



# Complete Genome Sequence of Human Oral Saccharibacterium “*Candidatus Nanosynbacter* sp. HMT352” Strain KC1

 Karissa L. Cross,<sup>a,b</sup> Dawn M. Klingeman,<sup>a</sup>  Mircea Podar<sup>a</sup>

<sup>a</sup>Biosciences Division, Oak Ridge National Laboratory, Oak Ridge, Tennessee, USA

<sup>b</sup>Department of Biological Sciences, Vanderbilt University, Nashville, Tennessee, USA

**ABSTRACT** “*Cand. Nanosynbacter* sp. HMT352” strain KC1 is an ectoparasitic saccharibacterium/TM7 that was co-isolated from a human saliva sample with its obligate bacterial host, *Schaalia odontolytica*. The genome of strain KC1 enables studies of the mechanisms and evolution of interspecies interactions and, for oral species, studies of their potential roles in health and disease.

Saccharibacteria (originally referred to as candidate division TM7) are a group of ubiquitous bacteria which, based on their reduced genomes, are inferred to be physiologically dependent on acquiring metabolic building blocks from the environment or directly from other organisms (1–4). Several saccharibacteria have been cultured from the human oral microbiota as obligate ectoparasites on various Actinobacteria (5–9). An isolate from wastewater foam, “*Cand. Mycosynbacter amalyticus*,” attaches to and lyses a variety of free-living Actinobacteria (10). The mechanisms by which saccharibacteria recognize suitable hosts, acquire nutrients, and (in many cases) lead to host lysis are still unknown. By using a targeted reverse-genomics approach, we isolated and cultured a human oral saccharibacterium, “*Cand. Nanosynbacter* sp. HMT352” strain KC1 (here referred to as HMT352-KC1) in association with its host, the actinobacterium *Schaalia odontolytica* strain ORNL 0103 (6, 11).

HMT352-KC1 and its host were cultured in 100 mL brain-heart infusion medium (BHI, Difco) for 3 days at 37°C, under a hypoxic atmosphere (93% N<sub>2</sub>, 5% CO<sub>2</sub>, and 2% O<sub>2</sub>). All downstream molecular procedures used commercial reagents and followed manufacturers’ protocols. Genomic DNA was extracted using a Quick-DNA Fungal/Bacterial Midiprep Kit (Zymo Research). A Nextera XT DNA Library Preparation Kit (Illumina, Inc.) was used to generate a library with an approximately 600-bp median insert size, based on a Bioanalyzer 2100 (Agilent Technologies). The library was sequenced (2 × 250-nucleotide reads) on a MiSeq instrument (Illumina, Inc.), generating 11.1 million paired-end reads. All subsequent bioinformatic analyses were conducted using software default settings unless otherwise specified. The reads were imported into KBase (12) and trimmed, based on quality scores, using Trimmomatic v.0.36 (13). The trimmed reads were assembled using MEGAHIT v1.2.9 (14) with the meta-sensitive setting and a minimum contig size of 2 kb. The contigs were binned based on nucleotide composition and coverage depth using MetaBAT2 v1.7 (15), resulting in three bins. Two of the bins contained contigs with high G+C% (>56%), corresponding to the genome of the *Schaalia* host, which had been previously sequenced (11). The third bin was represented by a single 678,346-bp contig with a G+C of 43% and was classified as Saccharibacteria based on GTDB-Tk v1.7.0 (16). Genes encoding proteins and RNAs were predicted and annotated using Prokka 1.14.0 (17) and the contig was imported into Geneious Prime 2021.0.1 (18) for final curation. Inspection of the genes at the contig ends revealed an identical region. Mapping of the sequencing reads to that region in Geneious enabled identification of the nucleotide position unique to each end. Based on this, one of the repeating regions was removed and the ends were joined, resulting in a circular 677,938-bp chromosome with a G+C content of 42.9%.

**Editor** Irene L. G. Newton, Indiana University, Bloomington

**Copyright** © 2022 Cross et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Mircea Podar, podarm@ornl.gov.

The authors declare no conflict of interest.

**Received** 17 December 2021

**Accepted** 28 January 2022

**Published** 10 February 2022

Final genome annotation was conducted using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v5.3 (19). The chromosome of HMT352-KC1 encodes 675 proteins, 42 tRNAs, one rRNA operon (23S, 16S, and 5.8S genes), and 2 noncoding RNAs (ncRNAs). The genome was compared with those of other human Saccharibacteria/TM7 using FastANI.1.2 (20). Based on an average pairwise nucleotide identity (ANI) of 83%, strain HMT352-KC1 is a closely related species to “*Cand. Nanosynbacter lyticus*” TM7X, the first isolated saccharibacterium (5).

**Data availability.** The “*Cand. Nanosynbacter* sp. HMT352” strain KC1 genome sequence has been deposited in GenBank under the accession number [CP089520](#). The version described in this paper is the first version, [CP089520.1](#). The Illumina reads have been deposited in SRA under the accession number [SRR17194431](#).

## ACKNOWLEDGMENTS

This research was funded by grant R01DE024463 from the National Institute of Dental and Craniofacial Research (NIDCR) of the US National Institutes of Health. Oak Ridge National Laboratory is managed by UT-Battelle, LLC, for the U.S. Department of Energy under contract DE-AC05-00OR22725.

