Development of vaccines against the

human immunodeficiency virus and

sexually transmitted infections gonorrhoea,

syphilis, Chlamydia, herpes simplex virus,

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Zika virus

Abstract: The success in preventing hepatitis B virus and human papillomavirus infections by means of vaccination paves the way for the development of other vaccines to prevent sexually transmitted infections (STIs) such as gonorrhoea, syphilis, chlamydia, herpes simplex virus, human immunodeficiency virus and Zika virus. The current status of vaccine development for these infections will be explored in this review.

The general principles for success include the need for prevention of latency, persistence and repeat infections. A reduction in transmission of STIs would reduce the global burden of disease. Therapeutic activity of vaccines against STIs would be advantageous over preventative activity alone, and prevention of congenital and neonatal infections would be an added benefit. There would be an added value in the prevention of long-term consequences of STIs. It may be possible to re-purpose 'old' vaccines for new indications. One of the major challenges is the determination of the target populations for STI vaccination.

Keywords: chlamydia, gonorrhoea, syphilis, herpes simplex virus, human immunodeficiency virus, Zika virus

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Introduction

According to the World Health Organisation (WHO), there were estimated to be more than 1 million sexually transmitted infections (STIs) acquired every day worldwide in 2016, and each year there are an estimated 357 million new infections with one of four STIs: chlamydia, gonorrhoea, syphilis and trichomoniasis.¹ Furthermore, antimicrobial drug resistance, especially for gonorrhoea, is a major threat to the successful control of STIs.

There are possibilities for reductions in STIs:

• Vaccination against *Neisseria meningitidis* serogroup B may offer a degree of protection against *Neisseria gonorrhoeae*.

- Candidate vaccines against syphilis, chlamydia, herpes simplex virus (HSV) and human immunodeficiency virus (HIV) are under development.
- Zika virus candidates are under development in the context of outbreak preparedness, the prevention of mosquito and sexual transmission, and the prevention of congenital infections.

Mechanisms of natural immunity and immune correlates of protection

Repeated gonorrhoea infections are common and can occur with the same strain or serotype, and the inability of natural infection to induce longlasting immunity hinders the development of

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All STIs	 Reach consensus on clinical trial endpointss Improve measuring endpoints, validating surrogate endpoints Strengthen clinical trial design, including sites Involve regulators; determine regulatory routes to licensure Promote phase I evaluation Establish systems to monitor outcomes in advance
HSV	 Define clinical endpoints: infection <i>versus</i> genital ulcer disease, and the role of shedding Understand potential risks/benefits of evaluation in high HIV prevalence areas
Chlamydia	 Define clinical endpoints: infection <i>versus</i> pelvic inflammatory disease Develop biomarkers, radiological tests, etc. for upper genital tract disease
Gonorrhoea	 Define clinical endpoints: infection <i>versus</i> pelvic inflammatory disease Develop biomarkers, radiological tests, etc. for upper genital tract disease Explore use of human male urethral challenge model in phase I/II clinical trials
Syphilis	Determine trial design, population and settingAssess possible evaluation as maternal immunisation
HIV, human imr	nunodeficiency virus; HSV, herpes simplex virus; STI, sexually transmitted infections.

 Table 1. Roadmap for facilitating clinical evaluation and vaccine introduction.⁹

vaccines against gonorrhoea.² Regarding syphilis, little is known about the correlates of immunity in humans and it is well recognised that individuals who acquire syphilis can be re-infected following treatment, and that this cycle can be repeated many times.³ At most, there may be a degree of partial protection following chlamydia infection.⁴ But, in general for STIs, natural infection does not generally protect against subsequent infections, and recurring infections are a feature.

Regarding the usefulness of immune correlates of protection, attempts to identify such correlates for HIV infection point to a broader repertoire of immune activation than simply antibody responses.^{5,6} There is no ideal animal model for studying sexually transmitted diseases, although the cotton rat, for example, has proved useful in the study of genital herpes simplex virus infection,⁷ and the nonhuman primate model is useful for studying HIV infection and acquired immunodeficiency syndrome (AIDS).⁸

Gottlieb and Johnston provide a very helpful guide with respect to clinical trials and the development of vaccines against sexually transmitted diseases (see Table 1).⁹

Issues related to the development of vaccines against STIs

The preventable burden of bacterial and viral STIs is difficult to estimate. The causes of this

uncertainty range from the reluctance of some individuals to seek diagnosis and treatment, asymptomatic infections rendering the individual unaware that they are infected, latency and persistence of infection, diagnostic tests not being performed and the over-use of antibiotics. These very reasons also help explain why clinical trial design for vaccines against STIs is problematic, and why target populations for vaccine implementation, once STI vaccines are licensed, are ill-defined.

As will be seen below, there is some progress in the development of vaccines against STIs that may have both preventative and therapeutic activity. The design of a clinical trial to test both preventative and therapeutic qualities of an STI vaccine would be based on the premise that the status of the subjects is known at the beginning of the trial, to distinguish infected from non-infected individuals. Such a dichotomy may not be easy to establish. It may be preferable to perform one trial in infected individuals and another in noninfected individuals. Some of these dilemmas will be explored below.

Gonococcus

Burden of disease

The WHO estimated the global prevalence and incidence of gonorrhoea in 2016.¹⁰ The 2016

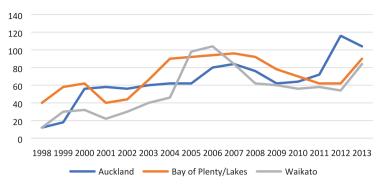


Figure 1. Gonorrhoea rates per 100,000 population in selected New Zealand regions, 1998–2014.

global prevalence estimate for gonorrhoea in women was 0.9% (95% uncertainty interval 0.7– 1.1) whereas in men it was 0.7% (95% uncertainty interval 0.5–1.1). There were 86.9 million total estimated incident cases of gonorrhoea globally (95% uncertainty interval 103.4–231.2 million).

