# Research Article

# Predictive Study of the Active Ingredients and Potential Targets of *Codonopsis pilosula* for the Treatment of Osteosarcoma via Network Pharmacology

Yu-Bao Gong <sup>(1)</sup>, <sup>1</sup> Shao-Jie Fu, <sup>2</sup> Zheng-Ren Wei, <sup>3</sup> and Jian-Guo Liu<sup>1</sup>

<sup>1</sup>Department of Orthopedics, The First Hospital, Jilin University, Changchun 130021, China <sup>2</sup>Department of Nephrology, The First Hospital, Jilin University, Changchun 130021, China <sup>3</sup>Department of Pharmacology, The Basic Medical School, Jilin University, Changchun, Jilin 130021, China

Correspondence should be addressed to Yu-Bao Gong; gongyb@jlu.edu.cn

Received 20 April 2020; Revised 29 October 2020; Accepted 25 May 2021; Published 7 June 2021

Academic Editor: Gauhar Rehman

Copyright © 2021 Yu-Bao Gong et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Osteosarcoma (OS) is the most common type of primary bone tumor in children and adults. Dangshen (*Codonopsis pilosula*) is a traditional Chinese medicine commonly used in the treatment of OS worldwide. However, the molecular mechanisms of Dangshen in OS remain unclear. Hence, in this study, we aimed to systematically explore the underlying mechanisms of Dangshen in the treatment of OS. Our study adopted a network pharmacology approach, focusing on the identification of active ingredients, drug target prediction, gene collection, gene ontology (GO) enrichment, Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment, and other network tools. The network analysis identified 15 active compounds in Dangshen that were linked to 48 possible therapeutic targets related to OS. The results of the gene enrichment analysis show that Dangshen produces a therapeutic effect in OS likely by regulating multiple pathways associated with DNA damage, cell proliferation, apoptosis, invasion, and migration. Based on the network pharmacology approach, we successfully predicted the active compounds and their respective targets. In addition, we illustrated the molecular mechanisms that mediate the therapeutic effect of Dangshen in OS. These findings may aid in the development of novel targeted therapies for OS in the future.

# 1. Introduction

Osteosarcoma (OS) is the most common type of cancer of the bones. It is a malignant tumor that primarily affects the long bones (e.g., legs), but it can also start in other bones. OS is rarely diagnosed in patients under five years of age, and the bimodal age-incidence curve peaks during the second decade of life (10–20 years old) and late adulthood (>40 years old) [1, 2]. As of 2019, approximately 560 children and adolescents are affected each year in the United States [1, 3], with a global incidence of 3.4 cases per one million people. Most OS patients present with metastatic disease, which contributes to its high morbidity and mortality rates worldwide. As of 2020, the standard treatment for OS is systemic chemotherapy [4], as the tumor is often resistant to radiation therapy. Surgical resection may be an option for patients diagnosed with locally noninvasive disease [5]. Most patients undergo multifaceted treatments that include preoperative chemotherapy, postoperative chemotherapy, surgical resection, and radiation therapy in rare cases [6]. Detectable metastases are present in only 20% of patients, and most of the remaining 80% of patients have undetectable micrometastases [7]. This makes it challenging to monitor disease progression and treatment response, which is why many physicians rely on long-term systemic chemotherapy [8]. However, the systemic chemotherapeutics needed to control the disease have serious adverse effects that further hinder the effective treatment of patients in the clinic [9].

In recent years, some hospitals have assessed the efficacy of traditional Chinese medicines as long-term treatments for OS. In some cases, orally administered Shenqi could decrease the growth, metastasis, and the number of chemotherapy-related side effects significantly in patients, especially those who received systemic chemotherapy [10, 11]. The major component of the Shenqi oral preparation is Dangshen, also known as (Codonopsis pilosula). Dangshen belongs to the family of Campanulaceae, a precious plant that grows at altitudes of 2000 meters in southern China [12]. The dry roots of this plant have been used for thousands of years in traditional Chinese medicine to treat qi and blood deficiencies, the loss of appetite, respiratory symptoms (e.g., cough, asthma, and shortness of breath), and cardiovascular problems (e.g., palpitations) [13]. Dangshen displays a variety of pharmacological effects on the circulatory system, immune system, digestive system, endocrine system, and reproductive system [14]. Dangshen has been shown to inhibit cancer growth in S180 tumorbearing mice, while enhancing the immune response, increasing spleen weight, promoting lymphocyte proliferation, and increasing natural killer (NK) cell activity [15, 16].

DNA damaging therapies are widely used as trigger molecules to study the signaling pathways of OS [17]. There has been a remarkable undertaking of investigations into the different signaling pathways involved in the pathogenesis of OS. Many signaling pathways, such as Wnt, PI3K/AKT, and JAK/STAT, reflect their specific roles in OS [18]. However, conventional research methods have been unable to fully elucidate the mechanisms of action. Nevertheless, the integration of bioinformatics and network pharmacology provides a practical approach to explore and verify the mechanisms of action [19]. Network pharmacology can systematically reveal the active components in drug molecules. In addition, network pharmacology can be used to predict the relationship between drug components and gene targets [20]. Therefore, in this study, we aimed to use network pharmacology to uncover the mechanisms by which Dangshen produces therapeutic effects in patients with OS, along with the associated signaling pathways.

## 2. Materials and Methods

2.1. Chemical Compounds in Dangshen. A flowchart of the study design is shown in Figure 1. The components of Dangshen were searched in the Traditional Chinese Medicine Systems Pharmacology (TCMSP) (http://tcmspw.com/ tcmsp.php) database [21] and Traditional Chinese Medicines Integrated Database (TCMID) (http://www.megabionet. org/tcmid/) [22]. TCMSP provides comprehensive information about components in Chinese herbs, while TCMID provides information on all aspects of traditional Chinese medicines, including herbs and herbal ingredients. Oral bioavailability (OB), which is the percentage of an orally administered drug that reaches the systemic circulation, reflects the degree of absorption and utilization of drugs in the body [23]. The drug-likeness (DL) value reflects the structural similarity between the compound and drug molecule, so the DL compounds are more likely to display suitable pharmacodynamic and pharmacokinetic properties [24]. Therefore, we selected the candidate compounds based on OB and DL properties. As suggested by the TCMSP database,  $OB \ge 30\%$  and  $DL \ge 0.18$  were used as the

screening criteria, and the compounds whose  $OB \ge 30\%$  and  $DL \ge 0.18$  were selected for subsequent experiments [25]. We searched the oral bioavailability of all the compounds of Dangshen on PubMed. If the OB data of some compounds were previously reported in related experiments, the real-world data of OB were used instead of silicon data, and the highest reported OB value was adopted. Otherwise, silicon data were used for OB. The TCMSP database calculated OB values by using OBioavail1.1. This model shows good potential in facilitating the prediction of oral bioavailability and can be applied in drug design.

