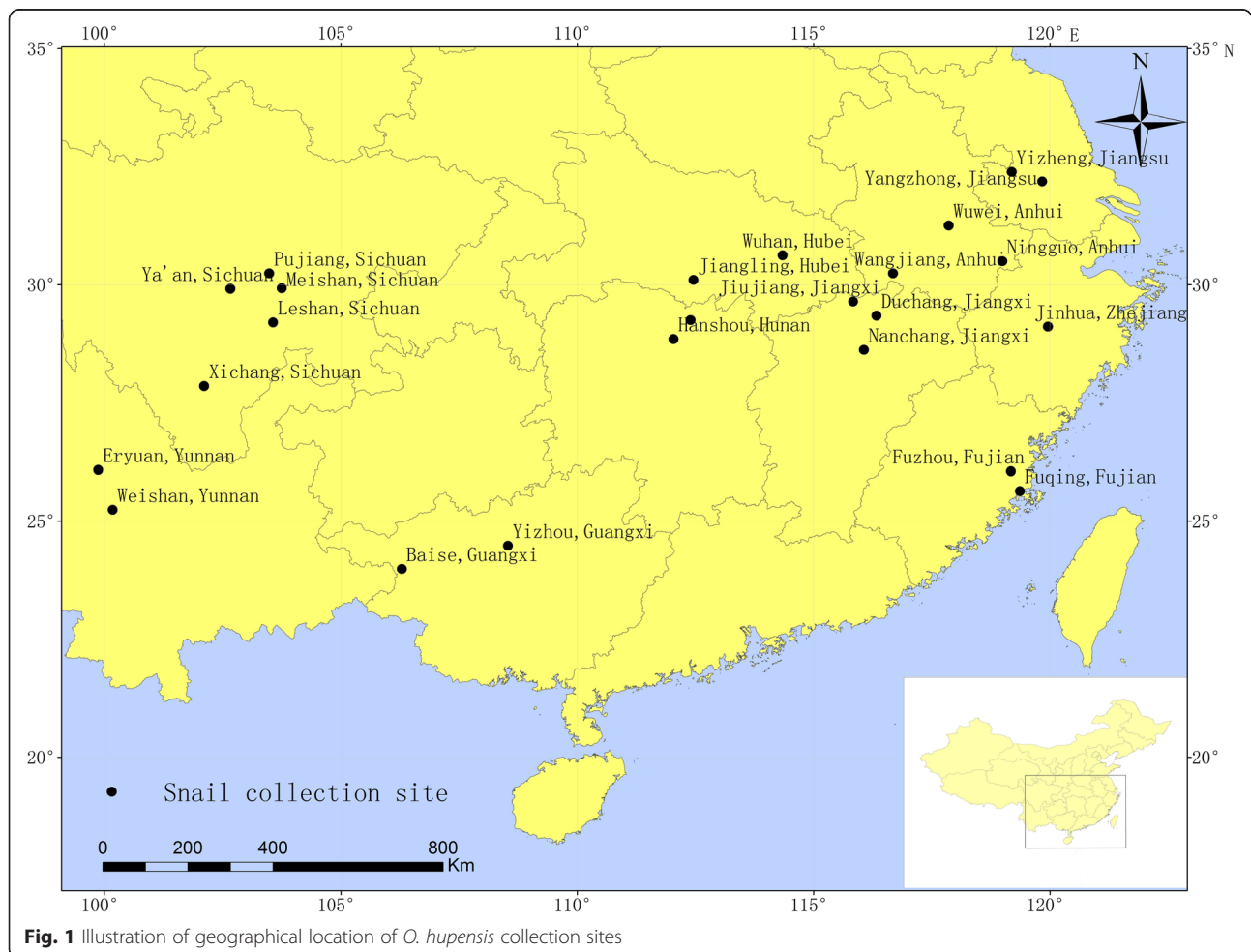
Full list of author information is available at the end of the article

Background

Schistosomiasis, caused by *Schistosoma japonicum*, remains one of the most prevalent parasitic diseases and effects severe socio-economic and public health losses in China [1, 2]. *Oncomelania hupensis* is the unique intermediate host of *S. japonicum*, which plays a critical role in the transmission of Schistosomiasis japonica [1, 3]. The geographical distribution of *O. hupensis* coincides with the endemic area of *S. japonicum* [4], which is mainly found throughout the southern region of the Yangtze River basin [5, 6]. As a result, significant genetic differentiation leads to the formation of multiple geographical populations of *O. hupensis* [3]. Coincident with the endemic area for schistosomiasis, *O. hupensis* has been mainly found in four types of ecological landscapes giving rise to subspecies including: (1) *O. h. hupensis* largely in the middle and lower reaches of the Yangtze River (among the provinces of Hunan, Hubei, Jiangxi, Anhui, Jiangsu and Zhejiang) (2) *O. h. robertsoni* in the mountainous region of Sichuan and Yunnan provinces (3) *O. h. guangxiensis* in the Karst landscape of Guangxi province and (4) *O. h. tangi* in the

southeastern coastal region of Fujian province [7, 8]. Interestingly, obvious morphological differences have been identified among individuals from the same regional population [9–11]. For example, *O. hupensis* from upstream of Miaohe basin, which contains regions of swamps and lakes, have a ribbed shell while those from downstream have a smooth shell [12].

Microsatellite DNA, known as short tandem repeat (STR) or simple sequence repeat (SSR), occurs throughout the eukaryotic genome. Differences in repetitive sequence numbers allow for high polymorphism due to the ubiquitous occurrence, high copy numbers, high heterozygosity and easy detection within population [13]. Along with other genome mark technology, it has been widely applied to research examining genetic diversity and serves as an important molecular marker [14–17]. At present, microsatellites have been isolated from many different organisms [18–20]. Specifically, from 128 molluscs, a total of 3,284 microsatellite sequences have been identified [21]. Although the microsatellite DNA library of *O. hupensis* was built recently [22], the microsatellite markers have



not been used extensively in population genetic structure studies and genome mapping of *O. hupensis* in P.R. China [23–25]. To deepen our knowledge on the genetic diversity of the intermediate host snail, we developed a novel multiplex PCR method to screen and analyze the genetic diversity of *O. hupensis* using microsatellites loci among the four various ecological landscape populations in mainland China.

Methods

Snail sampling

A total of 24 populations of *O. hupensis* were sampled from four ecological landscape populations in mainland China covering: (1) the region of swamps and lakes in the middle and lower reaches of the Yangtze River, (2) the mountainous region of the Sichuan and Yunnan provinces, (3) the littoral hill part of the Fujian province and (4) the karst landscape of Guangxi autonomous region (Fig. 1, Table 1).

