

Importance of long non-coding RNAs in the pathogenesis, diagnosis, and treatment of myocardial infarction

Maryam Zolfaghari Dehkharhani ^{a,*}, Safa Mousavi ^a, Nazanin Kianifard ^b, Amin Fazlzadeh ^a, Hamid Parsa ^b, Ali Tavakoli Pirzaman ^c, Andarz Fazlollahpour-Naghibi ^c

^a School of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran

^b School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

^c School of Medicine, Babol University of Medical Sciences, Babol, Iran



ARTICLE INFO

Keywords:

Long non-coding RNAs
Myocardial infarction
Therapeutic
Diagnostic

ABSTRACT

Myocardial infarction (MI), a major global cause of mortality and morbidity, continues to pose a significant burden on public health. Despite advances in understanding its pathogenesis, there remains a need to elucidate the intricate molecular mechanisms underlying MI progression. Long non-coding RNAs (lncRNAs) have emerged as key regulators in diverse biological processes, yet their specific roles in MI pathophysiology remain elusive. Conducting a thorough review of literature using PubMed and Google Scholar databases, we investigated the involvement of lncRNAs in MI, focusing on their regulatory functions and downstream signaling pathways. Our analysis revealed extensive dysregulation of lncRNAs in MI, impacting various biological processes through diverse mechanisms. Notably, lncRNAs act as crucial modulators of gene expression and signaling cascades, functioning as decoys, regulators, and scaffolds. Furthermore, studies identified the multifaceted roles of lncRNAs in modulating inflammation, apoptosis, autophagy, necrosis, fibrosis, remodeling, and ischemia-reperfusion injury during MI progression. Recent research highlights the pivotal contribution of lncRNAs to MI pathogenesis, offering novel insights into potential therapeutic interventions. Moreover, the identification of circulating lncRNA signatures holds promise for the development of non-invasive diagnostic biomarkers. In summary, findings underscore the significance of lncRNAs in MI pathophysiology, emphasizing their potential as therapeutic targets and diagnostic tools for improved patient management and outcomes.

1. Introduction

Myocardial infarction (MI) arises from thrombus formation obstructing an artery or complications within a bypass graft, leading to acute myocardial ischemia, heart failure, and mortality [1,2]. Clinically, MI presents with severe chest pain, dyspnea, nausea, and sweating, accompanied by complications such as angina, transient ischemic episodes, arrhythmias, and congestive heart failure, all posing significant risks to patient survival. A comprehensive clinical approach for suspected MI includes tailored management, supportive care, and secondary prevention strategies, with prognosis influenced by factors like symptom acuity, comorbidities, and treatment response [3]. Acute MI, the severest form of coronary artery disease, accounts for millions of deaths annually globally, with mortality rates declining due to lifestyle changes and evidence-based therapies. However, MI remains a substantial global health burden, affecting millions of people annually and

imposing significant economic costs [4–6].

Coronary artery obstruction diminishes myocardial blood supply, leading to ischemic damage. Despite advances in revascularization therapies, understanding the intricate molecular mechanisms underlying MI is ongoing [7,8]. Recently, the non-coding genome segment gained recognition for its role in diverse physiological and pathological processes [9,10]. Long non-coding RNAs (lncRNAs) attract interest for their critical involvement in gene regulation and cellular homeostasis [11–13]. Emerging evidence indicates lncRNAs' significant implications in MI development, regulating key biochemical pathways in heart function, ischemic injury, and remodeling at transcriptional, post-transcriptional, and epigenetic levels [14–17]. The involvement of lncRNAs in cardiac remodeling is a key mechanism via which they contribute to the onset and progression of MI [14,18]. Their involvement in cardiac remodeling is pivotal in MI progression, influencing fibrosis, hypertrophy, and contractility compromise post-ischemia,

* Corresponding author.

E-mail address: maryam.zd35@gmail.com (M. Zolfaghari Dehkharhani).

through regulation of signaling pathways like fibrosis, hypertrophy, apoptosis, and angiogenesis [19–21].

Furthermore, lncRNAs hold promise as diagnostic biomarkers for MI due to their remarkable stability and presence in physiological fluids like blood and urine [17,22]. Altered expression of specific lncRNAs in MI patients suggests their potential as non-invasive and highly sensitive diagnostic tools. Identifying and validating lncRNA signatures could aid in early diagnosis, risk assessment, and prognosis of MI, facilitating timely intervention and personalized therapeutic strategies [23,24]. Beyond diagnostics, lncRNAs emerge as attractive targets for therapeutic interventions in MI, offering the ability to modulate pathological pathways. Targeting specific lncRNAs, which regulate diverse biological processes, holds potential for reducing myocardial damage, preserving cardiac function, and promoting cardiac repair post-MI. Strategies such as antisense oligonucleotides, small interfering RNAs (siRNAs), and genome editing technologies offer avenues for modifying lncRNA production or function [25–27].

This review aims to evaluate the intricate role of lncRNAs in MI pathogenesis, exploring their molecular influence on cardiac remodeling. We will also assess the diagnostic potential of lncRNAs as non-invasive MI biomarkers and discuss their therapeutic implications. Specifically, we'll focus on the benefits and challenges of targeting specific lncRNAs in MI treatment.

2. Long non-coding RNAs

LncRNAs are transcripts exceeding 200 nucleotides in length that lack protein-coding capacity. Initially considered as non-functional transcriptional noise, recent data highlight their diverse regulatory roles across biological processes, employing multiple modes of action [11,28,29]. Despite lower expression levels compared to messenger RNAs (mRNAs), lncRNAs exhibit tissue and cell type-specific expression patterns [30]. While conservation is generally lower compared to protein-coding genes, certain lncRNAs display substantial conservation, indicating conserved functional elements [31]. Genomic composition includes exons, introns, and regulatory elements, often located near protein-coding genes or overlapping with their transcriptional units [32]. Functionally, lncRNAs reside in the nucleus and cytoplasm, regulating gene expression through epigenetic, transcriptional, and post-transcriptional mechanisms [33]. At the epigenetic level, lncRNAs interact with chromatin-modifying complexes, leading to histone modifications such as methylation and acetylation [34]. In transcriptional regulation, they engage with regulatory proteins and RNA polymerases to either promote or inhibit target gene transcription. Post-transcriptionally, lncRNAs influence mRNA stability, translation efficiency, and protein activity and localization [35]. Well-known examples include Xist lncRNA, which facilitates X chromosome inactivation by altering chromatin structure [36]; HOTAIR lncRNA, involved in recruiting histone modification complexes such as the Polycomb repressive complex 2 (PRC2) for H3K27 methylation and interacting with genes targeted for H3K4 demethylation via the LSD1/CoREST/REST complex [37]; and MALAT1 lncRNA, which regulates alternative splicing by binding to SR splicing proteins [38]. Genomic and molecular methodologies have led to significant progress in identifying and analyzing lncRNAs, revealing their importance in cellular homeostasis and disease progression [39,40]. Further investigation is crucial for identifying therapeutic targets and strategies.

3. LncRNAs and cardiovascular diseases

LncRNAs are key regulators in a wide range of biological processes, including cardiovascular disease. They influence gene expression through mechanisms such as chromatin modification, transcriptional, and post-transcriptional regulation [11,41,42]. Over the past decade, research has underscored the indispensable role of lncRNAs in cardiovascular development and function [43], with myocardial tissues

displaying significant expression of several lncRNAs [44–46]. These molecules exhibit dynamic transcription patterns during cardiac myocyte development, differentiation, and maturation stages [47–49]. Studies have elucidated their capacity to control the expression of various genes within cardiomyocytes [50,51]. Furthermore, aberrant expression of specific lncRNAs has been linked to diverse cardiovascular disorders (Fig. 1) [52,53].

Cardiac development is a complex process requiring precise gene expression control [54]. Extensive research has identified key lncRNAs essential for heart development and function [55]. For example, Braveheart (Bvht) lncRNA promotes cardiovascular lineage commitment from stem cells; its depletion in mouse embryonic stem cells downregulates genes crucial for mesodermal and cardiac development, impeding cardiovascular differentiation [56]. Bvht has no human homologue. Fetal-lethal non-coding developmental regulatory RNA (FENDRR), another important lncRNA in cardiogenesis, is crucial for septum and valve establishment during embryonic heart development; mice lacking FENDRR exhibit significant cardiac abnormalities, highlighting its role in cardiovascular morphogenesis [57]. In the adult heart, lncRNAs regulate homeostasis and stress responses [58]. MALAT1 lncRNA, highly expressed in endothelial cells lining blood arteries, contributes to angiogenesis, particularly after MI, aiding in cardiac tissue restoration [59–61]. Under stress conditions like ischemia, the cardioprotective lncRNA H19 expression increases, supporting cardiomyocyte survival [62].

Numerous lncRNAs significantly influence key cellular mechanisms in the cardiovascular system, including inflammation, oxidative stress, metabolism, and apoptosis, all crucial factors in cardiovascular disorder development [63–65]. For example, CHRF lncRNA inhibits inflammation by sequestering miR-489, reducing its inhibitory effect on secretory leukocyte protease inhibitor (SLPI) [66]. Cardiomyocyte apoptosis plays a pivotal role in MI and heart failure; several pro-apoptotic lncRNAs are promising therapeutic targets [67]. Increased expression of MI-associated transcript (MIAT) during myocardial ischemia/reperfusion injury activates apoptosis via interaction with miR-22-3p and subsequent SP1 activation [68,69]. Another example is linc-p21, which promotes cardiomyocyte apoptosis by activating the p53 signaling pathway; its suppression enhances heart function in animal models of MI [70,71]. Metabolic lncRNAs like LIPCAR can be dysregulated, leading to energy shortages in cardiovascular disorders [52,72]. LncRNAs play a crucial role in atrial fibrillation (AF) development [73], with fibrosis being a significant event in its progression. H19 accelerates fibrosis by sequestering let-7 and upregulating TGF-β1 signaling [74]. Recent studies have identified lncRNAs associated with AF, including CHAER, FENDRR, and Bvht, which contribute to arrhythmogenesis due to abnormal expression [75,76]. Recent transcriptomic advancements have illuminated lncRNA functions in biology and disease. Their precise regulatory roles in development, stress response, inflammation, and metabolism underscore their significance as contributors and potential therapeutic targets in cardiovascular disorders.

3.1. LncRNA molecular mechanisms of action in myocardial infarction

Recent studies underscore the pivotal role of lncRNAs as regulators of gene expression and cellular mechanisms in both the onset and progression of MI. These mechanisms, including inflammation, fibrosis, and impaired regeneration, are elucidated in detail in Table 1. It is increasingly evident that lncRNAs modulate diverse biological processes via signal transduction, decoy activity, guidance, and scaffolding functions (see Fig. 2) [77].

