DOI: 10.1111/cpr.13040

REVIEW





The complex roles of neutrophils in APAP-induced liver injury

Huiting Guo^{1,2} | Shiwei Chen^{1,2} | Mingjie Xie^{1,2} | Cheng Zhou^{1,2} | Min Zheng^{1,2}

¹The State Key Laboratory for Diagnosis and Treatment of Infectious Diseases, The First Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou, China

²Collaborative Innovation Center for Diagnosis and Treatment of Infectious Diseases, Hangzhou, China

Correspondence

Cheng Zhou and Min Zheng, The State Key Laboratory of Infectious Disease Diagnosis and Treatment, The First Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou 310003, China. Emails: zhoucheng0113@zju.edu.cn; minzheng@zju.edu.cn

Funding information

National Nature Science Foundation of China, Grant/Award Number: No. U20A20348; the National Nature Science Foundation of China, Grant/Award Number: 81871646; the Self-Topic Science Foundation of State Key Laboratory for Diagnosis and Treatment of Infectious Diseases

Abstract

Acetaminophen (APAP) is a widely applied drug for the alleviation of pain and fever, which is also a dose-depedent toxin. APAP-induced acute liver injury has become one of the primary causes of liver failure which is an increasingly serious threat to human health. Neutrophils are the major immune cells in human serving as the first barrier against the invasion of pathogen. It has been reported that neutrophils patriciate in the occurrence and development of APAP-induced liver injury. However, evolving evidences suggest that neutrophils also contribute to tissue repair and actively orchestrate resolution of inflammation. Here, we addressed the complex roles in APAP-induced liver injury on the basis of brief introduction of neutrophil's activation, recruitment and migration.

1 | INTRODUCTION

Acetaminophen (APAP) is a kind of widely used drug for the treatment of pain and fever. It is reported that its overdose can cause serious liver failure.¹ In Europe and the United States, APAP-induced acute liver injury (ALI) has become one of the primary causes of liver failure and thus arises widespread concern and research in the world.² Abundant evidences have suggested that both innate and adaptive immune cells, such as macrophages, neutrophils and T cells, played critical roles in APAP-induced ALI.^{3,4} It is now clear that neutrophils have important homeostatic functions in various organ systems including liver,^{5,6} but what the role of neutrophils plays in APAP-induced ALI still remains elusive.^{7,8} Neutrophils, a type of polymorphonuclear leukocytes, are the first-line guardians of the innate immune system. Normally, neutrophils are not activated and move slowly in peripheral blood circulation without direction. The half-life of neutrophils is only 6-7 hours.⁹ Once the pathogen invades or the endogenous stimulants releases, the pattern recognition receptors can recognize pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs). Then, inflammatory reaction occurs and neutrophils are activated.^{10,11} The activated neutrophils in peripheral circulation will tend to move towards the injury site. The migration of neutrophils involves rolling, activation and adhesion.^{12,13} They then participate in the inflammatory process through phagocytosis, degranulation and extracellular traps (NETs).¹⁴ On the one hand, neutrophils are drawn from the blood to sterility inflammatory site, which contribute to wound healing. On the other hand, their prolonged half-life, release of granule proteins such as myeloperoxidase (MPO), neutrophil serine proteases (NSPs), lipocalin 2 (LCN2), human neutrophil peptide (HNP), excessive infiltration and uncontrolled activation may lead to destruction of normal tissue structures and serious inflammations.¹⁵ Therefore, how neutrophils play in APAP-induced ALI has aroused great controversy. In this review, we outline the neutrophil's

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2021 The Authors. Cell Proliferation Published by John Wiley & Sons Ltd.

2 | NEUTROPHIL'S ACTIVATION, RECRUITMENT AND MIGRATION IN APAP-INDUCED LIVER INJURY

2.1 | DAMPs

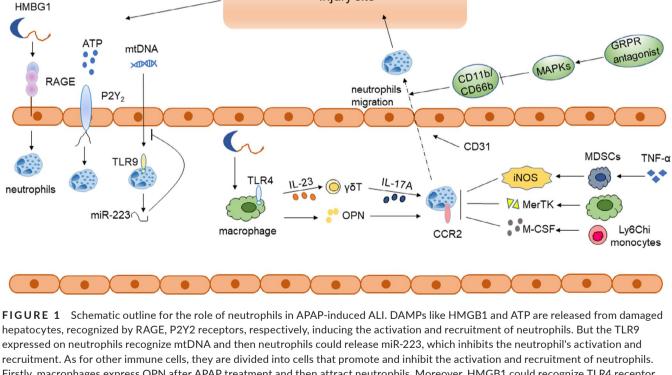
VII FV

DAMPs are key signals which induce cell death in sterile inflammation including high-mobility group box 1(HMGB1), ATP, ADP, IL-1, IL-33, heat shock proteins (HSPs) and so on. Epithelial HMGB1 triggered recruitment of neutrophils but not macrophages through its receptor RAGE, finally inducing necrosis after APAP treatment¹⁶ (Figure 1). In addition, HMGB1 could also mediate neutrophil infiltration via HMGB1-TLR4-IL-23-IL-17A axis¹⁷ (Figure 1). After the administration of APAP, wild-type C57BL/6 mice released abundant ATP and following increased the expression of P2Y2 receptors, which was required for the liver infiltration of neutrophils and subsequent liver injury¹⁸ (Figure 1). Mitochondrial DNA (mtDNA) released by damaged hepatocytes is another important stimulus which activates neutrophils via binding of Toll-like receptor 9 (TLR9). However, mtDNA/TLR9 could also limit neutrophil overactivation through the negative feedback pathway of microRNA-223 (miR-223)¹⁹ (Figure 1).

