## MINI REVIEW

Reproductive Medicine and Biology

WILEY

# Genetic counseling prior to assisted reproductive technology

Yukiko Katagiri<sup>1,2,3</sup> 🕑 | Yuko Tamaki<sup>1,2,3</sup>

<sup>1</sup>Department of Obstetrics and Gynecology, Faculty of Medicine, Toho University, Tokyo, Japan

<sup>2</sup>Division of Clinical Genetics, Toho University Omori Medical Center, Tokyo, Japan

<sup>3</sup>Reproduction Center, Toho University Omori Medical Center, Tokyo, Japan

#### Correspondence

Yukiko Katagiri, Department of Obstetrics and Gynecology, Faculty of Medicine, Toho University, 6-11-1 Omori-nishi, Ota-ku, Tokyo 143-8541, Japan. Email: yukikonk@med.toho-u.ac.jp

### Abstract

Background: Reproductive medicine deals with fertility and is closely related to heredity. In reproductive medicine, it is necessary to provide genetic information for the patients prior to assisted reproductive technology (ART). Japan Society for Reproductive Medicine (JSRM) requires doctors involved in reproductive medicine to have standard knowledge of reproductive genetics and knowledge of reproductive medicine, which is covered in their publication, "required knowledge of reproductive medicine."

Methods: With the aim of providing straightforward explanations to patients in the clinical situation at pre-ART counseling, we provide the following five topics, such as (a) risk of birth defects in children born with ART, (b) chromosomal abnormalities, (c) Y chromosome microdeletions (YCMs), (d) possible chromosomal abnormal pregnancy in oligospermatozoa requiring ICSI (intracytoplasmic sperm injection), and (e) epigenetic alterations.

Main findings: The frequency of chromosome abnormalities in infertile patients is 0.595%-0.64%. YCMs are observed in 2%-10% of severe oligospermic men. High incidence of spermatozoa with chromosomal abnormalities has been reported in advanced oligospermia and asthenozoospermia that require ICSI. Some epigenetic alterations were reported in the children born with ART.

Conclusion: Certain genetic knowledge is important for professionals involved in reproductive medicine, even if they are not genetic experts.

#### KEYWORDS

chromosomal abnormalities, epigenetics, genetic counseling, intracytoplasmic sperm injection, Y chromosome microdeletions

#### | INTRODUCTION 1

Since the birth of Louise Brown, the first IVF baby in 1978, currently, more than 1.7 million cycles of assisted reproductive technology (ART) are carried out annually worldwide, and more than 400 000 children are born by ART although the data for some countries and regions are not included.<sup>1,2</sup> ART is widely used as a standard infertility treatment. However, the major difference between reproductive medicine and other medical care is that reproductive medicine is related to the birth of the next generation. Although ART has been performed safely, there are several genetic challenges. It is appropriate to provide genetic information prior to the start of ART, and specific counseling is required for patients with or likely to have genetic changes.<sup>3-7</sup>

Genetic counseling is a medical practice that provides and supports appropriate genetic information for patients/family

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2020 The Authors. Reproductive Medicine and Biology published by John Wiley & Sons Australia, Ltd on behalf of Japan Society for Reproductive Medicine.

Reproductive Medicine and Biolc

members of hereditary diseases or those who have the possibility to be able to decide and act on life plan choices by themselves.<sup>8</sup> In genetic counseling, it is performed step by step, such as whether it is a chromosome issue or a gene issue. In addition to genetic counseling in infertility treatment, it is important to clarify whether it is the cause of infertility, whether it is related to infertility treatment, or whether it may affect the born child or not. For genetic counseling in reproductive medicine, it is appropriate to respond before starting ART.

Common genetic counseling is often initiated from a disease, whereas in genetic counseling in reproductive medicine, healthy couples often undergo genetic testing as an infertility screening test. Genetic change is recognized, and they will see counseling. Compared to the frequency of genetic disorders in female infertile patients who experience menstruation, the rate of genetic disorders is high in male infertility.<sup>1,9-18</sup> So, genetic testing is recommended for male infertility, which may be often the origin of genetic counseling. Genetic information provision/counseling in reproductive medicine provides typical genetic information in reproduction. There are five general categories of genetic subjects. (a) risk of birth defects in children born with ART, (b) chromosomal abnormalities, (c) Y chromosome microdeletions (YCMs), (d) possible chromosomal abnormal pregnancy in oligospermatozoa requiring ICSI (intracytoplasmic sperm injection), and (e) epigenetic alterations. We review five issues of typical genetic subjects for pre-ART counseling, with the aim of providing straightforward explanations to patients in the clinical situation.

# 2 | RISK OF BIRTH DEFECTS IN CHILDREN BORN WITH ART

Congenital deficits in ART children include two problems: the child's own problems and the risk of maternal perinatal complications.

The general frequency of apparent morphological abnormalities is about 2% in newborns.<sup>19</sup> In addition, congenital anomalies are about 3%-4% with mental retardation and genetic disorders.<sup>19</sup> Regarding the association between ART pregnancy and congenital anomalies, there are both reports of a 1.37-fold increased birth defects in ART,<sup>20</sup> though there is no statistically significant difference between ART and natural conception.<sup>21-23</sup> There are also reports that techniques such as ICSI and TESE do not increase birth defects.<sup>24,25</sup> In the reports that ART children are at increased risk of major congenital malformations and chromosomal abnormalities compared with children born after natural conception, the risk is mainly due to the risk factors for the father and the mother. It is interpreted that it is not due to ART but due to the background of couples who require ART.<sup>26</sup>

In maternal perinatal complications after ART, it has been reported to increase the risk of gestational diabetes (GDM) (OR: 1.99, 95% CI: 1.69-2.36), gestational hypertension (OR: 2.58, 95% CI: 2.11-3.15), preeclampsia (OR: 1.49, 95% CI: 1.12-1.98), intrahepatic cholestasis of pregnancy (ICP) (OR: 2.86, 95% CI: 2.39-3.42),

placenta previa (OR: 2.23, 95% CI: 1.79-2.78), placental abruption (OR: 5.06, 95% CI: 2.83-9.06), preterm premature rupture of membranes (pPROM) (OR: 3.05, 95% CI: 2.48-3.74), placental adherence (OR: 2.37, 95% CI: 1.90-2.95), postpartum hemorrhage (OR: 2.72, 95% CI: 2.18-3.41), and polyhydramnios (OR: 1.79, 95% CI: 1.26-2.53).<sup>27-30</sup>

It has been reported that a link suggested between ART technology and birthweight, and maternal complications such as preeclampsia and placenta previa. It has been clarified that the birthweight of a baby born by fresh embryo transfer is lighter than that of a naturally pregnant baby. On the other hand, babies born by frozen-thawed embryo transfer are heavier than babies born by natural conception.<sup>31,32</sup>

Imudia et al reported that serum E2 > 3,450 pg/ml significantly increased the risk (OR: 9.40, 95% CI: 3.22-27.46) of low birthweight infants in 292 singleton pregnancies by fresh embryo transfer.<sup>33</sup> Pereira et al analyzed 4071 singleton pregnancies by fresh embryo transfer and reported that serum E2 > 2500 pg/ml was an independent risk factor (OR: 10.8, 95% CI: 9.2-12.5) for the birth of low birthweight infants.<sup>34</sup> High estrogen levels during fresh embryo transfer are thought to be responsible for low birthweight.