The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

## REFERENCES

- Hugenholtz P, Goebel BM, Pace NR. 1998. Impact of culture-independent studies on the emerging phylogenetic view of bacterial diversity. *J Bacteriol* 180:4765–4774. <https://doi.org/10.1128/JB.180.18.4765-4774.1998>.
- Hugenholtz P, Tyson GW, Webb RI, Wagner AM, Blackall LL. 2001. Investigation of candidate division TM7, a recently recognized major lineage of the domain Bacteria with no known pure-culture representatives. *Appl Environ Microbiol* 67:411–419. <https://doi.org/10.1128/AEM.67.1.411-419.2001>.
- Albertsen M, Hugenholtz P, Skarshewski A, Nielsen KL, Tyson GW, Nielsen PH. 2013. Genome sequences of rare, uncultured bacteria obtained by differential coverage binning of multiple metagenomes. *Nat Biotechnol* 31:533–538. <https://doi.org/10.1038/nbt.2579>.
- Starr EP, Shi S, Blazewicz SJ, Probst AJ, Herman DJ, Firestone MK, Banfield JF. 2018. Stable isotope informed genome-resolved metagenomics reveals that Saccharibacteria utilize microbially-processed plant-derived carbon. *Microbiome* 6:122. <https://doi.org/10.1186/s40168-018-0499-z>.
- He X, McLean JS, Edlund A, Yooshef S, Hall AP, Liu SY, Dorrestein PC, Esquenazi E, Hunter RC, Cheng G, Nelson KE, Lux R, Shi W. 2015. Cultivation of a human-associated TM7 phylotype reveals a reduced genome and epibiotic parasitic lifestyle. *Proc Natl Acad Sci U S A* 112:244–249. <https://doi.org/10.1073/pnas.1419038112>.
- Cross KL, Campbell JH, Balachandran M, Campbell AG, Cooper SJ, Griffen A, Heaton M, Joshi S, Klingeman D, Leys E, Yang Z, Parks JM, Podar M. 2019. Targeted isolation and cultivation of uncultivated bacteria by reverse genomics. *Nat Biotechnol* 37:1314–1321. <https://doi.org/10.1038/s41587-019-0260-6>.
- Bor B, Bedree JK, Shi W, McLean JS, He X. 2019. Saccharibacteria (TM7) in the human oral microbiome. *J Dent Res* 98:500–509. <https://doi.org/10.1177/0022034519831671>.
- Chipashvili O, Utter DR, Bedree JK, Ma Y, Schulte F, Mascarin G, Alayyoubi Y, Chouhan D, Hardt M, Bidlack F, Hasturk H, He X, McLean JS, Bor B. 2021. Epibiotic Saccharibacteria suppresses gingival inflammation and bone loss in mice through host bacterial modulation. *Cell Host Microbe* 29:1649–1662.e7. <https://doi.org/10.1016/j.chom.2021.09.009>.
- Bor B, Collins AJ, Murugkar PP, Balasubramanian S, To TT, Hendrickson EL, Bedree JK, Bidlack FB, Johnston CD, Shi W, McLean JS, He X, Dewhirst FE. 2020. Insights obtained by culturing Saccharibacteria with their bacterial hosts. *J Dent Res* 99:685–694. <https://doi.org/10.1177/0022034520905792>.
- Batinovic S, Rose JJA, Ratcliffe J, Seviour RJ, Petrovski S. 2021. Cocultivation of an ultrasmall environmental parasitic bacterium with lytic ability against bacteria associated with wastewater foams. *Nat Microbiol* 6:703–711. <https://doi.org/10.1038/s41564-021-00892-1>.
- Podar NA, Klingeman D, Miranda-Sanchez F, Dewhirst FE, Podar M. 2021. Draft genome sequence of *Schaalia odontolytica* strain ORNL0103, a symbiont of “*Candidatus* Saccharibacteria” HMT352. *Microbiol Resour Announc* 10:e0079321. <https://doi.org/10.1128/MRA.00793-21>.
- Arkin AP, Cottingham RW, Henry CS, Harris NL, Stevens RL, Maslov S, Dehal P, Ware D, Perez F, Canon S, Sneddon MW, Henderson ML, Riehl WJ, Murphy-Olson D, Chan SY, Kamimura RT, Kumari S, Drake MM, Brettin TS, Glass EM, Chivian D, Gunter D, Weston DJ, Allen BH, Baumohl J, Best AA, Bowen B, Brenner SE, Bun CC, Chandonia JM, Chia JM, Colasanti R, Conrad N, Davis JJ, Davison BH, DeJongh M, Devoid S, Dietrich E, Dubchak I, Edirisinghe JN, Fang G, Faria JP, Frybarger PM, Gerlach W, Gerstein M, Greiner A, Gurtowski J, Haun HL, He F, Jain R, et al. 2018. KBase: the United States Department of Energy Systems Biology Knowledgebase. *Nat Biotechnol* 36:566–569. <https://doi.org/10.1038/nbt.4163>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Li D, Luo R, Liu CM, Leung CM, Ting HF, Sadakane K, Yamashita H, Lam TW. 2016. MEGAHIT v1.0: a fast and scalable metagenome assembler driven by advanced methodologies and community practices. *Methods* 102:3–11. <https://doi.org/10.1016/j.jmeth.2016.02.020>.
- Kang DD, Li F, Kirton E, Thomas A, Egan R, An H, Wang Z. 2019. MetaBAT 2: an adaptive binning algorithm for robust and efficient genome reconstruction from metagenome assemblies. *PeerJ* 7:e7359. <https://doi.org/10.7717/peerj.7359>.
- Chaumeil PA, Mussig AJ, Hugenholtz P, Parks DH. 2019. GTDB-Tk: a toolkit to classify genomes with the Genome Taxonomy Database. *Bioinformatics* 36:1925–1927. <https://doi.org/10.1093/bioinformatics/btz848>.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28:1647–1649. <https://doi.org/10.1093/bioinformatics/bts199>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Jain C, Rodriguez RL, Phillipy AM, Konstantinidis KT, Aluru S. 2018. High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nat Commun* 9:5114. <https://doi.org/10.1038/s41467-018-07641-9>.