2.2 | Interplay with other immune cells

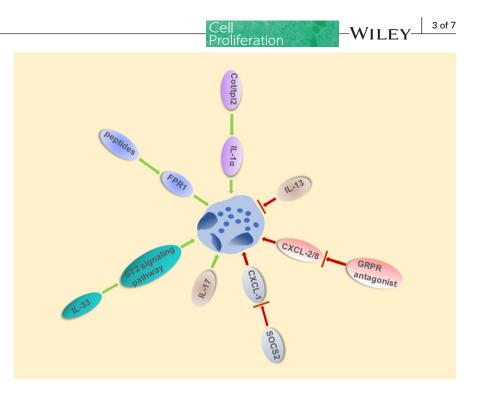
2.2.1 | Macrophages

Numerous pro-inflammatory factors such as chemokines, IL-1 β and TNF- α are released by activated liver macrophages, thus further aggravating inflammation and increasing neutrophils inflow.²⁰ In addition, osteopontin (OPN), another pro-inflammatory cytokine related to liver cell necrosis, was rarely expressed in Kupffer cells normally, but its expression increased significantly in hepatic macrophages at 6 hours after APAP administration, which attracted neutrophils to hepatic injury sites and caused massive liver necrosis (Figure 1).²¹ This result was further demonstrated in OPN-/- mice model, which exhibited less neutrophil infiltration and reduced expression of pro-inflammatory cytokines in liver, such as IL-1 α and TNF- α .²² Macrophages could also induce the generation of IL-17-producing



Injury site

recruitment. As for other immune cells, they are divided into cells that promote and inhibit the activation and recruitment of neutrophils. Firstly, macrophages express OPN after APAP treatment and then attract neutrophils. Moreover, HMGB1 could recognize TLR4 receptor expressed on macrophages, release IL-23 to induce the release of IL-17A in $\gamma\delta$ T cells and finally activate and recruit neutrophils. However, Ly6C^{hi} monocytes inhibit the activation and recruitment of neutrophils via CCR2 and M-CSF pathways. Macrophages could also generate MerTK to inhibit neutrophils. Furthermore, TNF- α /LPS MDSCs could express iNOS to decrease the intrahepatic neutrophils infiltration and induce the apoptosis of activated neutrophil. Activated and recruited neutrophils then migrate to the site of injury to alleviate the inflammation and induce tissue repair. GRPR antagonist could downregulate the expression of CD11b/CD66b via activating MAPKs pathways, which significantly influence the migration of neutrophils to the injury site FIGURE 2 Chemokine and Cytokine influence activation, recruitment and migration of neutrophils in APAP-induced ALI. Various chemokine and cytokine influence the activation and recruitment of neutrophils in APAP-induced ALI. Mitochondria-derived formyl peptides recruit neutrophils to sites of liver necrosis via FPR1. Similarly, IL-33, IL-17 and activated Cot/tpl2 induced neutrophils infiltration whereas IL-13 significantly attenuates APAP-induced liver injury via decreased neutrophils recruitment. SOCS2 and GRPR antagonist can also inhibited infiltration of neutrophils via downregulating CXCL-1 and CXCL-2/8, respectively



 $\gamma\delta$ T cells via the HMGB1-TLR4-IL-23 pathway, enhancing the neutrophil infiltration and liver injury (Figure 1).¹⁷ However, some studies showed opposite results that macrophages could also inhibit the activation and recruitment of neutrophils. In APAP-induced ALI mice model, injection of bone marrow-derived macrophages significantly reduced HMGB1 translocation, infiltrating neutrophils and hepatocytes necrosis.²³ It was also reported that increased MerTK+ macrophages could inhibit the continuous necrosis in APAP-induced ALI by suppressing the activation of neutrophils and accelerating their clearance²⁴ (Figure 1).

2.2.2 | Monocytes

Ly6C^{hi} monocytes controlled the activation and recruitment of neutrophils via CCR2(C-C motif chemokine receptor 2) and M-CSF pathways.²⁵ In the livers of CCR2-/- mice, a significant increase in neutrophils could be observed at 24 hours following APAP administration. Moreover, the ablation of Ly6C^{hi} monocytes and their MoMF (monocyte-derived macrophages) descendants leaded to a profound increase in neutrophil levels²⁶ (Figure 1).