DNA preparation

Ten to 20 *O. hupensis* samples were randomly chosen from each site, fed for 1 week and identified as infected

or non-infected with *S. japonicum* by observation of cercariae emerging from the snails. Only non-infected snails were used in this study. After removal of the gut and digestive glands from the soft parts of the snails, the 30 mg muscle tissues from the pleopod of a single snail were digested for 3 hours at 56 °C with proteinase K (Amresco Inc. Solon, OH, USA) followed by the standard DNA extraction procedure [26] using mollusc DNA Kit (Omega, USA).

PCR amplification and detection of PCR products

The microsatellite DNA polymorphic loci were selected and evaluated from previous microsatellite loci library [22]. Two rounds of multiplex PCR reaction were developed including four microsatellite loci in each one, which were identified by different lengths and fluorescence peaks of 6-FAM, VIC, NED and PET labeled by (Sigma-aldrich London, UK). Primer sequences and information are summarized in Table 2.

The multiplex PCRs were developed using the Type-it Microsatellite PCR Kit (QiaGen, London, UK) with a 25 µl reaction system, including 2x Type-it Multiplex

Table 1 Location of *O. hupensis* collection

Collection site(Code)	Geomorphic feature	No. samples	Collection date	Longitude	Latitude
Ningguo, Anhui(AHNG)	swamps and lakes	17	09/12/2012	30.5022° N	118.9891° E
Wangjiang, Anhuui(AHWJ)	swamps and lakes	20	09/12/2012	30.2423° N	116.2814° E
Wuwei, Anhui(AHWW)	swamps and lakes	18	09/12/2012	31.2571° N	117.8573° E
Jiangling, Hubei(HBJL)	swamps and lakes	18	06/14/2013	31.1034° N	112.4631° E
Wuhan, Hubei(HBWH)	swamps and lakes	17	05/11/2012	30.6749° N	114.3865° E
Hanshou, Hunan(HNHS)	swamps and lakes	16	03/18/2013	28.8592° N	112.0378° E
Nanxian, Hunan(HNNX)	swamps and lakes	11	03/18/2013	29.2581° N	112.3972° E
Yizheng, Jiangsu(JSYZ)	swamps and lakes	19	04/21/2013	32.3911° N	119.1914° E
Yangzhong, Jiangsu(JSYZ)	swamps and lakes	18	04/21/2013	32.1942° N	119.8353° E
Duchang, Jiangxi(JXDC)	swamps and lakes	19	04/14/2012	29.3562° N	116.3324° E
Jiujiang, Jiangxi(JXJJ)	swamps and lakes	15	04/14/2012	29.6517° N	115.8356 °E
Nanchang, Jiangxi(JXNC)	swamps and lakes	14	04/14/2012	28.6252° N	116.0642°E
Jinhua, Zhejiang(ZJH)	swamps and lakes	16	06/23/2012	29.1044° N	120.0052° E
Yaan, Sichuan(SCYA)	Mountains	17	09/25/2012	29.8931° N	102.6651° E
Leshan, Sichuan(SCLS)	Mountains	16	09/25/2012	29.1722° N	103.5759° E
Meishan, Sichuan(SCMS)	Mountains	19	09/25/2012	29.8788° N	104.0949° E
Xichang, Sichuan(SCXC)	Mountains	20	09/27/2012	27.8632° N	102.1134° E
Pujiang, Sichuan(SCPJ)	Mountains	15	09/27/2012	30.2412° N	103.4897° E
Eryuan, Yunnan(YNEY)	Mountains	15	03/21/2013	26.0852° N	112.0371° E
Weishan, Yunnan(YNWS)	Mountains	12	03/21/2013	31.2573° N	117.8574° E
Baise, Guangxi(GXBS)	Karst	9	03/22/2013	23.9829° N	106.1678° E
Yizhou, Guangxi(GXYZ)	Karst	18	03/22/2013	24.4792° N	108.5362° E
Fuqing, Fujian/ FJFQ)	Coastal hills	20	04/17/2012	25.6374° N	119.3652° E
Fuzhou, Fujian(FJFZ)	Coastal hills	17	04/17/2012	25.9911° N	119.1674° E

Table 2 Primers of the 8 microsatellite loci in *O. hupensis*

Locus	Primer sequence (5' → 3')	Repeat motif	Annealing temperature/(°C)	Allele size from field snails (bp)	NO. of multiplex PCR	GenBank accession No.
T1-10	Pf: TCACTCGGGTGAATGCT Pr: TTTGTTACTGATGGTGGC	(GA) ₃₈	55	173–259	1	GU204080
T4-25	Pf: CAATAGTTCGACTCGGAAGA Pr: CGAGGTATGGCGTTGCTT	(CT) ₃₅	52	142–228	1	GU204084
T4-22	Pf: TATCCAAGAAGCCGAAAC Pr: GAGGAAAGCGAGGTAAGA	(CA) ₁₀	50	224–256	1	GU204083
D11	Pf: TTCAGTTGTCTTATTCGTG Pr: TAGATGTTCACTGGTTTGTC	(TG) ₁₇	55	141–192	1	GU204223
T5-11	Pf: ACGCCAGTCTTGGTGCA Pr: TACTTGGGCAGAAGGGTT	(GT) ₁₄	55	153–210	2	GU204092
T6-17	Pf: GCTGTCCTTTTACCAACTGC Pr: TATCAAAGGATTATGCCGAG	(AC) ₈	55	192–248	2	GU204108
A18	Pf: GCCGATGATACAAGACCC Pr: GAGAATCTCCAGGCACGC	(CT) ₁₈	60	131–256	2	GU204047
C22	Pf: CGGTACATCTGGATAGTGG Pr: TGCGAAACAGTTGCAGACAC	(CA) ₂₁	62	185–239	2	GU204145

PCR Master Mix 12.5 µl, 10x primer mix 2 µl including four primers in each mix, template DNA 2 µl with less than 200 ng then add RNase-free water to 25 µl. The reaction conditions for PCR amplification were as follows: 95 °C, 5 min; 95 °C, 30 s, 60 °C, 60 s; 72 °C, 30 s, 30 cycles; 65 °C, 30 min for final extension. 1 µl of the PCR product was mixed with 0.6 µl of ROX and 8.4 µl ultrapure Hi-Di formamide, denatured at 95 °C for 5 min and detected using automatic genetic analyzer (3730XL, ABI, USA).