It has been reported that the incidence of hypertensive disorder of pregnancy (HDP) is increased in pregnancy by frozen embryo transfer.<sup>35,36</sup> According to an analysis based on Japan's 2014 ART database, gestational hypertension nephropathy was 1.43 times (95% Cl: 1.14-1.80) and placenta accreta was 6.91 times (95% Cl: 2.87-16.66) in frozen embryo transfer under the hormone replacement cycle compared with the natural cycle.<sup>37</sup> It has been reported that the corpus luteum-derived factor does not work under the hormone replacement cycle, and it becomes poorly adapted to changes in hemodynamics associated with pregnancy, which may increase the incidence of preeclampsia.<sup>38</sup>

It has been pointed out that placenta previa, residual placenta, and postpartum bleeding increase in pregnancy by in vitro fertilization. Smoking, endometriosis, and endometrial thickness are cited as independent placenta previa risks, and endometrial thickness at transplant is 2.02 times (95% CI: 1.12-3.65, P = .02) at 9-11 mm compared with less than 9 mm, 3.74 times (95% CI: 1.90-7.34, P < .01) for 12 mm or more, and it has been reported that the incidence of placenta previa increases.<sup>39</sup> The thickness of the endometrium at the time of transplant is related to the risk of placenta previa, and it is considered important to adjust the endometrium to obtain an appropriate endometrial thickness.

The association between ART and complications includes various confounding factors such as the background of spermatogenic disorders, ovulation affects, aging, and district have an effect. There is also increasing evidence that infertility is an independent risk factor for obstetric complications and perinatal adverse outcomes without the addition of ART.<sup>23</sup> Therefore, technique-related risks cannot be independently eliminated. ART is a safe alternative for couples who are otherwise unable to conceive, but the risk requires thorough evaluation and counseling before ART is performed.<sup>27,40</sup>

#### CHROMOSOMAL ABNORMALITIES 3

The frequency of chromosome abnormalities in the general population is approximately 0.65% in screening tests for newborns, and chromosomal abnormalities in infertile patients are 0.595% in women and 0.64% in men.<sup>9-11</sup> Especially for couples who are treated with ART, the incidences of chromosomal abnormalities are 1.2%-2.1% in female and 1.1%-6.1% in male.<sup>1,12</sup> In addition, it is observed in 3.8%-18.4% in severe oligospermia and 14.7%-35% in azoospermia<sup>13,14</sup> (Table 1).<sup>1,9-14,41</sup> Chromosomal abnormalities include an euploidy and structural abnormalities.

### 3.1 | Aneuploidy

An euploidy is an abnormality in the number of chromosomes with a large or small number of chromosomes, and the mechanism due to meiotic nondisjunction. Patients undergoing infertility treatment are adults of sexual maturity, and it is necessary to consider separately the chromosomal abnormalities that are taken up as problems of the patient themselves and the chromosomal abnormalities that are taken up as problems of the resulting embryos, fetuses, and neonates. Aneuploidy includes autosomal aberrations and sex chromosome aberrations. Sex chromosomal aneuploidy is the most common abnormality among human aneuploidy, especially in infertile male patients.

Klinefelter's syndrome (KS) is the most frequent observed sex chromosomal abnormality, with an estimated frequency of 1:500 to 1:1000 men.<sup>11,42</sup> KS has an extra X chromosome (genotype XXY) instead of the usual male sex complement (genotype XY). The classic form of KS, which is present in the 80%-90% of the cases, is defined by a 47,XXY karyotype resulting from the aneuploidy of the sex chromosomes, whereas higher-grade aneuploidies (eg, 48, XXXY or 48, XXYY), structurally abnormal X chromosome (eg, 47, iXq,Y), or mosaicisms (eg, 47,XXY/46,XY) make up approximately in the remaining 10%-20% of cases.<sup>42</sup> For any genotype, hypogonadism is a common symptom in KS.43 The prevalence of KS rises to 3%-4% among infertile males and 10%-12% in non-obstructive azoospermia (NOA).<sup>11</sup> This may indicate that the rise of the KS might be related to the parental meiotic alterations. The recurrence rate is low due to chromosomal insemination during gametogenesis. KS patients have a phenotype, which is extremely variable,43-47 but without any obvious facial dysmorphology that makes them indistinguishable from the boys with normal karyotype.<sup>45</sup> It is rarely diagnosed in childhood and adolescence and is often diagnosed by infertility examination. The mean age of diagnosis is in the mid-30s reproductive age.<sup>42</sup> Because spermatogenesis has affected in KS, surgical correction for spermatozoa is required frequently. It has been reported that many KS patients could conceive a child with TESE (testicular sperm extraction)/ICSI, and the offspring were healthy with normal karyotypes.<sup>48-50</sup> The risk of ART for patients with KS is not high.<sup>51</sup> Sperm retrieval rates (SRRs) in KS adults are approximately 50%-70% with TESE and micro-TESE, which are higher than those of other NOA cases.<sup>49,52,53</sup> However, the SRR in KS patients decreases with aging.<sup>49,52,54</sup> In the exception of sex chromosome abnormalities in men, there is 47,XYY syndrome, which has an incidence of 0.1% of male births.<sup>18</sup>

Turner syndrome (TS) is one of the most common sex chromosomal abnormalities in women. Turner syndrome is a monosomy of the X chromosome, typically 45, X, and includes structural abnormalities such as i (Xq), Xp-, Yp-, and various mosaics. Although it is a disease with a high miscarriage rate, it is present at 0.05%-0.125% in female birth. Turner's syndrome may be diagnosed in early childhood due to skeletal signs such as valgus elbow and fourth metacarpal shortening, soft tissue signs such as pterygium and lymphedema, visceral malformations such as aortic constriction and renal malformation, congenital lymphedema, and sensorineural hearing loss. However, it is often diagnosed as short stature or amenorrhea after puberty. In many cases, premature ovarian insufficiency (POI) has already occurred in TS, at the time of infertility treatment, because of homologous chromosome pairing failure at meiosis.55,56 The rate of spontaneous pregnancy is about 2%-5% in TS.<sup>57</sup> It has been reported healthy offspring from TS with their own oocvtes.<sup>58</sup> Trisomy X (47,XXX genotype) is also one of the most common female

	Female (%)	Male (%)				Reference's
		Total	KS	Autosomal t	Rob	No.
General population	0.85	0.85	0.1- 0.2	0.25	0.1	19
Infertility	0.595	0.64		0.5-1.0	0.8	9-11
Couples in ART	1.5	1.1				1,9-12
Couples with ICSI	2.1	6.1				1,9-12
Severe oligospermia	-	5-7	2-5		3.4	13,14
Azoospermia	-	10-15	5-10			13,14

Note: The reason that the proportion of chromosomal aberration in the general population is higher than that in infertile patients may be that those with severe clinical symptoms are not included in infertile patients.