2.2.3 | Myeloid-derived suppressor cells (MDSCs)

Myeloid-derived suppressor cells (MDSCs), a heterogenous population of immune cells from myeloid lineage, can be enlarged during various pathological conditions, such as cancer and inflammatory diseases. Treated with various cytokines, bone marrow-derived MDSCs can be differentiated to various types of cells with different functions. With APAP administration, tumour necrosis factor alpha/LPS-primed MDSCs (TNF- α /LPS MDSCs) could express iNOS to decrease the excessive intrahepatic neutrophil infiltration and induce the apoptosis of activated neutrophils, showing the strongest hepatic protective effect²⁷ (Figure 1).

2.3 | Chemokines and Cytokines

In addition to the interaction between immune cells contributes a lot in APAP-induced hepatitis, some chemokines and cytokines also play an indispensable role. It was demonstrated that chemokines and mitochondria-derived formyl peptides collaborated to recruit neutrophils to sites of liver necrosis via CXC chemokine receptor 2 (CXCR2) and formyl peptide receptor 1 (FPR1), respectively (Figure 2).²⁸ As an important inhibitor of CXCL-1, SOCS2 is able to control the activation and recruitment of neutrophils. In SOCS2-/mice treated with APAP, the expression of the neutrophil-active chemokine CXCL-1 increased significantly, inducing more neutrophil recruitment and liver necrosis (Figure 2).²⁹ Besides, in vivo, gastrin-releasing peptide receptor (GRPR) antagonist inhibited both CXCL2-induced neutrophil migration and activation through the downregulation of CD11b and CD62L. GRPR could also induce activation of MAPKs (p38 and ERK1/2) and downregulation expression of CD11b and CD66b, which significantly inhibited the adhesion and migration of neutrophils. In vitro, it decreased CXCL8-driven neutrophil recruitment independently of CXCR2³⁰ (Figure 2).

Some researchers believed that the cytokines released through inflammasome pathway also affected neutrophil's activation and recruitment. Imaeda et al mentioned that inhibition of the Nalp3 pathway with Aspirin and the knockout of Nalp3 in mice could decrease the infiltration of neutrophils and thus alleviate APAP-induced liver injury.^{31,32} However, C David Williams et al³³ put forward the opposite results that after APAP treatment, mice deficient for each component of the Nalp3 inflammasome (caspase-1, ASC and Nalp3) had similar neutrophil recruitment and IL FY-

liver injury when compared with that of WT mice, which appeared the limit effect of Nalp3 inflammasome. It is notable that these two studies all showed that plasma levels of IL-1 β protein are very minor even after APAP overdose. Moreover, Williams et al treated animals with pharmacological levels of murine recombinant IL-1ß and found that it had no effect on liver injury, which is consistent with IL-1R-/- mice were not protected against APAP hepatotoxicity. These data together indicate that this minor increase in IL-1^β induced by APAP overdose did not aggravate liver injury. In addition, no increase expression of IL-18 was observed in APAPinduced liver injury.³³ Furthermore, Williams et al repeat the aspirin pre-treatment experiment and revealed that like many other interventions against neutrophils, Aspirin cannot protect liver from APAP-induced injury through detecting HMGB1, K-18 and other APAP metabolism bookmarks, which were absent in Imaeda study.³³⁻³⁵ As for the time point. Williams et al³³ took their experiments at 24 hours instead of 12h in the Imaeda study, which should be taken into consideration when comparing their results. Above all, Nalp3 inflammasome signalling might not be a major pathway involved in APAP hepatotoxicity.

Proliferation

Other interleukins also play important roles in the activation and recruitment of neutrophils. The deficiency of IL-13³⁶ aggravated liver damage due to the neutrophils infiltration induced by injurious downstream events whereas absence of IL-17³⁷ significantly attenuated APAP-induced liver injury via decreasing neutrophils recruitment (Figure 2). Acetaminophen overdose caused massive release of IL-33 which can activate and recruit neutrophils, inducing liver injury by IL-33/ST2 signalling pathway.³⁸ In APAP-induced ALI, activated Cot/tpl2 mediated the production of IL-1 α and IL-1 β , and the recruitment of neutrophils in response to DAMPs was largely dependent on IL-1 α^{39} (Figure 2).

CD31, also known as platelet endothelial cell adhesion molecule-1 (PECAM-1/ CD31), has pivotal function in the migration and clearance of neutrophils. The absence of CD31 could significantly alleviate APAP-induced liver injury compared with WT mice.⁴⁰

2.4 | Time points of APAP overdose treatment

Neutrophils migrate into liver tissue in mice in the early stage of APAP-induced liver injury occurrence(6,12 hours), and most were located at the healthy part of liver.^{41,42} Then, the injury began to aggravate until it reached the peak at 24 hours with most of neutrophils migrated into the necrosis area. Subsequently, in the recovery phase of liver injury, neutrophils also began to decrease.