Signaling is crucial in lncRNA-mediated effects during MI. LncRNAs act as signaling molecules, responsive to pathogenic stimuli, and modulate gene expression and signaling pathways through cis or trans activities [78,79]. For instance, the downregulation of lncRNA FAF in cardiomyocytes under ischemia-hypoxia and MI conditions regulates FGF9/FGFR2 expression, inhibiting apoptosis via the PI3K/Akt survival

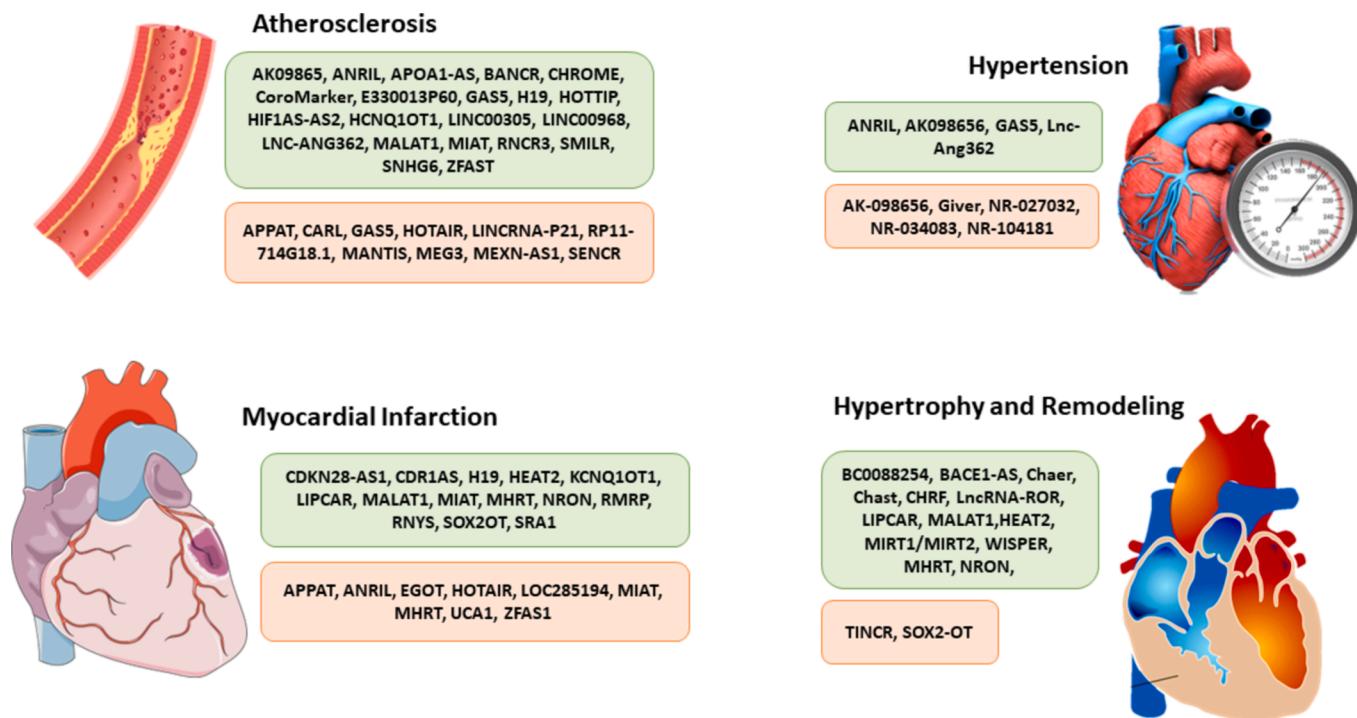


Fig. 1. Cardiovascular disease processes encompass the participation of long noncoding RNAs. There is a growing body of evidence indicating that long noncoding RNAs (lncRNAs) play a significant role as response and regulatory molecules in cardiac pathophysiologies. Green boxes showed upregulation lncRNAs and red boxes showed down regulation lncRNAs. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

pathway [80]. Similarly, reduced expression of lncRNA SLC8A1-AS1 in MI patients prevents myocardial injury by downregulating SLC8A1 and activating the cGMP-PKG pathway, reducing pro-inflammatory cytokine production and infarct size [81].

Decoy utilization represents a significant strategy through which lncRNAs exert cardioprotective effects. Acting as molecular decoys or competitive endogenous RNAs (ceRNAs), lncRNAs interact with microRNAs (miRNAs) and mRNAs, thereby regulating gene expression [82]. For example, the lncRNA CARL serves as a ceRNA for miR-539, leading to the downregulation of the anti-apoptotic factor PHB2. By sequestering miR-539, CARL increases PHB2 levels, suppressing mitochondrial fission and apoptosis post-myocardial infarction [83]. Similarly, the lncRNA APF interacts with miR-188-3p, relieving suppression on the autophagy regulator ATG7. This interaction enhances autophagic flux, protecting cardiomyocytes from injury after myocardial infarction [84]. Additionally, lncRNAs regulate necrosis by employing decoy mechanisms. Binding of necrosis-related factor (NRF) to miR-873 inhibits miR-873, relieving repression on necrotic mediators RIPK1 and RIPK3. This reduces cardiomyocyte necrosis and infarct size in ischemia/reperfusion injury models [85].

In addition to their signaling and decoy functions, lncRNAs can guide the recruitment of epigenetic modifiers and transcription factors to specific genomic loci, enabling precise transcriptional regulation in MI contexts [86]. For example, the lncRNA Kcnq1ot1 recruits DNMT1 to the promoter region of the cell viability regulator RUNX3, stimulating cardiac microvascular endothelial cell proliferation and inflammation in MI [87]. Similarly, the lncRNA CPR interacts with DNMT3A to methylate MCM3's promoter, enhancing cardiomyocyte proliferation and repair post-MI [88]. Additionally, lncRNA Sarrah forms a triple helix structure with the NRF2 gene promoter, recruiting transcriptional activators CRIP2 and p300 to enhance NRF2 activity, thereby improving cardiomyocyte survival in MI models [89].

Additionally, lncRNAs serve as molecular scaffolds, participating in regulatory complex formation for transcriptional and epigenetic control in MI due to their secondary structures [90,91]. For instance, the

lncRNA Ahit interacts with SUZ12, a core component of PRC2, facilitating its transport to the MEF2A promoter region, initiating repression via H3K27 methylation to regulate cardiac hypertrophy [92]. Similarly, ANRIL assembles histone modifier complexes with WDR5 and HDAC3, increasing reactive oxygen species (ROS) levels and promoting phenotypic switching in vascular smooth muscle cells, contributing to coronary artery disease (CAD) progression and MI risk [93]. Furthermore, many investigated lncRNAs function as non-invasive biomarkers, advancing MI diagnosis, prognosis prediction, and therapeutic monitoring. For instance, circulating levels of Zinc finger antisense 1 (ZFAS1) and Cdr1 antisense (CDR1AS) show notable sensitivity and specificity in predicting acute MI [91]. Assessing ZFAS1 levels holds potential for early MI detection [91].

In summary, recent findings highlight the substantial role of lncRNAs in MI, operating through diverse molecular mechanisms such as signaling, decoy, guide, and scaffold activities. In this capacity, they precisely modulate genes, signaling pathways, epigenetic modifiers, and transcriptional complexes associated with infarction, inflammation, fibrosis, and regeneration. Understanding these mechanisms and their targets could lead to innovative therapeutic interventions against MI by enhancing or inhibiting specific functions.

3.2. LncRNAs and secondary changes of myocardial infarction

MI is characterized by acute ischemia in cardiac tissue, leading to cardiomyocyte death. This initial injury triggers subsequent processes such as ischemia-reperfusion injury, fibrosis, cardiac remodeling, and ultimately heart failure, significantly affecting prognosis and recovery [112]. Recent research highlights the regulatory functions of lncRNAs in these processes through various molecular pathways. Ischemia-reperfusion injury (IRI) occurs when blood flow is reintroduced to ischemic tissue, paradoxically leading to cell death [113]. Several lncRNAs, such as NRF, regulate IRI by acting as decoys for miR-873, alleviating repression on genes like RIPK1 and RIPK3, involved in necroptosis post-IRI. Inhibition of NRF exacerbates IRI by increasing

Table 1

LncRNAs whose functions and target genes have been characterized in myocardial infarction.

LncRNA	Aim	Action	Ref
H19	miR-675/PPAR α	Regulate cell death and IR injury	[94959697]
	miR-103/107/FADD	Regulate programmed Necrosis and Myocardial I/R Injury	
	miR-877-3p/Bcl-2	Inhibit the mitochondrial apoptotic pathway in myocardial	
	miR-675/VEGF/ICAM-1	Mediate the cardioprotective effect of MSC-derived exosomes and promoted the angiogenesis of MI.	
Meg3	RNA-binding protein FUS (fused in sarcoma)	regulates cardiomyocyte apoptosis in myocardial infarction	[98]
GAS5	miR-21 and PDCD4	Regulates PDCD4 expression and mediates myocardial infarction-induced cardiomyocytes apoptosis via targeting Mir-21	[99]
TUG1	miR-590 and FGF1	TUG1 knockdown suppresses cardiac fibrosis after myocardial infarction	[100]
NEAT1	mitogen-activated protein kinase	Promotes myocardial ischemia-reperfusion injury via activating the MAPK signaling pathway	[101]
XIST	JAK2 and CDC42	LncRNA XIST appears to be a risk factor for AMI likely through its ability to regulate JAK2 and CDC42 gene expressions.	[102]
Sox2OT	miR-23b	Sox2OT augments heart dysfunction by facilitating the release of reactive oxygen species (ROS) in septic cardiomyopathy	[103]
ANRIL	WDR5-HDAC3	Form protein complexes and increase ROS level.	[93]
Ahit	SUZ12	Downregulating the expression of MEF2A and preventing cardiac hypertrophy through epigenomic modulation.	[92]
ZFAS1	SERCA2a	Inhibitor of SERCA2a and limits systolic function during MI.	[104]
Airn	Igf2bp2	Affects the translation of Igf2bp2, silencing Airn can increase apoptosis and affect the physiological function of cardiomyocytes.	[105]
CPR	MCM3	Interact and recruit DNMT3A to the CpG island of MCM3 promoter, then inhibit cardiomyocyte proliferation and cardiac function after MI	[88]
Kcnq1ot1	RUNX3	Recruits DNMT1 to the RUNX3 promoter region and regulates CMEC viability and inflammatory response during MI.	[87]
NRF	miR-873/RIPK1-RIPK3	Regulate cardiomyocyte necrosis and myocardial injury in I/R.	[85]
CARL	miR-539/PHB2	Inhibit mitochondrial fission and myocardial apoptosis in MI.	[83]
APF	miR-188-3p/ATG7	Regulates autophagic program and autophagic cell death after MI.	[84]
CAIF	p53-mediated myocardin	Repress autophagic cell death and alleviate MI.	[106]
Sarrah	NRF2	Recruit CRIP2 and p300 to form complex and regulate cardiomyocytes apoptosis in MI.	[89]
SLC8A1-AS1	SLC8A1/cGMP-PKG	Reduce infarct size and ischemia damage in MI.	[81]
FAF	FGF9/FGFR2/PI3K/Akt	Regulate myocardial cell apoptosis in MI.	[80]
UCA1	miR-873-5p/XIAP	Improve the level of antiapoptotic protein and cardiac protection	[107]
KLF3-AS1	mir-138-5p/SIRT1	Inhibit cell pyroptosis and attenuate MI progression.	[108]
Wisper	TIAR	Reduce the development of myocardial fibrosis after MI and prevent adverse remodeling.	[25]
LncRNA AC005332.7	miR-331-3p/CCND2	Inhibited Ferroptosis to Alleviate Acute Myocardial Infarction	[109]
Gm47283	miR-706/ Ptgs2	Suppression of lncRNA Gm47283 attenuates myocardial infarction via miR-706/ Ptgs2/ferroptosis axis	[110]
LncRNA 554	TGF- β 1 signaling pathway	Regulate CFs migration and ECM expression following MI.	[111]

necrotic cell death [85]. Similarly, H19 inhibits miR-103/107 to upregulate FADD, reducing cardiomyocyte necrosis during IRI by limiting formation of the RIPK1/RIPK3 necosome [95]. Downregulation of FAF in the hypoxic myocardium decreases FGF9/FGFR2 signaling and increases apoptosis [80]. These findings demonstrate the protective roles of lncRNAs against IRI by modulating necrotic and apoptotic pathways.

