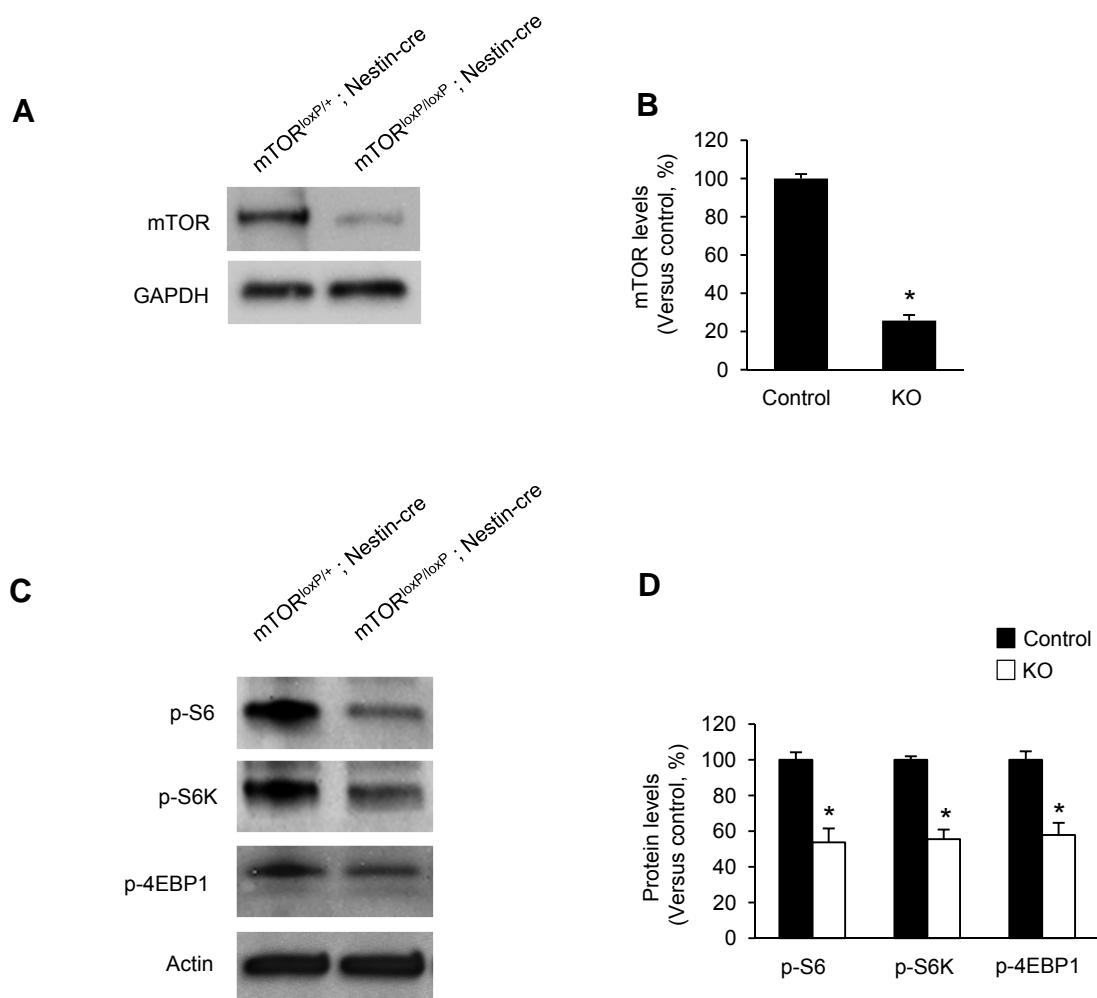