Some human's data showed that the phagocytic capacity and ROS production of neutrophils in patients with APAP-induced liver injury were all increased during the recovery period (day 1-day 4) which indicated that activation of neutrophils occurred after injury peaks. But there were also differences between human and mouse data. The difference was that the expression of CD11b is increased in peripheral blood neutrophils of mice, but not in humans, which may be caused by species variation and injury time.⁴²

3 | COMPLEX ROLES OF NEUTROPHILS

Abundant researches uncovered the pivotal roles of neutrophils in APAP-induced liver injury, a few researchers believe that neutrophils contribute a lot in aggravating APAP-induced liver damage. It was reported that neutrophils depletion by RB6-8C5 (an anti-Gr-1 antibody) moderated APAP-induced ALI, which indicated neutrophils induced liver injury. Yuko Ishida et al also confirmed this result. In vivo, they found improved survival rate in neutropenic WT mice and CXC chemokine receptor 2 (CXCR2)-deficient mice when compared with WT mice under the same dose of APAP treatment. In vitro, isolated human neutrophils were toxic to HepG2 cells when cocultured via direct contact with HepG2 cells and the CXCR2-FPR1-signalling pathway.^{28,43} Researchers further confirmed that neutrophil activation occurred secondary to the initial liver injury induced by APAP. They found that resolvins which can prevent and reduce the infiltration of neutrophils in the inflammation site could extend the therapeutic window after APAP administration. Collectively, these results indicate that neutrophils mediate (at least partially) the hepatotoxic effects of oral acetaminophen.44,45

However, some researchers hold different views. Lawson et al reported that neutrophils participate in necrotic debris removal instead of affecting the pathogenesis of APAP-induced ALI directly.⁴⁶ It was further proved by Williams, C. D et al⁴⁷ that upregulation of CD11b or reactive oxygen was not observed on neutrophils isolated from the liver after APAP administration. Moreover, hepatic neutrophils accumulation and activation caused by high pharmacological doses of IL-1beta do not worsen APAP-induced ALI.⁴⁸ Other researchers showed that β 2 integrins(CD11/CD18) is essential for neutrophils in the transmigration and adherence steps in mice liver. But treatment with anti-CD18 monoclonal antibody had no protective effects on liver damage after APAP administration during the 24h time period, suggesting that neutrophils do not contribute to the initiation or progression of APAP-induced ALI.⁴¹ Clapperton et al⁴⁹ further showed a neutrophil defect in ALF due to overdose of paracetamol, which is complement-dependent, but has nothing to do with serum complement, and may be related to complement receptors.

In addition, with the growing evidence that neutrophils actively control the regression of inflammation and contribute to tissue repair, the perception of the indiscriminate killers of neutrophils seems to be changed. Some researchers have also proposed the repair role of neutrophils in APAP-induce ALI. Researchers found evidence that APAP treatment could lead to the delay of previously activated neutrophil apoptosis, which is beneficial to the repair of injured liver tissue. Freitas, M. et al⁵⁰ also confirmed this view for the longevity of neutrophils was found to be prolonged after APAP administration, which helps tissues heal and resolve inflammation. Mechanically, neutrophils play a pivotal role in liver repair by promoting the conversion of pro-inflammatory Ly6C^{hi}CX₃CR1^{lo} monocyte/macrophage phenotype into prodegradable Ly6C^{lo}CX₂CR1^{hi} macrophages.⁵¹ Furthermore, reactive oxygen species (ROS), mainly expressed by neutrophils in APAP-induced liver injury, are important mediators that trigger

-WILEY-

the repair phenotype transformation of macrophages to promote liver healing.⁵² Recent studies further revealed that blocking platelet CLEC-2 signalling increased the production of tumour necrosis factor alpha (TNF- α), which then further accelerated the recruitment of reparative hepatic neutrophils, thereby promoting liver recovery from acute APAP-induced injuries.⁵³ These studies all strongly confirmed the indispensable role of neutrophils in tissue repair and regeneration.

4 | CONCLUSION

As outlined in this review, we review the neutrophil's activation, recruitment, migration and the complex roles in APAP-induced liver injury. There are different views about the roles of neutrophils involved in APAP-induce liver injury. Some researchers have shown that neutrophils aggravate APAP-induced liver damage, while others demonstrated that neutrophils are not involved in this damage. Several studies revealed that neutrophils even have repair functions in this liver hepatitis model. These guite controversial opinions might be related to time point of experimental observation. The timing of APAP treatment should be taken into account because the pathophysiology of APAP-induced liver injury consists of several different stages which suggests that we should not use a single time point as the experimental observation point. It is necessary to investigate injury events at several time points within 24 hours and evaluate regeneration within 96 hours.⁵⁴ Besides, different susceptibility of various mouse strains to APAP hepatotoxicity has been confirmed which need to be taken into consideration.⁵⁵ What's more, off-target effects in drugs or biological reagents used in APAP relevant researches should not be ignored.⁵⁶ Thus, in order to clarify this elusive question, further studies need to be more detailed concerning the impact on APAP metabolism and intracellular signalling events of different intervention strategies.⁵⁴