2.2. Compounds of Dangshen and Their Targets. PubChem (https://pubchem.ncbi.nlm.nih.gov/) is an open chemistry database of the National Institutes of Health (NIH) [26]. This database serves as an important source of chemical information, including chemical structures, biological activities, chemical and physical properties, and safety [27]. We imported the compounds filtered from Dangshen into Pub-Chem and obtained their 3D molecular structure files in SDF format. Structural information is necessary for predicting the targets of compounds, so the compounds without precise structural details were removed from the analysis.

PharmMapper (http://www.lilab-ecust.cn/pharmm apper/check.html) is a freely accessed web server that uses the pharmacophore mapping approach to identify potential small-molecule targets [28, 29]. We imported the 3D structural files in SDF format into PharmMapper and selected the pharmacophore model with a pKd value  $\geq 6.0$ . In our study, the top matched 50 targets were selected as the potential targets of each compound.

2.3. Collection of Gene Targets in OS. The human genes associated with OS were gathered from OMIM (Online Mendelian Inheritance in Man, https://omim.org/) and GeneCards (https://www.genecards.org/). OMIM is an authoritative and comprehensive database of human genes and genetic phenotypes [30], while GeneCards is an integrative database that provides information on all predicted and annotated human genes [31]. The search term "osteosarcoma" was used to retrieve the OS targets from both databases.

2.4. Therapeutic Targets of Dangshen in OS. We screened the active compounds of Dangshen and obtained their target genes. We also gathered the OS-related genes. The potential therapeutic targets were identified from the shared genes mentioned above.

2.5. Protein-Protein Interaction (PPI) Data. The therapeutic targets were imported into the STRING database to obtain their interaction relationship. STRING (https://string-db. org/, version 11.0) is a database that contains known and predicted protein-protein interactions, and it collects the information using bioinformatics strategies [32]. The species were limited to "Homo sapiens," and the PPIs with confidence scores >0.4 were selected for this study.



FIGURE 1: Schematic illustration showing the network pharmacology study of Dangshen (*Codonopsis pilosula*) for the treatment of osteosarcoma (OS).

2.6. Target Organs. Data about the organ targets were collected from the BioGPS (http://biogps.org) database. BioGPS is an extensible and customizable genetic annotation portal that enables researchers to acquire distributed genetic annotation resources [33]. To reveal the underlying mechanisms of Dangshen in OS, median gene expression levels were used as the standard to screen for organs with high expression of the therapeutic targets. 2.7. Network Construction. The network of active compounds and therapeutic targets was constructed by linking the compounds and therapeutic targets to understand the complex interactions between the compounds of Dangshen and the therapeutic targets of OS. The network of therapeutic targets and organs was established by linking therapeutic targets and their distribution in organs to clarify the relationship between the therapeutic targets and organs with increased expression of the target. The therapeutic targets' PPI network was built by linking the therapeutic targets to their interacting targets. Next, Cytoscape version 3.7.2 (http://www.cytoscape.org/) was used to present the networks mentioned above, which is a software program for network visualization [34]. Lastly, NetworkAnalyzer [35] was used to calculate three topological parameters of each node in the network, including the degree, betweenness centrality, and closeness centrality [36].

2.8. GO and KEGG Pathway Enrichment Analysis. To learn more about the role of therapeutic targets involved in the biological process (BP), cell component (CC), and molecular function (MF), we used the Gene Ontology (GO) database (http://geneontology.org/) to clarify the possible biological mechanisms [37]. The Kyoto Encyclopedia of Genes and Genomes (KEGG) (https://www.kegg.jp/) is a database for extracting biological information about functional classification, annotation, and enriched pathways of various genes [38]. In this study, we used an R-package-Bioconductor clusterProfiler to perform the GO and KEGG enrichment analysis. The R-package-Bioconductor clusterProfiler is widely used to automate the biological term classification and enrichment analysis of gene clusters [39].

#### 3. Results

3.1. Chemical Compounds of Dangshen. Using the keyword search in TCMSP and TCMID, a total of 134 components of Dangshen were identified, including flavonoids, steroids, alkaloids, glycosides, and triterpenes. According to the OB and DL characteristics of the ingredients, 25 screened compounds were chosen for the next experiments. As structural information is essential for predicting the targets of a compound, ten compounds without 3D structural information were discarded. Finally, 15 compounds were determined as possible active compounds whose characteristics are listed in Table 1.

*3.2. Dangshen Compound Targets.* We obtained the top 50 matched targets for each potential active compound from PharmMapper. These targets were regarded as the potential targets of Dangshen (Supplementary Table S1).

3.3. Collection of Gene Targets for OS. "Osteosarcoma" was used as the keyword to retrieve the OS targets from OMIM and GeneCards databases. A total of 2,079 genes were retrieved from the two databases (Supplementary Table S2).

3.4. Therapeutic Targets of Dangshen for OS. The targeted genes of Dangshen and OS were obtained. Using the shared genes described above, 48 possible therapeutic targets were obtained, and the features are listed in Table 2.

3.5. Active Compound-Therapeutic Target Network. The active compound-therapeutic target network is depicted in Figure 2. This network demonstrates the complicated relationship between the compounds and therapeutic targets, including 65 total nodes (15 compound nodes, 48 therapeutic target nodes, one Dangshen node, and one OS node) and 204 edges. In Figure 2, the therapeutic targets are represented by green ovals, Dangshen is represented by a blue quadrangle, OS is represented by a red hexagon, active compounds are represented by yellow triangles, and the sizes of compound nodes were proportional to their degree. The three with the highest degree of the compound nodes were Frutinone A (degree = 16), Perlolyrine (degree = 14), and Glycitein (degree = 13). The three compounds were more likely to show significant therapeutic activity against OS.

3.6. Therapeutic Target-PPI Network. The PPI network of the therapeutic targets is shown in Figure 3, including 48 nodes and 304 edges. NetworkAnalyzer was employed to calculate three topological features of the 48 targets to identify the key nodes in the network (Table 2). The median values of the degree, node betweenness, and closeness were 10, 0.047, and 0.549, respectively. The nodes with "degree >10," "node betweenness >0.047," and "node closeness >0.63" were considered to be the key targets. Hence, 20 genes were identified as central targets of Dangshen against OS, including TP53, HSP90AA1, CCND1, AR, ERBB2, MDM2, IGF1R, DICER1, CCNE1, SOD2, among others.

3.7. Therapeutic Target-Organ Network. The organs with high expression of each therapeutic target were collected via BioGPS (Supplementary Table S3). The therapeutic targetorgan network, shown in Figure 4, is used to delineate the relationship between therapeutic targets and the organs that highly express these targets, including 132 nodes (58 therapeutic target nodes and 84 organ nodes) and 2,031 edges. The color shade of the organ node is proportional to its degree, as shown in Figure 4. These findings demonstrate that many therapeutic targets are highly expressed in tissues, such as the thyroid, retina, pituitary, and pineal gland, and on the surface of antigens, including CD33, CD34, and CD56.

