



Short communication

A Transfer of Silver Nanoparticles from Pregnant Rat to Offspring

Yeonjin Lee¹, Jonghye Choi¹, Pilje Kim², Kyunghee Choi², Suhyon Kim³, Woochan Shon⁴ and Kwangsik Park¹

¹College of Pharmacy, Dongduk Women's University, Seoul 136-714

²National Institute of Environmental Research, Incheon 404-708

³Korea Testing and Research Institute, Gimpo 415-871

⁴University of Ulsan College of Medicine, Seoul 680-749, Korea

(Received July 2, 2012; Revised September 1, 2012; Accepted September 4, 2012)

Silver nanoparticles (size: 7.9 ± 0.95 nm, dosage: 250 mg/kg) were orally administered to pregnant rats. At 4 days after parturition, four pups were randomly selected (one pup from one dam) and silver level in liver, kidney, lung and brain was determined by ICP-MS and electron microscope. As results, silver nanoparticles highly accumulated in the tissues of the pups. Silver level in the treated group was 132.4 ± 43.9 ng/g in the kidney (12.3 fold compared to control group), 37.3 ± 11.3 ng/g in the liver (7.9 fold), 42.0 ± 8.6 ng/g in the lung (5.9 fold), and 31.1 ± 4.3 ng/g in the brain (5.4 fold). This result suggested that the possible transfer of silver nanoparticles from pregnant dams to the fetus through mainly placenta.

Key words: Silver nanoparticles, Pregnant rats, Placenta transfer, Accumulation in pups

With a rapid increase of nanoparticle applications, there is an urgent need for toxicologists to study nanotoxicity on the risk assessment of nanoproducts. Many consumer products of silver nanoparticles (AgNPs) including medicines, water disinfectants, room sprays and fabrics may release nanoparticles that can be exposed to the human body (Benn *et al.*, 2010). The exposed AgNPs can be absorbed into blood circulation, and accumulate in tissues including the liver, kidney, spleen and other organs (Kim *et al.*, 2008; Tang *et al.*, 2009). Recently, serum kinetics and organ distribution of AgNPs by oral administration or intravenous injection have been studied. Bioavailability was reported to be about 4% in 10 mg/kg dosage (Park *et al.*, 2011). Although the bioavailability of AgNPs through oral administration is very low, long term treatments may allow accumulation of nanoparticles in body organs. Furthermore, AgNPs administered to pregnant animals were found to be transferred to the fetus (Kulvietis *et al.*, 2011). However, data on the possible transfer to fetus through the placenta barrier or other possible pathways are still limited. In this study, transfer of AgNPs from pregnant female rats to offspring was investigated.

The AgNPs coated with citrate were kindly provided by

ABC NANOTECH (Daejeon, Korea). The size of the AgNPs was estimated to be 7.9 ± 0.95 nm based on the TEM image and the mean particle surface area was found to be 7.53×10^2 nm²/particle; whereas the mean particle mass and mean particle volume were found to be 2×10^{-17} g and 1.9×10^3 nm³, respectively. Sprague Dawley (SD) female rats ($n > 4$) were treated by oral administration of AgNPs (250 mg/kg) for 14 days before mating, during the mating period and gestation period, and finally 4 days after parturition. SD male rats ($n > 4$) were also treated with AgNPs (250 mg/kg) for 14 days before mating and during the mating period. The doses and treatment regimen was based on the OECD Test Guideline 422: Combined Repeated Dose Toxicity Study with the Reproduction/Developmental Toxicity Screening Tests, which recommends three dose-groups. In this study, the highest dosage of 250 mg/kg was selected among the three dosages (250, 125, and 62.5 mg/kg). Mating was confirmed by the microscopic finding of sperm in the vaginal exudate. Data regarding the reproductive toxicity test were rule out in this study. Offspring were sacrificed 4 days after birth, and the organs of pups including the liver, kidney, lung and brain were obtained (total 4 offsprings, one offspring was chosen from one pregnant rat, respectively). All the animals used in this study were taken care of in accordance with the principles outlined in the "Guide for the Care and Use of Laboratory Animals" issued by the Animal Care and Committee of NVRQS (National Veterinary Research and Quarantine Service).

Correspondence to: Kwangsik Park, College of Pharmacy, Dongduk Women's University, #23-1, Wolgok-dong, Seongbuk-gu, Seoul 136-714, Korea
E-mail: kspark@dongduk.ac.kr

For the analysis of total silver, the samples (approximately 200 mg of each tissue) were digested in a mixed solution of 70% HNO₃ (7 ml) and 30% H₂O₂ (1 ml) using the microwave digestion system (Milestone, Sorisole, Italy) under conditions of high temperature and pressure. After dilution of the acidic digested preparation with deionized water (1 : 11), the concentration of silver was analyzed using the ICP-MS (Elan6100/Perkin Elmer, USA) at Korean Basic Science Institute (KBSI, Seoul, Korea). Liver tissue was fixed with glutaraldehyde and osmium tetroxide, and observed with a transmission electron microscope (JEOL, JEM01200 EX II). Student *t*-test was performed for the statistical analysis.