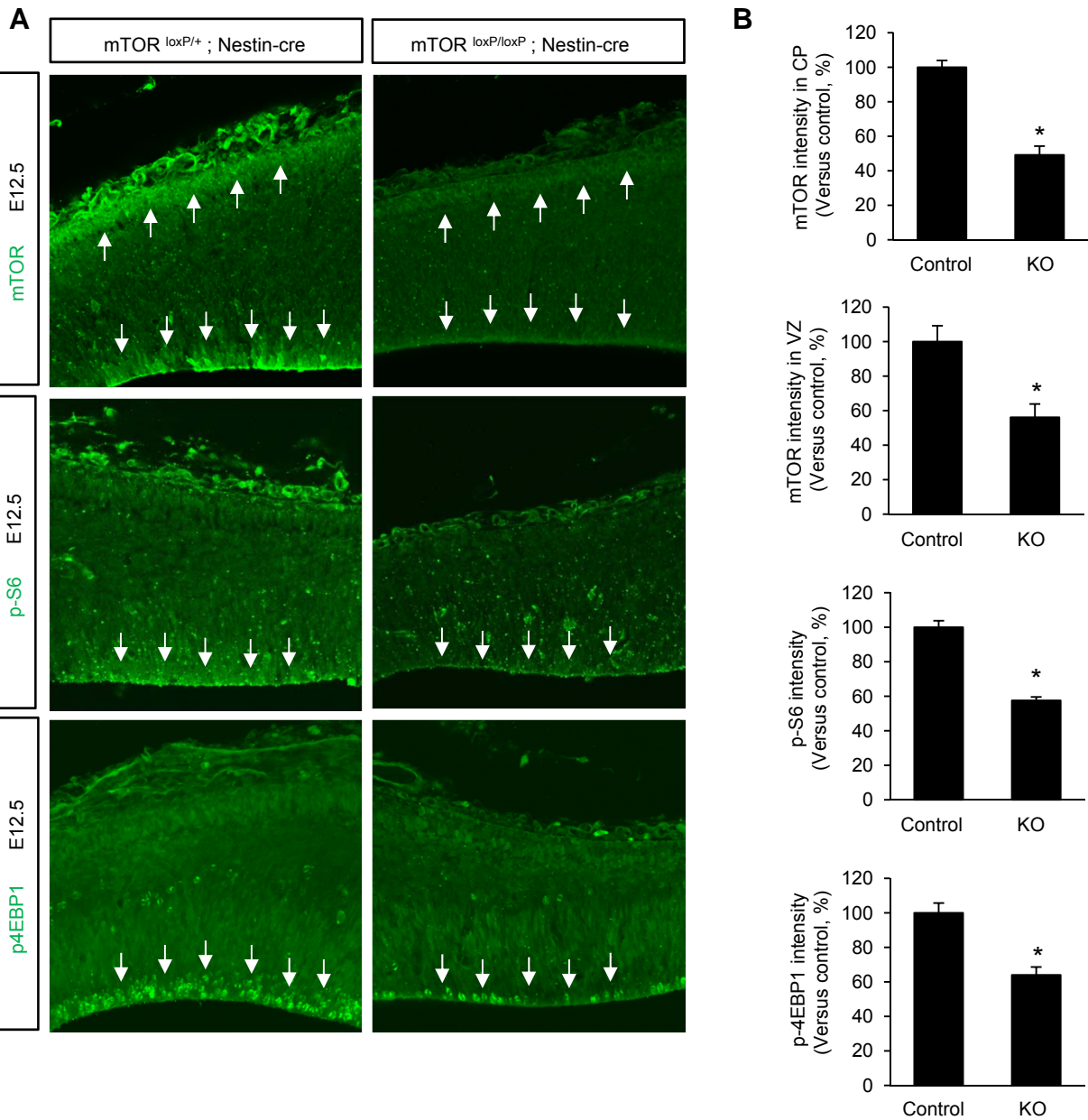