3.8. GO and KEGG Pathway Enrichment. To illuminate the complex mechanisms of Dangshen against OS, we conducted analyses of the GO biological process (BP), cell component (CC), and molecular function (MF) for the 48 therapeutic targets. The top ten biological processes, cell components, and molecular functions are shown in Figures 5(a), 6(a), and 7(a), respectively. The relationship between the genes and biological processes, cell component, and molecular function targets is depicted in Figures 5(b), 6(b), and 7(b), respectively. The details of the GO enrichment analysis of BP, CC, and MF are listed in Supplementary Tables S4–S6, respectively.

KEGG pathway enrichment analysis was performed to explore the underlying mechanisms of Dangshen against OS further. As shown in Supplementary Table S7 and Figure 8, there are 69 primary pathways that participate in Dangshen against OS with p < 0.05. These 69 pathways involve human Evidence-Based Complementary and Alternative Medicine

Compound	MF	Structure	MW	OB (%)	DL	HL
Stigmasterol	$C_{29}H_{48}O$	fx1	412.77	43.83	0.76	5.57
Stigmast-7-enol	C29H50O	fx2	414.79	37.42	0.75	6.28
Luteolin	$C_{15}H_{10}O_{6}$	fx3	286.25	32.00	0.25	15.94
11-Hydroxyrankinidine	$C_{20}H_{24}N_2O_4$	fx4	356.46	40.00	0.66	10.80
Perlolyrine	$C_{16}H_{12}N_2O_2$	fx5	264.30	65.95	0.27	12.62
Glycitein	$C_{16}H_{12}O_5$	fx6	284.28	50.48	0.24	16.32
Spinasterol	$C_{29}H_{48}O$	fx7	412.77	42.98	0.76	5.32
Frutinone A	$C_{16}H_8O_4$	fx8	264.24	65.90	0.34	19.10
Poriferasta-7,22E-dien-3beta-ol	$C_{29}H_{48}O$	fx9	412.77	42.98	0.76	5.48
7-Methoxy-2-methyl isoflavone	C <sub>17</sub> H <sub>14</sub> O <sub>3</sub>	fx10	266.31	42.56	0.20	16.89
5-α-Stigmastan-3,6-dione	$C_{29}H_{48}O_2$	fx11	428.77	33.12	0.79	5.19
3-β-Hydroxymethyllenetanshiquinone	$C_{18}H_{14}O_{4}$	fx12	294.32	32.16	0.41	22.51
Zinc03978781	C <sub>29</sub> H <sub>48</sub> O	fx13	412.77	43.83	0.76	5.79
Taraxerol	C <sub>30</sub> H <sub>50</sub> O	fx14	426.80	38.40	0.77	2.07
Stigmasterone	$C_{29}H_{46}O$	fx15	410.75	45.40	0.76	5.65

TABLE 1: Characteristics of the active ingredients.

TABLE 2: Characteristics of the 48 therapeutic targets.

Target	Name	Degree	Betweenness centrality	Closeness centrality
TP53	Cellular tumor antigen p53	40	0.19161047	0.87037037
HSP90AA1	Heat shock protein HSP 90-alpha	33	0.08748708	0.77049180
CCND1	G1/S-specific cyclin-D1	30	0.05438164	0.73437500
AR	Androgen receptor	29	0.08088557	0.72307692
MDM2	E3 ubiquitin-protein ligase Mdm2	28	0.05726816	0.70149254
ERBB2	Receptor tyrosine-protein kinase erbB-2	28	0.05040772	0.71212121
IGF1R	Insulin-like growth factor-1 receptor	27	0.04394018	0.70149254
DICER1	Endoribonuclease Dicer	24	0.03867798	0.66197183
CCNE1	G1/S-specific cyclin-E1	19	0.01018846	0.61842105
THBS1	Thrombospondin-1	19	0.01547887	0.61038961
SOD2	Superoxide dismutase [Mn], mitochondrial	18	0.01677310	0.61038961
XRCC6	ATP-dependent DNA helicase 2 subunit 1	16	0.02818547	0.59493671
MAPK9	Mitogen-activated protein kinase 9	16	0.01100001	0.59493671
E2F1	Transcription factor E2F1	15	0.00460006	0.58750000
MUC1	Mucin-1	15	0.01251038	0.58024691
CUL1	Cullin-1	14	0.02049293	0.56626506
RAC1	Ras-related C3 botulinum toxin substrate 1	14	0.04760817	0.58750000
B2M	Beta-2-microglobulin	14	0.01143842	0.58024691
MPO	Myeloperoxidase	14	0.01314265	0.57317073
HDAC6	Histone deacetylase 6	12	0.00411776	0.56626506
MAP2K2	Dual specificity mitogen-activated protein kinase kinase 2	12	0.00552768	0.55294118
EZR	Ezrin	12	0.00234926	0.57317073
PAX6	Paired box protein Pax-6	12	0.00636814	0.55952381
TP73	Tumor protein p73	10	0.00045360	0.54022989
BMPR2	Bone morphogenetic protein receptor type 2	10	0.00217280	0.55294118
RPS3	40S ribosomal protein S3	9	0.00443058	0.53409091
HDAC8	Histone deacetylase 8	9	0.00168382	0.54022989
POLB	DNA polymerase $\beta$	9	0.00620249	0.54651163
FOLH1	Glutamate carboxypeptidase 2	9	0.00471413	0.54651163
SMAD1	Mothers against decapentaplegic homolog 1	8	0.00048566	0.52808989
ATIC	Bifunctional purine biosynthesis protein PURH	8	0.00650462	0.52222222
S100A6	Protein S100-A6	8	0.00018501	0.53409091
NR3C2	Mineralocorticoid receptor	7	0.00397370	0.52808989
EIF2AK2	Interferon-induced, double-stranded RNA-activated protein kinase	7	0.00917844	0.52808989
RPA1	Replication protein A 70 kDa DNA-binding subunit	6	0.00034097	0.51648352
SATB2	DNA-binding protein SATB2	5	0.00026981	0.49473684
CDCA8	Borealin	5	0	0.49473684
FLI1	Friend leukemia integration 1 transcription factor	5	0.00061402	0.50000000
NHP2L1	Human recombinant protein P01	4	0.00020331	0.47000000
CAMK2A	Calcium/calmodulin-dependent protein kinase type II $\alpha$ chain	4	0	0.46078431

TABLE	2:	Continued.
-------	----	------------

Target	Name	Degree	Betweenness centrality	Closeness centrality
HSD11B1	Corticosteroid 11- $\beta$ -dehydrogenase isozyme 1	4	0.00067245	0.44761905
ASS1	Argininosuccinate synthase	4	0	0.49473684
FAP	Seprase	4	0.00011563	0.44339623
CANT1	Soluble calcium-activated nucleotidase 1	3	0.00036617	0.44761905
BCL9	B-cell CLL/lymphoma 9 protein	3	0	0.48453608
CSDE1	Cold shock domain-containing protein E1	3	0.00053191	0.41592920
BRD7	Bromodomain-containing protein 7	2	0	0.47959184
SRGAP2	SLIT-ROBO rho GTPase-activating protein 2	1	0	0.37301587



FIGURE 2: Active compounds-therapeutic targets network. Yellow triangles represent the active compounds from Dangshen, while green ovals represent the therapeutic targets. The size of the triangles is directly proportional to their degree.