Pre-clinical vaccine development

Lipooligosaccharide being the most abundant molecule expressed on the gonococcal surface and contributing significantly to pathogenesis, is an obvious target for vaccine development.¹¹ The promising candidate known as TMCP2 has been shown to elicit bactericidal IgG, and reduced colonisation levels of gonococci in experimentally infected mice whilst also accelerating clearance by each of two different gonococcal strains. Other approaches in pre-clinical development include outer membrane vesicles (see below) and purified protein subunit vaccines¹²

MeNZB™

Bioinformatic analysis has been performed to assess the similarity of a prototype *Neisseria meningitidis* serogroup B vaccine known as MeNZB[™] OMV (outer membrane vesicle) and antigens from a four-component *N. meningitidis* serogroup B vaccine known as Bexsero[®], to gonococcal proteins.¹³ Rabbits were immunised with the OMV component or three recombinant antigens of Bexsero[®], and western blot and enzyme-linked immunosorbent assay (ELISA) were used to assess the generation of antibodies recognising *Neisseria gonorrhoeae*. Serum from humans immunised with Bexsero[®] was investigated to assess the nature of the antigonococcal response. There was found to be a high level of sequence identity between MeNZB[™] OMV and Bexsero[®] OMV antigens and gonococcal proteins. Bexsero[®] induced antibodies in humans that recognise gonococcal proteins.

During and after the use of MeNZBTM in New Zealand during a meningococcal outbreak between 2004 and 2006, there was observed to be a simultaneous decline in reported cases of gonorrhoea.¹⁴ Figure 1 shows the impact of MeNZBTM on gonorrheal infections in three regions of New Zealand arising from the use of MeNZBTM during 2004–2006.¹⁵ No other STIs described in national surveillance reports declined during this period.

The anti-gonococcal antibodies induced by MeNZB-like OMV proteins could explain the previously seen decrease in gonococcal cases following MeNZB vaccination.¹³

Modelling

To explore the potential impact of vaccination against *N. meningitidis* on *N. gonorrhoeae* in the United States (US), a decision-analysis modelling exercise was performed.¹⁶ The authors modelled the theoretical impact of a US four-component *N. meningitidis* serogroup B (4CMenB) vaccination programme on gonorrhoea outcomes. A decision-analysis model was populated using published healthcare utilisation and cost data. A two-dose adolescent vaccination campaign was assumed, with protective immunity starting at age 15 years and an assumed base-case efficacy against gonorrhoea of 20%. One of the key outcome measures was a reduction in gonorrhoea and HIV infections.

The model predicts that without vaccination, a theoretical US adolescent cohort would experience 844,000 gonorrhoea infections (95% credible interval 439,200–1,399,000) over a lifetime.¹⁶ Without vaccination, gonorrhoea infections would increase the number of HIV cases by 557. The model also predicts that adolescent vaccination with 4CMenB would prevent 83,167 (95% credible interval 44,600–134,6000) gonorrhoea infections and decrease the number of HIV infections by 55 (95% credible interval 2–129) per vaccinated birth cohort in the USA. The authors conclude that, even with a low effectiveness against gonorrhoea, for example 20%, a US vaccination programme against serogroup B meningococcal disease using 4CMenB would substantially reduce the number of gonorrhoea infections.

Challenges and next steps

An understanding of how *Neisseria meningitidis* group B OMV vaccines work against *Neisseria* gonorrhoeae could enhance the development pathways of vaccines against gonorrhoea.¹⁷

The challenges of vaccine development for gonorrhoea are highlighted in Table 1 and include the use of human challenge models. The next step for this field is to await the results of studies using 4CMenB against gonorrhea and other OMVbased vaccines.^{17–19}

Syphilis

Burden of disease

The WHO estimated the global prevalence and incidence of syphilis in 2016.¹⁰ The 2016 global prevalence estimate for syphilis in women was 0.5% (95% uncertainty interval 0.4–0.6), whereas in men it was 0.5% (95% uncertainty interval 0.4–0.6). There were 6.3 million total estimated incident cases of syphilis globally (95% uncertainty interval 5.5–7.1 million).

Pre-clinical vaccine development

A prototype syphilis vaccine was developed by James Miller as early as 1973.²⁰ The difficulties ahead are foretold in his conclusion: 'Conclusive evidence that *Treponema pallidum* immobilization (TPI) antibody is not associated with the immune response was provided by the finding that 8 of the 11 immune rabbits challenged 1 year after vaccination had no TPI antibody before challenge and

failed to develop immobilizing antibody during the 3-month period of observation after challenge'. Nevertheless, the experiment was successful in that rabbits immunised with a non-infectious y-irradiated *T. pallidum* were completely protected against disease for at least 1 year in the face of *T. pallidum* infectious challenge.

Possible targets include selected subsets of the T. pallidum repeat (Tpr) protein family (targeting susceptibility and persistence), and the treponemal adhesin protein pallilysin Tp0751 (targeting dissemination).^{21,22} A novel approach has been the use of non-infectious Borrelia burgdorferi as an effective carrier to deliver and elicit a specific host response to T. pallidum antigens such as those expressed by tp0897 (tprK) and tp0435 genes.²³ Plasmid DNA encoding T. pallidum flagellin FlaB3 has been used as a candidate vaccine for the evaluation of immunogenicity and protection against dissemination.²⁴ Subsequent intradermal challenge in rabbits showed a significant reduction in the bacterial burden in blood, liver, spleen and testicles in the rabbits vaccinated with plasmid DNA-encoded flagellin (FlaB3).

Table 2 shows the key issues and implications that need to be considered during the pre-clinical development phase of candidate syphilis vaccines.

Challenges and next steps

Because syphilis transmission occurs by contact with the infectious primary chancre or through secondary lesions, prevention or attenuation of these lesions is a necessary requirement for a syphilis vaccine, according to a recent review of the subject by Caroline Cameron.²⁵ This is because of the need for such a vaccine to either eliminate, or at least reduce, person-to-person transmission. It is also necessary for the vaccine to prevent dissemination of the organism within the infected host, repeat infections, persistence and latency.^{21,22}

Once a suitable candidate or candidates are identified, it will be time to move to the clinical phase of testing in humans. However, with the cloud of the Tuskegee Study, in which the subjects were not informed as to the true purpose of the study and in which subjects with syphilis were not treated, there will be ethical questions related to safety and treatment to be answered before clinical trials begin.²⁶ Table 2. Key issues needing to be assessed during the process of syphilis pre-clinical vaccine development.