Abbreviations: Autosomal t, autosomal translocation; KS, Klinefelter's syndrome; Rob, Robertsonian's translocation.

TABLE 1 Possibility that either couple is a chromosome carrier<sup>1,9-14,19</sup>

abnormal



FIGURE 1 Types of segregation at meiosis in reciprocal balanced translocation. Balanced translocation chromosomes can segregate 2:2 (ie, two chromosomes go to each pole) and 3:1 (ie, leading gametes with 22 or 24 chromosomes). There are three types of 2:2 segregation, described as alternate, adjacent 1, and adjacent 2. Both adjacent 1 segregation and adjacent 2 segregation yield unbalanced gametes

1

unbalanced

chromosomal abnormalities, occurring in approximately 0.1% of female births. The disease presents with a variable phenotype caused by the presence of an extra X chromosome.<sup>59</sup> Pubertal onset and sexual development are usually normal in trisomy X; however, there have been cases of POI.<sup>59-61</sup> Sex chromosome aneuploidy should be the most common cause of POI.<sup>55,56,59,60,62</sup>

mutual exchange

#### 3.2 Structural abnormalities

separation

Chromosomal structural abnormalities include reciprocal translocation and the Robertsonian translocation. Reciprocal translocations occur when heterologous chromosomes are cleaved or rearranged. Reciprocal translocation includes balanced and unbalanced types. An example of a breakpoint in a chromosome test is shown in the figure (Figure 1). Translocations usually occur only between two chromosomes. All-arm reciprocal translocation between homologous chromosomes is impossible to acquire a live child.<sup>63</sup> The person does not affect the phenotype unless there is an overall excess or deficiency, and the carrier is healthy. So, a balanced reciprocal translocation refers to a translocation in which the gene is missing or negligible and the phenotype is normal. The frequency of reciprocal translocation is generally about 0.25%, but it is found in 0.5%-1.0% in infertile men.<sup>1,9-12,64</sup> When an unbalanced gamete is subjected

### TABLE 2 Subgroup of human chromosome

47

Group	Chromosome No.	Length	Location of centromere
A	1-3	Long	Metacentric chromosome or submetacentric chromosome
В	4-5	Long	Submetacentric chromosome
С	6-12, X	Moderate	Submetacentric chromosome
D	13-15	Moderate	Acrocentric chromosome
E	16-18	Relatively short	Metacentric chromosome or submetacentric chromosome
F	19-20	Short	Metacentric chromosome
G	21-22, Y	Short	Acrocentric chromosome

to fertilization, an embryo with an abnormal chromosome number is formed. Although there are 16 karyotypes of gametes, there are nine types that can be born. Because of the chromosomal imbalance, the meiosis of gametogenesis stops in the middle and may exhibit spermatogenic dysfunction. In fact, half of the embryos are of normal karyotype or balanced type. The proportion of chromosomal imbalance in gametes subjected to fertilization has decreased.63 Couples with reciprocal translocations may experience poorer ART results than couples without chromosomal abnormalities, because

X	metacentric chromosome	the centromere is located approximately in the center of the chromosome and the lengths of the two arms (short arm and long arm) are approximately equal
X	submetacentric chromosome 2, 4, 5, 6, 7, 8, 9, 10, 11, 12, 17, 18, X,	the centromere is located slightly off the center of the chromosome and the short arm can be easily distinguished from the long arm, but the short arm is not extremely short
X	acrocentric chromosome 13, 14, 15, 21, 22, Y	the centromere is close to the end of the chromosome and the short arm is extremely short

FIGURE 2 Chromosome types in human. Based on the length and centromere location, all chromosomes are classified as metacentric chromosomes, submetacentric chromosomes, and acrocentric chromosomes



**FIGURE 3** An example of karyotype in the Robertsonian translocation. The Robertsonian translocation is written as "rob" or "der." The Robertsonian translocation results in 45 chromosomes of which two of the acrocentric chromosomes in groups D<sup>13-15</sup> and G<sup>21,22</sup> translocate and lose their short arms

most chromosomally abnormal embryos are spontaneously culled as arrested growth or implantation failure.<sup>65</sup> The ratio of natural selection is due to the size of the translocation segment.

Chromosomes have centromeres that lie between short arms and long arms as a boundary. Chromosomes are classified from Group A to Group G according to their size and centromere location (Table 2). The chromosomes in Group D and Group G, whose short arms are extremely short, are called acrocentric chromosomes (Figure 2).<sup>66</sup> The Robertsonian translocation is present in 0.1% of the general population and 0.8% of male infertile patients.<sup>16</sup> It refers to an acentric chromosome excluding the Y chromosome, in which two have lost their short arms due to the translocation and have 45 chromosomes (Figure 3). Most of the Robertson translocations are rob (13; 14), rob (14; 21), and rob (21; 21), and others are rare. If one of the couples has a Robertsonian translocation and the other has a normal karyotype, six karyotypes' embryos are produced (Figure 4). The probability that

a child of trisomy will be born is higher when the mother is a translocation carrier than when the father is a carrier (Table 3). As with reciprocal translocation, gametogenesis is impaired due to meiotic arrest, resulting in an increase in normal or balanced embryo. Among couples who have a Robertsonian translocation in a male, the probability that a chromosomally abnormal embryo will continue to be pregnant is lower than that in a woman with a translocation.<sup>18</sup>

# 4 | Y CHROMOSOME MICRODELETIONS

Along with chromosomal abnormalities, another genetic factor for male infertility is YCMs.<sup>67</sup> Several YCMs have been reported to be involved in male infertility.<sup>68-73</sup> Among them, azoospermia factor (AZF), a gene related to spermatogenesis, is present in the long arm of the Y chromosome.<sup>74-77</sup> AZF microdeletion is observed in 2%-10% of severe oligospermic men and in 5%-15% of non-obstructive azoospermic men.<sup>74,78-80</sup> AZF has been classified into three areas: a, b, and c,<sup>76</sup> and AZFc deletion is a maximum frequency of 80%, and the frequency of AZFa is 0.5%-4%, 1%-5% in the AZFb region, and 1%-3% in the AZFb + c region.<sup>73</sup> However, it has been revealed that there are five palindrome structures in the long arm of the Y chromosome.<sup>75,81-84</sup> The palindrome structure has a homologous and co-directional set structure in its base sequence, and deletion occurs as a result of pathological recombination between sets.<sup>81</sup> For example, recombination between P5 and proximal P1 results in AZFb, and recombination between P5 and distal P1 results in AZFb + c (Figure 5). In AZFa deletion, the histological phenotype is Sertoli cell-only syndrome (SCO),<sup>75</sup> and in AZFb deletion, it is maturation arrest.<sup>85</sup> In cases with AZFa deletion and/or AZFb deletion, the possibility of sperm recovery is unlikely even if testicular sperm extractions performed. So, AZF is used to evaluate the possibility of sperm collection. Although there are various theories in the evaluation of the AZFc region,<sup>85,86</sup> the possibility of sperm recovery can be expected to be about 70% even in the case 138

	balanced		unbalanced			
			unbalanced chr 14		unbalanced chr 21	
gamete karyotype after the first meiosis in a carrier	14 20	21)	21)	21	21)	14
fertilization	×	×	×	×	×	×
gamete karyotype after the first meiosis in a person with normal karyotype	14 21	14 21	14 21		14 (21)	14 21
karyotypes of embryos						
	normal	carrier	trisomy 14	monosomy 14	trisomy 21	monosomy 21
birth rate of a child with a chromosomal abnormality when the wife is a carrier	36%	54%			10%	
ex. a case with rob(14;21)	14	normal chromosome	14 21	normal chromosor	me 21	21) rob(14;21)