Analysis of microsatellite diversity

The accurate length of amplified fragments of microsatellite DNA loci were determined using Geneious software (Version 7.0.6) and subsequently exported as an Excel table. The raw data in the table were converted into a recognized format by Arlequin and Genepop using the toolkit of the Excel microsatellite toolkit. The data format which fits for Popgene were acquired by DataTrans 1.0. Various parameters of genetic difference within populations include: number of alleles (N_a), number of efficient alleles (N_e), inbreeding coefficient (F_{is}), expected heterozygosity (H_e) and observed heterozygosity (H_o) were calculated. The degree of Hardy-Weinberg equilibrium (HWE) and linkage disequilibrium (LD) were tested with Genepop 4.1.10. The frequency of null alleles within every population was calculated in Genepop. The index of genetic variation between populations (F_{st}), gene flow (N_m) and genetic distance [$F_{st}/(1-F_{st})$] were determined using Arlequin [27]. The correlation between genetic distance and geographical distance were tested with Mantel regression. Analysis of

molecular variance (AMOVA) was processed through Popgene software, clustering analysis was determined by unweighted pair group method with arithmetic means (UPGMA) and the phylogenetic tree was modified with TreeView [28]. The polymorphism information content (PIC) was calculated according to the formula previously described [28].

Results

Gene scan

From the 24 populations of *O. hupensis* sampled, 396 specimens were scanned at the genetic level across eight polymorphic loci of microsatellite DNA. The lengths of amplified fragments for a total of 6,196 microsatellite DNA loci were obtained.

Genetic differences within populations

Results obtained from the analysis of the 24 populations of *O. hupensis* showed that the number of alleles per locus ranged from 29 to 70 with an average of 45.625, and that of effective alleles were 18.5 to 45.8 with an average of 27.4. The GXYZ and HNHS populations had the minimum and maximum average N_a values, respectively. The average H_e within populations ranged from 0.888 to 0.974, and the average H_o ranged from 0.331 to 0.57. The populations with the highest and lowest H_o values were HNHS and GXYZ, respectively. The average PIC for all populations of *O. hupensis* was 0.940 (Tables 3, 4 and 5).

Significant deviation from Hardy-Weinberg equilibrium (HWE) was observed: 47 out of 192 (24.48 %)

Table 3 Coefficients of genetic diversity of *O. hupensis* at different loci (the populations of landscape of swamps and lakes)

Populations	Index	Microsatellite loci								Total
		T1-10	T4-25	D11	T4-22	T5-11	T6-27	A18	C22	
AHNG	<i>Na</i>	13	12	7	8	14	9	11	10	10.500
	<i>He</i>	0.863	0.815	0.806	0.774	0.927*	0.847*	0.929*	0.941*	0.863
	<i>Ho</i>	0.412	0.706	0.188	0.706	0.882	0.588	0.071	0.222	0.472
	<i>PIC</i>	0.948	0.938	0.913	0.902	0.927	0.932	0.948	0.949	0.932
AHWJ	<i>Na</i>	13	15	4	2	8	6	8	1	7.125
	<i>He</i>	0.918*	0.936	0.406	0.258	0.749	0.549	0.777	0.000	0.574
	<i>Ho</i>	0.588	0.471	0.000	0.059	0.765	0.133	0.200	0.104	0.317
	<i>PIC</i>	0.967	0.927	0.987	0.923	0.937	0.927	0.914	0.972	0.944
AHWW	<i>Na</i>	6	21	9	10	11	10	15	17	12.375
	<i>He</i>	0.810	0.963*	0.856	0.860	0.898	0.849	0.914*	0.936	0.886
	<i>Ho</i>	0.091	0.444	0.353	0.278	0.389	0.611	0.412	0.647	0.403
	<i>PIC</i>	0.943	0.923	0.938	0.912	0.924	0.972	0.916	0.976	0.937
HBJL	<i>Na</i>	12	19	15	10	12	14	13	13	13.500
	<i>He</i>	0.913*	0.961*	0.939	0.904	0.879	0.938*	0.895	0.930	0.920
	<i>Ho</i>	0.417	0.647	0.357	0.294	0.471	0.706	0.750	0.529	0.521
	<i>PIC</i>	0.947	0.933	0.937	0.890	0.927	0.928	0.968	0.972	0.939
HBWH	<i>Na</i>	12	19	16	12	15	13	19	18	15.500
	<i>He</i>	0.944	0.961*	0.956*	0.903	0.949*	0.924	0.966*	0.966*	0.946
	<i>Ho</i>	0.272	0.533	0.467	0.667	0.533	0.733	0.733	0.600	0.567
	<i>PIC</i>	0.991	0.896	0.922	0.917	0.958	0.921	0.970	0.927	0.938
HNHS	<i>Na</i>	16	21	15	16	17	8	20	18	16.375
	<i>He</i>	0.952*	0.974*	0.927	0.907*	0.952*	0.798	0.962*	0.956*	0.929
	<i>Ho</i>	0.250	0.750	0.438	0.813	0.733	0.750	0.500	0.688	0.615
	<i>PIC</i>	0.956	0.973	0.974	0.932	0.941	0.931	0.952	0.938	0.950
HNNX	<i>Na</i>	7	10	7	6	9	9	12	10	8.750
	<i>He</i>	0.801	0.913	0.853	0.844*	0.887	0.810	0.942*	0.892	0.868
	<i>Ho</i>	0.091	0.818	0.200	0.364	0.636	0.545	0.500	0.909	0.508
	<i>PIC</i>	0.936	0.976	0.926	0.927	0.956	0.912	0.951	0.936	0.941
JSYZ	<i>Na</i>	7	18	10	12	13	10	12	13	11.875
	<i>He</i>	0.909*	0.961*	0.806	0.924*	0.926	0.905*	0.915	0.937	0.910
	<i>Ho</i>	0.333	0.733	0.385	0.667	0.500	0.500	0.143	0.571	0.479
	<i>PIC</i>	0.897	0.918	0.973	0.899	0.973	0.948	0.940	0.918	0.933
JSYZJZ	<i>Na</i>	6	21	8	13	16	11	18	17	13.750
	<i>He</i>	0.817	0.954*	0.859	0.894	0.910	0.889*	0.943*	0.938	0.901
	<i>Ho</i>	0.111	0.722	0.412	0.611	0.500	0.611	0.500	0.611	0.510
	<i>PIC</i>	0.949	0.972	0.936	0.879	0.910	0.980	0.938	0.938	0.938
JXDC	<i>Na</i>	7	21	7	11	16	10	12	14	12.250
	<i>He</i>	0.890	0.968*	0.800	0.890	0.945*	0.761	0.908	0.922	0.886
	<i>Ho</i>	0.143	0.733	0.385	0.467	0.867	0.533	0.133	0.667	0.491
	<i>PIC</i>	0.982	0.936	0.926	0.919	0.928	0.979	0.914	0.935	0.943
JXJJ	<i>Na</i>	5	14	8	9	11	7	11	16	10.125
	<i>He</i>	0.803	0.957*	0.902	0.887	0.931	0.481	0.950*	0.957*	0.859
	<i>Ho</i>	0.167	0.545	0.667	0.636	0.727	0.455	0.500	0.818	0.564