Issue	Implication
Number of vaccine administrations required to achieve maximal immunity	Compliance and vaccination scheduling
Duration of immunity induced	Protection against repeated frequent exposure
Cross-protection against diverse strains	Need for a global vaccine approach
Appropriate multi-valent vaccine	Multi-component approach more likely to be successful
Adjuvant selection and optimisation	For immunogenicity in HIV-infected individuals
HIV, human immunodeficiency virus.	

Chlamydia

Burden of disease

The WHO has estimated the global prevalence and incidence of chlamydia in $2016.^{10}$ The 2016 global prevalence estimate for chlamydia in women was 3.8% (95% uncertainty interval 3.3-4.5) whereas in men it was 2.7% (95% uncertainty interval 1.9-3.7). There were 127.2 million total estimated incident cases of chlamydia globally (95% uncertainty interval 95.1-165.9 million).

Pre-clinical and clinical vaccine development

Data related to protective immune responses at the cervical mucosa, that could potentially limit chlamydia infection and reinfection, have been used to inform vaccine approaches and biomarkers.²⁷ It was found that cytokines involved with humoral type 1 interferon and Th17 responses are associated with susceptibility to *Chlamydia trachomatis* whilst cytokines involved in Th1 polarisation, recruitment and activation are associated with protection against ascension and reinfection.

Trials in mice and koalas indicate that the major outer membrane protein (MOMP) is a highly recognised antigenic target and may be preferable to whole cell targets, which have their limitations.²⁸ Whilst sterilising immunity is the ultimate goal, vaccine-induced partial immunity preventing upper genital tract infection and inflammation may be cost-effective when compared with current screening and treatment strategies.²⁹ Current vaccine candidate development is shown in Table 3.³⁰

Challenges and next steps

The conclusions and recommendations of a National Institute of Allergy and Infectious Diseases workshop entitled 'Chlamydia vaccines: the way forward' highlight some of the challenges in chlamydia vaccine development. Zhong *et al.* state:³⁹

- Although preliminary modelling suggests that even a partially protective *C. trachomatis* (Ct) vaccine may be cost-effective, more data are needed regarding progression of Ct infection to upper genital tract sequelae and burden of Ct-associated disease, especially in lower- and middle-income countries, to define better the potential worldwide impact of a Ct vaccine.
- Clinical testing of a Ct vaccine is feasible; however, choice of clinical trial endpoints warrants further investigation and discussion. Blood biomarkers and other novel approaches for identifying upper genital tract infection and inflammation in women would be useful for defining endpoints for vaccine efficacy studies as well as disease burden.
- Although the immunological basis for protection from Ct infection and disease has been well studied, key issues such as the role of antibody still need to be clarified. A relative consensus was reached that putative Ct vaccines should generate Ct-specific CD_4 T cells targeting genital epithelial cells, combined with a strong antibody response.
- Further analysis is needed on the utility of several mouse models available to test candidate vaccines. Harmonization these models such that candidate vaccines can be

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Candidate name/identifier (sponsor)	Pre-clinical	Phase I	Reference
MOMP-VD4 neutralising antibodies (Statens Serum Institut)		X	Olsen <i>et al.</i> ^{31,33} ; Bøje <i>et al.</i> ³²
Intranasal MOMP nano-emulsion (NanoBio Corp)	Х		Fattom ³⁴
MOMP + Pmps (Pan-Provincial Vaccine Enterprise Inc. and British Columbia CDC)	X		Karunakaran <i>et al.</i> ³⁵
cSAP TLR7 agonist with UV-killed <i>Chlamydia</i> (Selecta Biosciences)	Х		Stary et al. ³⁶
Vaxonella platform (<i>Salmonella</i> vector) (Prokarium)	х		Garmory et al. ³⁷
Live attenuated (plasmid-deficient) trachoma vaccine (NIH/NIAID)		Х	Kari <i>et al</i> . ³⁸

Table 3. Current Ct vaccine candidate development.³⁰

CDC, Centre for Disease Control; Ct, *Chlamydia trachomatis*; MOMP, major outer membrane protein; NIH, National Institutes of Health; NIAID, National Institute of Allergy and Infectious Diseases; pmps, polymorphic membrane proteins; UV, ultra violet.

compared across laboratories with respect to important clinical endpoints of product indications would be valuable.

 Although intramuscular immunization has worked effectively for preventing cervical human papillomavirus infection, it is unclear whether a Ct vaccine can be similarly administered to achieve protection, given the need for robust local T call immunity. An effective *Chlamydia* vaccine may need to induce strong transmucosal immunity with resident memory T cells in the genital tract

Self-adjuvanting biodegradable nanoparticles may be an option for Ct vaccine delivery.⁴⁰

HSV

Burden of disease

The WHO has estimated the global prevalence and incidence of herpes simplex virus type 2 infection.⁴¹ The estimated number of individuals aged 15–49 years who were living with herpes simplex virus type 2 infection worldwide in 2003 was 536 million, or roughly 16% of the world's population in this age group.

Pre-clinical vaccine development

Pre-clinical development of HSV vaccines utilises well-established animal models to test and screen promising candidate vaccines.⁴² The mouse model unfortunately does not recapitulate human infection, because genital reactivation does not occur and mortality is high in initial infection. The guinea pig model mimics infection in humans although promising vaccines in guinea pigs have not translated into efficacious vaccines in humans. Candidates in pre-clinical development include: a trivalent glycoprotein (gD/gC/gE), a live-attenuated vaccine, mutated HSV-1 and HSV-2, an inactivated HSV-2 in MPL/alum, an HSV-1 glycoprotein B lentiviral vector and an intranasal recombinant HSV-1 gB vaccine.