Cardiac fibrosis, characterized by excessive extracellular matrix (ECM) accumulation, impairs heart function post-MI [114]. Numerous lncRNAs regulate fibrosis. For instance, MIAT activates fibroblasts and collagen production via the TGF- β 1/Smad3 pathway; fibrotic responses decrease with MIAT knockdown [115]. Mirt2 promotes fibroblast proliferation and collagen synthesis via the ERK1/2 pathway [116]. Conversely, Lethe inhibits fibrosis by acting as a decoy for miR-188-3p, relieving suppression of activin receptor type-1 (Acvr1), thus reducing fibroblast proliferation and ECM deposition [117]. These findings highlight lncRNAs' direct impact on fibrosis development by regulating fibroblast activation, ECM metabolism, and tissue remodeling signaling pathways.

Heart failure (HF) post-MI entails reduced cardiac function due to structural and functional heart changes [118]. Various lncRNAs influence HF uniquely. For instance, Chaer mediates cardiac dysfunction by competitively binding miR-139-5p, relieving inhibition of its target, the profibrotic cytokine CTGF [119,120]. Cardiopulmonary resuscitation stimulates cardiomyocyte proliferation and guards against HF by recruiting DNMT3A, suppressing the cell cycle inhibitor MCM3 [88]. In HF, Mhrt exacerbates decompensation by suppressing miR-133/135 and enhancing calcineurin/NFAT signaling, contributing to hypertension [121]. Conversely, Fendrr maintains cardiac performance by stabilizing histone methylation and preserving potassium channel function [57].

These findings underscore lncRNAs' regulatory roles in HF progression and regression, influenced by contextual factors. Cardiac remodeling, pivotal in post-MI recovery, involves complex structural, biochemical, and molecular myocardial alterations [122]. LncRNAs significantly contribute to remodeling. For example, Chaer activates CTGF/TGF- β 1 pathways, promoting remodeling [119]. ANRIL forms complexes with WDR5 and HDAC3, enhancing ROS signaling implicated in maladaptive remodeling [93]. Mhrt modulates calcineurin/NFAT signaling to promote hypertension [123]. The lncRNA Bvht induces epigenetic modifications in cardiac specification, crucial for postnatal cardiac growth [56]. Overall, lncRNAs are key regulators of secondary alterations post-MI, including IRI, fibrosis, HF, and remodeling, acting through various pathways influenced by fibroblast activity, ECM deposition, signaling pathways, epigenetic changes, and microRNA activity.

4. LncRNAs and roles as novel biomarkers of myocardial infarction

The management of MI has historically depended on the assessment of protein-based biomarkers, such as cardiac troponins and natriuretic peptides, which yield critical insights into myocardial injury and cardiac dysfunction, respectively. These established biomarkers have proven their clinical utility in the diagnosis, risk stratification, and monitoring of patients with MI. Nevertheless, the pursuit of novel biomarkers that can enhance the accuracy and timeliness of MI detection, as well as improve prognostic predictions, has prompted the investigation of alternative molecular markers, including lncRNAs [124].

Numerous studies have identified heart-specific lncRNAs with abnormal expression post-MI, suggesting their potential as non-invasive

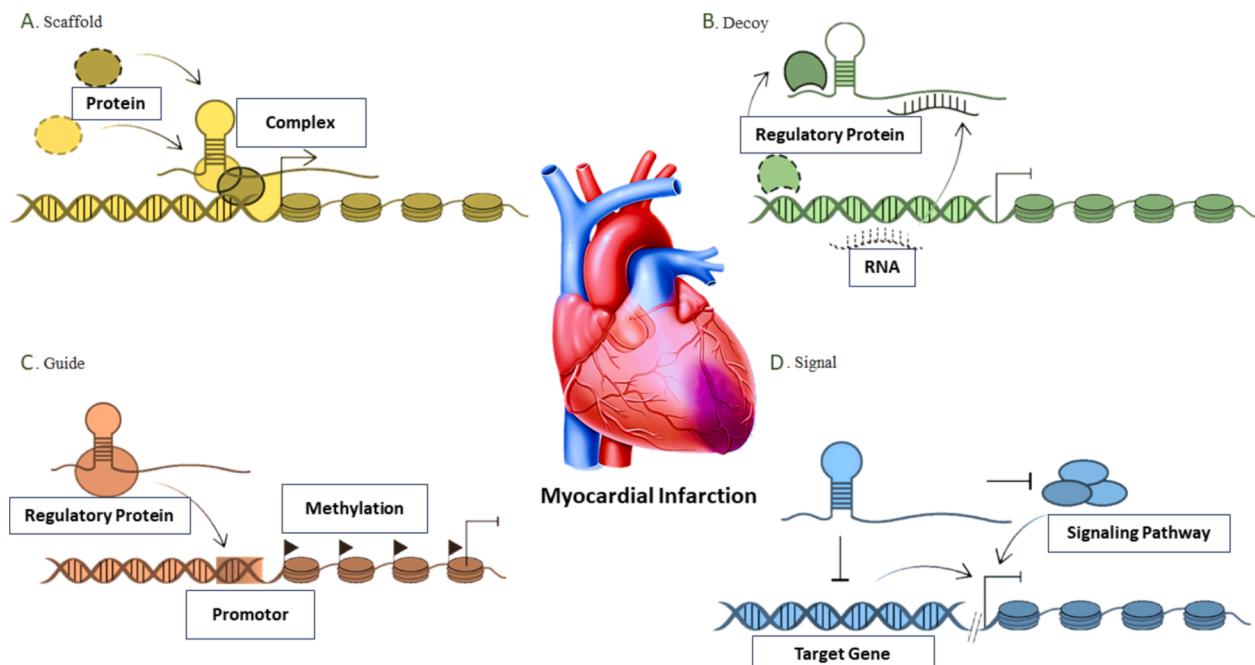


Fig. 2. Four distinct lncRNA molecular mechanisms of action for myocardial infarction are modeled. (A) Scaffold. It's possible that lncRNAs function as scaffolds to join proteins in biological control. (B) Decoy. When interacting with regulatory proteins or RNAs that are not their targets, lncRNAs can function as sponges. (C) Guide. Through molecular interactions, lncRNAs can assemble and bind regulatory proteins or DNAs, directing the resulting complexes to specified destinations. (D) Signal. By controlling signaling pathways in a *cis*- or *trans*-way, lncRNAs participate in the regulation of gene expression and react to particular stimuli.

MI biomarkers [22]. Upregulation of MIAT, detected as early as three hours post-symptom onset and persisting up to 10 days, shows promise in distinguishing MI patients from controls [125,126]. Cardiac apoptosis-related long non-coding RNA (CARL), predominantly expressed in the heart, exhibits significantly elevated plasma levels in MI compared to healthy and unstable angina individuals, indicating potential as an early MI biomarker needing further validation [127]. Integrating cardiac lncRNAs enhances diagnostic efficacy; a panel including CART1, MIAT, MIATv2, and LIPCAR surpasses troponin testing alone, reaching over 98 % accuracy with clinical data inclusion [24,126]. The metabolic lncRNA LIPCAR plays a key role in maintaining energy balance, which is disrupted during MI. Increased LIPCAR levels post-MI, along with MIAT, aid early detection and provide insights into infarction biology [72,115,128]. H19, maternally expressed and responsive to stressors, shows elevated plasma levels during MI, indicating diagnostic potential and predicting unfavorable outcomes [129]. Recent research also implicates additional heart-specific lncRNAs like SCHLAc, PEGASUS, and lnc-Ang392 in MI development, offering innovative biomarker potential. Research has demonstrated that the long non-coding RNA, NEAT1, plays a significant role in myocardial ischemia-reperfusion injury through the activation of the MAPK pathway. The suppression of NEAT1 results in a decrease in pro-inflammatory cytokines such as TNF- α , IL-1 β , and IL-6, which are known to stimulate the production of matrix metalloproteinases (MMPs) [101,130]. Another study, exosomal NEAT1, miR-204, and MMP-9 displayed potent biomarkers for diagnosis of acute ST-segment elevation myocardial infarction [131].

Although in early stages, findings highlight the increasing variety of MI-associated lncRNAs with potential diagnostic implications [112,132]. HIF1A-AS2, the antisense transcript of HIF1 α , is inducible under hypoxia and negatively regulates HIF-1 α mRNA expression [133]. HIF-1 α , a crucial transcription factor, modulates oxygen homeostasis, vascular remodeling, and angiogenesis, pivotal in ischemic heart disease and heart failure pathophysiology [134,135]. HIF1A-AS2 is proposed as a potential early diagnostic biomarker for acute MI with high sensitivity [136]. Additionally, lncRNAs TTY15 and HULC show significant

differential expression in acute MI patients versus healthy controls, suggesting their utility as novel biomarkers alongside conventional markers [137]. Zhu et al. investigated the prognostic diagnostic value of N1LR and SNHG1 in acute MI, indicating their potential to reflect disease progression [138]. Unlike typical protein biomarkers, lncRNAs offer advantageous characteristics. Their compact nature ensures stability in bodily fluids, making them suitable as circulating biomarkers [139]. Prompt release of lncRNAs into peripheral circulation upon cardiomyocyte death allows detection before observable structural changes on imaging. Dynamic analysis of lncRNA profiles plays a crucial role in disease monitoring. Future research elucidating mechanisms and verifying clinical value could enhance early MI diagnosis using lncRNAs, either individually or in tailored signatures. Integrating these technologies into diagnostic algorithms could improve MI management with minimally invasive, highly sensitive, and specific methods. Ongoing efforts focus on refining clinically significant lncRNA biomarker panels, evaluating effectiveness in diverse populations, and developing point-of-care applications [140]. In summary, lncRNAs present a promising avenue for advancing precision cardiovascular care by enabling prompt MI detection.