But undeniably, the studies of neutrophils also cast light in the prevention and treatment of APAP-induced ALI. Neutrophil elastase (NE) is a pro-inflammatory protein that secreted by activated neutrophils. The combination treatment of NAC (N-acetyl-L-cysteine) and sivelestat (the inhibitor of NE) significantly attenuated liver damage. This strategy is more effective than NAC monotherapy and might provide new options for the treatment of APAP-ALI.⁵⁷ Besides, A large number of studies have shown that some molecules and signal pathways were involved in reducing APAP-induced liver damage by inhibiting the recruitment and infiltration of neutrophils, such as Berberine (BBR),⁵⁸ Geniposide (GP),⁵⁹ BLT1 signalling,⁶⁰ Glycyrrhetinic acid (GA),⁶¹ Citral,⁶² lemongrass essential oil (LGO),⁶³ SmKI-1⁶⁴ and β -d-glucan.⁶⁵ Therefore, it has been proposed that neutrophils can be considered as a target for the treatment of APAP-induced ALI. Through clarification on the relationship of neutrophils with APAP-induced inflammation may provide new strategies for the prevention and treatment of APAP-induced ALI.

ACKNOWLEDGEMENTS

The study was supported by Grants from the National Nature Science Foundation of China, No. U20A20348; the National Nature Science Foundation of China, No. 81871646; the Self-Topic Science Foundation of State Key Laboratory for Diagnosis and Treatment of Infectious Diseases.

CONFLICTS OF INTEREST

The authors declare no financial conflicts of interests.

Proliferation

AUTHOR CONTRIBUTIONS

Huiting Guo wrote the manuscript and prepared figures; Min Zheng, Cheng Zhou, Shiwei Chen and Mingjie Xie provided expert comments and edits. All authors reviewed the manuscript.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ORCID

Min Zheng D https://orcid.org/0000-0001-6159-9879

REFERENCES

- Barbier-Torres L, Iruzubieta P, Fernández-Ramos D, et al. The mitochondrial negative regulator MCJ is a therapeutic target for acetaminophen-induced liver injury. *Nat Commun.* 2017;8:2068. https://doi.org/10.1038/s41467-017-01970-x
- Bernal W, Auzinger G, Dhawan A, Wendon J. Acute liver failure. Lancet. 2010;376:190-201. https://doi.org/10.1016/S0140 -6736(10)60274-7
- Laskin DL, Pendino KJ. Macrophages and inflammatory mediators in tissue injury. Annu Rev Pharmacol Toxicol. 1995;35:655-677. https://doi.org/10.1146/annurev.pa.35.040195.003255
- Wang X, Sun R, Chen Y, Lian ZX, Wei H, Tian Z. Regulatory T cells ameliorate acetaminophen-induced immune-mediated liver injury. *Int Immunopharmacol.* 2015;25:293-301. https://doi.org/10.1016/j. intimp.2015.02.008
- Casanova-Acebes M, Nicolás-Ávila JA, Li Jackson LY, et al. Neutrophils instruct homeostatic and pathological states in naive tissues. J Exp Med. 2018;215:2778-2795. https://doi.org/10.1084/ jem.20181468
- Puga I, Cols M, Barra CM, et al. B cell-helper neutrophils stimulate the diversification and production of immunoglobulin in the marginal zone of the spleen. *Nat Immunol.* 2011;13:170-180. https:// doi.org/10.1038/ni.2194
- Yan M, Huo Y, Yin S, Hu H. Mechanisms of acetaminopheninduced liver injury and its implications for therapeutic interventions. *Redox Biol.* 2018;17:274-283. https://doi.org/10.1016/j. redox.2018.04.019
- Chen D, Ni HM, Wang L, et al. p53 up-regulated modulator of apoptosis induction mediates acetaminophen-induced necrosis and liver injury in mice. *Hepatology*. 2019;69:2164-2179. https://doi. org/10.1002/hep.30422
- Hsu LC, Enzler T, Seita J, et al. IL-1beta-driven neutrophilia preserves antibacterial defense in the absence of the kinase IKKbeta. *Nat Immunol.* 2011;12:144-150. https://doi.org/10.1038/ni.1976
- Oliveira C, Navarro-Xavier R, Anjos-Vallota E, et al. Effect of plant neutrophil elastase inhibitor on leucocyte migration, adhesion and cytokine release in inflammatory conditions. Br J Pharmacol.

6 of 7

 Proliferation

 2010;161:899-910.
 https://doi.org/10.1111/j.1476-5381.2010.