An early HSV type 2 vaccine, whilst shown to induce high levels of neutralising antibodies, nevertheless did not exhibit protection against HSV type 2 infection.⁴³ A possible explanation for this lies in the ability of HSV to become latent and to reactivate.44 Further experiments with plasmid DNA encoding full-length glycoprotein D from HSV type 2 (gD2), secreted gD2 or cytosolic gD2, when evaluated in mice and guinea pigs, whilst showing some evidence of reduction of acute disease and subsequent recurrent disease, neither resulted in improved virus clearance from the inoculation site nor significantly reduced recurrent disease when used as a therapeutic vaccine.45 HSV vaccine under development are shown in Table 4.46

More recently, intramuscular vaccination with a live-attenuated HSV vaccine (VC2) was shown to stimulate vaginal IL-17A dependent antimicrobial peptide immune responses in a guinea pig model of genital HSV type 2 infection.⁴⁷ The

Table 4. HSV candidate vaccines in development.46

Candidate name	Developer	Platform antigens	Phase I	Phase II
GEN-003	Genocea	Therapeutic bivalent subunit vaccine gD2/ICP4 with Matrix M2 adjuvant		Х
HerpV	Agenus	32 35-mer peptides, complexed with HSP, QS-21 adjuvant		Х
Codon optimised polynucleotide	Admedus	DNA vaccine: gD2 codon		Х
VCL-HB01/HM01	Vical	DNA vaccine gD2+/-UL46Vaxfectin		Х
HSV529	Sanofi	Replication-defective HSV-2	Х	
HSV, herpes simplex v	irus.			

live-attenuated virus had been engineered with disabling mutations in the protein that otherwise allows the virus to enter axons. The responses were described as robust and protective of animals from acquiring any appreciable disease after exposure to a highly virulent HSV type 2 strain.

Clinical vaccine development

The candidate GEN-003 therapeutic bivalent subunit vaccine for HSV type 2 has been shown to be clinically effective at doses of $60\mu g/50\mu g$ and $60\mu g/75\mu g$ in reducing viral shedding for up to 1 year in adults with symptomatic genital HSV type 2 infection.^{48,49} Lesion rate reductions were also observed at between 31% and 69%, although lesion rates also decreased in the placebo arm at 62%.

Controversies and next steps in herpes simplex virus vaccine development

HSV vaccine development has not been without controversy.⁵⁰ It was reported in 2018 that a US professor had injected individuals with his experimental live HSV type 2 vaccine, which had not been cleared from the regulatory viewpoint and which he administered without the necessary safety precautions.

One promising approach uses a vaccine to prime (with a subunit HSV vaccine) and a chemoattractant (intravaginal/topical imiquimod) to pull immune cells into the genital tract, the socalled prime and pull strategy, which shows that the strategy is able to decrease recurrent HSV more effectively than vaccine alone.⁵¹

HIV

Burden of disease

The WHO states that, since the beginning of the HIV/AIDs epidemic, 75 million people have been infected with HIV and approximately 32 million have died as a result of HIV.⁵² Furthermore, globally there were estimated to be 37.9 million people (95% confidence interval 32.7–44.0 million) living with HIV at the end of 2018 and an estimated 0.8% of adults aged 15–49 years are thought to be living with HIV.

Progress of vaccine development to date: pre-clinical and clinical

Some examples of HIV vaccine candidates under pre-clinical and early clinical development are shown in Table 5.

A vaccine that had therapeutic as well as preventative properties could be a worthwhile approach. The following are some examples of therapeutic vaccine trials currently underway:

 Profectus BioSciences: an exploratory randomised, 2-arm (1:1), double-blind, placebo-controlled trial evaluating the safety and efficacy of an HIV-1 multi-antigen plasmid DNA (HIV-MAG pDNA) vaccine prime in combination with an interleukin-12 plasmid DNA (IL-12 pDNA) adjuvant delivered by *in vivo* electroporation followed by a recombinant vesicular stomatitis virus vector containing the HIV-1 gag gene (rVSV HIV gag) booster vaccine in Table 5. Examples of HIV vaccine candidates under pre-clinical and early clinical development.

Candidate/mechanism	Reference
Envelope domains: antibodies neutralising HIV through binding to the envelope domains	Simek <i>et al</i> . ⁵³
HIV envelope trimers: prime boost	Rerks-Ngarm <i>et al</i> . ⁵⁴
Germline stimulating immunogens: to initiate the generation of broadly neutralising antibodies	Jardine <i>et al</i> . ⁵⁵
Broadly neutralising antibodies: passive immunisation	Moldt <i>et al</i> . ⁵⁶
Broadly neutralising antibodies: genetic delivery by incorporation into persistent viral vectors such as adeno associated virus	Lewis <i>et al.</i> ⁵⁷
Delivery of payloads by replicating viral vectors	Parks <i>et al.</i> ⁵⁸
HIV, human immunodeficiency virus.	

subjects on combination anti-retroviral therapy (cART) who started therapy during acute or early HIV infection.⁵⁹

- National Institute of Allergy and Infectious Diseases (NIAID): evaluation of the use of ALVAC vCP1452 vaccine in combination with IL-2 to increase HIV-specific immune responses in HIV-infected patients. ALVAC vCP1452 vaccine is a recombinant canarypox HIV vaccine that is administered as a monthly intramuscular injection. The IL-2 is self-administered as a daily subcutaneous injection at a low, non-toxic dose (2 million units).⁶⁰
- Sharon Ridler (University of Pittsburgh): evaluation of an investigational dendritic cell HIV vaccine made from a person's own white blood cells.⁶¹
- NIAID: an open label study to evaluate safety, tolerability, and immune response of a six-plasmid multiclade HIV-1 DNA vaccine, VRC-HIVDNA016-00-VP. The hypothesis is that this regimen will be safe for human administration and elicit immune responses to HIV-1 clade B Gag, Pol and Nef proteins, as well as clades A, B and C Env proteins. The primary objective is to evaluate the safety and tolerability in humans of the investigational vaccine and secondary objectives are to evaluate the immunogenicity of the vaccine as measured by intracellular cytokine staining (ICS) in the 4weeks after the second or

third dose of vaccine and the social impact of participating in an HIV-1 vaccine trial.⁶²

- NIAID: comparison of the safety, tolerability and immunogenicity of CH505TF gp120 produced from stably transfected cells to CH505TF gp120 produced from transiently transfected cells in healthy, HIV-1-uninfected adult participants.⁶³
- GlaxoSmithKline (GSK): study designed to determine whether administration of the GSK Biologicals HIV vaccine 732462 can lead to a reduction in viral load, and impact on the course of human immunodeficiency virus type 1 (HIV-1) infection. In HIV-1 infected persons who have not yet started antiretroviral therapy (ART), such a vaccine would potentially lead to a delay in the initiation of treatment.⁶⁴
- NIAID, University of California (Los Angeles), Inovio Pharmaceuticals: safety, immunogenicity and anti-reservoir activity of an electroporation-administered HIV DNA vaccine encoding GAG, POL and ENV proteins with IL-12 plasmid in HIVinfected adults on anti-retroviral therapy.⁶⁵
- NIAID: a rollover study of those who participated in study A5058s: now a phase II trial to evaluate the ability of vaccineinduced helper and CTL responses to control viremia in the absence of anti-retroviral therapy.⁶⁶
- French National Institute for Health and Medical Research-French National Agency

for Research on AIDS and Viral Hepatitis (Inserm-ANRS): EVHA T01 is an international, phaseI/II, multicentre, multi-stage, double-blind study that will evaluate at least three experimental arms compared with placebo control in HIV-1 infected participants to see if one or more has a clinically relevant impact on the control of viral replication. The three interventions are: GTU-multiHIV B-clade vaccine plus MVA HIV-B HIV vaccine; GTU-multiHIV B-clade vaccine plus MVA HIV-B HIV vaccine plus vedolizumab; vedolizumab.⁶⁷