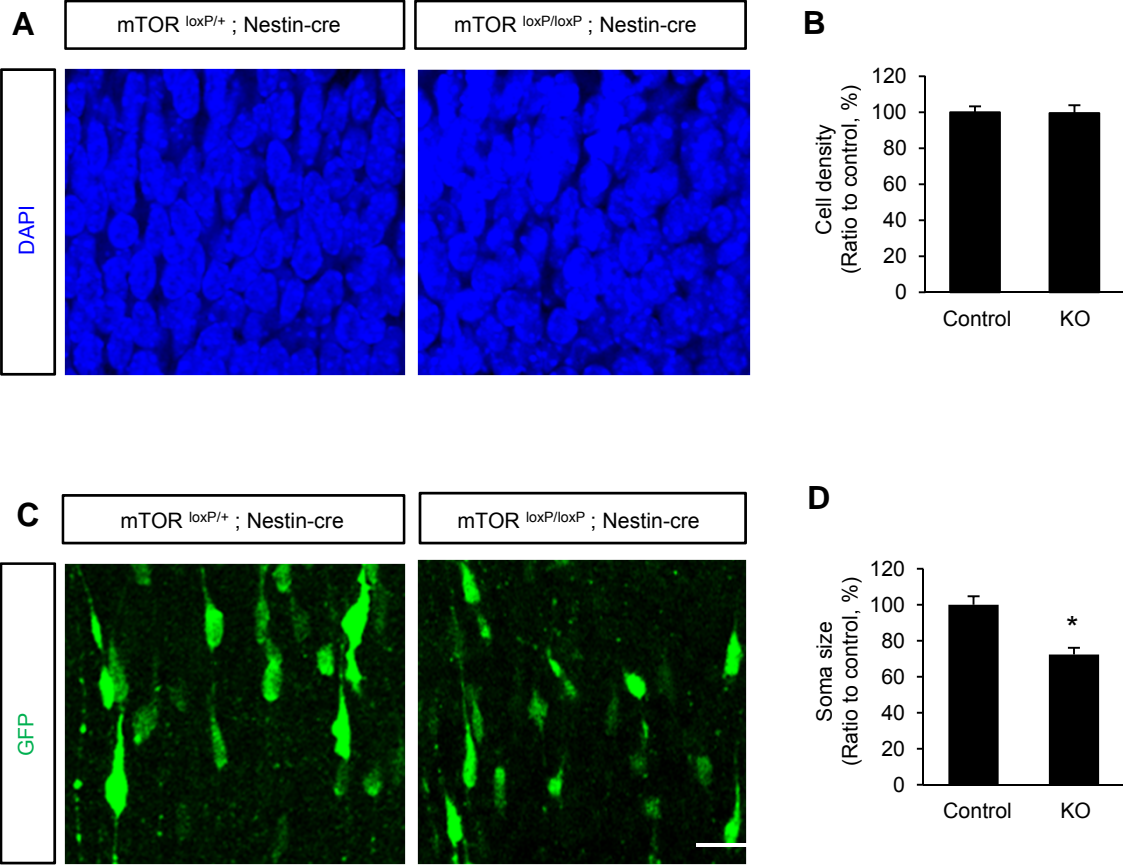
Supplemental Figure 1



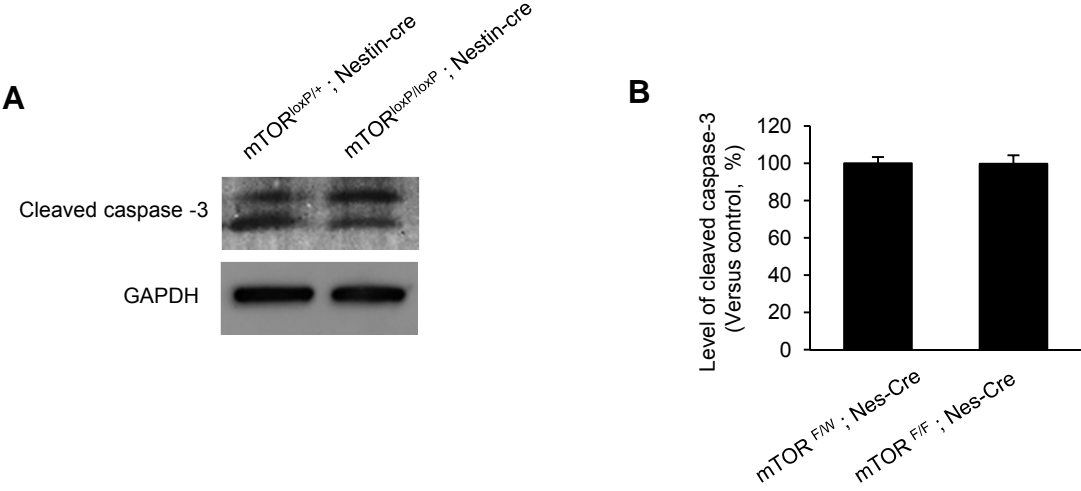
Supplemental Figure 2



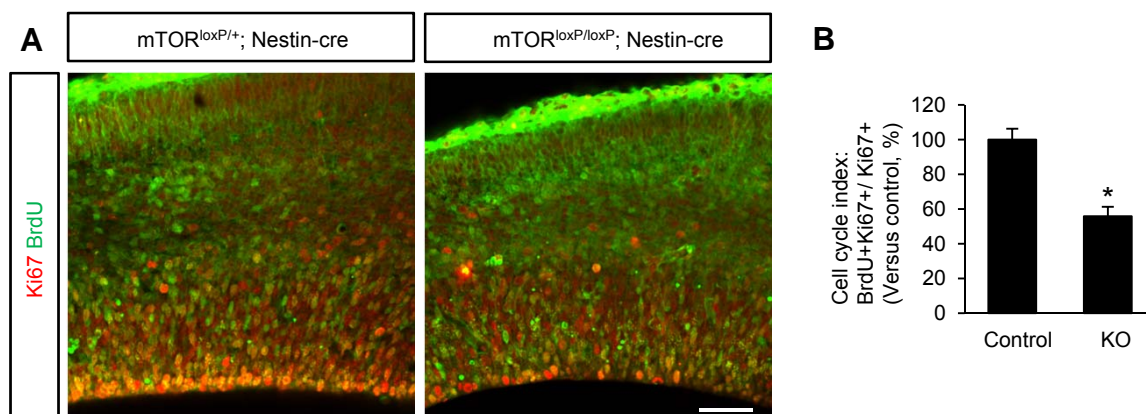