5. LncRNAs and roles as therapeutic options of myocardial infarction

It is indicated that lncRNAs play vital roles in MI progression and may serve as therapeutic targets [141]. Dysregulated lncRNAs following MI have been identified through microarray profiling and RNA sequencing. For example, MIAT was upregulated in a mouse MI model and in ischemia mimetic-treated cardiac microvascular endothelial cells. MIAT knockdown reduced cell death and improved angiogenesis in mice after MI by regulating microvascular dysfunction [142,143]. Similarly, CARL was upregulated post-MI in mice [127], and its knockdown decreased cardiomyocyte apoptosis and infarct size by inhibiting miR-539, which targets phagocytic receptor TIM-1 [83]. LncRNAs also influence other processes crucial for MI recovery, including cardiomyocyte apoptosis, angiogenesis, and cardiac fibrosis [14]. NRF

regulated cardiomyocyte necrosis and cardiac repair post-MI in mice; NRF knockdown worsened heart failure by increasing cardiomyocyte necrosis PUNISHER modulated cardiomyocyte apoptosis in response to ischemic injury by affecting p53 transcriptional activity [144]. MALAT1 lncRNA regulated post-MI angiogenesis by targeting pro-angiogenic miR-26a [145]. CHRF inhibition promoted cardiac fibrosis after MI in mice by sponging anti-fibrotic miR-489 [66]. Other lncRNAs like H19, ANRIL, and LIPCAR are implicated in cardiomyocyte apoptosis and death after MI [146,147]. The cardioprotective lncRNA FENDRR improved cardiac function and reduced infarct size by promoting angiogenesis [148].

LncRNAs also regulate inflammatory responses post-myocardial infarction, impacting cardiac damage. For instance, Apela inhibits inflammation by modulating macrophage activation through JNK and NF- κ B signaling [149,150]. ANRIL, upregulated after MI, regulates cellular proliferation and apoptosis via epigenetic mechanisms [151]. MCM3AP-AS1, an antisense lncRNA to MCM3AP, when silenced, upregulates miR-24-3p, accelerating proliferation and migration of vascular endothelial cells in MI rat models by reducing EIF4G2 expression [152,153]. MIR4435-2HG, implicated in MI-induced myocardial injury and cardiomyocyte apoptosis, interacts with miR-125a-5p during myocardial ischemia-reperfusion, suggesting a therapeutic target for mitigating ischemia-reperfusion-induced myocardial injury [154]. Conversely, LIPCAR, downregulated post-MI, regulates cardiac fibrosis and apoptosis [155]. These transcripts may offer novel biomarkers and potential avenues for optimizing MI treatment. Maternally expressed gene 3 (Meg3), a long non-coding RNA, is a significant regulator of various biological mechanisms [156]. This lncRNA has been shown to influence cardiomyocyte apoptosis during myocardial infarction (MI) by forming a complex with p53-induced FUS. Wu et al.'s research highlights the potential of specifically targeting this Meg3 complex in cardiomyocytes using the adeno associated virus serotype 9 (AAV9) system as a therapeutic strategy for MI in preclinical studies [98]. Growth arrest-specific 5 (GAS5) is a non-coding RNA molecule that exhibits elevated levels in cells undergoing growth arrest [157]. This RNA is implicated in various cellular processes, such as cell cycle arrest, proliferation, and apoptosis [158]. Research has revealed that GAS5 downregulates miR-21, and this effect is counteracted by miR-21 mimics. Interestingly, GAS5 acts as a competing endogenous RNA (ceRNA) for miR-21, leading to the upregulation of PDCD4 expression in a hypoxia/reoxygenation (H/R) model. Furthermore, GAS5 stimulates PDCD4 expression while inhibiting the PI3K/AKT signaling pathway. The regulatory role of GAS5 on PDCD4 expression in cardiomyocyte apoptosis induced by myocardial infarction (MI) is mediated through miR-21 targeting, indicating its potential as a therapeutic target for MI treatment [99]. Furthermore, long non-coding RNA taurine-upregulated gene 1 (TUG1) is one of the first identified lncRNAs associated with human disease, which actively involved in various physiological processes, including regulating genes at epigenetics, transcription, post-transcription, translation, and posttranslation [159]. Knockdown of TUG1 suppressed cell viability and migration and improved collagen production of TGF- β 1 treated cardiac fibroblasts. TUG1 served as a sponge for miR-590 and FGF1 is a direct target of miR-590. TUG1 expression was increased in acute myocardial infarction tissue and cardiac fibroblasts treated with TGF- β 1. TUG1 knockdown suppressed the biological process of cardiac fibroblasts treated with TGF- β 1 by sponging miR-590 [100].

The therapeutic potential of lncRNAs in myocardial infarction relies on strategies to normalize their expression. For upregulated lncRNAs exacerbating injury, such as MIAT, CARL, and ANRIL, methods like RNA interference and antisense oligonucleotides could inhibit their function. Conversely, delivering mimetics of downregulated cardioprotective lncRNAs through adeno-associated viruses or lipid nanoparticles may improve outcomes. CRISPR-Cas9 genome editing also offers potential for permanently normalizing dysregulated lncRNAs [160]. In summary, lncRNAs present an emerging and promising avenue for therapeutic

development in myocardial infarction. An increasing number of dysregulated lncRNAs have been identified post-MI, playing pivotal roles in processes like cardiomyocyte death, cardiac fibrosis, angiogenesis, and inflammation through diverse molecular mechanisms. Therapeutic targeting of lncRNAs holds great promise for enhancing clinical outcomes after MI [14,161].

6. LncRNAs and current challenges and future prospects

LncRNAs have emerged as influential factors in various cardiovascular disorders and show potential as diagnostic and therapeutic tools for MI. However, several obstacles hinder their clinical implementation. A significant challenge lies in identifying lncRNAs with strong diagnostic or prognostic potential among numerous candidates. Their selective expression patterns at tissue and cellular levels require comprehensive profiling of healthy and pathological cardiac tissues across different cell types. Bioinformatics analysis aids in identifying highly expressed and differentially expressed lncRNAs, but validation in separate patient cohorts is essential to ensure reliability and accuracy. Effective lncRNA biomarkers must surpass recognized protein biomarkers in both sensitivity and specificity [162]. Detecting lncRNAs in minimally invasive samples like blood presents an additional diagnostic challenge, as tissue expression levels may not correlate with plasma or serum levels. Robust techniques for isolating and quantifying circulating lncRNAs are needed for therapeutic applications. Standardized protocols for sample processing and normalization in qRT-PCR or RNA-seq pose further challenges, with factors like hemolysis potentially introducing confounding effects [163].

A major challenge in therapy is delivering therapeutic agents precisely to the cardiac region, hindered by the large size and negative charge of lncRNAs, which restricts passive cellular entry. Potential solutions include using viral vectors, lipid nanoparticles, conjugation with cardiac troponins, or direct administration during surgical procedures, though preventing off-target effects is crucial. Enhancing lncRNA stability against nucleases through chemical modifications like 2'-O-methylation is essential [164]. Another hurdle is addressing the temporary effects of pharmacological inhibitors like siRNAs or antisense oligonucleotides. Utilizing CRISPR-Cas9 for permanent restoration of dysregulated lncRNAs may offer a remedy, but concerns about specificity and off-target effects persist. Novel approaches ensuring continuous lncRNA expression from synthetic vectors or gene editing techniques need development while minimizing potential negative consequences like inflammation [165]. Integrating lncRNA analysis with omics data and machine learning methods shows promise in identifying signatures with enhanced diagnostic accuracy. LncRNAs can contribute to uncovering molecular mechanisms and discovering new therapeutic targets. Advances in nanotechnology and biomaterials offer potential for precise and prolonged lncRNA delivery. Utilizing three-dimensional modeling and structure determination techniques to identify essential features for lncRNA functionality would greatly aid rational drug design.

7. Conclusion

In summary, recent research highlights the aberrant expression patterns of lncRNAs in MI and their regulatory roles in vital physiological mechanisms like proliferation, apoptosis, and inflammation. Promising circulating and tissue-specific lncRNAs emerge as novel non-invasive biomarkers for MI detection and prognosis assessment. However, extensive clinical validation studies are essential to firmly establish the clinical utility of lncRNA signatures for MI diagnosis. Further investigation is necessary to deepen our understanding of the molecular mechanisms driving lncRNA dysregulation and their functional roles in MI pathogenesis. Integrating lncRNA biomarkers into routine clinical practice could revolutionize MI management, offering cost-effective, expedited, and recurrent patient monitoring. Moreover, therapeutic

targeting of lncRNAs holds substantial promise for improving post-MI clinical outcomes, but it requires comprehensive exploration of tissue-specific delivery methods and thorough assessment of safety and efficacy for lncRNA-based medicines. Strategies aimed at normalizing dysregulated lncRNA expression present a promising opportunity to mitigate this significant contributor to global disease burden and mortality, warranting further exploration.

Funding

There is no funding for this study.

Informed consent

Not applicable.

Research Data Policy and Data Availability Statements

All data are available in manuscript.

CRediT authorship contribution statement

Maryam Zolfaghari Dehkharghani: Writing – review & editing, Writing – original draft, Validation, Supervision, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. **Safa Mousavi:** Writing – review & editing, Writing – original draft, Methodology, Data curation, Conceptualization. **Nazanin Kianifard:** Writing – review & editing, Writing – original draft, Methodology. **Amin Fazl-zadeh:** Writing – review & editing, Investigation, Data curation. **Hamid Parsa:** Writing – review & editing, Writing – original draft, Methodology, Investigation. **Ali Tavakoli Pirzaman:** Writing – review & editing, Writing – original draft, Methodology, Investigation. **Andarz Fazlolahpour-Naghibi:** Writing – review & editing, Writing – original draft, Methodology, Investigation.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgment

None.