 00924 x
 00924 x

- 11. Mantovani A, Cassatella MA, Costantini C, Jaillon S. Neutrophils in the activation and regulation of innate and adaptive immunity. *Nat Rev Immunol.* 2011;11:519-531. https://doi.org/10.1038/nri3024
- Segel GB, Halterman MW, Lichtman MA. The paradox of the neutrophil's role in tissue injury. *J Leukoc Biol*. 2011;89:359-372. https:// doi.org/10.1189/jlb.0910538
- Kobayashi SD, Voyich JM, Burlak C, DeLeo FR. Neutrophils in the innate immune response. Arch Immunol Ther Exp (Warsz). 2005;53:505-517.
- 14. Kolaczkowska E, Kubes P. Neutrophil recruitment and function in health and inflammation. *Nat Rev Immunol.* 2013;13:159-175. https://doi.org/10.1038/nri3399
- Mishalian I, Granot Z, Fridlender ZG. The diversity of circulating neutrophils in cancer. *Immunobiology*. 2017;222:82-88. https://doi. org/10.1016/j.imbio.2016.02.001
- Huebener P, Pradere JP, Hernandez C, et al. The HMGB1/RAGE axis triggers neutrophil-mediated injury amplification following necrosis. J Clin Invest. 2019;130:1802. https://doi.org/10.1172/JCl12 6975
- Wang X, Sun R, Wei H, Tian Z. High-mobility group box 1 (HMGB1)-Toll-like receptor (TLR)4-interleukin (IL)-23-IL-17A axis in drug-induced damage-associated lethal hepatitis: interaction of gammadelta T cells with macrophages. *Hepatology*. 2013;57:373-384. https://doi.org/10.1002/hep.25982
- Ayata CK, Ganal SC, Hockenjos B, et al. Purinergic P2Y(2) receptors promote neutrophil infiltration and hepatocyte death in mice with acute liver injury. *Gastroenterology*. 2012;143:1620-1629.e1624. https://doi.org/10.1053/j.gastro.2012.08.049
- He Y, Feng D, Li M, et al. Hepatic mitochondrial DNA/Toll-like receptor 9/MicroRNA-223 forms a negative feedback loop to limit neutrophil overactivation and acetaminophen hepatotoxicity in mice. *Hepatology*. 2017;66:220-234. https://doi.org/10.1002/ hep.29153
- Krenkel O, Mossanen JC, Tacke F. Immune mechanisms in acetaminophen-induced acute liver failure. *Hepatobiliary Surg Nutr.* 2014;3:331-343. https://doi.org/10.3978/j.issn.2304-3881. 2014.11.01
- Srungaram P, Rule JA, Yuan HJ, et al. Plasma osteopontin in acute liver failure. Cytokine. 2015;73:270-276. https://doi.org/10.1016/j. cyto.2015.02.021
- He CY, Liang B, Fan X, et al. The dual role of osteopontin in acetaminophen hepatotoxicity. *Acta Pharmacol Sin.* 2012;33:1004-1012. https://doi.org/10.1038/aps.2012.47
- Starkey Lewis P, Campana L, Aleksieva N, et al. Alternatively activated macrophages promote resolution of necrosis following acute liver injury. *J Hepatol.* 2020;73:349-360. https://doi.org/10.1016/j.jhep.2020.02.031
- Triantafyllou E, Pop OT, Possamai LA, et al. MerTK expressing hepatic macrophages promote the resolution of inflammation in acute liver failure. *Gut.* 2018;67:333-347. https://doi.org/10.1136/gutjn l-2016-313615
- Graubardt N, Vugman M, Mouhadeb O, et al. Ly6C(hi) monocytes and their macrophage descendants regulate neutrophil function and clearance in acetaminophen-induced liver injury. *Front Immunol*. 2017;8:626. https://doi.org/10.3389/fimmu.2017.00626
- Zigmond E, Samia-Grinberg S, Pasmanik-Chor M, et al. Infiltrating monocyte-derived macrophages and resident kupffer cells display different ontogeny and functions in acute liver injury. J Immunol. 2014;193:344-353. https://doi.org/10.4049/jimmunol.1400574
- 27. Hsu CY, Lin YC, Chang LY, et al. Therapeutic role of inducible nitric oxide synthase expressing myeloid-derived suppressor cells in acetaminophen-induced murine liver failure. *Front Immunol.* 2020;11:574839. https://doi.org/10.3389/fimmu.2020.574839