Supplemental Figure 3



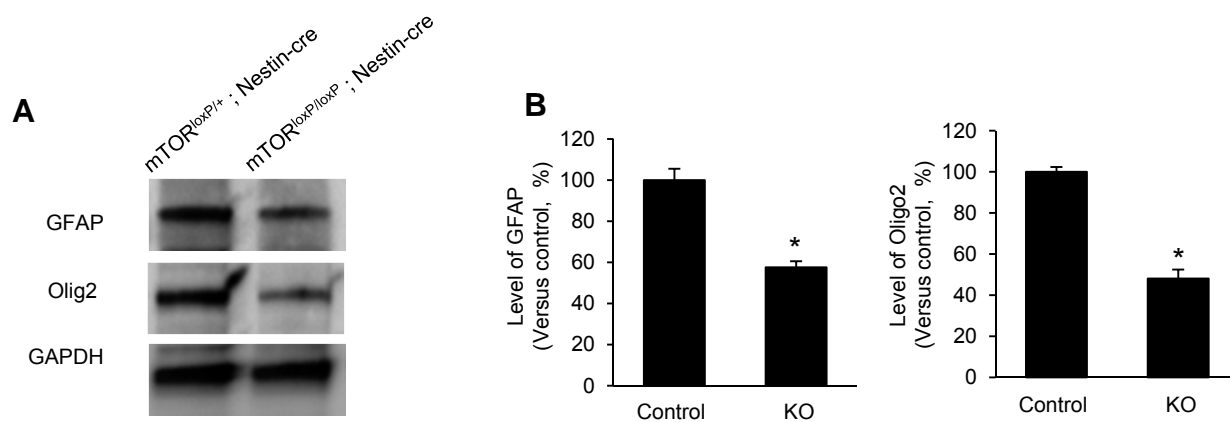
Supplemental Figure 4



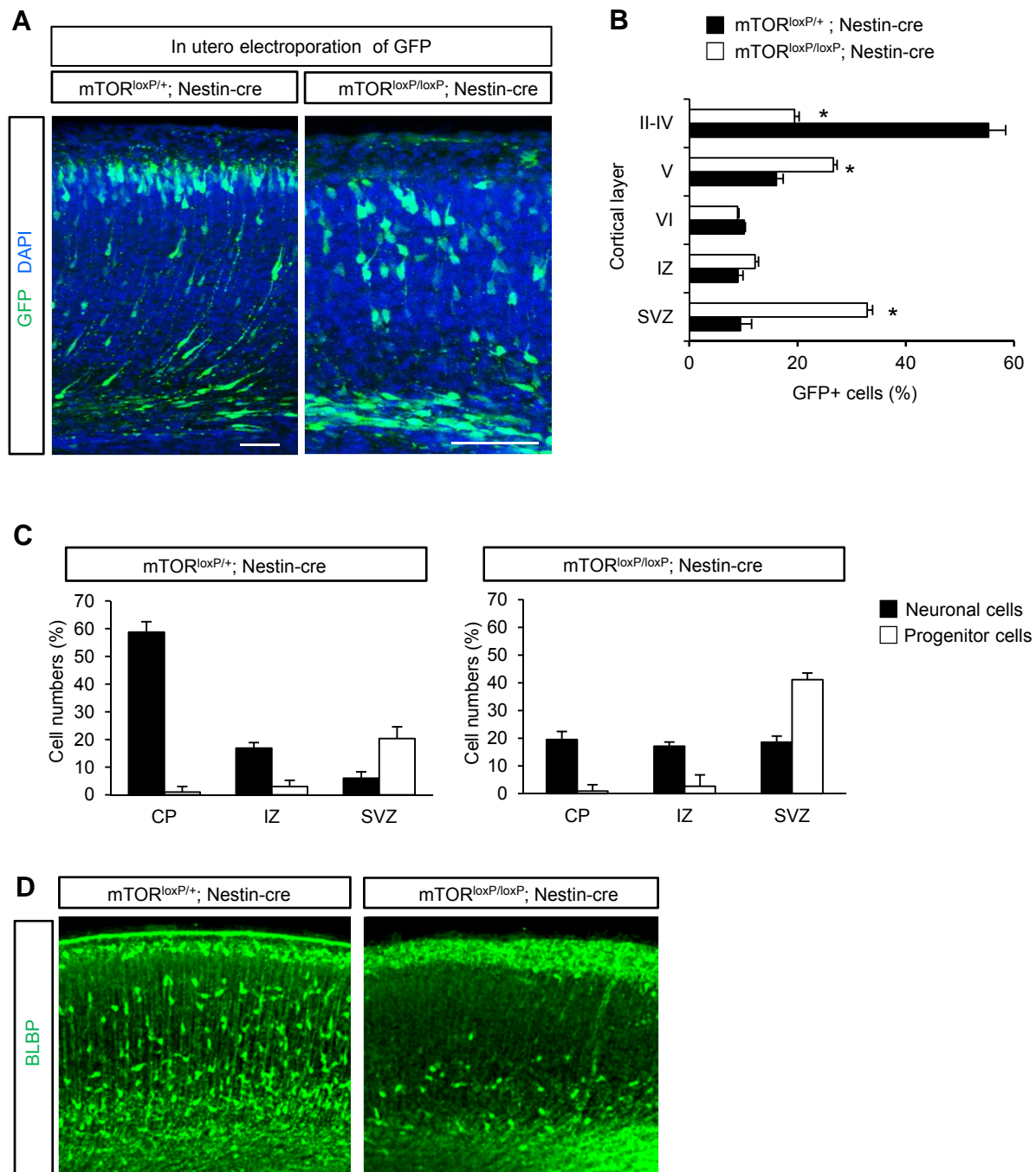
Supplemental Figure 5