Table 3 Coefficients of genetic diversity of *O. hupensis* at different loci (the populations of landscape of swamps and lakes) (Continued)

JXNC	PIC	0.968	0.973	0.927	0.898	0.918	0.977	0.927	0.963	0.947
	Na	6	17	9	8	9	7	7	12	9.375
	He	0.911*	0.993*	0.908*	0.869*	0.915	0.824	0.856	0.948	0.903
	Ho	0.200	0.889	0.500	0.333	0.444	0.778	0.111	0.667	0.490
ZJJH	PIC	0.953	0.911	0.890	0.915	0.937	0.967	0.917	0.967	0.932
	Na	3	14	1	7	16	0	12	6	7.375
	He	0.800	0.940*	0.000	0.764	0.948*	0.000	0.915	0.720	0.636
	Ho	0.000	0.625	-	0.563	0.813	-	0.500	0.438	0.490
	PIC	0.946	0.927	0.917	0.908	0.918	0.952	0.978	0.962	0.939

- Relevant data unavailable

*Statistically significant deviation from Hardy-Weinberg equilibrium ($P < 0.01$)**Table 4** Coefficients of genetic diversity of *O. hupensis* at different loci (the populations of landscape of mountains)

Populations	Index	Microsatellite loci								Total
		T1-10	T4-25	D11	T4-22	T5-11	T6-27	A18	C22	
SCLS	Na	13	12	7	8	14	9	11	10	10.500
	He	0.863	0.815	0.806	0.774	0.927	0.847	0.929	0.941*	0.863
	Ho	0.412	0.706	0.188	0.706	0.882	0.588	0.071	0.222	0.472
	PIC	0.948	0.927	0.971	0.909	0.929	0.972	0.927	0.938	0.945
SCMS	Na	15	15	12	10	16	10	21	14	14.125
	He	0.925*	0.924	0.892	0.863	0.941	0.865	0.964*	0.899	0.909
	Ho	0.563	0.700	0.474	0.263	0.850	0.350	0.550	0.650	0.550
	PIC	0.983	0.924	0.912	0.965	0.901	0.908	0.967	0.961	0.944
SCPJ	Na	6	9	6	3	8	5	9	2	6.000
	He	0.748	0.883	0.800	0.446	0.763	0.580	0.742	0.667	0.704
	Ho	0.308	0.769	0.385	0.077	0.538	0.500	0.385	0.000	0.370
	PIC	0.981	0.959	0.923	0.932	0.972	0.971	0.927	0.940	0.951
SCXC	Na	3	8	4	2	4	1	4	5	3.875
	He	0.567	0.816	0.743	0.067	0.395	0.000	0.559	0.618	0.471
	Ho	0.000	0.467	0.800	0.067	0.400	-	0.067	0.733	0.362
	PIC	0.974	0.979	0.890	0.910	0.969	0.918	0.976	0.978	0.949
SCYA	Na	9	13	5	3	6	4	7	0	5.875
	He	0.869*	0.909	0.756	0.536	0.732	0.538	0.802	0.000	0.643
	Ho	0.688	0.938	0.750	0.267	0.375	0.500	0.250	-	0.538
	PIC	0.916	0.928	0.910	0.912	0.890	0.935	0.979	0.966	0.957
YNEY	Na	8	9	0	4	3	2	4	1	3.875
	He	0.818	0.846	0.000	0.251	0.191	0.667	0.251	0.000	0.378
	Ho	0.133	0.333	-	0.133	0.067	0.000	0.067	-	0.107
	PIC	0.972	0.899	0.926	0.930	0.929	0.927	0.972	0.967	0.941
YNWS	Na	6	8	6	2	7	6	7	1	5.375
	He	0.779	0.862	0.801	0.159	0.833	0.500	0.848	0.000	0.598
	Ho	0.333	0.750	0.500	0.000	0.667	0.417	0.727	-	0.485
	PIC	0.954	0.901	0.927	0.915	0.928	0.926	0.981	0.958	0.946

- Relevant data unavailable

*Statistically significant deviation from Hardy-Weinberg equilibrium ($P < 0.01$)

Table 5 Coefficients of genetic diversity of *O. hupensis* at different loci (the populations of landscape of karst and coastal hills)

Populations	Index	Microsatellite loci								Total
		T1-10	T4-25	D11	T4-22	T5-11	T6-27	A18	C22	
GXBS	<i>Na</i>	0	3	4	2	4	3	3	5	3.000
	<i>He</i>	0.000	0.601	0.739	0.667	0.788	0.503	0.582	0.739*	0.577
	<i>Ho</i>	-	0.556	0.444	0.000	0.000	0.667	0.111	0.222	0.286
	<i>PIC</i>	0.957	0.898	0.918	0.904	0.944	0.920	0.972	0.971	0.936
GXYZ	<i>Na</i>	3	1	1	0	0	1	2	1	1.25
	<i>He</i>	0.506	0.000	0.000	0.000	0.000	0.000	0.315	0.000	0.103
	<i>Ho</i>	0.063	-	-	-	-	-	0.375	-	0.055
	<i>PIC</i>	0.946	0.912	0.937	0.901	0.891	0.921	0.969	0.964	0.931
FJFZ	<i>Na</i>	9	8	7	1	5	5	6	0	5.125
	<i>He</i>	0.861	0.698	0.861	0.000	0.705	0.714	0.754	0.000	0.574
	<i>Ho</i>	0.364	0.692	0.364	-	0.538	0.769	0.231	-	0.493
	<i>PIC</i>	0.947	0.914	0.925	0.921	0.923	0.931	0.902	0.978	0.930
FJFQ	<i>Na</i>	10	10	6	5	12	4	13	6	8.250
	<i>He</i>	0.786	0.832	0.864	0.498	0.826	0.800*	0.805	0.377	0.724
	<i>Ho</i>	0.222	0.158	0.167	0.444	0.842	0.667	0.842	0.053	0.424
	<i>PIC</i>	0.886	0.960	0.927	0.908	0.922	0.907	0.908	0.922	0.918

- Relevant data unavailable

*Statistically significant deviation from Hardy-Weinberg equilibrium ($P < 0.01$)

possible single exact locus tests ($P < 0.01$). No significant linkage disequilibrium was found between all pairs of the eight loci examined ($P < 0.01$), which indicated the independent behaviour of all loci. Analysis with Genepop software showed the possible occurrence of null alleles, which may lead to deviations from HWE and result in exaggerated levels of genetic differentiation [26, 29, 30]. Null alleles may be due to flank sequence variation decreasing primer annealing efficiency, allele drop out or DNA quality [23, 31].