There are open questions regarding therapeutic HIV vaccines: would the same vaccine serve for prevention of both sexual and maternal transmission, would the same vaccine serve for both developing and developed countries, would a therapeutic vaccine be limited to those with antiretroviral failure, would a therapeutic vaccine decrease transmission?

Ongoing phase IIb/III trials

Using the vaccinia vector with insertion of the HIV envelope protein gp160, the first phaseI human vaccine trial was performed in France in 1986.68 This did not demonstrate efficacy against HIV. A degree of efficacy was not demonstrated until 2009 in the phaseIII RV144 trial.54 This consisted of a priming with a canarypox vector (containing gag, pol and nef genes) followed by a boost with gp120, and demonstrated an efficacy of 31%. Since then, this approach continues to be explored. Meanwhile, the only other approach under consideration uses a mosaic vaccine candidate.⁶⁹ This involves a public-private partnership with Janssen, the NIAID of the NIH, the HIV Vaccine Trials Network, and the US Army Medical Research and Development Command. One important aim of this mosaic approach is to address the global genetic diversity of HIV.

Challenges of vaccine development

The advent, deployment, uptake and impact of anti-retroviral medications is changing the HIV landscape, especially by dramatically reducing sexual transmission.^{70,71} Maternal transmission to the unborn baby is also potentially completely controllable with these medications, and this in turn has future positive implications for sexual

transmission by the time these infants reach a dulthood. $^{72}\,$

In this context, is there a need for an HIV vaccine? If one were available, to whom would it be administered? What efficacy would be acceptable? Would its use detract from the successful use of anti-retroviral medications?

Next steps

The next steps involve progressing both vaccine approaches: the prime-boost approach and the mosaic approach, completing further clinical trials, deciding what would be an 'acceptable' efficacy, and planning how to distribute a successful vaccine to those who need it the most.

Zika virus

Burden of disease

At the beginning of 2020, Zika virus is quiescent; the current burden is the large cohort of infants who were infected maternally in 2015–2017.⁷³

Progress of vaccine development to date

Whilst being transmitted by mosquitos, Zika virus is also established as a sexually transmitted infection,⁷⁴ and Zika virus RNA has been demonstrated to be commonly present in the semen of males with symptomatic Zika virus infection, with persistence in some for more than 6 months.⁷⁵ Zika virus vaccines under development are summarised in Table 6 (WHO 2019).⁷⁶

Challenges and next steps in vaccine development

Whilst there is a healthy Zika virus vaccine pipeline, questions over the precise role of such a vaccine remain. Would the main purpose be to prevent vector-borne Zika virus infection, or sexually transmitted Zika virus infection, or both? Would it be used in routine vaccination or reserved for use in outbreaks? And more fundamentally, can a correlate of protection be developed and what is the best development pathway?

The development of vaccines able to prevent both vertical transmission and congenital Zika virus syndrome is challenging, and clinical trials for

Candidate	Platform	Immunogen	Adjuvant
GLS-5700	DNA	prME	None
AGS-v	Peptide	Mosquito salivary proteins	ISA-51
MV-Zika	Recombinant viral vector	prME	None
mRNA-1325	mRNA	prME	None
VRC-ZKADNA085-00-VP and 090-00-VP	DNA	prME	None
ZIKV PIV	Inactivated whole target organism	Whole virus	Alum
PIZV or TAK-426	Inactivated whole target organism	Whole virus	Alum

Table 6. Zika virus vaccines under development (WH0

MV, measles virus; PIV, purified inactivated virus; PIZV, purified inactivated Zika vaccine; prME, pre-membrane and envelope ; WHO, World Health Organisation; ZIKV, Zika virus.

testing vaccine efficacy in relation to foetus protection are going to take time.⁷⁷

Delivery of protection against STIs

How will it be possible to deliver protection to those at risk of STIs, to target populations most in need? The first clue lies in HIV, with the facilitation of testing in hard-to-reach groups.⁷⁸ Programmes like this could readily be adapted to vaccination rather than testing. The next clue lies in the need for education and raising of awareness in 'implementers' of progammes.⁷⁹ Programmes for education and raising awareness have a universal relevance to vaccination against all forms of sexually transmitted infections. The third clue lies in being capable of rapidly measuring the impact and success of a vaccination programme against STIs.⁸⁰

The predicted levels of coverage required for protection by putative vaccines against the STIs gonorrhoea, syphilis, chlamydia, herpes simplex virus, human immunodeficiency virus and Zika virus vary widely according to epidemiology, burden of disease, extent and frequency of transmission, silent infection, ability to cause repeated infections and of course the quality of the vaccine response, and no one model can be applied. In an outbreak situation such as occurred with Zika virus, it is likely that high coverage levels would be required. For maternal transmission, then the coverage could potentially be more targeted. And for sometimes silent and often common infections, universal coverage would be ideal. Could delivery of vaccines against STIs be performed in a cost-effective way? Based on current delivery models for human papillomavirus vaccine and current screening for STIs, and taking account of regional incidence data, the conclusions of a large costing study are that increased investment and innovative financing are required, especially in low- and middle-income countries, and that there is a need for synergising with other health programmes.⁸¹ And the conclusion of a costeffectiveness study into a Ct vaccination programme in the US is that it would result in increased costs to the healthcare system but with the benefit of significantly averting morbidity and mortality.⁸²

Conclusion

Whilst there are considerable costs associated with (i) further research and development into vaccines against STIs and (ii) their future delivery and deployment, nevertheless, in the context of antimicrobial resistance and the potential for resistance against anti-virals, and the potential benefits, further investment in vaccines against STIs is warranted.