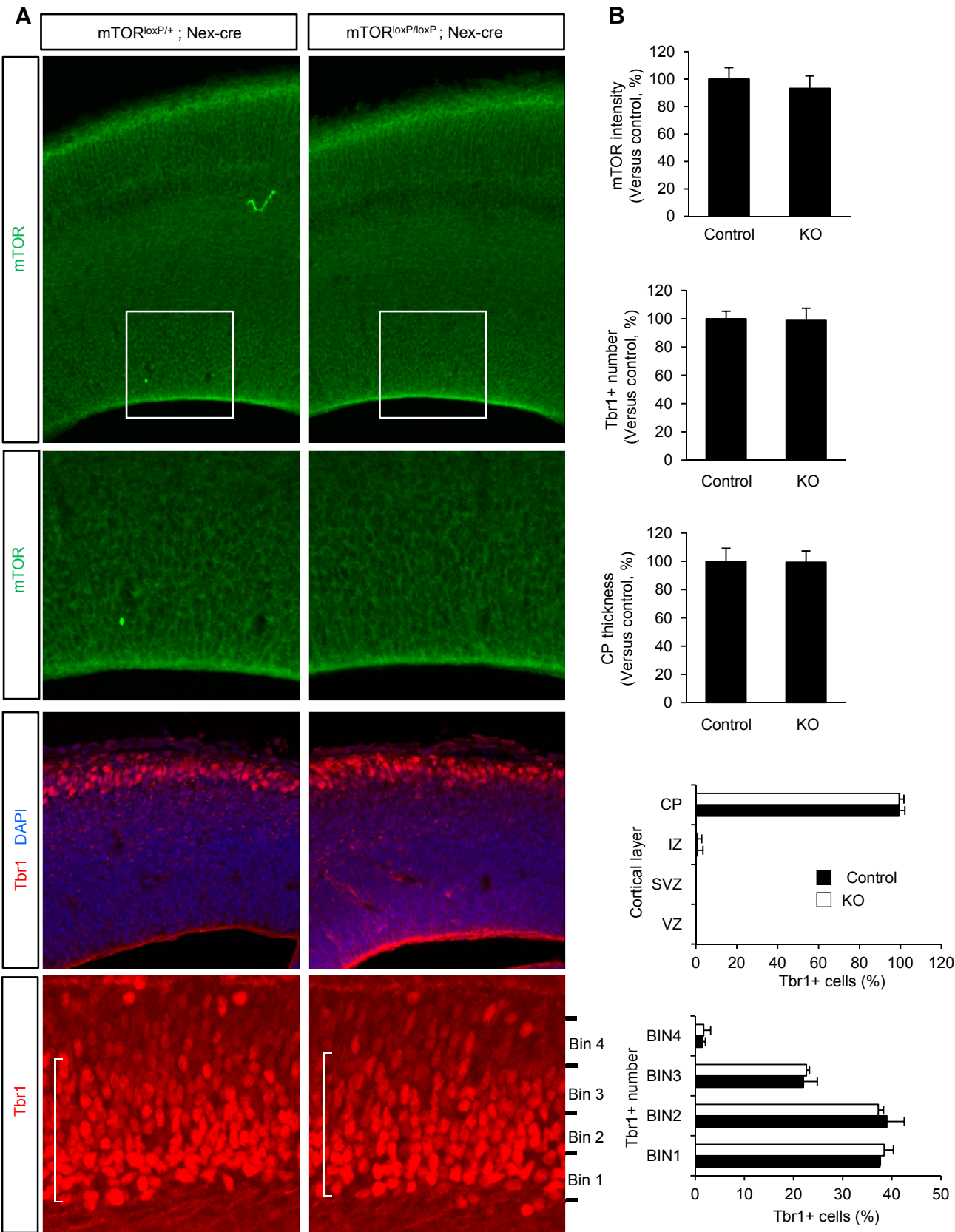
Supplemental Figure 6



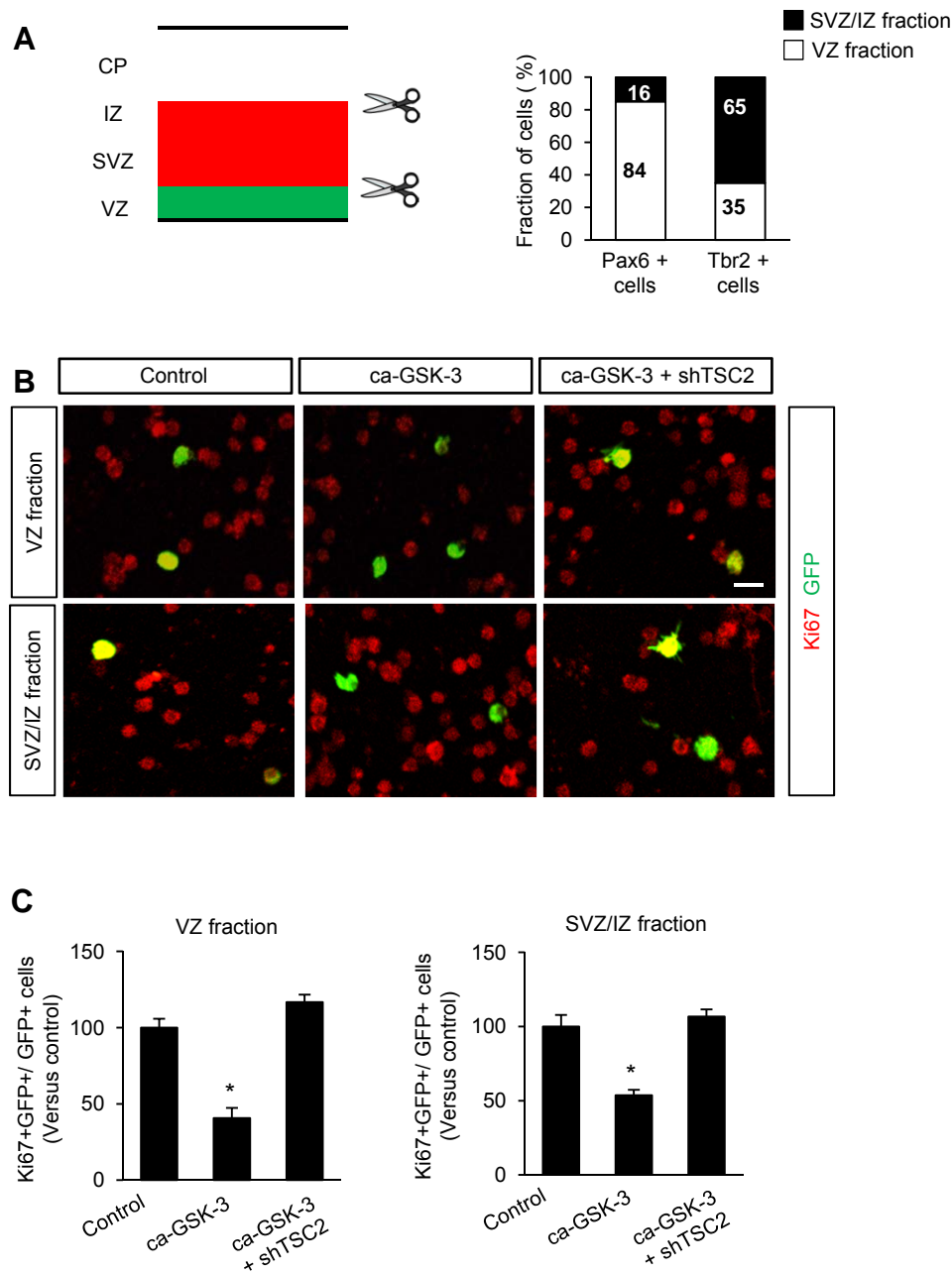
Supplemental Figure 7



Supplemental Figure 8



Supplemental Figure 9



Supplemental Figure legends

Supplemental Figure 1. The levels of mTOR and phosphorylated targets in *mTor^{loxP/loxP}; Nestin-cre* mice