References

- [1] P. Golforoush, D.M. Yellon, S.M. Davidson, Mouse models of atherosclerosis and their suitability for the study of myocardial infarction, *Basic Res. Cardiol.* 115 (2020) 1–24.
- [2] G.A. Mensah, G.A. Roth, V. Fuster, The global burden of cardiovascular diseases and risk factors: 2020 and beyond, *J. Am. Coll. Cardiol.* 74 (20) (2019) 2529–2532.
- [3] S.N. Fathima, An Update on Myocardial Infarction, in: D. Kumar (Ed.), *Curr Res Trend Med Sci Technol*, Scripown Publications2021, pp. 1–33.
- [4] M. Nichols, N. Townsend, P. Scarborough, M. Rayner, Cardiovascular disease in Europe 2014: epidemiological update, *Eur. Heart J.* 35 (42) (2014) 2950–2959.
- [5] R.W. Yeh, S. Sidney, M. Chandra, M. Sorel, J.V. Selby, A.S. Go, Population trends in the incidence and outcomes of acute myocardial infarction, *New Engl. J. Med.* 362 (23) (2010) 2155–2165.
- [6] W.S. Weintraub, S.R. Daniels, L.E. Burke, B.A. Franklin, D.C. Goff Jr, L. L. Hayman, D. Lloyd-Jones, D.K. Pandey, E.J. Sanchez, A.P. Schram, Value of primordial and primary prevention for cardiovascular disease: a policy statement from the American Heart Association, *Circulation* 124 (8) (2011) 967–990.
- [7] X. Tian, B. Zhou, Coronary vessel formation in development and regeneration: origins and mechanisms, *J. Mol. Cell. Cardiol.* 167 (2022) 67–82.
- [8] B.A. Potz, A.B. Parulkar, R.M. Abid, N.R. Soda, F.W. Sellke, Novel molecular targets for coronary angiogenesis and ischemic heart disease, *Coron. Artery Dis.* 28 (7) (2017) 605.
- [9] J. Li, J. Yang, P. Zhou, Y. Le, Z. Gong, The biological functions and regulations of competing endogenous RNA, *Yi Chuan: Hereditas.* 37 (8) (2015) 756–764.
- [10] Y. Guo, F. Luo, Q. Liu, D. Xu, Regulatory non-coding RNA s in acute myocardial infarction, *J. Cell Mol. Med.* 21 (5) (2017) 1013–1023.
- [11] J. Cao, The functional role of long non-coding RNAs and epigenetics, *Biol. Proced. Online.* 16 (1) (2014) 1–13.
- [12] X. Jiang, R. Lei, Extracellular lncRNAs secreted and absorbed by cardiomyocytes, *J. Cell. Biochem.* 124 (2023) 785–796.
- [13] J.K. Dhanoa, R.S. Sethi, R. Verma, J.S. Arora, C.S. Mukhopadhyay, Long non-coding RNA: its evolutionary relics and biological implications in mammals: a review, *J. Anim. Sci. Technol.* 60 (2018) 1–10.
- [14] C. Chen, Y. Tang, H. Sun, X. Lin, B. Jiang, The roles of long noncoding RNAs in myocardial pathophysiology, *Biosci. Rep.* 39 (11) (2019). BSR20190966.
- [15] Q. Li, Z. Li, Z. Fan, Y. Yang, C. Lu, Involvement of non-coding RNAs in the pathogenesis of myocardial ischemia/reperfusion injury, *Int. J. Mol. Med.* 47 (4) (2021) 1.
- [16] P. Lu, F. Ding, Y.K. Xiang, L. Hao, M. Zhao, Noncoding RNAs in cardiac hypertrophy and heart failure, *Cells* 11 (5) (2022) 777.
- [17] H. Almaghrbi, R. Giordo, G. Pintus, H. Zayed, Non-coding RNAs as biomarkers of myocardial infarction, *Clin. Chim. Acta* 540 (2023) 117222.
- [18] N. Kim, W.-Y. Chung, J.-Y. Cho, The role and medical prospects of long non-coding RNAs in cardiovascular disease, *Heart Fail. Rev.* 28 (6) (2023) 1437–1453.
- [19] S. Shen, H. Jiang, Y. Bei, J. Xiao, X. Li, Long non-coding RNAs in cardiac remodeling, *Cell. Physiol. Biochem.* 41 (5) (2017) 1830–1837.
- [20] C. Humeres, N.G. Frangogiannis, Fibroblasts in the infarcted, remodeling, and failing heart, *JACC Basic Transl. Sci.* 4 (3) (2019) 449–467.
- [21] A. Al-Masri, Apoptosis and long non-coding RNAs: Focus on their roles in Heart diseases, *Pathol. Res. Pract.* 251 (2023) 154889.
- [22] X. Zhang, Z. Chen, J. Zang, C. Yao, J. Shi, R. Nie, G. Wu, LncRNA-mRNA co-expression analysis discovered the diagnostic and prognostic biomarkers and potential therapeutic agents for myocardial infarction, *Aging (Albany NY)* 13 (6) (2021) 8944.
- [23] R. Mir, I. Elfaki, N. Khullar, A.A. Waza, C. Jha, M.M. Mir, S. Nisa, B. Mohammad, T.A. Mir, M. Maqbool, Role of selected miRNAs as diagnostic and prognostic biomarkers in cardiovascular diseases, including coronary artery disease, myocardial infarction and atherosclerosis, *J. Cardiovasc. Dev. Dis.* 8 (2) (2021) 22.
- [24] C. Sun, H. Jiang, Z. Sun, Y. Gui, H. Xia, Identification of long non-coding RNAs biomarkers for early diagnosis of myocardial infarction from the dysregulated coding-non-coding co-expression network, *Oncotarget* 7 (45) (2016) 73541.
- [25] R. Micheletti, I. Plaisance, B.J. Abraham, A. Sarre, C.-C. Ting, M. Alexanian, D. Marie, D. Maison, M. Nemir, R.A. Young, The long noncoding RNA Wisper controls cardiac fibrosis and remodeling, *Sci. Transl. Med.* 9 (395) (2017) eaai9118.
- [26] F. Zhang, X. Fu, M. Kataoka, N. Liu, Y. Wang, F. Gao, T. Liang, X. Dong, J. Pei, X. Hu, Long noncoding RNA Cfast regulates cardiac fibrosis, *Mol. Ther. Nucleic Acids* 23 (2021) 377–392.
- [27] S. Haemmig, V. Simion, D. Yang, Y. Deng, M.W. Feinberg, Long noncoding RNAs in cardiovascular pathology, diagnosis, and therapy, *Curr. Opin. Cardiol.* 32 (6) (2017) 776.
- [28] H. Zhu, J. Ouyang, J.-L. Chen, Function and regulation of long non-coding RNAs in tumorigenesis and host innate immunity – a review, *Acta Microbiol. Sinica.* 55 (7) (2015) 801–812.
- [29] L. Wu, S. Liu, H. Qi, H. Cai, M. Xu, Research progress on plant long non-coding RNA, *Plants* 9 (4) (2020) 408.
- [30] L. Chen, Y.-H. Zhang, X. Pan, M. Liu, S. Wang, T. Huang, Y.-D. Cai, Tissue expression difference between mRNAs and lncRNAs, *Int. J. Mol. Sci.* 19 (11) (2018) 3416.
- [31] P. Johnsson, L. Lipovich, D. Grandér, K.V. Morris, Evolutionary conservation of long non-coding RNAs: sequence, structure, function, *Biochim. Biophys. Acta Gen. Subj.* 1840 (3) (2014) 1063–1071.
- [32] S. Kannan, D. Chernikova, I.B. Rogozin, E. Poliakov, D. Managadze, E.V. Koonin, L. Milanesi, Transposable element insertions in long intergenic non-coding RNA genes, *Front. Bioeng. Biotechnol.* 3 (2015) 71.
- [33] K. Zhang, Z.-M. Shi, Y.-N. Chang, Z.-M. Hu, H.-X. Qi, W. Hong, The ways of action of long non-coding RNAs in cytoplasm and nucleus, *Gene* 547 (1) (2014) 1–9.
- [34] J. Chen, Y. Xue, Emerging roles of non-coding RNAs in epigenetic regulation, *Sci. China Life Sci.* 59 (2016) 227–235.
- [35] C. Zhang, B. Han, T. Xu, D. Li, The biological function and potential mechanism of long non-coding RNAs in cardiovascular disease, *J. Cell Mol. Med.* 24 (22) (2020) 12900–12909.
- [36] N. Brockdorff, J.S. Bowness, G. Wei, Progress toward understanding chromosome silencing by Xist RNA, *Genes Deve.* 34 (11–12) (2020) 733–744.
- [37] B. Cai, X. Song, J. Cai, S. Zhang, HOTAIR: a cancer-related long non-coding RNA, *Neoplasma* 61 (4) (2014) 379–391.
- [38] D. Meseure, S. Vacher, F. Lallemand, K.D. Alsibai, R. Hatem, W. Chemlali, A. Nicolas, L. De Koning, E. Pasman, C. Callens, Prognostic value of a newly identified MALAT1 alternatively spliced transcript in breast cancer, *Br. J. Cancer* 114 (12) (2016) 1395–1404.
- [39] K. Palos, L.a. Yu, C.E. Railey, A.C. Nelson Dittrich, A.D. Nelson, Linking discoveries, mechanisms, and technologies to develop a clearer perspective on plant long noncoding RNAs, *Plant Cell.* 35(6) (2023) 1762–1786.
- [40] M. Huarte, The emerging role of lncRNAs in cancer, *Nat. Med.* 21 (11) (2015) 1253–1261.
- [41] R. Begolli, N. Sideris, A. Giakountis, LncRNAs as chromatin regulators in cancer: from molecular function to clinical potential, *Cancers* 11 (10) (2019) 1524.
- [42] C.-F. Yeh, Y.-C.-E. Chang, C.-Y. Lu, C.-F. Hsuan, W.-T. Chang, K.-C. Yang, Expedition to the missing link: Long noncoding RNAs in cardiovascular diseases, *J. Biomed. Sci.* 27 (2020) 1–16.
- [43] Y. Wang, X. Sun, The functions of lncRNA in the heart, *Diabetes Res. Clin. Pract.* 168 (2020) 108249.
- [44] L. Kurian, A. Aguirre, I. Sancho-Martinez, C. Benner, T. Hishida, T.B. Nguyen, P. Reddy, E. Nivet, M.N. Krause, D.A. Nelles, Identification of novel long