Roadmap: general principles

- Need for prevention of latency, persistence and repeat infections;
- 2) Reduction in transmission of STIs would reduce the global burden;
- 3) Therapeutic activity advantageous over preventative activity alone;

- 4) Prevention of congenital and neonatal infections also an added benefit;
- 5) Added value of the prevention of long-term consequences of STIs;
- 6) It may be possible to re-purpose 'old' vaccines for new indications;
- 7) Challenge of determining the target populations for STI vaccination

Conflict of interest statement

EDG McIntosh is an employee of Merck Sharp & Dohme (MSD), which manufactures vaccines. The views expressed herein are his own and do not necessarily reflect the views of Imperial College or MSD.

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References

- World Health Organization. Sexually transmitted infections, https://www.who.int/news-room/factsheets/detail/sexually-transmitted-infections-(stis) (2019). (accessed 14 June 2019).
- 2. Vincent LR and Jerse AE. Biological feasibility and importance of a gonorrhoea vaccine for global public health. *Vaccine* 2019; 37: 7419–7426.
- Cameron CE and Lukehart SA. Current status of syphilis vaccine development: needs, challenges, prospects. *Vaccine* 2014; 32: 1602–1609.
- Batteiger BE, Xu F, Johnson RE, et al. Protective immunity to Chlamydia trachomatis genital infection: evidence from human studies. *J Infect Dis* 2010; 201(Suppl. 2): S178–S189.
- Biasin M, Trabattoni D, Rossigno JF, et al. Immune correlates of protection against HIV infection and how to elicit them. Mucosal Immunology 2017; 10: 827–828.
- Veazey RS, Pilch-Cooper HA, Hope TJ, et al. Prevention of SHIV transmission by topical IFN-β treatment. *Mucosal Immunol* 2016; 9: 1568–1536.
- 7. Yim KC, Carroll CJ, Tuyama A, *et al.* The cotton rat provides a novel model to study genital herpes

infection and to evaluate preventive strategies. \Im Virol 2005; 79: 14632–14639.

- Veazey RS and Lackner AA. Nonhuman primate models and understanding the pathogenesis of HIV infection and AIDS. *ILAR J* 2017; 58: 160–171.
- 9. Gottlieb SL and Johnston C. Future prospects for new vaccines against sexually transmitted infections. *Curr Opin Infect Dis* 2017; 30: 77–86.
- Rowley J, Vander Hoorn S, Korenromp E, et al. Chlamydia, gonorrhoea, trichomoniasis and syphilis: global prevalence and incidence estimates, 2016. Bull World Health Organ 2019; 97: 548–562.
- Gulati S, Pennington MW, Czerwinski A, et al. Preclinical efficacy of a lipooligosaccharide peptide mimic candidate gonococcal vaccine. *mBio* 2019; 10: e02552-19.
- Gottlieb SL, Jerse AE, Delany-Moretlwe S, et al. Advancing vaccine development for gonorrhoea and the global STI roadmap. Sex Health 2019; 16: 426–432.
- Semchenko EA, Tan A, Borrow R, et al. The serogroup B meningococcal vaccine Bexsero[®] elicits antibodies to Neisseria gonorrhoeae. Clin Infect Dis 2019; 69: 1101–1111.
- Petousis-Harris H. Impact of meningococcal group B OMV vaccines, beyond their brief. *Hum* Vaccin Immunother 2017; 14: 1058–1063.
- Institute of Environmental Science and Research Ltd. Sexually transmitted infections in New Zealand: annual surveillance report 2014. Porirua (New Zealand), https://surv.esr.cri.nz/ surveillance/annual_sti.php?we_objectID=4844 (2015). (accessed 9 September 2019).
- Régnier S and Huels J. Potential impact of vaccination against Neisseria meningitidis on Neisseria gonorrhoeae in the United States: results from a decision-analysis model. *Hum Vaccin Immunother* 2014; 10: 3737–3745.
- 17. Petousis-Harris H and Radcliff FJ. Exploitation of Neisseria meningitidis group B OMV vaccines against N. gonorrhoeae to inform the development and deployment of effective gonorrhoea vaccines. *Front Immunol* 2019; 10: 683.
- ClinicalTrials.gov. Identifier NCT04094883. Study to assess gonorrhoeae immune responses induced by a *N. meningitidis* vaccine (4CMenB), https://clinicaltrials.gov/ct2/show/NCT04094883. (accessed 20 September 2019).
- 19. Australian New Zealand Clinical Trials Registry. ANZTR identifier ACTRN12619001478101.

MenGO: does the licensed meningococcal vaccine Bexsero[®] provide cross-protection against gonorrhoea in gay and bisexual men? https://www.anzctr.org.au/Trial/Registration/TrialReview.aspx?id=376715. (accessed 31 October 2020).

- Miller JN. Immunity in experimental syphilis. VI. Successful vaccination of rabbits with Treponema pallidum, Nichols strain, attenuated by irradiation. *J Immunol* 1973; 110: 1206–1215.
- Lithgow KV and Cameron CE. Vaccine development for syphilis. *Expert Rev Vaccines* 2017; 16: 37–44.
- 22. Kao WA, Petrosova H, Ebady R, *et al.* Identification of Tp0751 (Pallilysin) as a Treponema pallidum vascular adhesin by heterologous expression in the Lyme disease spirochete. *Sci Rep* 2017; 7: 1538.
- 23. Parveen N, Fernandez MC, Haynes AM, *et al.* Non-pathogenic borrelia burgdorferi expressing Treponema pallidum TprK and Tp0435 antigens as a novel approach to evaluate syphilis vaccine candidates. *Vaccine* 2019; 37: 1807–1818.
- 24. Zheng K, Zu M, Xiao Y, *et al.* Immunogenicity and protective efficacy against Treponema pallidum in New Zealand rabbits immunized with plasmid DNA encoding flagellin. *Emerg Microbes Infect* 2018; 7: 177.
- Cameron C. Syphilis vaccine development: requirements, challenges, and opportunities. Sex Transm Dis 2018; 45: S17–S19.
- Centre for Disease Control and Prevention. The Tuskegee timeline, https://www.cdc.gov/tuskegee/ timeline.htm. (accessed 13 August 2019).
- Poston TB, Lee DE, Darville T, et al. Cervical cytokines associated with *Chlamydia trachomatis* susceptibility and protection. *J Infect Dis* 2019; 20: 330–339.
- Phillips S, Quigley BL and Timms P. Seventy years of Chlamydia vaccine research – limitations of the past and directions for the future. *Front Microbiol* 2019; 10: 70.
- 29. Poston TB and Darville T. Chlamydia trachomatis: protective adaptive responses and prospects for a vaccine. *Curr Top Microbiol Immunol* 2018; 412: 217–237.
- Poston TB, Gottlieb SL and Darville T. Status of vaccine research and development of vaccines for Chlamydia trachomatis infection. *Vaccine* 2017; 37: 7289–7294.
- 31. Olsen AW, Follmann F, Erneholm K, *et al.* Protection against Chlamydia trachomatis infection