(A) Western blot shows mTOR protein is almost completely eliminated in E15.5 brain tissues of *mTor^{loxP/loxP}; Nestin-cre* mice. (B) Quantification of (A). The mTOR level was decreased by 76 % in *mTor^{loxP/loxP}; Nestin-cre* brains compared to controls. Data were shown as relative changes vs. control. Control: *mTor^{loxP/+}; Nestin-cre*. KO: *mTor^{loxP/loxP}; Nestin-cre*. * $p < 0.05$. (C) Phosphorylation of mTOR target proteins is inhibited in *mTor^{loxP/loxP}; Nestin-cre* brains. Western blotting using E15.5 brain lysates showed that the levels of the phosphorylated targets were reduced by approximately 50%. (D) Quantification of (C). Control: *mTor^{loxP/+}; Nestin-cre*. KO: *mTor^{loxP/loxP}; Nestin-cre*. * $p < 0.05$.

Supplemental Figure 2. mTOR deletion in *mTor^{loxP/loxP}; Nestin-cre* mice

(A) mTOR expression in control and *mTor^{loxP/loxP}; Nestin-cre* brains. mTOR expression was assessed by immunostaining of E12.5 brain samples. mTOR is expressed in the VZ/SVZ and the cortical plate in control tissues, but the expression was reduced in *mTor^{loxP/loxP}; Nestin-cre* brains. Similar patterns were observed in brain samples immunostained with p-4EBP1 and p-S6 antibodies. Arrow indicates the regions that highly express the mTOR components. (B) The fluorescence intensities of mTOR, phospho-S6, and phospho-4EBP1 immunostaining shown in (A) were quantified by ImageJ. Data were presented as relative changes vs. control. Control: *mTor^{loxP/+}; Nestin-cre*. KO: *mTor^{loxP/loxP}; Nestin-cre*. * $p < 0.05$.

Supplemental Figure 3. Cell density and size in mTOR-deleted brains

(A) Cell density is not changed in *mTor^{loxP/loxP}; Nestin-cre* brains. E15.5 control and mTOR-deleted brains were stained with DAPI. (B) Quantification of (A). There is no significant difference in cell density between control and mTOR-deleted brains. Data were shown as relative changes vs. control. Control:

mTor^{loxP/+}; *Nestin-cre*. KO: *mTor*^{loxP/loxP}; *Nestin-cre*. (C) A GFP construct was electroporated in utero into E13.5 control and *mTor*^{loxP/loxP}; *Nestin-cre* brains. The sizes of GFP-positive cells were assessed three days later. Scale bar, 30 μ m. (D) Quantification of (C). Control: *mTor*^{loxP/+}; *Nestin-cre*. KO: *mTor*^{loxP/loxP}; *Nestin-cre*. * $p < 0.05$.

Supplemental Figure 4. Levels of cleaved-caspase 3 in *mTor*^{loxP/loxP}; *Nestin-cre* mice

(A) The levels of cleaved-caspase 3 were assessed in control and *mTor*^{loxP/loxP}; *Nestin-cre* brains by Western blotting. (B) Quantification of (A). No significant changes in cleaved caspase-3 levels were found between controls and *mTor*^{loxP/loxP}; *Nestin-cre* brain samples. Data were shown as relative changes vs. control. Control: *mTor*^{loxP/+}; *Nestin-cre*. KO: *mTor*^{loxP/loxP}; *Nestin-cre*.

Supplemental Figure 5. mTOR-deleted neural progenitors exhibit lengthened cell cycle

(A) To analyze cell cycle length, E14.5 control and *mTor*^{loxP/loxP}; *Nestin-cre* mice were pulse-labeled with BrdU for 1 hour and then brains were collected for immunostaining with BrdU (green) and Ki67 (red) antibodies. Scale bar, 50 μ m. (B) Quantification of cell cycle index. The labeling index of cell cycle length is defined as the fraction of BrdU- and Ki67-positive cells in total Ki67-positive cells in the brain. The results were shown as relative changes vs. control. The decreased labeling index suggests prolonged cell cycles in mTOR-deficient neural progenitors. Control: *mTor*^{loxP/+}; *Nestin-cre*. KO: *mTor*^{loxP/loxP}; *Nestin-cre*. * $p < 0.05$.