- noncoding RNAs underlying vertebrate cardiovascular development, *Circulation* 131 (14) (2015) 1278–1290.
- [45] M. Touma, X. Kang, Y. Zhao, A.A. Cass, F. Gao, R. Biniwale, G. Coppola, X. Xiao, B. Reemtsen, Y. Wang, Decoding the long noncoding RNA during cardiac maturation: a roadmap for functional discovery, *Circulation Cardiovasc Genet.* 9 (5) (2016) 395–407.
- [46] Z. Tang, Y. Wu, Y. Yang, Y.-C.-T. Yang, Z. Wang, J. Yuan, Y. Yang, C. Hua, X. Fan, G. Niu, Comprehensive analysis of long non-coding RNAs highlights their spatio-temporal expression patterns and evolutionary conservation in *Sus scrofa*, *Sci. Rep.* 7 (1) (2017) 43166.
- [47] C. He, H. Hu, K.D. Wilson, H. Wu, J. Feng, S. Xia, J. Churko, K. Qu, H.Y. Chang, J. C. Wu, Systematic characterization of long noncoding RNAs reveals the contrasting coordination of cis-and trans-molecular regulation in human fetal and adult hearts, *Circulat. Cardiovasc Genet.* 9 (2) (2016) 110–118.
- [48] Y. Li, J. Zhang, C. Huo, N. Ding, J. Li, J. Xiao, X. Lin, B. Cai, Y. Zhang, J. Xu, Dynamic organization of lncRNA and circular RNA regulators collectively controlled cardiac differentiation in humans, *EBioMed.* 24 (2017) 137–146.
- [49] J. Beermann, D. Kirste, K. Iwanov, D. Lu, F. Kleemil, R. Kumarswamy, K. Schimmel, C. Bär, T. Thum, A large shRNA library approach identifies lncRNA Ntep as an essential regulator of cell proliferation, *Cell Death Differ.* 25 (2) (2018) 307–318.
- [50] C.-P. Chang, P. Han, Epigenetic and lncRNA regulation of cardiac pathophysiology, *Biochim. Biophys. Acta Gen. Subj.* 1863 (7) (2016) 1767–1771.
- [51] Y. Li, W. Du, R. Zhao, J. Hu, H. Li, R. Han, Q. Yue, R. Wu, W. Li, J. Zhao, New insights into epigenetic modifications in heart failure, *Front Biosci (landmark Ed).* 22 (2017) 230–247.
- [52] Z. Zhang, W. Gao, Q.-Q. Long, J. Zhang, Y.-F. Li, D.-C. Liu, J.-J. Yan, Z.-J. Yang, L.-S. Wang, Increased plasma levels of lncRNA H19 and LIPCAR are associated with increased risk of coronary artery disease in a Chinese population, *Sci. Rep.* 7 (1) (2017) 7491.
- [53] S. Ounzain, F. Burdet, M. Ibberson, T. Pedrazzini, Discovery and functional characterization of cardiovascular long noncoding RNAs, *J. Mol. Cell. Cardiol.* 89 (2015) 17–26.
- [54] G. Wang, B. Wang, P. Yang, Epigenetics in congenital heart disease, *J. Am. Heart Assoc.* 11 (7) (2022) e025163.
- [55] L.E. Philippen, E. Dirkx, P.A. da Costa-Martins, L.J. De Windt, Non-coding RNA in control of gene regulatory programs in cardiac development and disease, *J. Mol. Cell. Cardiol.* 89 (2015) 51–58.
- [56] C.A. Klattenhoff, J.C. Scheuermann, L.E. Surface, R.K. Bradley, P.A. Fields, M. L. Steinhauser, H. Ding, V.L. Butty, L. Torrey, S. Haas, Braveheart, a long noncoding RNA required for cardiovascular lineage commitment, *Cell* 152 (3) (2013) 570–583.
- [57] P. Grote, L. Wittler, D. Hendrix, F. Koch, S. Währisch, A. Beisaw, K. Macura, G. Bläss, M. Kellis, M. Werber, The tissue-specific lncRNA Fendrr is an essential regulator of heart and body wall development in the mouse, *Dev. Cell* 24 (2) (2013) 206–214.
- [58] C. Li, Y.-Q. Ni, H. Xu, Q.-Y. Xiang, Y. Zhao, J.-K. Zhan, J.-Y. He, S. Li, Y.-S. Liu, Roles and mechanisms of exosomal non-coding RNAs in human health and diseases, *Signal Transduct. Target. Ther.* 6 (1) (2021) 383.
- [59] G. Arun, D. Aggarwal, D.L. Spector, MALAT1 long non-coding RNA: Functional implications, *Noncoding RNA.* 6 (2) (2020) 22.
- [60] D. Li, C. Zhang, J. Li, J. Che, X. Yang, Y. Xian, X. Li, C. Cao, Long non-coding RNA MALAT1 promotes cardiac remodeling in hypertensive rats by inhibiting the transcription of MyoD, *Aging (Albany NY)* 11 (20) (2019) 8792.
- [61] B. Chen, L. Luo, X. Wei, D. Gong, Z. Li, S. Li, W. Tang, L. Jin, M1 bone marrow-derived macrophage-derived extracellular vesicles inhibit angiogenesis and myocardial regeneration following myocardial infarction via the MALAT1/MicroRNA-25-3p/CDC42 Axis, *Oxid. Med. Cell. Longev.* 2021 (2021).
- [62] Y. Wang, X. Sun, X. Sun, The functions of long non-coding RNA (lncRNA) H19 in the heart, *Heart Lung Circ.* 31 (3) (2022) 341–349.
- [63] R. Nadhan, C. Isidoro, Y.S. Song, D.N. Dhanasekaran, Signaling by lncRNAs: Structure, cellular homeostasis, and disease pathology, *Cells.* 11 (16) (2022) 2517.
- [64] L.W. Harries, Long non-coding RNAs and human disease, *Biochem. Soc. Trans.* 40 (4) (2012) 902–906.
- [65] C. Iaconetti, C. Gareri, A. Polimeni, C. Indolfi, Non-coding RNAs: the “dark matter” of cardiovascular pathophysiology, *Int. J. Mol. Sci.* 14 (10) (2013) 19987–20018.
- [66] K. Wang, F. Liu, L.-Y. Zhou, B. Long, S.-M. Yuan, Y. Wang, C.-Y. Liu, T. Sun, X.-J. Zhang, P.-F. Li, The long noncoding RNA CHRF regulates cardiac hypertrophy by targeting miR-489, *Circ. Res.* 114 (9) (2014) 1377–1388.
- [67] D.D. Singh, Y. Kim, S.A. Choi, I. Han, D.K. Yadav, Clinical significance of MicroRNAs, long non-coding RNAs, and CircRNAs in cardiovascular diseases, *Cells.* 12 (12) (2023) 1629.
- [68] W. Xiong, Y. Qu, H. Chen, J. Qian, Insight into long noncoding RNA-miRNA-mRNA axes in myocardial ischemia-reperfusion injury: the implications for mechanism and therapy, *Epigenomics* 11 (15) (2019) 1733–1748.
- [69] X. Zhou, W. Zhang, M. Jin, J. Chen, W. Xu, X. Kong, lncRNA MIAT functions as a competing endogenous RNA to upregulate DAPK2 by sponging miR-22-3p in diabetic cardiomyopathy, *Cell Death Dis.* 8 (7) (2017) e2929-e.
- [70] S. Sweta, T. Dudnakova, S. Sudheer, A.H. Baker, R. Bhushan, Importance of long non-coding RNAs in the development and disease of skeletal muscle and cardiovascular lineages, *Front. Cell Dev. Biol.* 7 (2019) 228.
- [71] G. Wu, J. Cai, Y. Han, J. Chen, Z.-P. Huang, C. Chen, Y. Cai, H. Huang, Y. Yang, Y. Liu, LincRNA-p21 regulates neointima formation, vascular smooth muscle cell proliferation, apoptosis, and atherosclerosis by enhancing p53 activity, *Circulation* 130 (17) (2014) 1452–1465.
- [72] M. Li, L.-F. Wang, X.-C. Yang, L. Xu, W.-M. Li, K. Xia, D.-P. Zhang, R.-N. Wu, T. Gan, Circulating long noncoding RNA LIPCAR acts as a novel biomarker in patients with ST-segment elevation myocardial infarction, *Med. Sci. Monit.* 24 (2018) 5064.
- [73] W. Wang, B. Tian, Z. Ning, X. Li, Research progress of lncRNAs in atrial fibrillation, *Mol. Biotechnol.* 64 (7) (2022) 758–772.
- [74] F. Guo, C. Tang, B. Huang, L. Gu, J. Zhou, Z. Mo, C. Liu, Y. Liu, lncRNA H19 drives proliferation of cardiac fibroblasts and collagen production via suppression of the miR-29a-3p/miR-29b-3p-VEGFA/TGF- β axis, *Mol. Cell* 45 (3) (2022) 122.
- [75] P. Yang, Y. Cao, H. Jian, H. Chen, Identification of Hub mRNAs and lncRNAs in atrial fibrillation using weighted co-expression network analysis with RNA-Seq data, *Front. Cell Dev. Biol.* 9 (2021) 722671.
- [76] Z. Ruan, X. Sun, H. Sheng, L. Zhu, Long non-coding RNA expression profile in atrial fibrillation, *Int. J. Clin. Exp. Path.* 8 (7) (2015) 8402.
- [77] Y. Devaux, J. Zangrandino, B. Schroen, E.E. Creemers, T. Pedrazzini, C.-P. Chang, G.W. Dorn, T. Thum, S. Heymans, C. network, Long noncoding RNAs in cardiac development and ageing, *Nat Rev Cardiol.* 12(7) (2015) 415–425.
- [78] P.J. Batista, H.Y. Chang, Long noncoding RNAs: cellular address codes in development and disease, *Cell* 152 (6) (2013) 1298–1307.
- [79] M.V. Koerner, F.M. Pauler, R. Huang, D.P. Barlow, The function of non-coding RNAs in genomic imprinting, *Development* 136 (11) (2009) 1771–1783.
- [80] H.J. Shi, M.W. Wang, J.T. Sun, H. Wang, Y.F. Li, B.R. Chen, Y. Fan, S.B. Wang, Z. M. Wang, Q.M. Wang, A novel long noncoding RNA FAF inhibits apoptosis via upregulating FGFR through PI3K/AKT signaling pathway in ischemia-hypoxia cardiomyocytes, *J. Cell. Physiol.* 234 (12) (2019) 21973–21987.
- [81] G.L. Guo, L.Q. Sun, M.H. Sun, H.M. Xu, lncRNA SLC8A1-AS1 protects against myocardial damage through activation of cGMP-PKG signaling pathway by inhibiting SLC8A1 in mice models of myocardial infarction, *J. Cell. Physiol.* 234 (6) (2019) 9019–9032.
- [82] M. Cesana, D. Cacchiarelli, I. Legnini, T. Santini, O. Stahdier, M. Chinappi, A. Tramontano, I. Bozzoni, A long noncoding RNA controls muscle differentiation by functioning as a competing endogenous RNA, *Cell* 147 (2) (2011) 358–369.
- [83] K. Wang, B. Long, L.-Y. Zhou, F. Liu, Q.-Y. Zhou, C.-Y. Liu, Y.-Y. Fan, P.-F. Li, CARL lncRNA inhibits anoxia-induced mitochondrial fission and apoptosis in cardiomyocytes by impairing miR-539-dependent PHB2 downregulation, *Nat. Commun.* 5 (1) (2014) 3596.
- [84] K. Wang, C.-Y. Liu, L.-Y. Zhou, J.-X. Wang, M. Wang, B. Zhao, W.-K. Zhao, S.-J. Xu, L.-H. Fan, X.-J. Zhang, APF lncRNA regulates autophagy and myocardial infarction by targeting miR-188-3p, *Nat. Commun.* 6 (1) (2015) 6779.
- [85] K. Wang, F. Liu, C. Liu, T. An, J. Zhang, L. Zhou, M. Wang, Y. Dong, N. Li, J. Gao, The long noncoding RNA NRF regulates programmed necrosis and myocardial injury during ischemia and reperfusion by targeting miR-873, *Cell Death Differ.* 23 (8) (2016) 1394–1405.
- [86] V.A. Moran, R.J. Perera, A.M. Khalil, Emerging functional and mechanistic paradigms of mammalian long non-coding RNAs, *Nucleic Acids Res.* 40 (14) (2012) 6391–6400.
- [87] Y. Wang, X. Yang, A. Jiang, W. Wang, J. Li, J. Wen, Methylation-dependent transcriptional repression of RUNX3 by KCNQ1OT1 regulates mouse cardiac microvascular endothelial cell viability and inflammatory response following myocardial infarction, *FASEB J.* 33 (12) (2019) 13145.
- [88] M. Ponnusamy, F. Liu, Y.-H. Zhang, R.-B. Li, M. Zhai, F. Liu, L.-Y. Zhou, C.-Y. Liu, K.-W. Yan, Y.-H. Dong, Long noncoding RNA CPR (cardiomyocyte proliferation regulator) regulates cardiomyocyte proliferation and cardiac repair, *Circulation* 139 (23) (2019) 2668–2684.
- [89] D.J. Trembinski, D.I. Bink, K. Theodorou, J. Sommer, A. Fischer, A. van Bergen, C.-C. Kuo, I.G. Costa, C. Schürmann, M.S. Leisegang, Aging-regulated anti-apoptotic long non-coding RNA Sarrab augments recovery from acute myocardial infarction, *Nat. Commun.* 11 (1) (2020) 2039.
- [90] M. Guttman, J.L. Rinn, Modular regulatory principles of large non-coding RNAs, *Nature* 482 (7385) (2012) 339–346.
- [91] D.M. Ribeiro, A. Zanzoni, A. Cipriano, R. Delli Ponti, L. Spinelli, M. Ballarino, I. Bozzoni, G.G. Tartaglia, C. Brun, Protein complex scaffolding predicted as a prevalent function of long non-coding RNAs, *Nucleic Acids Res.* 46 (2) (2018) 917–928.
- [92] J. Yu, Y. Yang, Z. Xu, C. Lan, C. Chen, C. Li, Z. Chen, C. Yu, X. Xia, Q. Liao, Long noncoding RNA Ahit protects against cardiac hypertrophy through SUZ12 (suppressor of Zeste 12 protein homolog)-mediated downregulation of MEF2A (myocyte enhancer factor 2A), *Circ. Heart Fail.* 13 (1) (2020) e006525.
- [93] C. Zhang, S. Ge, W. Gong, J. Xu, Z. Guo, Z. Liu, X. Gao, X. Wei, S. Ge, lncRNA ANRIL acts as a modular scaffold of WDR5 and HDAC3 complexes and promotes alteration of the vascular smooth muscle cell phenotype, *Cell Death Dis.* 11 (6) (2020) 435.
- [94] H. Luo, J. Wang, D. Liu, S. Zang, N. Ma, L. Zhao, L. Zhang, X. Zhang, C. Qiao, The lncRNA H19/miR-675 axis regulates myocardial ischemic and reperfusion injury by targeting PPAR α , *Mol. Immunol.* 105 (2019) 46–54.
- [95] J.-X. Wang, X.-J. Zhang, Q. Li, K. Wang, Y. Wang, J.-Q. Jiao, C. Feng, S. Teng, L.-Y. Zhou, Y. Gong, MicroRNA-103/107 regulate programmed necrosis and myocardial ischemia/reperfusion injury through targeting FADD, *Circ. Res.* 117 (4) (2015) 352–363.
- [96] X. Li, S. Luo, J. Zhang, Y. Yuan, W. Jiang, H. Zhu, X. Ding, L. Zhan, H. Wu, Y. Xie, lncRNA H19 alleviated myocardial I/R injury via suppressing miR-877-3p/Bcl-2-mediated mitochondrial apoptosis, *Mol. Ther. Nucleic Acids* 17 (2019) 297–309.
- [97] P. Huang, L. Wang, Q. Li, X. Tian, J. Xu, J. Xu, Y. Xiong, G. Chen, H. Qian, C. Jin, Atorvastatin enhances the therapeutic efficacy of mesenchymal stem cells-derived