**FIGURE 4** Karyotypes of gametes and embryos in couples with the Robertsonian syndrome. If one in the couple has a Robertsonian translocation and the other has a normal karyotype, six karyotypes are produced [Colour figure can be viewed at wileyonlinelibrary.com]

Balanced translocation	Mother is a carrier of translocation (%)	Father is a carrier of translocation (%)
rob(13;13)	100	100
rob(13;14)	<1	Rare
rob(14;21)	10	2.4
rob(21;21)	100	100
rob(21;22)	6.8	<2.9

**TABLE 3** The incidence of trisomychildren born, whom with translocation,from the balanced Robertsoniantranslocation carriers

*Note*: The risk of trisomy in a child is higher when the mother is a translocation carrier than when the father is a translocation carrier.

of complete deletion in the AZFc region.<sup>74</sup> Although a gr/gr region deletion exists in the AZFc, no significant correlation was observed between the gr/gr region deletion and spermatogenesis in the Japanese population.<sup>85</sup> Although men with severe spermatogenic disorder have been able to raise their children by ICSI, if the spermatogenic disorder is due to the AZF microdeletion, and a boy is delivered by the contribution of ICSI, YCMs are inherited and similar genetic aberrations may be transmitted. In addition, microdeletions may newly occur or expand, and the range of inherited microdeletions may be expanded, resulting in a worsening of spermatogenic disorder than the father.<sup>86</sup> It is important to fully explain them before ART.

# 5 | POSSIBILITY OF CHROMOSOMAL ABNORMAL PREGNANCY IN OLIGOSPERMIA REQUIRING ICSI

Since 1992, the development of intracytoplasmic sperm injection (ICSI) has rejoiced couples with infertility and especially those affected by severe male factor infertility.<sup>87</sup> There are two concerns about the safety of ICSI. The first is the possibility of fertilization operations affecting the embryo, and the second is an increase in birth defects due to the use of severe oligo-, astheno-, and/or teratozoospermia. The former is denied, while the latter increases some



FIGURE 5 AZF deletions by palindromic structures. In the long arm of the Y chromosome, there are five palindromic structures. The palindromic structure has a homologous and co-directional set structure in its base sequence, and deletion occurs as a result of pathological recombination between sets. For example, recombination between P5 and proximal P1 results in AZFb, and recombination between P5 and distal P1 results in AZFb + c

chromosomal abnormalities.<sup>9,20,21,88-90</sup> Prenatal diagnosis of post-ICSI pregnancy has reported 2.96% of chromosomal abnormalities, 1.39% of structural abnormalities such as parental translocations, 1.58% of de novo chromosomal abnormalities, 0.63% of sex chromosomal abnormalities, 0.5% of autosomal aneuploidies, and 0.44% of structural abnormalities, which were not derived from the parent. These chromosomal abnormalities occur 3-5 times the general frequency.<sup>9</sup> As the background of the increase in the numerical chromosomal abnormalities, aging of the wife is considered as a confounding factor. However, in advanced oligospermia and asthenozoospermia that require ICSI, the reason is the high incidence of spermatozoa with chromosomal abnormalities. The risk of congenital malformation is 7.1% for ICSI and 4.0% for the general population (OR: 1.99, 95% CI: 1.87-2.11).<sup>91</sup> It has been reported that when sperm is damaged by oxidative stress, sperm DNA is damaged. The percentage of sperm with this damaged DNA is called the DNA fragmentation index (DFI). A high sperm DFI (over 15%) will increase the miscarriage rate.<sup>92,93</sup>

#### 6 **EPIGENETIC ALTERATIONS**

Epigenetics is a mechanism found in mammals that changes the gene expression without changing the DNA sequence.<sup>94,95</sup> Although in many genes, the expression control is the same regardless of whether it is derived from the father or the mother, the genome imprinting is an epigenetic phenomenon in which the expression of the gene derived from the father is different from the expression of the gene of the mother. For some genes called imprinted genes, only genes from either the father or mother are expressed, other genes are suppressed, and the genes function normally in that combination. An imprinted gene is due to inactivation by methylation. The gene to be expressed is not expressed, or the gene to be suppressed is expressed, or two chromosomes that should be received one by one from both parents are inherited from one parent only (uniparental disomy), imprint gene on / off does not go well, and disease develops. It has been reported imprint abnormalities, such as Prader-Willi syndrome, Angelman syndrome, Beckwith-Wiedemann syndrome, Silver-Russell syndrome, and retinal cell tumor. Epigenetics can also affect fetal development, birthweight, and insulin resistance and cardiovascular disease.96 Imprint abnormalities have been reported to be more common in children born by ART than in children born by natural conception.<sup>97,98</sup> In ART pregnancy, it is clear that the neonatal birthweight increases after blastocyst transfer rather than cleavage embryo transfer, and after frozen-thawed embryo transfer compared with fresh embryo transfer.<sup>99,100</sup> Imprinting occurs mainly at the stage of gametogenesis, fertilization, and early embryo development.<sup>95,101-107</sup> Reproductive Medicine and Biology

So, there are concerns that the effects of reproductive medicine on epigenetics such as ovarian stimulation, in vitro maturation, in vitro fertilization, culture conditions, and cryopreservation are affected. However, a pilot study has found that DNA methylation errors in imprinted genes in children born after ART have not been apparent,<sup>108</sup> and infertility treatment does not cause imprint abnormalities, but patient background required infertility treatment is involved in epigenetic changes.<sup>109-111</sup> On the other hand, most ART-related mutations in pre- and postnatal methylation occur independently of embryo culture, and the epigenetic birth-related changes associated with ART are largely resolved by adulthood. There is no direct evidence that ART-related mutations in E. coli affect development and health.<sup>112</sup> These suggest that epigenetics is involved with ART. Further research is needed to avoid the risk of epigenetic changes due to ART and to confirm that ART is not associated with child epigenetic changes.<sup>113-115</sup>

# 7 | FOR MORE INFORMATION

There is a technique called preimplantation genetic testing (PGT) that has been made possible by the developments of reproductive technologies and genetic analysis.<sup>116</sup> PGT is a method of genetically evaluating an embryo by performing an embryo biopsy prior to transfer to the uterus. There are three categories of preimplantation diagnosis: preimplantation genetic testing for monogenic / single gene defects (PGT-M), preimplantation genetic for diagnosing embryonic chromosomal structural abnormalities against the background of recurrent miscarriage of translocation carriers testing for structural rearrangement (PGT-SR), and preimplantation genetic testing for aneuploidy (PGT-A) for the purpose of embryo transfer without chromosomal abnormalities, especially chromosomal numerical abnormalities, with the aim of improving implantation rates and reducing miscarriage rates. Since indication and operation rules differ depending on the countries or regions, counseling for them varies depending on the rules in the countries or regions. The most important thing seems to be the provision of medical services without any disadvantage for clients who need medical technology. Counseling is required to accurately understand the information that most patients need.