- Marques PE, Amaral SS, Pires DA, et al. Chemokines and mitochondrial products activate neutrophils to amplify organ injury during mouse acute liver failure. *Hepatology*. 2012;56:1971-1982. https:// doi.org/10.1002/hep.25801
- Monti-Rocha R, Cramer A, Gaio Leite P, et al. SOCS2 Is critical for the balancing of immune response and oxidate stress protecting against acetaminophen-induced acute liver injury. *Front Immunol.* 2018;9:3134. https://doi.org/10.3389/fimmu.2018.03134
- Czepielewski RS, Jaeger N, Marques PE, et al. GRPR antagonist protects from drug-induced liver injury by impairing neutrophil chemotaxis and motility. Eur J Immunol. 2017;47:646-657. https://doi. org/10.1002/eji.201646394
- Woolbright BL, Jaeschke H. Role of the inflammasome in acetaminophen-induced liver injury and acute liver failure. *J Hepatol.* 2017;66:836-848. https://doi.org/10.1016/j.jhep.2016.11.017
- Imaeda AB, Watanabe A, Sohail MA, et al. Acetaminophen-induced hepatotoxicity in mice is dependent on TIr9 and the Nalp3 inflammasome. J Clin Invest. 2009;119:305-314. https://doi.org/10.1172/ JCI35958
- Williams CD, Antoine DJ, Shaw PJ, et al. Role of the Nalp3 inflammasome in acetaminophen-induced sterile inflammation and liver injury. *Toxicol Appl Pharmacol.* 2011;252:289-297. https://doi. org/10.1016/j.taap.2011.03.001
- Bauer I, Vollmar B, Jaeschke H, et al. Transcriptional activation of heme oxygenase-1 and its functional significance in acetaminopheninduced hepatitis and hepatocellular injury in the rat. J Hepatol. 2000;33:395-406. https://doi.org/10.1016/s0168-8278(00)80275-5
- Cover C, Liu J, Farhood A, et al. Pathophysiological role of the acute inflammatory response during acetaminophen hepatotoxicity. *Toxicol Appl Pharmacol*. 2006;216:98-107. https://doi.org/10.1016/j. taap.2006.04.010
- Yee SB, Bourdi M, Masson MJ, Pohl LR. Hepatoprotective role of endogenous interleukin-13 in a murine model of acetaminopheninduced liver disease. *Chem Res Toxicol.* 2007;20:734-744. https:// doi.org/10.1021/tx600349f
- Lee HC, Liao CC, Day YJ, Liou JT, Li AH, Liu FC. IL-17 deficiency attenuates acetaminophen-induced hepatotoxicity in mice. *Toxicol Lett.* 2018;292:20-30. https://doi.org/10.1016/j.toxlet.2018.04.021
- Antunes MM, Araújo AM, Diniz Ariane B, et al. IL-33 signalling in liver immune cells enhances drug-induced liver injury and inflammation. *Inflamm Res.* 2018;67:77-88. https://doi.org/10.1007/ s00011-017-1098-3
- Sanz-Garcia C, Ferrer-Mayorga G, González-Rodríguez Á, et al. Sterile inflammation in acetaminophen-induced liver injury is mediated by Cot/tpl2. J Biol Chem. 2013;288:15342-15351. https://doi. org/10.1074/jbc.M112.439547
- Cheng GY, Jiang Q, Deng AP, et al. CD31 induces inflammatory response by promoting hepatic inflammatory response and cell apoptosis. Eur Rev Med Pharmacol Sci. 2018;22:7543-7550. https://doi. org/10.26355/eurrev_201811_16296
- Lawson JA, Farhood A, Hopper RD, Bajt ML, Jaeschke H. The hepatic inflammatory response after acetaminophen overdose: role of neutrophils. *Toxicol Sci.* 2000;54:509-516. https://doi.org/10.1093/toxsci/54.2.509
- Williams CD, Bajt ML, Sharpe MR, McGill MR, Farhood A, Jaeschke H. Neutrophil activation during acetaminophen hepatotoxicity and repair in mice and humans. *Toxicol Appl Pharmacol*. 2014;275:122-133. https://doi.org/10.1016/j.taap.2014.01.004
- Ishida Y, Kondo T, Kimura A, et al. Opposite roles of neutrophils and macrophages in the pathogenesis of acetaminophen-induced acute liver injury. *Eur J Immunol*. 2006;36:1028-1038. https://doi. org/10.1002/eji.200535261
- Patel SJ, Luther J, Bohr S, et al. A novel resolvin-based strategy for limiting acetaminophen hepatotoxicity. *Clin Transl Gastroenterol*. 2016;7:e153. https://doi.org/10.1038/ctg.2016.13