- exosomes in acute myocardial infarction via up-regulating long non-coding RNA H19, *Cardiovasc. Res.* 116 (2) (2020) 353–367.
- [98] H. Wu, Z.-A. Zhao, J. Liu, K. Hao, Y. Yu, X. Han, J. Li, Y. Wang, W. Lei, N. Dong, Long noncoding RNA Meg3 regulates cardiomyocyte apoptosis in myocardial infarction, *Gene Ther.* 25 (8) (2018) 511–523.
- [99] X.-H. Zhou, H.-X. Chai, M. Bai, Z. Zhang, LncRNA-GASS5 regulates PDCD4 expression and mediates myocardial infarction-induced cardiomyocytes apoptosis via targeting MiR-21, *Cell Cycle* 19 (11) (2020) 1363–1377.
- [100] Q. Sun, M. Luo, Z. Gao, X. Han, Z. Yan, S. Xie, H. Zhao, H. Sun, TUG1 knockdown suppresses cardiac fibrosis after myocardial infarction, *Mamm. Genome* 32 (2021) 435–442.
- [101] X.J. Du, J. Wei, D. Tian, C. Yan, P. Hu, X. Wu, W. Yang, X. Hu, NEAT1 promotes myocardial ischemia-reperfusion injury via activating the MAPK signaling pathway, *J. Cell. Physiol.* 234 (10) (2019) 18773–18780.
- [102] P.-F. Zheng, L.-Z. Chen, P. Liu, H.-W. Pan, A novel lncRNA-miRNA-mRNA triple network identifies lncRNA XIST as a biomarker for acute myocardial infarction, *Aging (Albany NY)* 14 (9) (2022) 4085.
- [103] L. Xuan, D. Fu, D. Zhen, D. Bai, L. Yu, G. Gong, Long non-coding RNA Sox2OT promotes coronary microembolization-induced myocardial injury by mediating pyroptosis, *ESC Heart Fail.* 9 (3) (2022) 1689–1702.
- [104] Y. Zhang, L. Jiao, L. Sun, Y. Li, Y. Gao, C. Xu, Y. Shao, M. Li, C. Li, Y. Lu, LncRNA ZFAS1 as a SERCA2a inhibitor to cause intracellular Ca²⁺ overload and contractile dysfunction in a mouse model of myocardial infarction, *Circ. Res.* 122 (10) (2018) 1354–1368.
- [105] M.R. Hosen, G. Militello, T. Weirick, Y. Ponomareva, S. Dassanayaka, J. B. Moore IV, C. Döring, M. Wysoczynski, S.P. Jones, S. Dimmeler, Airn regulates Igf2bp2 translation in cardiomyocytes, *Circ. Res.* 122 (10) (2018) 1347–1353.
- [106] C.-Y. Liu, Y.-H. Zhang, R.-B. Li, L.-Y. Zhou, T. An, R.-C. Zhang, M. Zhai, Y. Huang, K.-W. Yan, Y.-H. Dong, LncRNA CAIF inhibits autophagy and attenuates myocardial infarction by blocking p53-mediated myocardin transcription, *Nat. Commun.* 9 (1) (2018) 29.
- [107] L. Sun, W. Zhu, P. Zhao, Q. Wang, B. Fan, Y. Zhu, Y. Lu, Q. Chen, J. Zhang, F. Zhang, Long noncoding RNA UCA1 from hypoxia-conditioned hMSC-derived exosomes: a novel molecular target for cardioprotection through miR-873-5p/XIAP axis, *Cell Death Differ.* 11 (8) (2020) 696.
- [108] Q. Mao, X.-L. Liang, C.-L. Zhang, Y.-H. Pang, Y.-X. Lu, LncRNA KLF3-AS1 in human mesenchymal stem cell-derived exosomes ameliorates pyroptosis of cardiomyocytes and myocardial infarction through miR-138-5p/Sirt1 axis, *Stem Cell Res Ther* 10 (1) (2019) 1–14.
- [109] R. Dai, X. Yang, W. He, Q. Su, X. Deng, J. Li, LncRNA AC005332.7 inhibited ferroptosis to alleviate acute myocardial infarction through regulating miR-331-3p/CCND2 axis, *Korean Circ J.* 53 (3) (2023) 151–167.
- [110] F. Gao, Y. Zhao, B. Zhang, C. Xiao, Z. Sun, Y. Gao, X. Dou, Suppression of lncRNA Gm47283 attenuates myocardial infarction via miR-706/Ptgs2/ferroptosis axis, *Bioengineering*. 13 (4) (2022) 10786–10802.
- [111] B. Luo, Z. He, S. Huang, J. Wang, D. Han, H. Xie, P. Liu, X. Zeng, D. Lu, Long non-coding RNA 554 promotes cardiac fibrosis via TGF-β1 pathway in mice following myocardial infarction, *Front. Pharmacol.* 11 (2020) 585680.
- [112] L. Xie, Q. Zhang, J. Mao, J. Zhang, L. Li, The roles of lncRNA in myocardial infarction: molecular mechanisms, diagnosis biomarkers, and therapeutic perspectives, *Front. Cell Dev. Biol.* 9 (2021) 680713.
- [113] S. Hashmi, S. Al-Salam, Acute myocardial infarction and myocardial ischemia-reperfusion injury: a comparison, *Int. J. Clin. Exp. Path.* 8 (8) (2015) 8786.
- [114] K. Maruyama, K. Imanaka-Yoshida, The pathogenesis of cardiac fibrosis: a review of recent progress, *Int. J. Mol. Sci.* 23 (5) (2022) 2617.
- [115] X. Qu, Y. Du, Y. Shu, M. Gao, F. Sun, S. Luo, T. Yang, L. Zhan, Y. Yuan, W. Chu, MIAT is a pro-fibrotic long non-coding RNA governing cardiac fibrosis in post-infarct myocardium, *Sci. Rep.* 7 (1) (2017) 42657.
- [116] F. Zhu, Q. Li, J. Li, B. Li, D. Li, Long noncoding Mirt2 reduces apoptosis to alleviate myocardial infarction through regulation of the miR-764/PDK1 axis, *Lab. Invest.* 101 (2) (2021) 165–176.
- [117] J. Deng, M. Guo, J. Xiao, Long Noncoding RNAs in Cardiovascular Development and Diseases, in: S. Jurja, J. Barciszewski (Eds.), *The Chemical Biology of Long Noncoding RNAs*, Springer2020, pp. 363–383.
- [118] J.A. Ezekowitz, P. Kaul, J.A. Bakal, P.W. Armstrong, R.C. Welsh, F.A. McAlister, Declining in-hospital mortality and increasing heart failure incidence in elderly patients with first myocardial infarction, *J. Am. Coll. Cardiol.* 53 (1) (2009) 13–20.
- [119] Z. Wang, X.-J. Zhang, Y.-X. Ji, P. Zhang, K.-Q. Deng, J. Gong, S. Ren, X. Wang, I. Chen, H. Wang, The long noncoding RNA Chaer defines an epigenetic checkpoint in cardiac hypertrophy, *Nat. Med.* 22 (10) (2016) 1131–1139.
- [120] C. Wang, D. Yang, C. Xu, H. Duan, MicroRNA-139-5p inhibits vascular endothelial cell viability and serves as a diagnostic biomarker in acute myocardial infarction patients, *Exp. Gerontol.* 152 (2021) 111453.
- [121] Y. Xu, Y. Luo, C. Liang, T. Zhang, LncRNA-Mhrt regulates cardiac hypertrophy by modulating the miR-145a-5p/KLF4/myocardin axis, *J. Mol. Cell. Cardiol.* 139 (2020) 47–61.
- [122] M.-M. Bostan, C. Stătescu, L. Anghel, I.-L. Ţerban, E. Cojocaru, R. Sascau, Post-myocardial infarction ventricular remodeling biomarkers—the key link between pathophysiology and clinic, *Biomolecules* 10 (11) (2020) 1587.
- [123] P. Han, W. Li, C.-H. Lin, J. Yang, C. Shang, S.T. Nurnberg, K.K. Jin, W. Xu, C.-Y. Lin, C.-J. Lin, A long noncoding RNA protects the heart from pathological hypertrophy, *Nature* 514 (7520) (2014) 102–106.
- [124] H.-J. Feistritzer, G. Klug, S.J. Reinstadler, M. Reindl, A. Mayr, J. Mair, B. Metzler, Novel biomarkers predicting cardiac function after acute myocardial infarction, *Br. Med. Bull.* 119 (1) (2016) 63–74.
- [125] M. Azat, X. Huojiahemaiti, R. Gao, P. Peng, Long noncoding RNA MIAT: A potential role in the diagnosis and mediation of acute myocardial infarction, *Mol. Med. Rep.* 20 (6) (2019) 5216–5222.
- [126] X.-M. Wang, X.-M. Li, N. Song, H. Zhai, X.-M. Gao, Y.-N. Yang, Long non-coding RNAs H19, MALAT1 and MIAT as potential novel biomarkers for diagnosis of acute myocardial infarction, *Biomed. Pharmacother.* 118 (2019) 109208.
- [127] L. Li, J.-J. Wang, H.-S. Zhang, LncRNA-CAR1 in a rat model of myocardial infarction, *Eur. Rev. Med. Pharmacol. Sci.* 22 (13) (2018) 4332–4340.
- [128] L. Yan, Y. Zhang, M. Wang, L. Wang, W. Zhang, Z.-R. Ge, Circulating LIPCAR is a potential biomarker of heart failure in patients post-acute myocardial infarction, *Exp. Biol. Med.* 246 (24) (2021) 2589–2594.
- [129] S. Safaei, M. Tahmasebi-Birgani, M. Bijanzadeh, S.M. Seyedian, Increased expression level of long noncoding RNA H19 in plasma of patients with myocardial infarction, *Int J Mol Cell Med.* 9 (2) (2020) 122.
- [130] N. Cui, M. Hu, R.A. Khalil, Biochemical and biological attributes of matrix metalloproteinases, *Prog. Mol. Biol. Transl. Sci.* 147 (2017) 1–73.
- [131] Z. Chen, Y. Yan, J. Wu, C. Qi, J. Liu, J. Wang, Expression level and diagnostic value of exosomal NEAT1/miR-204/MMP-9 in acute ST-segment elevation myocardial infarction, *IUBMB Life* 72 (11) (2020) 2499–2507.
- [132] X. Wu, L. Sun, Z. Wang, Identification of lncRNA competitively regulated subpathways in myocardial infarction Corrigendum in 10.3892/etm.2019.8210, *Exp Ther Med.* 17(4) (2019) 3041–3046.
- [133] F. Zheng, J. Chen, X. Zhang, Z. Wang, J. Chen, X. Lin, H. Huang, W. Fu, J. Liang, W. Wu, The HIF-1α antisense long non-coding RNA drives a positive feedback loop of HIF-1α mediated transactivation and glycolysis, *Nat. Commun.* 12 (1) (2021) 1341.
- [134] C. Corrado, S. Fontana, Hypoxia and HIF signaling: one axis with divergent effects, *Int. J. Mol. Sci.* 21 (16) (2020) 5611.
- [135] G.L. Semenza, Hypoxia-inducible factor 1 and cardiovascular disease, *Annual Rev Physiol.* 76 (2014) 39–56.
- [136] E. Tayae, E. Amr, A. Zaki, D. Elkaffash, LncRNA HIF1A-AS2: a potential biomarker for early diagnosis of acute myocardial infarction and predictor of left ventricular dysfunction, *BMC Cardiovasc. Disord.* 23 (1) (2023) 135.
- [137] J. Xie, W. Liao, W. Chen, D. Lai, Q. Tang, Y. Li, Circulating long non-coding RNA TTY15 and HULC serve as potential novel biomarkers for predicting acute myocardial infarction, *BMC Cardiovasc. Disord.* 22 (1) (2022) 86.
- [138] W. Zhu, L. Luo, G. Ye, J. Ou, Potential diagnostic value of NILR and SNHG1 in acute myocardial infarction, *BMC Med. Genomics* 16 (1) (2023) 71.
- [139] O. Beylerli, I. Gareev, A. Sufianov, T. Ilyasova, Y. Guang, Long noncoding RNAs as promising biomarkers in cancer, *Noncoding RNA Res.* 7 (2) (2022) 66–70.
- [140] L. Ma, J. Cao, L. Liu, Q. Du, Z. Li, D. Zou, V.B. Bajic, Z. Zhang, LncBook: a curated knowledgebase of human long non-coding RNAs, *Nucleic Acids Res.* 47 (D1) (2019) D128–D134.
- [141] S.-F. Huang, X.-F. Peng, L. Jiang, C.Y. Hu, W.-C. Ye, LncRNAs as therapeutic targets and potential biomarkers for lipid-related diseases, *Front. Pharmacol.* 12 (2021) 729745.
- [142] Y. Chen, S. Li, Y. Zhang, M. Wang, X. Li, S. Liu, D. Xu, Y. Bao, P. Jia, N. Wu, The lncRNA Malat1 regulates microvascular function after myocardial infarction in mice via miR-26b-5p/Mfn1 axis-mediated mitochondrial dynamics, *Redox Biol.* 41 (2021) 101910.
- [143] T. Lucas, A. Bonauer, S. Dimmeler, RNA therapeutics in cardiovascular disease, *Circ. Res.* 123 (2) (2018) 205–220.
- [144] Y. Yang, M. Li, Y. Liu, Z. Wang, X. Fu, X. He, Q. Wang, X.X. Li, H. Ma, K. Wang, L. Zou, J.X. Wang, T. Yu, The lncRNA punisher regulates apoptosis and mitochondrial homeostasis of vascular smooth muscle cells via targeting miR-664a-5p and OPA1, *Oxid. Med. Cell. Longev.* 2022 (2022) 5477024.
- [145] Y. Luo, H. Xu, Z. Yang, X. Lin, F. Zhao, Y. Huang, Y. Wang, X. Yang, H. Li, L. Wang, M. Wen, S. Xian, Long non-coding RNA MALAT1 silencing elevates microRNA-26a-5p to ameliorate myocardial injury in sepsis by reducing regulator of calcineurin 2, *Arch. Biochem. Biophys.* 715 (2022) 109047.
- [146] Y.M. Ding, E.C. Chan, L.C. Liu, Z.W. Liu, Q. Wang, J.L. Wang, X.P. Cui, F. Jiang, X. S. Guo, Long noncoding RNAs: important participants and potential therapeutic targets for myocardial ischaemia reperfusion injury, *Clin. Exp. Pharmacol. Physiol.* 47 (11) (2020) 1783–1790.
- [147] H. Zhou, B. Wang, Y.X. Yang, Q.J. Jia, A. Zhang, Z.W. Qi, J.P. Zhang, Long noncoding RNAs in pathological cardiac remodeling: a review of the update literature, *Biomed. Res. Int.* 2019 (2019) 7159592.
- [148] P. Grote, L. Wittler, D. Hendrix, F. Koch, S. Währisch, A. Beisaw, K. Macura, G. Bläss, M. Kellis, M. Werber, B.G. Herrmann, The tissue-specific lncRNA Fendrr is an essential regulator of heart and body wall development in the mouse, *Dev. Cell* 24 (2) (2013) 206–214.
- [149] L. Jin, Q. Li, J. Li, Y. Pan, J. Zou, X. Wu, Z. Wang, Apela inhibits systemic and renal inflammatory reactions in mice with type I cardiorenal syndrome, *FASEB J.* 35 (10) (2021) e21907.
- [150] Y. Pan, Q. Li, H. Yan, J. Huang, Z. Wang, Apela improves cardiac and renal function in mice with acute myocardial infarction, *J. Cell Mol. Med.* 24 (18) (2020) 10382–10390.
- [151] Q. Huang, M. Pan, J.P. Zhou, F. Yin, Overexpression of long non-coding RNA ANRIL promotes post-ischaemic angiogenesis and improves cardiac functions by targeting Akt, *J. Cell Mol. Med.* 24 (12) (2020) 6860–6868.
- [152] C. Yang, J. Zheng, Y. Xue, H. Yu, X. Liu, J. Ma, L. Liu, P. Wang, Z. Li, H. Cai, The effect of MCM3AP-AS1/miR-211/KLF5/AGGF1 axis regulating glioblastoma angiogenesis, *Front. Mol. Neurosci.* 10 (2018) 437.
- [153] K. Chen, M. Xi, Q. Huang, H. Wu, G. Lu, S. Song, W. Shi, Long non-coding RNA MCM3AP antisense RNA 1 silencing upregulates microRNA-24-3p to accelerate