# 8 | CONCLUSION

The genetic counseling is important for couples undergoing infertility treatment to understand the genetic background and unclear points of ART. It is important for the couples to know in advance the risk of birth defects and chromosomal abnormalities that are born with a certain probability, and it is also necessary for the medical staffs who provide reproductive techniques to understand that as well. The genetic counseling is often provided by genetic experts; however, reproductive staffs also require standard knowledge of genetics. Prior to ART, patients should be able to receive standard information about the genetics of ART equally and accurately.

### ACKNOWLEDGMENTS

This paper was presented as a special lecture at the 36th Annual Meeting of the Japanese Society of Fertilization and Implantation. The authors thank for the opportunity to speak.

### DISCLOSURES

*Conflict of interest*: The authors declare no conflict of interest. *Human/Animal rights*: This article does not contain any studies with human and animal subjects performed by any of the authors.

# ORCID

Yukiko Katagiri D https://orcid.org/0000-0002-0052-896X

#### REFERENCES

- Adamson GD, de Mouzon J, Chambers GM, et al. International Committee for Monitoring Assisted Reproductive Technology: world report on assisted reproductive technology, 2011. *Fertil Steril.* 2018;110:1067-1080. https://doi.org/10.1016/j.fertn stert.2018.06.039
- Seifer DB. Relevance of International Committee for Monitoring Assisted Reproductive Technology (ICMART) Registry report 2011. Fertil Steril. 2018;110:1032-1033. https://doi.org/10.1016/j. fertnstert.2018.07.011
- Soini S, Ibarreta D, Anastasiadou V, et al. The interface between assisted reproductive technologies and genetics: technical, social, ethical and legal issues. *Eur J Hum Genet*. 2006;14:588-645.
- 4. Kaariainen H. Assisted reproduction and genetics. *Eur J Hum Genet*. 2006;14:505.
- European Society of Human Genetics; European Society of Human Reproduction and Embryology. The need for interaction between assisted reproduction technology and genetics: recommendations of the European Societies of Human Genetics and Human Reproduction and Embryology. *Hum Reprod.* 2006;21:1971-1973.
- European Society of Human Genetics; European Society of Human Reproduction and Embryology. The need for interaction between assisted reproduction technology and genetics. Recommendations of the European Societies of Human Genetics and Human Reproduction and Embryology. *Eur J Hum Genet*. 2006;14:509-511.
- The Japanese Association of Medical Sciences. Guidelines for genetic tests and diagnoses in medical practice. 2011. http://jams. med.or.jp/guideline/genetics-diagnosis\_e.pdf. Accessed January 5, 2020.
- Hsu LYF. Prenatal diagnosis of chromosomal abnormalities thought amniocentesis. In: Milunsky A, Ed. *Genetic Disorders and the Fetus*, 4th ed. Baltimore: Johns Hopkins University Press; 1998. pp. 179-248.
- Bonduelle M, Van Assche E, Joris H, et al. Prenatal testing in ICSI pregnancies: incidence of chromosomal anomalies in 1586 karyotypes and relation to sperm parameters. *Hum Reprod*. 2002;17:2600-2614. https://doi.org/10.1093/humrep/17.10.2600
- Jacobs PA, Browne C, Gregson N, Joyce C, White H. Estimates of the frequency of chromosome abnormalities detectable in unselected newborns using moderate levels of banding. J Med Genet. 1992;29:103-108. https://doi.org/10.1136/jmg.29.2.103
- Riccaboni A, Lalatta F, Caliari I, Bonetti S, Somigliana E, Ragni G. Genetic screening in 2,710 infertile candidate couples for assisted reproductive techniques: results of application of Italian guidelines

for the appropriate use of genetic tests. *Fertil Steril*. 2008;89:800-808. https://doi.org/10.1016/j.fertnstert.2007.04.032

- Fu L, Xiong DK, Ding XP, et al. Genetic screening for chromosomal abnormalities and Y chromosome microdeletions in Chinese infertile men. J Assist Reprod Genet. 2012;29:521-527. https://doi. org/10.1007/s10815-012-9741-γ
- Pylyp LY, Spinenko LO, Verhoglyad NV, Zukin VD. Chromosomal abnormalities in patients with oligozoospermia and non-obstructive azoospermia. J Assist Reprod Genet. 2013;30:729-732. https:// doi.org/10.1007/s10815-013-9990-4
- Chiba K, Enatsu N, Fujisawa M. Management of non-obstructive azoospermia. *Reprod Med Biol.* 2016;15:165-173. https://doi. org/10.1007/s12522-016-0234-z
- Alberto F, Florina R, Valentina G, Daniela Z, Giandomenico P, Carlo F. Male infertility: role of genetic background. *Reprod Biomed Online*. 2007;14:734-745.
- Bor P, Hindkjaer J, Kølvraa S, Ingerslev HJ. Y-chromosome microdeletions and cytogenetic findings in unselected ICSI candidates at a Danish fertility clinic. J Assist Reprod Genet. 2002;19:224-231.
- 17. Dohle GR, Halley DJ, Van Hemel JO, et al. Genetic risk factors in infertile men with severe oligozoospermia and azoospermia. *Hum Reprod.* 2002;17:13-16.
- Gardner RJ, Sutherland GR. Chromosome Abnormalities and Genetic Counseling, 2nd ed. New York, NY: Oxford University Press; 1996:1478.
- Milunsky A, Milunsky JM. Genetic counceling; Preconception, prenatal and perinatal. In: Milunsky A, Milunsky JM, eds. *Genetic Disorders and the Fetus*, 6th edn. West Sussex, UK: Wiley-Blackwell; 2010:1-62.
- Wen J, Jiang J, Ding C, et al. Birth defects in children conceived by in vitro fertilization and intracytoplasmic sperm injection: a meta-analysis. *Fertil Steril.* 2012;97:1331-1337.e1-4. https://doi. org/10.1016/j.fertnstert.2012.02.053
- Rimm AA, Katayama AC, Katayama KP. A meta-analysis of the impact of IVF and ICSI on major malformations after adjusting for the effect of subfertility. J Assist Reprod Genet. 2011;28:699-705. https://doi.org/10.1007/s10815-011-9583-z
- 22. Farhi J, Fisch B. Risk of major congenital malformations associated with infertility and its treatment by extent of iatrogenic intervention. *Pediatr Endocrinol Rev.* 2007;4:352-357.
- Okun N, Sierra S, Douglas Wilson R, et al. Pregnancy outcomes after assisted human reproduction. J Obstet Gynaecol Can. 2014;36:64-83. https://doi.org/10.1016/S1701-2163(15)30685-X
- Fedder J, Loft A, Parner ET, Rasmussen S, Pinborg A. Neonatal outcome and congenital malformations in children born after ICSI with testicular or epididymal sperm: a controlled national cohort study. *Hum Reprod.* 2013;28:230-240. https://doi.org/10.1093/ humrep/des377
- 25. Sutcliffe AG, Saunders K, McLachlan R, et al. A retrospective 474 case-control study of developmental and other outcomes in a 475 cohort of Australian children conceived by intracytoplasmic sperm injection compared with a similar group in the United Kingdom. *Fertil Steril.* 2003;79:512-516.
- Imbar T, Tsafrir A, Lev-Sagie A, Hurwitz A, Laufer N, Holzer H. Assisted reproduction technologies and the risk of fetal, chromosomal and genetic malformations. *Harefuah*. 2006;145(3):223-228, 243-4.
- Qin JB, Sheng XQ, Wu D, et al. Worldwide prevalence of adverse pregnancy outcomes among singleton pregnancies after in vitro fertilization/intracytoplasmic sperm injection: a systematic review and meta-analysis. Arch Gynecol Obstet. 2017;295:285-301. https://doi.org/10.1007/s00404-016-4250-3
- Karami M, Jenabi E, Fereidooni B. The association of placenta previa and assisted reproductive techniques: a meta-analysis.