- Smith GS, Nadig DE, Kokoska ER, Solomon H, Tiniakos DG, Miller TA. Role of neutrophils in hepatotoxicity induced by oral acetaminophen administration in rats. *J Surg Res.* 1998;80:252-258. https:// doi.org/10.1006/jsre.1998.5441
- Connolly MK, Ayo D, Malhotra A, et al. Dendritic cell depletion exacerbates acetaminophen hepatotoxicity. *Hepatology*. 2011;54:959-968. https://doi.org/10.1002/hep.24429
- Williams CD, Bajt ML, Farhood A, Jaeschke H. Acetaminopheninduced hepatic neutrophil accumulation and inflammatory liver injury in CD18-deficient mice. *Liver Int*. 2010;30:1280-1292. https:// doi.org/10.1111/j.1478-3231.2010.02284.x
- Williams CD, Farhood A, Jaeschke H. Role of caspase-1 and interleukin-1beta in acetaminophen-induced hepatic inflammation and liver injury. *Toxicol Appl Pharmacol.* 2010;247:169-178. https:// doi.org/10.1016/j.taap.2010.07.004
- Clapperton M, Rolando N, Sandoval L, Davies E, Williams R. Neutrophil superoxide and hydrogen peroxide production in patients with acute liver failure. *Eur J Clin Invest*. 1997;27:164-168. https://doi.org/10.1046/j.1365-2362.1997.920640.x
- Freitas M, Costa V, Ribeiro D, et al. Acetaminophen prevents oxidative burst and delays apoptosis in human neutrophils. *Toxicol Lett.* 2013;219:170-177. https://doi.org/10.1016/j.toxlet.2013.03.007
- Yang W, Tao Y, Wu Y, et al. Neutrophils promote the development of reparative macrophages mediated by ROS to orchestrate liver repair. *Nat Commun.* 2019;10:1076. https://doi.org/10.1038/s4146 7-019-09046-8
- Takada H, Mawet E, Shiratori Y, et al. Chemotactic factors released from hepatocytes exposed to acetaminophen. *Dig Dis Sci.* 1995;40:1831-1836. https://doi.org/10.1007/BF02212709
- Chauhan A, Sheriff L, Hussain M, et al. The platelet receptor CLEC-2 blocks neutrophil mediated hepatic recovery in acetaminophen induced acute liver failure. *Nat Commun.* 2020;11:1939. https://doi. org/10.1038/s41467-020-15584-3
- Jaeschke H, Ramachandran A. Mechanisms and pathophysiological significance of sterile inflammation during acetaminophen hepatotoxicity. *Food Chem Toxicol.* 2020;138:111240. https://doi. org/10.1016/j.fct.2020.111240
- Harrill AH, Ross PK, Gatti DM, Threadgill DW, Rusyn I. Populationbased discovery of toxicogenomics biomarkers for hepatotoxicity using a laboratory strain diversity panel. *Toxicol Sci.* 2009;110:235-243. https://doi.org/10.1093/toxsci/kfp096
- Jaeschke H, Liu J. Neutrophil depletion protects against murine acetaminophen hepatotoxicity: another perspective. *Hepatology*. 2007;45:1588-1589. https://doi.org/10.1002/hep.21549
- 57. Raevens S, Van Campenhout S, Debacker PJ, et al. Combination of sivelestat and N-acetylcysteine alleviates the inflammatory

response and exceeds standard treatment for acetaminopheninduced liver injury. *J Leukoc Biol.* 2020;107:341-355. https://doi. org/10.1002/JLB.5A1119-279R

 Zhao Z, Wei Q, Hua W, Liu Y, Liu X, Zhu Y. Hepatoprotective effects of berberine on acetaminophen-induced hepatotoxicity in mice. *Biomed Pharmacother*. 2018;103:1319-1326. https://doi. org/10.1016/j.biopha.2018.04.175

Proliferation

- 59. Yang S, Kuang G, Jiang R, et al. Geniposide protected hepatocytes from acetaminophen hepatotoxicity by down-regulating CYP 2E1 expression and inhibiting TLR 4/NF-kappaB signaling pathway. *Int Immunopharmacol.* 2019;74:105625. https://doi.org/10.1016/j. intimp.2019.05.010
- Kojo K, Ito Y, Eshima K, et al. BLT1 signalling protects the liver against acetaminophen hepatotoxicity by preventing excessive accumulation of hepatic neutrophils. *Sci Rep.* 2016;6:29650. https:// doi.org/10.1038/srep29650
- Yang G, Zhang L, Ma L, et al. Glycyrrhetinic acid prevents acetaminophen-induced acute liver injury via the inhibition of CYP2E1 expression and HMGB1-TLR4 signal activation in mice. *Int Immunopharmacol.* 2017;50:186-193. https://doi.org/10.1016/j. intimp.2017.06.027
- Uchida NS, Silva-Filho SE, Cardia GFE, et al. Hepatoprotective effect of citral on acetaminophen-induced liver toxicity in mice. *Evid Based Complement Alternat Med.* 2017;2017:1796209. https://doi.org/10.1155/2017/1796209
- Uchida NS, Silva-Filho SE, Aguiar RP, et al. Protective effect of Cymbopogon citratus essential oil in experimental model of acetaminophen-induced liver injury. Am J Chin Med. 2017;45:515-532. https://doi.org/10.1142/S0192415X17500318
- Morais SB, Figueiredo BC, Assis NRG, et al. Schistosoma mansoni SmKI-1 serine protease inhibitor binds to elastase and impairs neutrophil function and inflammation. *PLoS Pathog.* 2018;14:e1006870. https://doi.org/10.1371/journal.ppat.1006870
- Toklu HZ, Sehirli AO, Velioglu-Ogunc A, Cetinel S, Sener G. Acetaminophen-induced toxicity is prevented by beta-D-glucan treatment in mice. *Eur J Pharmacol*. 2006;543:133-140. https://doi. org/10.1016/j.ejphar.2006.05.033

How to cite this article: Guo H, Chen S, Xie M, Zhou C, Zheng M. The complex roles of neutrophils in APAP-induced liver injury. *Cell Prolif.* 2021;54:e13040. <u>https://doi.org/10.1111/</u>cpr.13040