Genetic differences among individuals

Fit and *Fst* values were 54.95 and 37.62 %, respectively. This suggests that genetic differences mainly exist within

populations rather than among those with unbalanced differentiation degrees (Table 6).

Mantel's test of regression showed that the correlation (41.97 %) between geographic distance and genetic distance among populations is positive ($R^2 = 0.1011$, $P < 0.05$) and genetic distribution of all populations accorded with the Isolation-by-distance Model (Fig. 2, Tables 7 and 8).

Genetic parameters of the four groups from different landscapes (i.e. lakes and marshes, high mountains, Karst and coastal Hills) showed that *Na* ranged from 2.063 to 11.452, *He* from 0.465 to 0.852 and *Ho* from 0.274 to 0.492. The group from the Karst landscape had the lowest value in all three indices, which indicated its low

Table 6 F-Statistics and gene flow for all loci

Locus	Sample Size	<i>Fis</i>	<i>Fit</i>	<i>Fst</i>	<i>Nm</i>
T1-10	396	0.6107	0.7534	0.3665	0.4321
T4-25	396	0.0569	0.3253	0.2846	0.6284
D11	396	0.3852	0.6297	0.3977	0.3786
T4-22	396	0.3883	0.6821	0.4803	0.2705
T5-11	396	0.0883	0.3750	0.3144	0.5451
T6-27	396	-0.0044	0.4410	0.4435	0.3138
A18	396	0.4368	0.6229	0.3304	0.5067
C22	396	0.2437	0.5459	0.3996	0.3756
Mean	396	0.2721	0.5459	0.3762	0.4146

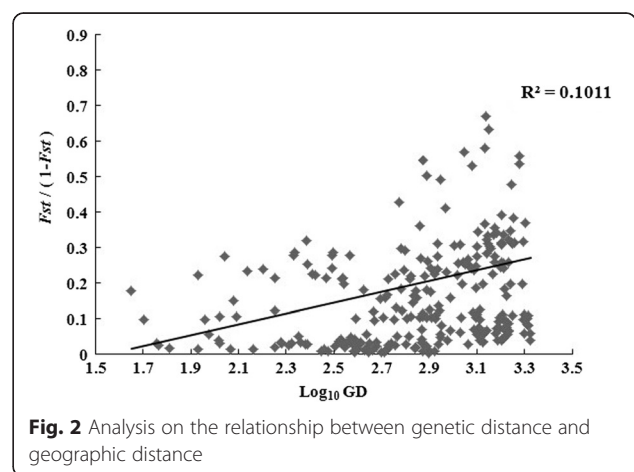


Fig. 2 Analysis on the relationship between genetic distance and geographic distance

Table 7 *F_{ST}* and geographic distance among paired *O. hupensis* populations of landscape of swamps and lakes

Population	AHWW	HBJL	HBWH	HNHS	HNNX	JSYZ	JSYZJZ	JXDC	JXJJ	JXNC	ZJJH	AHNG	AHWJ
AHWW		536.297	344.514	625.695	574.347	178.616	214.554	257.933	264.471	340.625	312.257	137.411	159.779
HBJL	0.013		192.208	145.904	95.03	693.51	744.317	386.605	332.681	389.57	738.131	633.471	409.929
HBWH	0.014	0.032		300.549	244.352	502.28	552.224	239.473	181.045	279.355	569.54	447.876	229.76
HNHS	0.008	0.015	0.008		57.273	794.893	839.558	425.374	383.178	397.899	776.932	701.923	479.599
HNNX	0.025	0.055	0.028	0.032		741.501	787.471	385.642	339.518	367.616	739.602	655.832	432.023
JSYZ	0.015	0.018	0.006	0.01	0.034		64.775	435.036	442.934	515.829	370.643	210.766	338.393
JSYZJZ	0.008	0.032	0.03	0.01	0.004	0.018		460.911	476.096	537.653	341.035	204.009	371.034
JXDC	0.02	0.006	0.029	0.013	0.043	0.017	0.012		58.495	85.413	354.412	287.437	104.501
JXJJ	0.023	0.037	-0.012	0.008	0.036	0.007	0.035	0.027		116.585	405.588	319.529	105.121
JXNC	0.011	0.022	0.01	0.015	0.034	0.007	0.017	0.014	0.015		384.737	352.827	189.904
ZJJH	0.042	0.042	0.027	0.024	0.07	0.039	0.038	0.032	0.028	0.027		179.742	341.546
AHNG	0.03	0.029	0.032	0.021	0.051	0.03	0.025	0.015	0.022	0.02	0.021		224.566
AHWJ	0.039	0.034	0.035	0.023	0.057	0.042	0.048	0.041	0.03	0.033	0.032	0.052	

Lower triangle and upper triangle represent *F_{st}* and geographic distance (GD) / km, respectively

differentiation degree. AMOVA displayed that variations of *O. hupensis* mainly exists among individuals, which accounted for 60.58 % of total variations, and that of among individuals within populations, among populations within groups and among groups were only 26.60, 8.04 and 4.78 %, respectively (Table 9). This suggests that there is no significant genetic differentiation among groups.

UPGMA cluster analysis for the 24 *O. hupensis* populations based genetic distance showed that the populations spread in the landscape of middle and lower reaches of Yangtze River (HBWH, JSYZ, JXNC, HNHS, JXJJ, AHWW, HBJL, JXDC, HNNX, JSYZJZ, ZJJH, AHNG and AHWJ) clustered together first and then gathered with the populations of high mountains (SCMS, SCYA, SCPJ, YNEY, SCLS, YNWS and SCXC), coastal hills (FJFQ and FJFZ) and Karst land form (GXBS and GXYZ) successively (Fig. 3).

Discussion

Oncomelania hupensis is the sole intermediate host for transmitting *Schistosoma japonicum* in mainland China [32], and it is widely distributed in the southern region of the Yangtze River valley. Significant genetic variations have developed in *O. hupensis* from different geographic populations due to their distribution range, complexity of breeding environment and geographical location.