FIGURE 3: PPI network of therapeutic targets. Hexagons represent the therapeutic targets, and the color shade of hexagons is directly proportional to their degree.

diseases, pathophysiological mechanisms, and signaling pathways. The top ten significantly enriched signaling pathways include the p53 signaling pathway, PI3K-Akt signaling pathway, neurotrophin signaling pathway, FoxO signaling pathway, Wnt signaling pathway, ErbB signaling pathway, TGF- $\beta$  signaling pathway, HIF-1 signaling pathway, sphingolipid signaling pathway, and MAPK signaling pathway. Many therapeutic targets are involved in these signaling pathways. Figure 9 depicts a concept map containing Dangshen and OS targets in the P53 signaling pathway, further demonstrating that Dangshen regulates key targets in this signaling pathway.

#### 4. Discussion

Osteosarcoma (OS) is the most common primary bone tumor found in the clinic [40]. It is characterized by high metastatic rates, poor prognoses, and high mortality rates [41]. Dangshen (*Codonopsis pilosula*) is a well-known herbal medicine, and traditional Chinese medicine (TCM) preparation, based on Dangshen, which has shown high efficacy in the treatment of OS [10, 11]. However, its pharmacological mechanisms remain unclear. In the present study, we used network pharmacology to explore the potential active compounds and underlying mechanisms of Dangshen against OS.



FIGURE 4: Therapeutic target-organs network. Red ovals represent the tissues with high expression levels of the targets, and the color shade of the red ovals is directly proportional to the degree. Green ovals represent therapeutic targets.

After applying the screening methods, we identified 15 active compounds and 48 potential therapeutic targets. The active compounds of Dangshen likely treat OS by regulating these targets. We identified two active compounds, stigmasterol and luteolin, that have been studied previously for their efficacy against OS. Stigmasterol is a phytosterol, which has been shown to exert anticancer, antipyretic, and immune-modulating properties [42-44]. Previously, Trouillas et al. showed that stigmasterol could decrease the proliferation of OS cells [45]. Luteolin is a flavonoid found in vegetables and fruits. It can inhibit the proliferation and induce the apoptosis of OS cells by effectively downregulating the expression of BCL-2, caspase-3, and survivin proteins levels, while upregulating BAX protein levels [46]. In addition, it can induce autophagy in U2OS cells and enhance the sensitivity of these cells to doxorubicin-mediated autophagy signaling [47].

From the therapeutic target-PPI network, the following targets showed larger degree values: TP53, HSP90AA1, CCND1, AR, ERBB2, MDM2, IGF1R, DICER1, CCNE1, and SOD2. These targets may play a major role in the therapeutic effect of Dangshen against OS. Over 70% of OS cases show structural variants or mutations in the TP53 gene [48]. TP53 is a transcription factor that stabilizes following genotoxic stress and induces the transcription of genes associated with cell apoptosis, cycle arrest, and metabolism; thereby, suppressing the development and progression of tumors [49, 50]. HSP90AA1, a 90-kDa heat shock protein [51], is an important target for cancer treatment because it can stabilize several cancer-related client proteins essential for tumor progression, such as AKT, PIM1, and HIF1A [52]. Some studies found that, in tumor biopsies, the absence of HSP90AA1 may serve as a biomarker of favorable outcomes

[53, 54]. CCND1 is a member of the cyclin family that encodes cyclin-D1. In addition, it plays a key role in cell cycle regulation [55]. There is substantial evidence showing that CCND1 plays an important role in the development of human cancers [56], including the migration and metastasis of OS [57]. The ERBB family of tyrosine kinases plays an important role in cell cycle regulation, cell proliferation, and cell movement [58]. Tumors that overexpress ERBB2 are less likely to respond to anticancer therapies [59]. Previously, Abdou et al. reported on the overexpression of ERBB2 in OS and its adverse prognostic features, including higher tumor grades [60]. In addition, Wang et al. reported that chimeric anti-caspase-6 and anti-ERBB2 antibodies reduced the metastatic potential of human OS cells [61]. These findings suggest that the therapeutic effect of Dangshen against OS is primarily mediated by cell apoptosis, cell cycle arrest, and the inhibition of tumor cell migration and metastasis.

Next, we performed the GO enrichment analysis and KEGG pathway enrichment analysis of the therapeutic targets. Based on the GO terms, the therapeutic targets showed a strong correlation with the biological processes, such as the G1/S transition of mitotic cell cycle, cell cycle G1/ S phase transition, regulation of cell cycle arrest, and the intrinsic apoptotic signaling pathway; cell components, such as cell leading edge, ruffle, ruffle membrane, and endocytic vesicle; and molecular functions, such as p53 binding, disordered domain specific binding, histone deacetylase binding, damaged DNA binding, and ATPase binding. Hence, the mechanism of action for Dangshen may include biological processes, molecular functions, and various cellular components. For example, imbalanced cell cycle regulation is characteristic of tumor cells, and functional defects in cell cycle checkpoints lead to genetic changes that lead to



FIGURE 5: Top ten significant biological process (BP) entries. (a): GO enrichment analysis of therapeutic targets for biological process. (b): Relationship between the therapeutic targets and biological process.





FIGURE 6: Top ten significant cell component (CC entries). (a): GO enrichment analysis of therapeutic targets for cell components. (b) Relationship between the therapeutic targets and cell components.



FIGURE 7: Continued.





FIGURE 7: Top ten significant molecular function (MF) entries. (a): GO enrichment analysis of therapeutic targets for molecular function. (b) Relationship between the therapeutic targets and molecular function.

tumor development and progression [62, 63]. In addition, the G1/S phase transition is the target of many anticancer drugs [64]. Apoptosis is a form of cell death that occurs upon receipt of internal or external death signals [65, 66]. In addition, Chaiyawat et al. reported that reduced expression of histone deacetylase-2 of HDAC2 is associated with dismal patient outcomes in OS [67]. Furthermore, Sun et al. reported that histone deacetylase-2 may stimulate the ATM/ p53 pathway, leading to DNA damage-mediated cell death in human OS cells [68]. In addition, Cao et al. found that overexpression of histone deacetylase-4 promotes the proliferation and invasion of OS cells [69].