Supplemental Figure 6. Levels of GFAP and Olig2 in *mTor*^{loxP/loxP}; *Nestin-cre* mice

(A) Using E15.5 control and *mTor*^{loxP/loxP}; *Nestin-cre* brain lysates, Western blotting was performed with GFAP and Olig2 antibodies. (B) Quantification of (A). The levels of GFAP and Olig2 were significantly decreased in *mTor*^{loxP/loxP}; *Nestin-cre* brains compared to control tissues. Data were shown as relative changes vs. control. Control: *mTor*^{loxP/+}; *Nestin-cre*. KO: *mTor*^{loxP/loxP}; *Nestin-cre*. * $p < 0.05$.

Supplemental Figure 7. Neuronal placement in $mTor^{F/F}; Nestin-cre$ mice

(A) E13.5 control and $mTor^{loxP/loxP}; Nestin-cre$ mice were electroporated in utero with a GFP plasmid. The brain samples of the mice were collected at E18 and then cortical placement of GFP-positive cells were visualized. Scale bar, 50 μ m. (B) Quantification of GFP-positive cells in the cerebral cortex. While GFP-positive cells in control brain were placed mostly in cortical plate, GFP cells in $mTor^{F/F}; Nestin-cre$ brains were found throughout the cerebral cortex. Control: $mTor^{loxP/+}; Nestin-cre$. KO: $mTor^{loxP/loxP}; Nestin-cre$. * $p < 0.05$. (C) Quantification of GFP-positive cell types after in utero electroporation of a GFP construct as described in (A). Control and $mTor^{loxP/loxP}; Nestin-cre$ brains were co-immunostained with GFP and MAP2 or GFP and Pax6 (or Tbr2). Most GFP-positive cells were MAP2-positive neurons, while GFP-positive cells in the VZ/SVZ were divided to MAP2-positive neurons and PAX6-positive (or Tbr2-positive) progenitors. Control: $mTor^{loxP/+}; Nestin-cre$. KO: $mTor^{loxP/loxP}; Nestin-cre$. * $p < 0.05$. (D) Control and $mTor^{loxP/loxP}; Nestin-cre$ brains were immunostained with a BLBP antibody. Radial glial fibers were formed normally in control tissues. However, the glial fibers were disrupted in $mTor^{loxP/loxP}; Nestin-cre$ brain samples.

Supplemental Figure 8. $mTor^{loxP/loxP}; Nex-cre$ mice at E14.5

(A) Effects of mTOR deletion using Nex-cre mice at early developmental stage, E14.5. Top panels: mTOR immunostaining in the cerebral cortex. mTOR expression was examined by immunostaining of E14.5 control and $mTor^{loxP/loxP}; Nex-cre$ brain samples. Second row panels: Higher magnification images of white square areas in top panels. Third row panels: Brain sections from E14.5 control ($mTor^{loxP/+}; Nex-cre$) and $mTor^{loxP/loxP}; Nex-cre$ mice were immunostained with Tbr1 antibody. Cells were counterstained by DAPI. Bottom panels: Higher magnification images of Tbr1-immunostained samples. (B) Quantification of (A). $mTor^{loxP/loxP}; Nex-cre$ mice did not show altered positioning, number, and layer thickness of Tbr1-positive cells, compared to controls. The fluorescence intensities of mTOR were

quantified by ImageJ. Data were presented as relative changes vs. control. Control: *mTor*^{loxP/+}; *Nex-cre*. KO: *mTor*^{loxP/loxP}; *Nex-cre*. * p<0.05.

Supplemental Figure 9. TSC2 knockdown suppresses the inhibitory effects of GSK-3 on Pax6-positive or Tbr2-positive progenitor proliferation

(A) Left panel: A diagram showing embryonic cerebral cortex. Tissues containing the ventricular zone (green) or the subventricular/intermediate zone (red) from E13.5 mouse brains were fractionated by brain slicing and micro-dissection. Pax6-positive progenitors are mostly distributed in the ventricular zone while a major portion of Tbr2-positive cells are localized within the subventricular zone and intermediate zones (Arai et al., 2011). Right panel: Percentages of Pax6-positive and Tbr2-positive cells in the VZ fraction and the SVZ/IZ fraction were quantified. Most Pax6-positive cells were found in the VZ fraction. A major population of Tbr2-positive cells was seen in the SVZ/IZ fraction. (B) TSC2 knockdown suppresses the inhibitory effects of GSK-3 on proliferation of PAX6-positive and Tbr2-positive cells. E13.5 neural progenitors from each fraction were cultured and transfected with control GFP, ca-GSK-3-GFP, or ca-GSK-3-GFP and shTSC2. Then, proliferating neural progenitors were assessed by Ki67 and GFP co-immunostaining. (C) The effects of GSK-3, shTSC2, and shGSK-3 on cell proliferation were assessed by quantifying the Ki67 and GFP double-positive cells in total GFP-positive cells. The results were shown as relative changes vs. control. * p<0.05.