In this research, The genetic differentiation of four different landscape groups of *O. hupensis* were studied through eight screened polymorphic microsatellite DNA loci. This information is pertinent because it further improve our understanding on the effect of genetic diversities on the distribution of *O. hupensis*. This will ultimately help boost our surveillance activities and also strengthen the control of schistosomiasis transmission in China. genetic indices were tested across eight

Table 8 *F_{ST}* and geographic distance among paired *O. hupensis* populations of landscape of mountains, karst and Coastal hills

Population	SCXC	SCMS	SCLS	SCPJ	SCYA	FJFZ	YNWS	GXBS	GXYZ	YNEY	FJFQ
SCXC		293.116	216.775	310.261	246.145	1741.353	369.633	626.605	773.059	315.439	1769.631
SCMS	0.115		85.336	44.757	109.864	1603.349	662.713	733.298	794.413	604.077	1637.681
SCLS	0.188	0.124		119.599	123.065	1607.142	584.379	665.163	745.687	531.676	1639.497
SCPJ	0.178	0.079	0.15		91.093	1636.835	678.755	776.807	839.083	614.433	1671.819
SCYA	0.154	0.077	0.107	0.099		1710.986	606.707	781.117	868.641	535.667	1744.806
FJFZ	0.248	0.192	0.31	0.242	0.24		1958.674	1340.025	1098.056	1981.6	50.253
YNWS	0.179	0.11	0.051	0.146	0.084	0.254		667.13	887.951	104.933	1979.866
GXBS	0.392	0.271	0.205	0.304	0.262	0.53	0.219		242.047	723.997	1354.27
GXYZ	1.503	1.039	1.409	1.425	1.2	2.095	1.561	3.319		931.896	1112.731
YNEY	0.187	0.099	0.168	0.123	0.127	0.266	0.106	0.362	1.413		2005.112
FJFQ	0.211	0.138	0.184	0.166	0.145	0.298	0.17	0.368	1.169	0.169	

Lower triangle and upper triangle represent *F_{st}* and geographic distance (GD) / km, respectively

Table 9 Analysis of molecular variance (AMOVA) for the *Oncomelania hupensis*

Source of variation	Degree of freedom	Sum of squares	Variance components	Percentage of variation/%
Among group	3	15.653	0.02386	4.78
Among populations within groups	20	35.115	0.04015	8.04
Among individuals within populations	333	189.196	0.13282	26.60
Within individuals	357	108.000	0.30252	60.58
Total	713	347.964	0.49935	

microsatellite DNA loci. The mean *Fis* value for the 24 populations examined was 0.272, indicating a deficiency of heterozygotes and frequent inbreeding within populations, which is likely due to the small range of activity of *O. hupensis*. A total of 47 microsatellite DNA loci deviated from the Hardy Weinberg Equilibrium demonstrating a serious lack of heterozygotes. Possible explanations that may account for this include: activities of migration and inbreeding, drug pressure, gene mutation and null alleles. However, it is currently unclear which one is the dominant factor contributing to this phenomenon [33]. No significant linkage disequilibrium was found between all pairs of the eight loci, clearly showing the independent behaviour of all loci. Null alleles were found at all eight polymorphic loci. This may be due to: 1) mismatching of

primer pairs: mutations in microsatellite DNA sites critical for binding with primers leads to abnormal amplification 2) losses of large alleles: the superiority of short alleles restrict amplification of long fragments or 3) differences in DNA quality: unevenness of templates character obstruct amplification in some loci [26, 31, 34]. Null alleles could implicate genetic diversity parameters for populations such as excess of homozygote individuals, reduction of *Ho* and *He* and increase of genetic distance and *Fis*; moreover, it leads to inaccuracy of parent analysis [30–37].

The abundance of the number of heterozygotes and the amount of genetic information in a population is directly proportional to the *PIC* value [38, 39]. Result shows that *PIC* was greater than 0.5 at every locus, and the mean

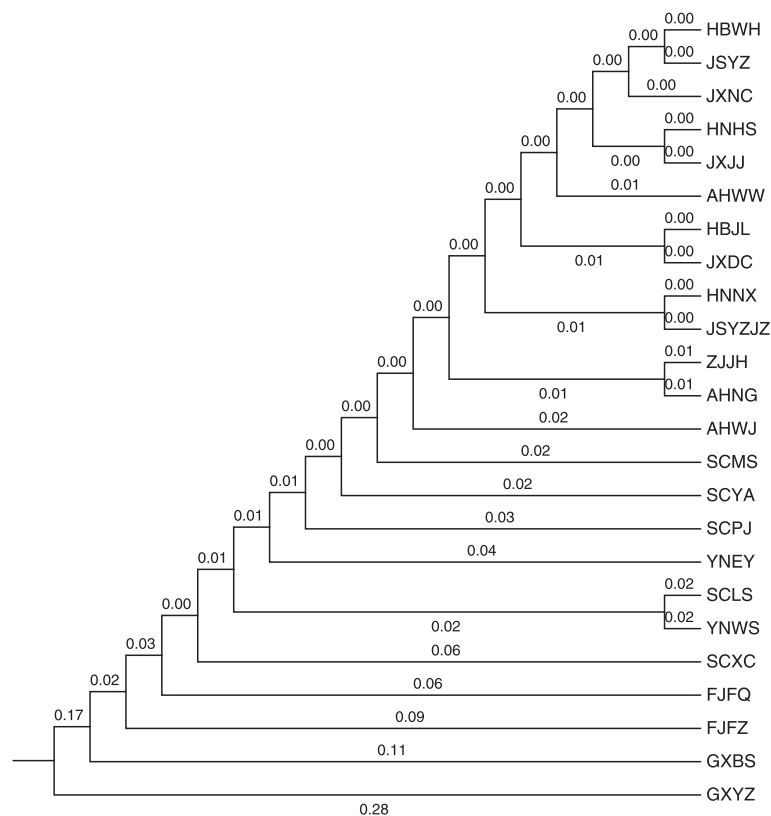


Fig. 3 UPGMA cluster analysis of 24 *O. hupensis* populations

value (0.947) from all populations was higher than (0.764) obtained from previous result [23]. This signifies that all the eight loci screened were highly polymorphic.

Furthermore, this study reveals that the average *Fst* for all loci was 0.376, which means that 37.6 % of genetic variation was among populations and 72.4 % was among individuals within populations. The analysis of AMOVA displayed that genetic variation among individuals (60.58 %) were far higher than that within populations (26.60 %), while among populations and among groups are (8.04 %) and (4.78 %) respectively. This implies that, genetic diversity is strongly derived from among-individuals rather than among-populations. However, the average *Fst* (0.376) and genetic variation among populations (8.04 %) were higher than values obtained from the previous results (0.048 and 4.8 %) respectively, revealing genetic variation among populations increased along with geographical distance [23]. The Mantel test demonstrated an apparent positive correlation between genetic distance and geographical distance. The genetic structure between geographical populations is embodied with some degree of independence. For example, the geographical distance between the HBWH and JSYZ populations located in the lake region was far, but with low degree of variation. This could possibly be related to the genetic differentiation principally being among individuals within populations rather than among geographic locations for the populations in Lakes and Marshes landscape.