and upper genital tract pathological changes by vaccine-promoted neutralizing antibodies directed to the VD4 of the major outer membrane protein. \mathcal{J} *Infect Dis* 2015; 212: 978–989.

- 32. Bøje S, Olsen AW, Erneholm K, et al. A multisubunit Chlamydia vaccine inducing neutralizing antibodies and strong IFN-y⁺ CMI responses protects against a genital infection in minipigs. *Immunol Cell Biol* 2016; 94: 185–195.
- Olsen AW, Lorenzen EK, Rosenkrands I, et al. Protective effect of vaccine promoted neutralizing antibodies against the intracellular pathogen Chlamydia trachomatis. *Front Immunol* 2017; 8: 1652.
- Fattom AI. Development of a nanoemulsionbased vaccine for Chlamydia infection. http:// grantome.com/grant/NIH/R43-AI134168-01A1. (accessed 9 September 2019).
- Karunakaran KP, Yu H, Jiang X, et al. Outer membrane proteins preferentially load MHC class II peptides: implications for a Chlamydia trachomatis T cell vaccine. Vaccine 2015; 33: 2159–2166.
- 36. Stary G, Olive A, Radovic-Moreno AF, *et al.* A mucosal vaccine against Chlamydia trachomatis generates two waves of protective memory T cells. *Science* 2015; 348: aaa8205.
- Garmory HS, Leckenby MW, Griffin KF, et al. Antibiotic-free plasmid stabilization by operatorrepressor titration for vaccine delivery by using live Salmonella enterica serovar typhimurium. *Infect Immun* 2005; 73: 2005–2011.
- Kari L, Whitmire WM, Olivares-Zavaleta N, et al. A live-attenuated Chlamydial vaccine protects against trachoma in non-human primates. J Exp Med 2011; 208: 2217–2223.
- Zhong G, Brunham RC, de la Maza L, et al. National institute of allergy and infectious diseases workshop report: 'Chlamydia vaccines: the way forward'. Vaccine 2019; 37: 7346–7354.
- 40. Sahu R, Verma R, Dixit S, *et al.* Future human Chlamydia vaccine: potential of self-adjuvanting biodegradable nanoparticles as safe vaccine delivery vehicles. *Expert Rev Vaccines* 2019; 17: 217–227.
- 41. Looker KJ, Garnett GP and Schmid GP. An estimate of the global prevalence and incidence of herpes simplex virus type 2 infection. *Bull World Health Organ* 2008; 86: 805–812.
- 42. World Health Organization. Status of vaccine research and development of vaccines for herpes simplex virus prepared for WHO PD-VAC, https://www.who.int/immunization/research/

meetings_workshops/HSV_vaccineRD_Sept2014. pdf. (accessed 9 September 2019).

- Corey L, Langenberg AG, Ashley R, et al. Recombinant glycoprotein vaccine for the prevention of genital HSV-2 infection. Two randomized controlled trials. *JAMA* 1999; 282: 331–340.
- Mascola JR. Herpes simplex virus vaccines why don't antibodies protect? *JAMA* 1999; 282: 379–380.
- Strasser JE, Arnold RL, Pachuk C, et al. Herpes simplex virus DNA vaccine efficacy: effect of glycoprotein D plasmid constructs. *J Infect Dis* 2000; 182: 1304–1210.
- Johnston C, Gottlieb SL and Wald A. Status of vaccine research and development of vaccines for herpes simplex virus. *Vaccine* 2016; 34: 2948–2952.
- 47. Stanfield BA, Rider PJF, Caskey J, et al. Intramuscular vaccination of guinea pigs with the live-attenuated human herpes simplex vaccine VC2 stimulates a transcriptional profile of vaginal TH17 and regulatory Tr1 responses. Vaccine 2018; 36: 2842–2849.
- Bernstein DI, Wald A, Warren T, et al. Therapeutic vaccine for genital herpes simplex virus-2 infection: findings from a randomized trial. J Infect Dis 2017; 215: 856–864.
- Van Wagoner N, Fife K, Leone PA, et al. Effects of different doses of GEN-003, a therapeutic vaccine for genital herpes simplex virus-2, on viral shedding and lesions: results of a randomized placebo-controlled trial. J Infect Dis 2018; 218: 1890–1899.
- Dyer O. FDA launches criminal investigation of unauthorised herpes vaccine trial. *BMJ* 2018; 361: k1753.
- Bernstein DI, Cardin RD, Bravo FJ, et al. Successful application of prime and pull strategy for a therapeutic HSV vaccine. NPJ Vaccines 2019; 4: 33.
- 52. World Health Organization. HIV/AIDS Summary of the global HIV epidemic 2018, https://www.who. int/gho/hiv/en/. (accessed 10 September 2019).
- 53. Simek MD, Rida W, Priddy FH, et al. Human immunodeficiency virus type 1 elite neutralizers: individuals with broad and potent neutralizing activity identified by using a high-throughput neutralization assay together with an analytical selection algorithm. J Virol 2009; 83: 7337–7348.
- 54. Rerks-Ngarm S, Pitisuttithum P, Nitayaphan S, *et al.* Vaccination with Alvac and AIDSVAX to

prevent HIV-1 infection in Thailand. New Engl J Med 2009; 361: 2209–2220.