J Matern Fetal Neonatal Med. 2018;31:1940-1947. https://doi. org/10.1080/14767058.2017.1332035

- Gasparri ML, Nirgianakis K, Taghavi K, Papadia A, Mueller MD. Placenta previa and placental abruption after assisted reproductive technology in patients with endometriosis: a systematic review and meta-analysis. Arch Gynecol Obstet. 2018;298:27-34. https://doi.org/10.1007/s00404-018-4765-x
- Zhu L, Zhang Y, Liu Y, et al. Maternal and live-birth outcomes of pregnancies following assisted reproductive technology: a Retrospective Cohort Study. *Sci Rep.* 2016;6:35141. https://doi. org/10.1038/srep35141
- Nakashima A, Araki R, Tani H, et al. Implications of assisted reproductive technologies on term singleton birth weight: an analysis of 25,777 children in the national assisted reproduction registry of Japan. *Fertil Steril*. 2013;99:450-455. https://doi.org/10.1016/j. fertnstert.2012.09.027
- Wennerholm UB, Henningsen AK, Romundstad LB, et al. Perinatal outcomes of children born after frozen-thawed embryo transfer: a Nordic cohort study from the CoNARTaS group. *Hum Reprod.* 2013;28:2545-2553. https://doi.org/10.1093/humrep/det272
- 33. Imudia AN, Awonuga AO, Doyle JO, et al. Peak serum estradiol level during controlled ovarian hyperstimulation is associated with increased risk of small for gestational age and preeclampsia in singleton pregnancies after in vitro fertilization. *Fertil Steril.* 2012;97:1374-1379.
- Pereira N, Elias RT, Christos PJ, et al. Supraphysiologic estradiol is an independent predictor of low birth weight in full-term singletons born after fresh embryo transfer. *Hum Reprod.* 2017;32:1410-1417. https://doi.org/10.1093/humrep/dex095
- Luke B. Pregnancy and birth outcomes in couples with infertility with and without assisted reproductive technology: with an emphasis on US population-based studies. *Am J Obstet Gynecol.* 2017;217:270-281. https://doi.org/10.1016/j.ajog.2017.03.012
- Roque M, Haahr T, Geber S, Esteves SC, Humaidan P. Fresh versus elective frozen embryo transfer in IVF/ICSI cycles: a systematic review and meta-analysis of reproductive outcomes. *Hum Reprod Update*. 2019;25:2-14. https://doi.org/10.1093/humupd/dmy033
- 37. Saito K, Kuwahara A, Ishikawa T, et al. Endometrial preparation methods for frozen-thawed embryo transfer are associated with altered risks of hypertensive disorders of pregnancy, placenta accreta, and gestational diabetes mellitus. *Hum Reprod.* 2019;34:1567-1575. https://doi.org/10.1093/humrep/dez079
- von Versen-Höynck F, Schaub AM, Chi YY, et al. Increased preeclampsia risk and reduced aortic compliance with in vitro fertilization cycles in the absence of a corpus luteum. *Hypertension*. 2019;73:640-649. https://doi.org/10.1161/HYPERTENSIONAHA.118.12043
- Rombauts L, Motteram C, Berkowitz E, Fernando S. Risk of placenta praevia is linked to endometrial thickness in a retrospective cohort study of 4537 singleton assisted reproduction technology births. *Hum Reprod.* 2014;29:2787-2793. https://doi.org/10.1093/ humrep/deu240
- Qin J, Liu X, Sheng X, Wang H, Gao S. Assisted reproductive technology and the risk of pregnancy-related complications and adverse pregnancy outcomes in singleton pregnancies: a meta-analysis of cohort studies. *Fertil Steril.* 2016;105:73-85.e1-6. https://doi.org/10.1016/j.fertnstert.2015.09.007
- Higurashi M, Iijima K, Ishikawa N, Hoshina H, Watanabe N. Incidence of major chromosome aberrations in 12,319 newborn infants in Tokyo. *Hum Genet*. 1979;46:163-172.
- Groth KA, Skakkebæk A, Høst C, Gravholt CH, Bojesen A. Clinical review: Klinefelter syndrome-a clinical update. J Clin Endocrinol Metab. 2013;98:20-30. https://doi.org/10.1210/jc.2012-2382
- Chang S, Skakkebæk A, Gravholt CH. Klinefelter Syndrome and medical treatment: hypogonadism and beyond. *Hormones (Athens)*. 2015;14:531-548. https://doi.org/10.14310/horm.2002.1622