The phylogenetic tree constructed by UPGMA also showed that populations in neighboring geographical locations generally cluster together, which was consistent with the Mantel test results. The cluster sequence of geographical populations showed us that the population from the karst landscape of Guangxi autonomous region maybe the most original one, then the population from the littoral hill part of the Fujian province, the population from the mountainous region of the Sichuan and Yunnan provinces and the population from the region of swamps and lakes in the middle and lower reaches of the Yangtze River, respectively. Regarding as the largest population spread throughout the middle and lower reaches of the Yangtze River [7], the populations from different provinces also crossed cluster, these include, between Hubei and Jiangsu, Hunan and Jiangxi, and Zhejiang and Anhui, which may be as a result of *O. hupensis* spreading along the river within the large population, or gene drifting for surged water flow in the lakes and marshes landscape [34]. Then this branch clustered with the populations of Sichuan and Yunnan province successively. Furthermore, the major branch clustered with the populations of Fujian and Guangxi province in turn, this agrees with the conclusion of four landscape populations relationships from previous studies using SSR-PCR [40] and DNA sequence markers [7, 41, 42].

Conclusion

This study has shown that the genetic diversity of *O. hupensis*, an important snail intermediate host of *S. japonicum* in China mainly originates from among-individuals rather than among-populations. It also reveals that the populations within subspecies have closer consanguinity than between subspecies in the mass, nevertheless, genetic variations exist within subspecies. These findings further provide important information on genetic structure of *O. hupensis* and strengthen our knowledge about diffusion trend and tracking to the source of *Oncomelania* in mainland China. Ultimately, these findings will help us develop more effective guidelines for controlling the spread and distribution of *Oncomelania* and consequently prevent the transmission of *Schistosomiasis* in China. Our data offers a better understanding of the genetic differentiation of *Oncomelania hupensis*, enhancing our ability to effective and efficient surveillance of *Schistosomiasis*.

Competing interests

The authors declare that they do not have competing interests.

Authors' contributions

WG, SL and BW conceived the study; WG, BW and SL performed the field collection, statistical analyses and wrote the manuscript; MA, BW, DR and XZ revised the manuscript and gave approval of the version to be published. All the authors read and approved the final version of the manuscript.

Acknowledgments

This study was supported by the National Natural Science Foundation of China (No. 81101280), the National Special Science and Technology Project for Major Infectious Diseases of China (Grant No. 2012ZX10004-220 and 2012ZX10004-201), Public Health Overseas Fund, Bureau of Health, Shanghai (No. GWHW201216), China-UK Global Health Support Programme (Grant No: GHSP-CS-OP1-01)

Author details

¹National Institute of Parasitic Diseases, Chinese Center for Disease Control and Prevention, Shanghai 200025, People's Republic of China. ²Key Laboratory of Parasite and Vector Biology, Ministry of Health, WHO Collaborating Center for Malaria, Schistosomiasis and Filariasis, Shanghai 200025, People's Republic of China. ³Department of Zoology, Federal University Lafia, P.M. B 146, Lafia, Nasarawa State, Nigeria. ⁴Wolfson Wellcome Biomedical Laboratories, Department of Zoology, Natural History Museum, Cromwell Road, London SW7 5BD, UK.

Received: 27 August 2015 Accepted: 12 January 2016

Published online: 20 January 2016

References

1. Wang L, Utzinger J, Zhou XN. Schistosomiasis control: experiences and lessons from China. *Lancet*. 2008;372:1793–5.
2. Li SZ, Luz A, Wang XH, Xu LL, Wang Q, Qian YJ, et al. Schistosomiasis in China: acute infections during 2005–2008. *Chin Med J (Engl)*. 2009;122:1009–14.
3. Utzinger J, Zhou XN, Chen MG, Bergquist R. Conquering schistosomiasis in China: the long march. *Acta Trop*. 2005;96:69–96.
4. Wang LD, Chen HG, Guo JG, Zeng XL, Hong XL, Xiong JJ, et al. A strategy to control transmission of *Schistosoma japonicum* in China. *N Engl J Med*. 2009;360:121–8.
5. Attwood SW, Upatham ES, Zhang YP, Yang ZQ, Southgate VR. A DNA-sequence based phylogeny for triculine snails (Gastropoda: Pomatiopsidae: Triculinae), intermediate hosts for *Schistosoma* (Trematoda: Digenea): Phylogeography and the origin of Neotricula. *J Zool*. 2004;262:47–56.