Based on the KEGG terms, the therapeutic targets for Dangshen against OS were primarily associated with the p53 signaling pathway, PI3K-Akt signaling pathway, FoxO signaling pathway, Wnt signaling pathway, and ErbB signaling pathway. P53 plays a critical role in cell cycle checkpoints regulation, DNA damage, and prevention of nonmalignant cells from developing malignant phenotypes [70, 71]. In addition, p53 is an essential regulator of epithelial-mesenchymal transition (EMT) [72], as it promotes the reversal of mesenchymal cells to the epithelial cell phenotype, which reduces the migration and invasion of cells [73]. Many anticancer drugs regulate the p53 signaling pathway. For example, theabrownin triggers DNA damage and induces

apoptosis in U2OS cells via p53 signaling activation [74]. Activation of the PI3K-Akt signaling pathway is also associated with cell proliferation and apoptosis of OS cells [75, 76], and the downregulation of AKT reduces cyclin-D1 levels, preventing cells from cycling from G1 to S [77]. The reduced expression of cyclin-D1 also leads to the inhibition of cell proliferation [78]. Simultaneously, AKT downregulates the expression of two essential proteins responsible for apoptosis, caspase-3 and caspase-8 [79]. Abnormal Wnt/  $\beta$ -catenin signaling is closely related to the formation, metastasis, and apoptosis of many cancers [80]. The upregulation of Wnt/ $\beta$ -catenin signaling was recently observed in OS [81]. As such, the WIF-1 protein, encoded by Wnt inhibitory factor-1 gene, is an important regulatory factor in the Wnt signaling pathway [82]. The WIF-1 gene combines with the Wnt protein to prevent Wnt signaling [83]. Previously, Li et al. reported on the downregulation of WIF-1 in OS cells [84]. Hence, the KEGG analysis revealed that Dangshen produces anticancer effects in OS through the regulation of several proteins, including MDM2, TP53, RAC1, ERBB2, and CCND1, which are all important mediators of various cellular signaling pathways. In addition, most therapeutic targets play their roles in multiple signaling pathways. In addition, most of the therapeutic targets play essential roles in multiple signaling pathways.



FIGURE 8: KEGG enrichment analysis for therapeutic targets.

Network pharmacology is an analytical method still in development worldwide. However, the method has some inherent flaws. For example, it heavily relies on existing resources of the databases, so it cannot analyze the compounds, targets, or mechanisms that have not been previously explored. Moreover, its predicted active ingredients, targets, and mechanisms of action are all purely theoretical, and there is a lack of experimental verification. Therefore, further clinical investigations are needed.



FIGURE 9: Modulation of the p53 signaling pathway by Dangshen. Targets associated with OS are in red and green, targets of Dangshen are in green, and other proteins in the pathway are in blue.

#### **5. Conclusions**

In this study, we explored the therapeutic mechanisms of Dangshen against OS through a network pharmacology approach. The therapeutic properties of Dangshen against OS arise from the regulation of biological pathways involved in the proliferation, apoptosis, invasion, and migration of cells, along with DNA damage. We believe these findings demonstrate the importance of understanding traditional Chinese medicines. The current study relied on data mining and analysis, and further clinical investigations are needed to verify the therapeutic mechanisms of Dangshen against OS.

# **Data Availability**

The data sets generated and analyzed during the present study are available from the corresponding author upon reasonable request.

# **Conflicts of Interest**

The authors declare that they have no conflicts of interest.

# Acknowledgments

This work was supported by the Science and Technology Development Project of Jilin Province (20190201081JC) and Health Special Project of Jilin Province (JLSWSRCZX2020-0053).

#### **Supplementary Materials**

Description of Supplementary Table S1: the top 50 matched targets for each potential active compound of Dangshen were gathered from PharmMapper, and these targets were regarded as the potential targets of Dangshen. The relationship between the compounds and their matched targets is listed in Supplementary Table S1. Description of Supplementary Table S2: the genes related to osteosarcoma were collected from GeneCards and OMIM databases. The genes related to osteosarcoma are listed in Supplementary Table S2. Description of Supplementary Table S3: the organs with high expression of each therapeutic target were collected via BioGPS. The relationship between the therapeutic targets and their high expressed organs is listed in Supplementary Table S3. Description of Supplementary Table S4: we used the Gene Ontology (GO) database to clarify the possible biological mechanisms. The details of the GO enrichment analysis of biological process are listed in Supplementary Table S4. Description of Supplementary Table S5: we used the Gene Ontology (GO) database to clarify the possible biological mechanisms. The details of the GO enrichment analysis of cell component are listed in Supplementary Table S5. Description of Supplementary Table S6: we used the Gene Ontology (GO) database to clarify the possible biological mechanisms. The details of the GO enrichment analysis of molecular function are listed in Supplementary Table S6. Description of Supplementary Table S7: we used the Kyoto Encyclopedia of Genes and Genomes (KEGG) database to extract biological information about functional classification, annotation, and enriched pathways of therapeutic targets. The details of the KEGG enrichment analysis are listed in Supplementary Table S7. . (*Supplementary Materials*)