Reproductive Medicine and Biology

- 44. Crawford D, Dearmun A. Klinefelter syndrome. Nurs Child Young People. 2017;29:19. https://doi.org/10.7748/ncyp.29.6.19.s21
- Bonomi M, Rochira V, Pasquali D, Balercia G, Jannini EA, Ferlin A. Klinefelter Italian Group (KING). Klinefelter syndrome (KS): genetics, clinical phenotype and hypogonadism. *J Endocrinol Invest*. 2017;40:123-134. https://doi.org/10.1007/s40618-016-0541-6
- Maiburg MC, Hoppenbrouwers AC, van Stel HF, Giltay JC. Attitudes of Klinefelter men and their relatives toward TESE-ICSI. J Assist Reprod Genet. 2011;28:809-8014.
- Palermo GD, Schlegel PN, Sills ES, et al. Births after intracytoplasmic injection of sperm obtained by testicular extraction from men with nonmosaic Klinefelter's syndrome. N Engl J Med. 1998;338:588-590.
- Miki T, Nagayoshi M, Takemoto Y, et al. Genetic risk of Klinefelter's syndrome in assisted reproductive technology. *Reprod Med Biol.* 2017;16:188-195. https://doi.org/10.1002/rmb2.12029
- Corona G, Pizzocaro A, Lanfranco F, et al. Sperm recovery and ICSI outcomes in Klinefelter syndrome: a systematic review and meta-analysis. *Hum Reprod Update*. 2017;23:265-275.
- Majzoub A, Arafa M, Al Said S, et al. Outcome of testicular sperm extraction in nonmosaic Klinefelter syndrome patients: what is the best approach? *Andrologia*. 2016;48:171-176.
- Shiraishi K, Matsuyama H. Klinefelter syndrome: from pediatrics to geriatrics. *Reprod Med Biol.* 2018;18:140-150. https://doi. org/10.1002/rmb2.12261
- Okada H, Goda K, Yamamoto Y, et al. Age as a limiting factor for successful sperm retrieval in patients with nonmosaic Klinefelter's syndrome. *Fertil Steril.* 2005;84:1662-1664.
- Borjian Boroujeni P, Sabbaghian M, Vosough Dizaji A, et al. Clinical aspects of infertile 47, XYY patients: a retrospective study. *Hum Fertil* (*Camb*). 2019;22:88-93. https://doi.org/10.1080/14647 273.2017.1353143
- 54. Kim IW, Khadilkar AC, Ko EY, Sabanegh ES Jr. 47, XYY Syndrome and male infertility. *Rev Urol.* 2013;15:188-196.
- Bouet PE, Godbout A, El Hachem H, et al. Fertility and pregnancy in Turner syndrome. J Obstet Gynaecol Can. 2016;38:712-718. https://doi.org/10.1016/j.jogc.2016.02.007
- Morgan T. Turner syndrome: diagnosis and management. Am Fam Physician. 2007;76:405-410.
- Hovatta O. Pregnancies in women with Turner's syndrome. Ann Med. 1999;31:106-110. https://doi.org/10.3109/0785389990 8998785
- Obata S, Tsuburai T, Shindo R, Aoki S, Miyagi E, Sakakibara H. Comprehensive medical treatment of women with Turner syndrome may improve pregnancy outcomes: a case report. *Clin Pediatr Endocrinol.* 2019;28:37-41. https://doi.org/10.1297/ cpe.28.37
- 59. Otter M, Schrander-Stumpel CT, Curfs LM. Triple X syndrome: a review of the literature. *Eur J Hum Genet*. 2010;18:265-271.
- 60. Tartaglia NR, Howell S, Sutherland A, Wilson R, Wilson L. A review of trisomy X (47, XXX). Orphanet J Race Dis. 2010;5:1-9.
- Sugawara N, Maeda M, Manome T, Nagai R, Araki Y. Patients with 47, XXX karyotype who experienced premature ovarian failure (POF): two case reports. *Reprod Med Biol.* 2013;12:193-195. https://doi.org/10.1007/s12522-013-0158-9
- Cohen O, Cans C, Mermet MA, Demongeot J, Jalbert P. Viability thresholds for partial trisomies and monosomies. A study of 1,159 viable unbalanced reciprocal translocations. *Hum Genet*. 1994;93:188-194.
- Nussbaum RL, McInnes RR. Thompson & Thompson Genetics in Medicine, 8th edn. Amsterdam, the Netherlands: Elsevier; 2017.
- 64. Babariya D, Fragouli E, Alfarawati S, Spath K, Wells D. The incidence and origin of segmental aneuploidy in human oocytes and preimplantation embryos. *Hum Reprod*. 2017;32:2549-2560. https://doi.org/10.1093/humrep/dex324

- 65. Therman E, Susman B, Denniston C. The nonrandom participation of human acrocentric chromosomes in Robertsonian translocations. *Ann Hum Genet*. 1989;53:49-65.
- Poongothai J, Gopenath TS, Manonayaki S. Genetics of human male infertility. *Singapore Med J.* 2009;50:336-347.
- 67. Brown GM, Furlong RA, Sargent CA, et al. Characterisation of the coding sequence and fine mapping of the human DFFRY gene and comparative expression analysis and mapping to the Sxrb interval of the mouse Y chromosome of the Dffry gene. *Hum Mol Genet*. 1998;7:97-107.
- Sun C, Skaletsky H, Birren B, et al. An azoospermic man with a de novo point mutation in the Y-chromosomal gene USP9Y. *Nat Genet*. 1999;23:429-432.
- Kuroda-Kawaguchi T, Skaletsky H, Brown LG, et al. The AZFc region of the Y chromosome features massive palindromes and uniform recurrent deletions in infertile men. *Nat Genet*. 2001;29:279-286.
- Tilford CA, Kuroda-Kawaguchi T, Skaletsky H, et al. A physical map of the human Y chromosome. *Nature*. 2001;409:943-945.
- 71. Repping S, Skaletsky H, Lange J, et al. Recombination between palindromes P5 and P1 on the human Y chromosome causes massive deletions and spermatogenic failure. *Am J Hum Genet*. 2002;71:906-922.
- Skaletsky H, Kuroda-Kawaguchi T, Minx PJ, et al. The male-specific region of the human Y chromosome is a mosaic of discrete sequence classes. *Nature*. 2003;423:825-837.
- Krausz C, Casamonti E. Spermatogenic failure and the Y chromosome. Hum Genet. 2017;136:637-655. https://doi.org/10.1007/ s00439-017-1793-8
- 74. Krausz C, Hoefsloot L, Simoni M, Tüttelmann F, European Academy of Andrology; European Molecular Genetics Quality Network. EAA/EMQN best practice guidelines for molecular diagnosis of Y-chromosomal microdeletions: state-of-the-art 2013. Andrology. 2014;2:5-19. https://doi.org/10.1111/j.2047-2927.2013.00173.x
- Gallego A, Rogel R, Luján S, Plaza B, Delgado F, Boronat F. AZF gene microdeletions: case series and literature review. Actas Urol Esp. 2014;38:698-702.
- Vogt PH, Edelmann A, Kirsch S, et al. Human Y chromosome azoospermia factors (AZF) mapped to different subregions in Yq11. *Hum Mol Genet*. 1996;5:933-943.
- Zhang YS, Dai RL, Wang RX, Zhang HG, Chen S, Liu RZ. Analysis of Y chromosome microdeletion in 1738 infertile men from northeastern China. *Urology*. 2013;82:584-588.
- Suganthi R, Vijesh VV, Vandana N, Benazir FA. Y choromosomal microdeletion screening in the workup of male infertility and its current status in India. Int. J Fertil Steril. 2014;7:253-266.
- Zhang F, Li L, Wang L, et al. Clinical characteristics and treatment of azoospermia and severe oligospermia patients with Y-chromosome microdeletions. *Mol Reprod Dev.* 2013;80:908-915.
- Lange J, Skaletsky H, van Daalen SK, et al. Isodicentric Y chromosomes and sex disorders as byproducts of homologous recombination that maintains palindromes. *Cell.* 1999;138:855-869.
- 81. Yen P. The fragility of fertility. Nat Genet. 2001;29:243-244.
- Repping S, Skaletsky H, Brown L, et al. Polymorphism for a 1.6-Mb deletion of the human Y chromosome persists through balance between recurrent mutation and haploid selection. *Nat Genet*. 2003;35:247-251.
- 83. Krausz C, Forti G, McElreavey K. The Y chromosome and male fertility and infertility. *Int J Androl*. 2003;26(2):70-75.
- 84. Fukushima M, Koh E, Choi J, Maeda Y, Namiki M, Yoshida A. Reevaluation of azoospermic factor c microdeletions using sequence-tagged site markers with confirmed physical positions from the GenBank database. *Fertil Steril.* 2006;85(4):965-971.
- Sin HS, Koh E, Shigehara K, et al. Features of constitutive gr/gr deletion in a Japanese population. *Hum Reprod.* 2010;25:2396-2403. https://doi.org/10.1093/humrep/deq191