- proliferation and migration of vascular endothelial cells in myocardial infarction rats by reducing EIF4G2, *Cell Cycle* 21 (7) (2022) 674–684.
- [154] X. Wang, L. Ren, S. Chen, Y. Tao, D. Zhao, C. Wu, Long non-coding RNA MIR4435-2HG/microRNA-125a-5p axis is involved in myocardial ischemic injuries, *Bioengineered.* 13 (4) (2022) 10707–10720.
- [155] R. Kumarswamy, C. Bauters, I. Volkmann, F. Maury, J. Fetisch, A. Holzmann, G. Lemesle, P. de Groote, F. Pinet, T. Thum, Circulating long noncoding RNA, LIPCAR, predicts survival in patients with heart failure, *Circ. Res.* 114 (10) (2014) 1569–1575.
- [156] T. Mondal, S. Subhash, R. Vaid, S. Enroth, S. Uday, B. Reinius, S. Mitra, A. Mohammed, A.R. James, E. Hoberg, MEG3 long noncoding RNA regulates the TGF- β pathway genes through formation of RNA-DNA triplex structures, *Nat. Commun.* 6 (1) (2015) 7743.
- [157] C. Schneider, R.M. King, L. Philipson, Genes specifically expressed at growth arrest of mammalian cells, *Cell* 54 (6) (1988) 787–793.
- [158] T. Kino, D.E. Hurt, T. Ichijo, N. Nader, G.P. Chrousos, Noncoding RNA gas5 is a growth arrest-and starvation-associated repressor of the glucocorticoid receptor, *Sci. Signal.* 3 (107) (2010) ra8-ra8.
- [159] C. Guo, Y. Qi, J. Qu, L. Gai, Y. Shi, C. Yuan, Pathophysiological functions of the lncRNA TUG1, *Curr. Pharm. Des.* 26 (6) (2020) 688–700.
- [160] V. Singh, D. Braddick, P.K. Dhar, Exploring the potential of genome editing CRISPR-Cas9 technology, *Gene* 599 (2017) 1–18.
- [161] W. Pan, Y. Zhu, X. Meng, C. Zhang, Y. Yang, Y. Bei, Immunomodulation by exosomes in myocardial infarction, *J. Cardiovasc. Transl. Res.* 12 (1) (2019) 28–36.
- [162] H. Wang, L. Shu, N. Niu, C. Zhao, S. Lu, Y. Li, H. Wang, Y. Liu, T. Zou, J. Zou, X. Wu, Y. Wang, Novel lncRNAs with diagnostic or prognostic value screened out from breast cancer via bioinformatics analyses, *PeerJ* 10 (2022) e13641.
- [163] G.E. Grieco, G. Sebastiani, D. Fignani, N. Brusco, L. Nigi, C. Formichi, G. Licata, M. Bruttini, R. D'Aurizio, C. Mathieu, C. Gysemans, F. Dotta, Protocol to analyze circulating small non-coding RNAs by high-throughput RNA sequencing from human plasma samples, *STAR Protocols.* 2 (3) (2021) 100606.
- [164] J.B. Pierce, H. Zhou, V. Simion, M.W. Feinberg, Long noncoding RNAs as therapeutic targets, *Adv. Exp. Med. Biol.* 1363 (2022) 161–175.
- [165] X. Xu, T. Wan, H. Xin, D. Li, H. Pan, J. Wu, Y. Ping, Delivery of CRISPR/Cas9 for therapeutic genome editing, *J. Gene Med.* 21 (7) (2019) e3107.