6. Zhou YB, Zhao GM, Peng WX. Spatial genetic correlation analyses of *Schistosoma japonicum* intermediate hosts within *Oncomelania hupensis* (Gastropoda: Rissooidea) from mainland China based on amplified fragment length polymorphisms. *Fudan Univ J Med Sci.* 2007;34:207–12.
7. Li SZ, Wang YX, Yang K, Liu Q, Wang Q, Zhang Y, et al. Landscape genetics: the correlation of spatial and genetic distances of *Oncomelania hupensis*, the intermediate host snail of *Schistosoma japonicum* in mainland China. *Geospat Health.* 2009;3:221–31.
8. Zhou YB, Yang MX, Zhao GM, Wei JG, Jiang QW. *Oncomelania hupensis* (Gastropoda: Rissooidea), intermediate host of *Schistosoma japonicum* in China: genetics, molecular phylogeny based on amplified fragment length polymorphisms. *Malacologia.* 2007;49:367–82.
9. Davis GM, Zhang Y, Hua GY, Spolsky C. Population genetics and systematic status of *Oncomelania hupensis* (Gastropoda: Pomatiopsidae) throughout China. *Malacologia.* 1995;37:133–56.
10. Li SZ, Wang Q, Qian YJ, Zhang Y, Zhou XN. Subspecies differentiation of *Oncomelania hupensis* in Mainland of China. *Chin J Schisto Control.* 2009;21:150–3.
11. Zhou YB, Jiang QW, Zhao GM, Yuan HC. Subspecies differentiation of *Oncomelania hupensis* from Mainland China. *Chin J Schisto Control.* 2007;19:485–7.
12. Shi CH, Wilke T, Davis GM, Xia MY, Qiu CP. Population genetics, micro-phylogeography, ecology, and infectivity of Chinese *Oncomelania hupensis* (Gastropoda: Rissooidea: Pomatiopsidae) in the Miao River system: is there a relationship to shell sculpture? *Malacologia.* 2002;44:333–47.
13. Schlotteröer C, Amos B, Tautz D. Conservation of polymorphic simple sequence loci in cetacean species. *Nature.* 1991;354:63–5.
14. Guo W, Shen ZR. The application of microsatellite DNA markers in entomology. *Biotechnology.* 2004;14:60–1.
15. Penaa HFJ, Vitaliano SN, Beltrameb MAV, Pereirac FEL, Gennaria SM, Soares RM. PCR-RFLP genotyping of *Toxoplasma gondii* from chickens from Espírito Santo state, Southeast region, Brazil: New genotypes and a new SAG3 marker allele. *Vet Parasitol.* 2013;192:111–7.
16. Webster BL, Webster JP, Gouvras AN, Garba A, Lamine MS, Diaw OT, et al. DNA 'barcoding' of *Schistosoma mansoni* across sub-Saharan Africa supports substantial within locality diversity and geographical separation of genotypes. *Acta Trop.* 2013;128:250–60.
17. Webster BL, Culverwell CL, Khamis IS, Mohammed KA, Rollinson D, Stothard JR. DNA barcoding of *Schistosoma haematobium* on Zanzibar reveals substantial genetic diversity and two major phylogenetic groups. *Acta Trop.* 2013;128:206–17.
18. Feng QQ, Sui ZH, Zhang XC, Kong FN. Study on microsatellite markers of *Gracilaria lemaneiformis*. *P Ocean Univ Chin.* 2010;40:77–81.
19. Zhang YP, Wang W, Su B, Fan ZY, Zhang HM, He TM. Screening and application on microsatellite makers of the giant panda. *Zoological Res.* 1995;16:301–6.
20. Zhang L, Fan Y, Ma YJ. Isolation of Microsatellite DNA and the Polymorphic Locus Screening from *Phlebotomus chinensis* (Diptera: Psychodidae). *Chin J Parasitol Parasit Dis.* 2009;27:503–7.
21. Li SZ, Wang YX, Ma YJ, Wang Q, Liu Q, Wu Y. Isolation and characterization of polymorphic microsatellite markers of *Oncomelania hupensis*. *Chin J Schisto Control.* 2010;22:122–5.
22. Li Z, Li SZ, Wang Q, Qian YJ, Liu Q, Yang P. Isolation and characterization of 15 new microsatellite markers in *Oncomelania hupensis*, the snail intermediate host of *Schistosoma japonicum* in mainland China. *Int J MolSci.* 2012;13:5844–50.
23. Zhou YB, Zhao GM, Wei JG, Jiang QW. Genetic diversity in 19 Chinese populations of *Oncomelania hupensis* (Gastropoda: Rissooidea) detected by simple sequence repeat-anchored polymerase chain reaction amplification. *Chin J Epidemiol.* 2007;28:859–62.
24. Li SZ, Zhang L, Liu Q, Lv S, Wang Q, Qian YJ. Study on the Genetic Differences among *Oncomelania hupensis* Population in Middle and Lower Reaches of Yangtze River using Microsatellite DNA Markers. *Chin J Parasitol Parasit Dis.* 2012;30:268–73.
25. Cui B, Guan W, You P, Li SZ. Fine-scale population genetic structure of *Oncomelania hupensis* based on microsatellite DNA markers. *Chin J Zoonoses.* 2014;30:701–8.
26. Ma L, Li SZ, Yang P, You P, Zhou XN. Comparison of five different methods of extracting genomic DNA from *Oncomelania hupensis*. *Chin J Pathogen Biol.* 2011;6:129–32.
27. Excoffier L, Laval G, Schneider S. Arlequin (version 3.0): an integrated software package for population genetics data analysis. *Evol Bioinform Online.* 2007;1:47–50.
28. Khan HA, Arif IA, Bahkali AH, Al Farhan AH, AlHomaidan AA. Bayesian, maximum parsimony and UPGMA models for inferring the phylogenies of antelopes using mitochondrial markers. *Evol Bioinform Online.* 2008;4:263–70.
29. DeSousa SN, Finkeldey R, Galling O. Experimental verification of microsatellite null alleles in Norway spruce (*Picea abies* [L.] Karst.): Implications for population genetic studies. *Plant Mol Biol Rep.* 2005;23:113–9.
30. Chapuis MP, Estoup A. Microsatellite null alleles and estimation of population differentiation. *Mol Biol Evol.* 2007;24:621–31.
31. Wen YF, Uchiyama K, Han WJ, Ueno S, Xie WD, Xu GB. Null alleles in microsatellite markers. *Bio diversity Science.* 2013;21:117–26.
32. Davis GM, Zhang Y, Guo YH, Spolsky C. Systematic status of *Oncomelania Hupensis* (Gastropoda: Pomatiopsidae) throughout China. *Stud Mar Sin.* 1997;39:89–95.
33. Zhou YB, Zhao GM, Wei JG, Jiang QW. Study on the genetic diversity among populations of schistosome intermediate hosts within *Oncomelania hupensis* (Gastropoda: Pomatiopsidae) in mainland China. *Chin J Epidemiol.* 2006;27:865–70.
34. Dakin EE, Avise JC. Microsatellite null alleles in parentage analysis. *Heredity.* 2004;93:504–9.
35. Lemer S, Rochel E, Planes S. Correction method for null alleles in species with variable microsatellite flanking regions, a case study of the black-lipped pearl oyster *Pinctada margaritifera*. *J Hered.* 2011;102:243–6.
36. Jones AG, Ardren WR. Methods of parentage analysis in natural populations. *Mol Ecol.* 2003;12:2511–23.
37. Moriguchi Y, Taira H, Tani N, Tsumura Y. Variation of paternal contribution in a seed orchard of *Cryptomeria japonica* determined using microsatellite markers. *Can J For Res.* 2004;34:1683–90.
38. Liao XL, Yu XM, Tan DQ, Tong JG. Microsatellite DNA analysis of genetic diversity of grass carp in Yangtze River system. *Acta Hydrobiol Sin.* 2005;29:113–9.
39. Yan LN, Zhang DX. Effects of sample size on various genetic diversity measures in population genetic study with microsatellite DNA markers. *Acta Zool Sin.* 2004;50:279–90.
40. Niu AO, Xiong YW. Studies on the genetic variation of *Oncomelania hupensis* with SSR-PCR. *Chin J Parasit Dis Control.* 2002;15:230–3.
41. Zhao QP, Jiang MS, Littlewood DT, Nie P. Distinct genetic diversity of *Oncomelania hupensis*, intermediate host of *Schistosoma japonicum* in mainland China as revealed by ITS sequences. *PLoS Negl Trop Dis.* 2010;4:e611.
42. Attwood SW, Ibaraki M, Saitoh Y, Nihei NJ, Janies DA. Comparative Phylogenetic Studies on *Schistosoma japonicum* and Its Snail Intermediate Host *Oncomelania hupensis*: Origins, Dispersal and Coevolution. *PLoS Negl Trop Dis.* 2015;9(7):e0003935. doi:10.1371/journal.pntd.0003935

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