## References

- D. D. Moore and H. H. Luu, "Osteosarcoma," Cancer Treatment and Research, vol. 162, pp. 65–92, 2014.
- [2] P. A. Meyers and R. Gorlick, "Osteosarcoma," *Pediatric Clinics of North America*, vol. 44, no. 4, pp. 973–989, 1997.
- [3] G. Ottaviani and N. Jaffe, "The epidemiology of osteosarcoma," in *Pediatric and Adolescent Osteosarcoma*, N. Jaffe, O. S. Bruland, and S. Bielack, Eds., Springer, Boston, MA, USA, 2010.
- [4] S. Ferrari and M. Serra, "An update on chemotherapy for osteosarcoma," *Expert Opinion on Pharmacotherapy*, vol. 16, no. 18, pp. 2727–2736, 2015.
- [5] S. Toki, E. Kobayashi, A. Yoshida et al., "A clinical comparison between dedifferentiated low-grade osteosarcoma and conventional osteosarcoma," *The Bone & Joint Journal*, vol. 101, no. 6, pp. 745–752, 2019.
- [6] J. L. Ferguson and S. P. Turner, "Bone cancer: diagnosis and treatment principles," *American Family Physician*, vol. 98, no. 4, pp. 205–213, 2018.
- [7] R. Sasaki, M. Osaki, and F. Okada, "MicroRNA-based diagnosis and treatment of metastatic human osteosarcoma," *Cancers*, vol. 11, no. 4, p. 553, 2019.
- [8] P. J. Messerschmitt, R. M. Garcia, F. W. Abdul-Karim, E. M. Greenfield, and P. J. Getty, "Osteosarcoma," *Journal of the American Academy of Orthopaedic Surgeons*, vol. 17, no. 8, pp. 515–527, 2009.
- [9] D. B. Glasser, J. M. Lane, A. G. Huvos, R. C. Marcove, and G. Rosen, "Survival, prognosis, and therapeutic response in osteogenic sarcoma. The memorial hospital experience," *Cancer*, vol. 69, no. 3, pp. 698–708, 1992.
- [10] T. Kuang and Y. Liu, "Application of Shenqi fuzheng injection in 30 cases of osteosarcoma with high dose neoadjuvant chemotherapy," *Journal of Chinese Oncology*, vol. 15, no. 05, pp. 473-474, 2009.
- [11] Y. Hu, H. Li, and Y. Zuo, "Effect of Shenqi Fuzheng Injection on immune function in patients with osteosarcoma high-dose chemotherapy," *China Continuing Medical Education*, vol. 10, no. 05, pp. 149–151, 2018.
- [12] J.-S. Su, F.-Y. Qin, Y. Liu, and Y. Zhang, "Four new polyynes from Codonopsis pilosula collected in Yunnan province, China," *Natural Product Research*, pp. 1–8, 2020.
- [13] Y.-P. Fu, L.-X. Li, B.-Z. Zhang et al., "Characterization and prebiotic activity in vitro of inulin-type fructan from Codonopsis pilosula roots," *Carbohydrate Polymers*, vol. 193, pp. 212–220, 2018.
- [14] Y.-F. Zou, Y.-Y. Zhang, Y.-P. Fu et al., "A polysaccharide isolated from codonopsis pilosula with immunomodulation effects both in vitro and in vivo," *Molecules*, vol. 24, no. 20, p. 3632, 2019.
- [15] J. Chen, L. Hu, H. Wu et al., "Codonopsis pilosula polysaccharides on tumor-bearing mice immune response and antitumor effect," *Chinese Journal of Cancer Prevention and Treatment*, vol. 22, no. 17, pp. 1357–1362, 2015.
- [16] H. Wang, H. Lin, J. Tan et al., "Codonopsis pharmacological effects and clinical application research progress," *World Latest Medicine Information*, vol. 19, no. 07, pp. 21-22+4, 2019.

- [17] C. M. Hattinger, M. P. Patrizio, S. Luppi et al., "Current understanding of pharmacogenetic implications of DNA damaging drugs used in osteosarcoma treatment," *Expert Opinion on Drug Metabolism & Toxicology*, vol. 15, no. 4, pp. 299–311, 2019.
- [18] P. Angulo, G. Kaushik, D. Subramaniam et al., "Natural compounds targeting major cell signaling pathways: a novel paradigm for osteosarcoma therapy," *Journal of Hematology* & Oncology, vol. 10, no. 1, p. 10, 2017.
- [19] B. Jain, U. Raj, and P. K. Varadwaj, "Drug target interplay: a network-based analysis of human diseases and the drug targets," *Current Topics in Medicinal Chemistry*, vol. 18, no. 13, pp. 1053–1061, 2018.
- [20] L. Shao and B. J. C. Zhang, "Traditional Chinese medicine network pharmacology: theory, methodology and application," *Chinese Journal of Natural Medicines*, vol. 11, no. 2, pp. 110–120, 2013.
- [21] J. Ru, P. Li, J. Wang et al., "TCMSP: a database of systems pharmacology for drug discovery from herbal medicines," *Journal of Cheminformatics*, vol. 6, no. 1, p. 13, 2014.
- [22] R. Xue, Z. Fang, M. Zhang, Z. Yi, C. Wen, and T. Shi, "TCMID: traditional Chinese Medicine integrative database for herb molecular mechanism analysis," *Nucleic Acids Research*, vol. 41, pp. D1089–D1095, 2013.
- [23] Z. Karami, M. R. Saghatchi Zanjani, N. Nasihatsheno, and M. Hamidi, "Improved oral bioavailability of repaglinide, a typical BCS Class II drug, with a chitosan-coated nanoemulsion," *Journal of Biomedical Materials Research*, vol. 108, 2019.
- [24] C. A. Lipinski, "Drug-like properties and the causes of poor solubility and poor permeability," *Journal of Pharmacological* and Toxicological Methods, vol. 44, no. 1, pp. 235–249, 2000.
- [25] B. Gong, Y. Kao, C. Zhang, H. Zhao, F. Sun, and Z. Gong, "Exploring the pharmacological mechanism of the herb pair "HuangLian-GanJiang" against colorectal cancer based on network pharmacology," *Evidence-Based Complementary and Alternative Medicine*, vol. 2019, Article ID 2735050, 12 pages, 2019.
- [26] S. Kim, P. A. Thiessen, E. E. Bolton et al., "PubChem substance and compound databases," *Nucleic Acids Research*, vol. 44, pp. D1202–D1213, 2016.
- [27] S. Kim, J. Chen, T. Cheng et al., "PubChem 2019 update: improved access to chemical data," *Nucleic Acids Research*, vol. 47, pp. D1102–D1109, 2019.
- [28] X. Wang, Y. Shen, S. Wang et al., "PharmMapper 2017 update: a web server for potential drug target identification with a comprehensive target pharmacophore database," *Nucleic Acids Research*, vol. 45, pp. W356–W360, 2017.
- [29] X. Wang, C. Pan, J. Gong, X. Liu, and H. Li, "Enhancing the enrichment of pharmacophore-based target prediction for the polypharmacological profiles of drugs," *Journal of Chemical Information and Modeling*, vol. 56, no. 6, pp. 1175–1183, 2016.
- [30] J. S. Amberger and A. Hamosh, "Searching online mendelian inheritance in man (OMIM): a knowledgebase of human genes and genetic phenotypes," *Current Protocols in Bioinformatics*, vol. 58, pp. 1–12, 2017.
- [31] G. Stelzer, N. Rosen, I. Plaschkes et al., "The GeneCards suite: from gene data mining to disease genome sequence analyses," *Current Protocols in Bioinformatics*, vol. 54, pp. 1–33, 2016.
- [32] D. Szklarczyk, A. L. Gable, D. Lyon et al., "STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets," *Nucleic Acids Research*, vol. 47, pp. D607–D613, 2019.