142

- Palermo G, Joris H, Devroey P, Van Steirteghem AC. Pregnancies after intracytoplasmic injection of single spermatozoon into an oocyte. *Lancet*. 1992;340:17-18.
- van Steirteghem A. Twenty years of in vitro fertilization: realization and questions for the future. Verh K Acad Geneeskd Belg. 2001;63:193-240; discussion 240–1.
- Jozwiak EA, Ulug U, Mesut A, Erden HF, Bahqeci M. Prenatal karyotypes of fetuses conceived by intracytoplasmic sperm injection. *Fertil Steril.* 2004;82:628-633.
- Gjerris AC, Loft A, Pinborg A, Christiansen M, Tabor A. Prenatal testing among women pregnant after assisted reproductive techniques in Denmark 1995-2000: a National Cohort Study. *Hum Reprod.* 2008;23:1545-1552.
- Lacamara C, Ortega C, Villa S, Pommer R, Schwarze JE. Are children born from singleton pregnancies conceived by ICSI at increased risk for congenital malformations when compared to children conceived naturally? A systematic review and meta-analysis. JBRA Assist Reprod. 2017;21:251-259. https://doi. org/10.5935/1518-0557.20170047
- Zeng H, Liu N, Fan X, Cai M, Xie H. Risk factors for early miscarriage among intrauterine singleton pregnancies after treatment with in vitro fertilization/intracytoplasmic sperm injection. *Zhong Nan Da Xue Xue Bao Yi Xue Ban*. 2016;41:815-820. https://doi. org/10.11817/j.issn.1672-7347.2016.08.007
- Xi D, Chen Y, Dai YT. Sperm DNA fragmentation index and the success rate of IVF/ICSI. Zhonghua Nan Ke Xue. 2016;22:77-81.
- Tachibana M. Epigenetics of sex determination in mammals. *Reprod* Med Biol. 2016;15:59-67.
- Morgan HD, Santos F, Green K, Dean W, Reik W. Epigenetic reprogramming in mammals. *Hum Mol Genet*. 2005;14:R47-58.
- Dopont C, Sifer A. A review of outcome data concerning children born following assisted reproductive technologies. *ISRN Obstet Gyecol.* 2012;2012:1-5.
- Hattori H, Hiura H, Kitamura A, et al. Association of four imprinting disorders and ART. *Clin Epigenetics*. 2019;11:21. https://doi. org/10.1186/s13148-019-0623-3
- Lubinsky M. An epigenetic association of malformations, adverse reproductive outcomes, and fetal origins hypothesis related effects. J Assist Reprod Genet. 2018;35:953-964. https://doi.org/10.1007/s10815-018-1197-2
- Litzky JF, Boulet SL, Esfandiari N, et al. Birthweight in infants conceived through in vitro fertilization following blastocyst or cleavage-stage embryo transfer: a national registry study. J Assist Reprod Genet. 2018;35:1027-1037. https://doi.org/10.1007/s1081 5-018-1168-7
- Anav M, Ferrières-Hoa A, Gala A, et al. Birth weight and frozen embryo transfer: state of the art. *Gynecol Obstet Fertil Senol*. 2018;46:489-496. https://doi.org/10.1016/j.gofs.2018.03.012
- Cortessis VK, Azadian M, Buxbaum J, et al. Comprehensive meta-analysis reveals association between multiple imprinting disorders and conception by assisted reproductive technology. J Assist Reprod Genet. 2018;35:943-952. https://doi.org/10.1007/s1081 5-018-1173-x

- Cox GF, Bürger J, Lip V, et al. Intracytoplasmic sperm injection may increase the risk of imprinting defects. Am J Hum Genet. 2002;71:162-164.
- 103. Allen C, Reardon W. Assisted reproduction technology and defects of genomic imprinting. *BJOG*. 2005;112:1589-1594.
- 104. Shiota K, Yamada S. Assisted reproductive technologies and birth defects. *Congenit Anom (Kyoto)*. 2005;45:39-43.
- Hartmann S, Bergmann M, Bohle RM, Weidner W, Steger K. Genetic imprinting during impaired spermatogenesis. *Mol Hum Reprod.* 2006;12:407-411.
- Sato A, Otsu E, Negishi H, Utsunomiya T, Arima T. Aberrant DNA methylation of imprinted loci in superovulated oocytes. *Hum Reprod.* 2007;22:26-35.
- Reik W, Dean W, Walter J. Epigenetic reprogramming in mammalian development. *Science*. 2001;293:1089-1093.
- Zheng H-Y, Shi X-Y, Wang L-L, Wu Y-Q, Chen S-L, Zhang L. Study of DNA methylation patterns of imprinted genes in children born after assisted reproductive technologies reveals no imprinting errors: a pilot study. *Exp Ther Med.* 2011;2:751-755.
- Ludwig M, Katalinic A, Gross S, Sutcliffe A, Varon R, Horsthemke B. Increased prevalence of imprinting defects in patients with Angelman syndrome born to subfertile couples. J Med Genet. 2005;42:289-291.
- Chamani IJ, Keefe DL. Epigenetics and female reproductive aging. Front Endocrinol (Lausanne). 2019;10:473. https://doi.org/10.3389/ fendo.2019.00473
- Kobayashi N, Miyauchi N, Tatsuta N, et al. Factors associated with aberrant imprint methylation and oligozoospermia. *Sci Rep.* 2017;10(7):42336. https://doi.org/10.1038/srep42336
- 112. Novakovic B, Lewis S, Halliday J, et al. Assisted reproductive technologies are associated with limited epigenetic variation at birth that largely resolves by adulthood. *Nat Commun.* 2019;10(1):3922. https://doi.org/10.1038/s41467-019-11929-9. PMID: 31477727
- 113. Wilkins-haug L. Assisted reproductive technology, congenital malformations, and epigenetic disease. *Clin Obstet Gynecol.* 2008;51(1):96-105. https://doi.org/10.1097/GRF.0b013e3181 61d25a
- Lidegaard Ø, Pinborg A, Andersen AN. Imprinting disorders after assisted reproductive technologies. *Curr Opin Obstet Gynecol.* 2006;18(3):293-296.
- 115. Katagiri Y, Shibui Y, Nagao K, Miura K, Morita M. Epigenetics in assisted reproductive technology. *Reprod Med Biol.* 2007;6(2):69-75. https://doi.org/10.1111/j.1447-0578.2007.00168.x
- Sueoka K. Preimplantation genetic diagnosis: an update on current technologies and ethical consideration. *Reprod Med Biol.* 2015;15:69-75.

How to cite this article: Katagiri Y, Tamaki Y. Genetic counseling prior to assisted reproductive technology. *Reprod Med Biol*. 2021;20:133–143. <u>https://doi.org/10.1002/</u> rmb2.12361