- 55. Jardine J, Julien JP, Menis S, *et al.* Rational HIV immunogen design to target specific germline B cell receptors. *Science* 2013; 340: 711–716.
- 56. Moldt B, Rakasz EG, Schultz N, et al. Highly potent HIV-specific antibody neutralization in vitro translates into effective protection against mucosal SHIV challenge in vivo. Proc Natl Acad Sci USA 2012; 109: 18921–18925.
- 57. Lewis AS, Chen R, Montefiori DC, et al. Generation of neutralizing activity against human immunodeficiency virus type 1 in serum by antibody gene transfer. J Virol 2002; 76: 8769–8775.
- Parks CL, Picker LJ and King CR. Development of replication-competent viral vectors for HIV vaccine delivery. *Curr Opin HIV AIDS* 2013; 8: 402–411.
- ClinicalTrials.gov. Identifier NCT01859325. Therapeutic vaccine for HIV, https://clinicaltrials. gov/ct2/show/NCT01859325. (accessed 20 March 2019).
- 60. ClinicalTrials.gov. Identifier NCT00056797. Therapeutic HIV vaccine and interleukin-2 to increase the immune system's response to HIV, https://clinicaltrials.gov/ct2/show/NCT00056797. (accessed 20 March 2019).
- ClinicalTrials.gov. Identifier NCT03758625. Comparison of dendritic cell-based therapeutic vaccine strategies for HIV functional cure (DC-HIV04), https://clinicaltrials.gov/ct2/show/ NCT03758625. (accessed 20 March 2019).
- ClinicalTrials.gov. Identifier NCT00089531. Candidate HIV vaccine, https://clinicaltrials.gov/ ct2/show/NCT00089531. (accessed 20 March 2019).
- 63. ClinicalTrials.gov. Identifier NCT03856996. A clinical trial in healthy, HIV-1-uninfected adult participants to compare the safety, tolerability and immunogenicity of CH505TF gp120 produced from stably transfected cells to CH505TF gp120 produced from transiently transfected cells, https://clinicaltrials.gov/ct2/ show/NCT03856996. (accessed 20 March 2019).
- ClinicalTrials.gov. Identifier NCT01218113. Efficacy and safety of GSK biologicals HIV vaccine in antiretroviral therapy (ART)-naïve HIV-1 infected persons, https://clinicaltrials.gov/ct2/show/ NCT01218113. (accessed 20 March 2019).
- 65. ClinicalTrials.gov. Identifier NCT03606213. Therapeutic vaccination in treated HIV disease,

https://clinicaltrials.gov/ct2/show/NCT03606213. (accessed 20 March 2019).

- ClinicalTrials.gov. Identifier NCT00050063. Effects of therapeutic HIV vaccination on control of HIV after discontinuation of anti-HIV drugs, https://clinicaltrials.gov/ct2/show/NCT00050063. (accessed 20 March 2019).
- ClinicalTrials.gov. Identifier NCT02972450. An HIV vaccine trial in individuals who started ART during primary or chronic infection (EHVAT01), https://clinicaltrials.gov/ct2/show/NCT02972450. (accessed 20 March 2019).
- Zagury D, Léonard R, Fouchard M, et al. Immunization against AIDS in humans. Nature 1987; 326: 249–250.
- ClinicalTrials.gov. Identifier NCT02919306. Safety and efficacy study of vaccine schedule with Ad26.Mos.HIV and MVA-Mosaic in human immunodeficiency virus (HIV)-infected adults, https://clinicaltrials.gov/ct2/show/NCT02919306 ?term=vaccine&cond=Human+Immunodeficien cy+Virus&draw=2&rank=1. (accessed 20 March 2019).
- Rodger AJ, Cambiano V, Bruun T, *et al.* Sexual activity without condoms and risk of HIV transmission in serodifferent couples when the HIV-positive partner is using suppressive antiretroviral therapy. *JAMA* 2016; 316: 171–181.
- Rodger AJ, Cambiano V, Bruun T, et al. Risk of HIV transmission through condomless sex in serodifferent gay couples with the HIV-positive partner taking suppressive antiretroviral therapy (PARTNER): final results of a multicentre, prospective, observational study. *Lancet* 2019; 393: 2428–2438.
- 72. Teasdale CA, Marais BJ and Abrams EJ. HIV: prevention of mother-to-child transmission. *BMB Clin Evid* 2011; 2011: 0909.
- 73. Staples JE, Martin SW and Fischer M. Zika: Travel-related infections. In: Brunette GW and Nemhauser JB (eds) *CDC Yellow Book 2020:*

Health Information for International Travel New York: Oxford University Press, 2019.

- 74. Counotte MJ, Kim CR, Wang J, *et al.* Sexual transmission of Zika virus and other flaviviruses: a living systematic review. *PLoS Med* 2018; 15: e1002611.
- Mead PS, Duggal NK, Hook SA, et al. Zika virus shedding in semen in symptomatic infected men. N Engl J Med 2018; 378: 1377–1385.
- World Health Organization. Welcome to the WHO vaccine trial tracker, https://docs.google. com/spreadsheets/d/19otvINcayJURCMg76xWO 4KvuyedYbMZDcXqbyJGdcZM/pubhtml (2019). (accessed 9 September 2020).
- Sakkas H, Bozidis P, Giannakopoulos X, et al. An update on sexual transmission of Zika virus. Pathogens 2018; 7: pii: E66.
- Mutch AJ, Lui CW, Dean J, et al. Increasing HIV testing among hard-to-reach groups: examination of RAPID, a community-based testing service in Queensland, Australia. BMC Health Serv Res 2017; 17: 310.
- 79. Siu JY, Lee A and Chan PKS. Schoolteachers' experiences of implementing school-based vaccination programs against human papillomavirus in a Chinese community: a qualitative study. *BMC Public Health* 2019; 19: 1514.
- Jacot-Guillarmod M, Pasquier J, Greub G, et al. Impact of HPV vaccination with Gardasil[®] in Switzerland. BMC Infect Dis 2017; 17: 790.
- Korenromp EL, Wi T, Resch S, et al. Costing of national STI program implementation for the global STI control strategy for the health sector, 2016-2021. PLoS One 2017; 12: e0170773.
- Ditkowsky J, Rahman A, Hammerschlag MR, et al. Cost-benefit analysis of a Chlamydia trachomatis vaccine program in adolescent girls in the United States. *J Pediatric Infect Dis Soc* 2018; 7: 296–302.

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