- [33] C. Wu, X. Jin, G. Tsueng, C. Afrasiabi, and A. I. Su, "BioGPS: building your own mash-up of gene annotations and expression profiles," *Nucleic Acids Research*, vol. 44, pp. D313–D316, 2016.
- [34] M. E. Smoot, K. Ono, J. Ruscheinski, P.-L. Wang, and T. Ideker, "Cytoscape 2.8: new features for data integration and network visualization," *Bioinformatics*, vol. 27, no. 3, pp. 431-432, 2011.
- [35] H. De Jong, J. Geiselmann, C. Hernandez, and M. Page, "Genetic Network Analyzer: qualitative simulation of genetic regulatory networks," *Bioinformatics*, vol. 19, no. 3, pp. 336–344, 2003.
- [36] K. Raman, N. Damaraju, and G. K. Joshi, "The organisational structure of protein networks: revisiting the centrality-lethality hypothesis," *Systems and Synthetic Biology*, vol. 8, no. 1, pp. 73–81, 2014.
- [37] Z. Ning and Z. Jiang, "GOVis, a gene ontology visualization tool based on multi-dimensional values," *Protein & Peptide Letters*, vol. 17, no. 5, pp. 675–680, 2010.
- [38] Z. Xing, C. Chu, L. Chen, and X. Kong, "The use of gene ontology terms and KEGG pathways for analysis and prediction of oncogenes," *Biochimica et biophysica acta*, vol. 1860, pp. 2725–2734, 2016.
- [39] G. Yu, L.-G. Wang, Y. Han, and Q.-Y. He, "ClusterProfiler: an R package for comparing biological themes among gene clusters," OMICS: A Journal of Integrative Biology, vol. 16, no. 5, pp. 284–287, 2012.
- [40] E. Simpson and H. L. Brown, "Understanding osteosarcomas," *Journal of the American Academy of Physician Assistants*, vol. 31, no. 8, pp. 15–19, 2018.
- [41] P. J. Grohar, K. A. Janeway, L. D. Mase, and J. D. Schiffman, "Advances in the treatment of pediatric bone sarcomas," *American Society of Clinical Oncology Educational Book*, vol. 37, pp. 725–735, 2017.
- [42] S. Etsebeth, C. Albrecht, and K. Pegel, "Beta-sitosterol and beta-sitosterol glucoside stimulate human peripheral blood lymphocyte proliferation: implications for their use as an immunomodulatory vitamin combination," *International Immunopharmacology*, vol. 18, no. 12, 1996.
- [43] P. J. Bouic and J. H. Lamprecht, "Plant sterols and sterolins: a review of their immune-modulating properties," *Alternative Medicine Review: A Journal of Clinical Therapeutic*, vol. 4, no. 3, pp. 170–177, 1999.
- [44] W.-P. Chen, C. Yu, P.-F. Hu, J.-P. Bao, J.-L. Tang, and L.-D. Wu, "Stigmasterol blocks cartilage degradation in rabbit model of osteoarthritis," *Acta biochimica Polonica*, vol. 59, no. 4, 2012.
- [45] P. Trouillas, C. Corbière, B. Liagre, J.-L. Duroux, and J.-L. Beneytout, "Structure-function relationship for saponin effects on cell cycle arrest and apoptosis in the human 1547 osteosarcoma cells: a molecular modelling approach of natural molecules structurally close to diosgenin," *Bioorganic & Medicinal Chemistry*, vol. 13, no. 4, pp. 1141–1149, 2005.
- [46] Y. Wang, D. Kong, X. Wang, X. Dong, Y. Tao, and H. Gong, "Molecular mechanisms of luteolin induced growth inhibition and apoptosis of human osteosarcoma cells," *Iranian Journal* of *Pharmaceutical Research: IJPR*, vol. 14, no. 2, pp. 531–8, 2015.
- [47] B. Zhang, X. Yu, and H. Xia, "The flavonoid luteolin enhances doxorubicin-induced autophagy in human osteosarcoma U2OS cells," *International Journal of Clinical and Experimental Medicine*, vol. 8, no. 9, 7 pages, Article ID 15190, 2015.

- [48] E. Thoenen, A. Curl, and T. J. P. Iwakuma, "TP53 in Bone and Soft Tissue Sarcomas," *Pharmacology & Therapeutics*, vol. 202, 2019.
- [49] A. Ranjan and T. Iwakuma, "Non-canonical cell death induced by p53," *International Journal of Molecular Sciences*, vol. 17, no. 12, p. 2068, 2016.
- [50] D. Lane and A. Levine, "p53 research: the past thirty years and the next thirty years," *Cold Spring Harbor Perspectives in Biology*, vol. 2, no. 12, Article ID a000893, 2010.
- [51] C. Yang, D. Huang, C. Ma et al., "Identification of pathogenic genes and transcription factors in osteosarcoma," *Pathology & Oncology Research*, vol. 26, pp. 1–8, 2019.
- [52] M. Taipale, D. F. Jarosz, and S. Lindquist, "HSP90 at the hub of protein homeostasis: emerging mechanistic insights," *Nature Reviews Molecular Cell Biology*, vol. 11, no. 7, pp. 515–528, 2010.
- [53] M. I. G. Ruiz, K. Floor, P. Roepman et al., "Integration of gene dosage and gene expression in non-small cell lung cancer, identification of HSP90 as potential target," *PLoS One*, vol. 3, no. 3, 2008.
- [54] Q. Cheng, J. T. Chang, J. Geradts et al., "Amplification and high-level expression of heat shock protein 90 marks aggressive phenotypes of human epidermal growth factor receptor 2 negative breast cancer," *Breast Cancer Research*, vol. 14, no. 2, p. R62, 2012.
- [55] M. Zhao, P. Xu, Z. Liu et al., "Dual roles of miR-374a by modulated c-Jun respectively targets CCND1-inducing PI3K/ AKT signal and PTEN-suppressing Wnt/β-catenin signaling in non-small-cell lung cancer," *Cell Death & Disease*, vol. 9, no. 2, pp. 78–17, 2018.
- [56] D.-G. Chen, B. Zhu, S.-Q. Lv et al., "Inhibition of EGR1 inhibits glioma proliferation by targeting CCND1 promoter," *Journal of Experimental & Clinical Cancer Research*, vol. 36, no. 1, p. 186, 2017.
- [57] Z. Li, C. Wang, G. Prendergast, and R. G. Pestell, "Cyclin D1 functions in cell migration," *Cell Cycle*, vol. 5, no. 21, pp. 2440–2442, 2006.
- [58] I. A. Pavlenko, L. E. Zavalishina, and P. E. Povilaitite, "HER2/ neu gene amplification as a mechanism of clonal heterogeneity in breast cancer," *Arkhiv Patologii*, vol. 81, no. 6, p. 49, 2019.
- [59] C. Gutierrez, R. Schiff, and L. medicine, "HER2: biology, detection, and clinical implications," *Archives of Pathology & Laboratory Medicine*, vol. 135, no. 1, pp. 55–62, 2011.
- [60] A. G. Abdou, M. Kandil, N. Y. Asaad et al., "The prognostic role of Ezrin and HER2/neu expression in osteosarcoma," *Applied Immunohistochemistry & Molecular Morphology*, vol. 24, no. 5, pp. 355–363, 2016.
- [61] L.-F. Wang, Y. Zhou, Y.-M. Xu et al., "A caspase-6 and anti-HER2 antibody chimeric tumor-targeted proapoptotic molecule decreased metastasis of human osteosarcoma," *Cancer Investigation*, vol. 27, no. 7, pp. 774–780, 2009.
- [62] B. Li, P. Zhou, K. Xu et al., "Metformin induces cell cycle arrest, apoptosis and autophagy through ROS/JNK signaling pathway in human osteosarcoma," *International Journal of Biological Sciences*, vol. 16, no. 1, pp. 74–84, 2020.
- [63] Z. A. Stewart, M. D. Westfall, and J. A. Pietenpol, "Cell-cycle dysregulation and anticancer therapy," *Trends in Pharmacological Sciences*, vol. 24, no. 3, pp. 139–145, 2003.
- [64] D. Catanzaro, E. Ragazzi, C. Vianello, L. Caparrotta, and M. MJNpc, "Effect of quercetin on cell cycle and cyclin expression in ovarian carcinoma and osteosarcoma cell lines," *SAGE Journal*, vol. 10, no. 8, 2015.

- [65] K. C. Zimmermann, C. Bonzon, and D. R. Green, "The machinery of programmed cell death," *Pharmacology & Therapeutics*, vol. 92, no. 1, pp. 57–70, 2001.
- [66] S.-I. Tanuma, Y. Shibui, T. Oyama, F. Uchiumi, and H. Abe, "Targeting poly (ADP-ribose) glycohydrolase to draw apoptosis codes in cancer," *Biochemical Pharmocology*, vol. 167, 2019.
- [67] P. Chaiyawat, D. Pruksakorn, A. Phanphaisarn, P. Teeyakasem, J. Klangjorhor, and J. Settakorn, "Expression patterns of class I histone deacetylases in osteosarcoma: a novel prognostic marker with potential therapeutic implications," *Modern Pathology*, vol. 31, no. 2, pp. 264–274, 2018.
- [68] D. Sun, M. Yu, Y. Li et al., "Histone deacetylase 2 is involved in DNA damage-mediated cell death of human osteosarcoma cells through stimulation of the ATM/p53 pathway," *FEBS Open Bio*, vol. 9, no. 3, pp. 478–489, 2019.
- [69] K. Cao, H. Wang, Y. Fang et al., "Histone deacetylase 4 promotes osteosarcoma cell proliferation and invasion by regulating expression of proliferating cell nuclear antigen," *Frontiers in Oncology*, vol. 9, 2019.
- [70] J. Cao, X. Liu, Y. Yang et al., "Decylubiquinone suppresses breast cancer growth and metastasis by inhibiting angiogenesis via the ROS/p53/Bai1 signaling pathway," *Angio*genesis, vol. 23, pp. 1–14, 2020.
- [71] V. K. Kashyap, N. Dan, N. Chauhan et al., "VERU-111 suppresses tumor growth and metastatic phenotypes of cervical cancer cells through the activation of p53 signaling pathway," *Cancer Letters*, vol. 470, pp. 64–74, 2020.
- [72] S.-H. Lee, S.-J. Lee, Y. S. Jung et al., "Blocking of p53-Snail binding, promoted by oncogenic K-Ras, recovers p53 expression and function," *Neoplasia*, vol. 11, no. 1, pp. 22–IN6, 2009.
- [73] E. Powell, D. Piwnica-Worms, and H. Piwnica-Worms, "Contribution of p53 to metastasis," *Cancer Discovery*, vol. 4, pp. 405–414, 2014.
- [74] W. Jin, L. Zhou, B. Yan et al., "Theabrownin triggersDNAdamage to suppress human osteosarcoma U2OScells by activating p53 signalling pathway," *Journal of Cellular and Molecular Medicine*, vol. 22, no. 9, pp. 4423–4436, 2018.
- [75] T. Wang, X. Gong, R. Jiang, H. Li, W. Du, and G. Kuang, "Ferulic acid inhibits proliferation and promotes apoptosis via blockage of PI3K/Akt pathway in osteosarcoma cell," *American Journal of Translational Research*, vol. 8, no. 2, pp. 968–80, 2016.
- [76] Z.-Z. Cjijoph, "Berberine induced apoptosis of human osteosarcoma cells by inhibiting phosphoinositide 3 kinase/protein kinase B (PI3K/Akt) signal pathway activation," *Iranian Journal of Public Health*, vol. 45, no. 5, p. 578, 2016.
- [77] D. Resnitzky, S. I. Reed, and c biology, "Different roles for cyclins D1 and E in regulation of the G1-to-S transition," *Molecular and Cellular Biology*, vol. 15, no. 7, pp. 3463–3469, 1995.
- [78] C. L. B. Kline, A. P. J. Van den Heuvel, J. E. Allen, V. V. Prabhu, D. T. Dicker, and W. S. El-Deiry, "ONC201 kills solid tumor cells by triggering an integrated stress response dependent on ATF4 activation by specific eIF2α kinases," *Science Signaling*, vol. 9, no. 415, 2016.
- [79] Y. W. Liu, T. Yang, L. Zhao et al., "Activation of Adenosine 2A receptor inhibits neutrophil apoptosis in an autophagy-dependent manner in mice with systemic inflammatory response syndrome," *Scientific Reports*, vol. 6, no. 1, pp. 33614–33626, 2016.
- [80] S.-M. Kim, E.-M. Kim, K.-Y. Ji et al., "TREM2 acts as a tumor suppressor in colorectal carcinoma through wnt1/β-catenin and erk signaling," *Cancers*, vol. 11, no. 9, p. 1315, 2019.

- [81] C. H. Lin, T. Ji, C.-F. Chen, and B. H. Hoang, "Wnt signaling in osteosarcoma," Advances in Experimental Medicine and Biology, vol. 804, pp. 33–45, 2014.
- [82] T. Malinauskas, A. R. Aricescu, W. Lu, C. Siebold, and E. Y. Jones, "Modular mechanism of Wnt signaling inhibition by Wnt inhibitory factor 1," *Nature Structural & Molecular Biology*, vol. 18, no. 8, pp. 886–893, 2011.
- [83] Q. Tang, H. Zhao, B. Yang et al., "WIF-1 gene inhibition and Wnt signal transduction pathway activation in NSCLC tumorigenesis," *Oncology Letters*, vol. 13, no. 3, pp. 1183–1188, 2017.
- [84] W. Li, Z. Meng, T. Zou et al., "MiR-374a activates Wnt/ β-catenin signaling to promote osteosarcoma cell migration by targeting WIF-1," *Pathology & Oncology*, vol. 26, pp. 1–7, 2018.