Pups were randomly selected from non-treated control dams and treated dams, respectively and tissue levels of AgNPs in pups (*n* = 4, one pup from one dam) was determined in liver, kidney, lung and brain. As shown in Table 1, accumulation of silver was observed in the tissues of respective organs in the offspring from the treated dams. The kidney seemed to be the main target for the accumulation of AgNPs and exhibited higher accumulation levels compared to the other tissues. Silver levels were 132.4 ± 43.9 ng/g in the kidney (12.3 fold compared to control group) and the levels were 37.3 ± 11.3 ng/g in the liver (7.9 fold), 42.0 ± 8.6 ng/g in the lung (5.9 fold), and 31.1 ± 4.3 ng/g in the brain (5.4 fold). The accumulated AgNPs were observed in the liver of pups by a transmission electron microscope (Fig. 1). This result suggested a possible transfer of AgNPs from pregnant dams to offspring mainly through the placenta or minor pathway of milk.

In previous publications (Takeda *et al.*, 2009; Menezes *et*

Table 1. Silver concentration in the organs of offspring (*n* = 4) sacrificed at 4 days after birth from dams treated with AgNPs (250 mg/kg) by oral administration of silver nanoparticles (unit : ng/g)

	Liver	Kidney	Lung	Brain
Control	4.7 ± 0.59	10.8 ± 1.4	7.1 ± 2.1	5.8 ± 2.5
AgNPs	37.3 ± 11.3*	132.4 ± 43.9*	42.0 ± 8.6*	31.1 ± 4.3*

*, Statistically significant compared to control value (*P* < 0.01).

al., 2011), titanium dioxide nanoparticles (25~70 nm) were administrated subcutaneously to pregnant mice at 3, 7, 10 and 14 days post-coitum and the nanoparticles were detected in the testes and brain of the offspring 6 weeks after birth. The authors suggested that nanoparticles may be delivered by a placental passage. They also hypothesized that the nanoparticles reached the fetal brain because they were administered before the full development of the blood-brain barrier. In our study, AgNPs (7.9 ± 0.95 nm) were also identified in the brain of offspring, which means that AgNPs may reach the brain before the blood-brain barrier is formed in the fetus or they may directly pass the barrier. The AgNPs used in this study are capped with hydrophilic citrate which is hydrophilic but it seems that they have some kind of mechanism to pass the cellular lipid membrane and other barriers. Further studies are needed to investigate the detailed mechanisms of the transport system of AgNPs.

REFERENCES

Benn, T., Cavanagh, B., Hristovski, K., Posner, J.D. and Wester-

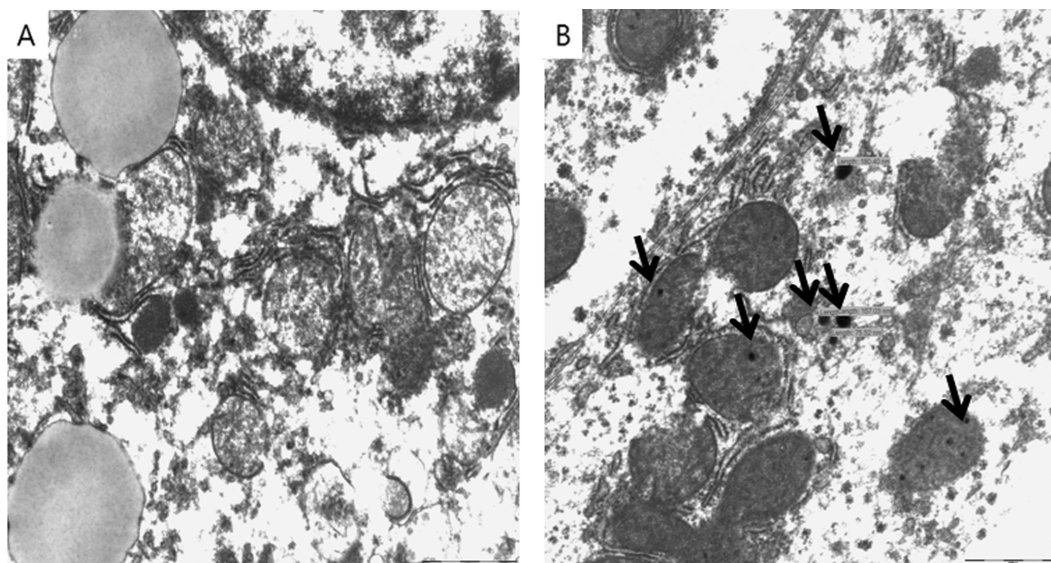


Fig. 1. Localization of AgNPs in the liver of offspring from pregnant dam treated with AgNPs by oral administration (250 mg/kg). Panel A and B are representative TEM images of 80 nm liver sections of offspring from the non-treated control group and the treated group, respectively. Representative AgNPs visible in the hepatocytes as black electron-dense spots were indicated by arrows.

- hoff, P. (2010). The release of nanosilver from consumer products used in the home. *J. Environ. Qual.*, **39**, 1875-1882.
- Kim, Y.S., Kim, J.S., Cho, H.S., Rha, D.S., Kim, J.M., Park, J.D., Choi, B.S., Lim, R., Chang, H.K., Chung, Y.H., Kwon, I.H., Jeong, J., Han, B.S. and Yu, I.J. (2008). Twenty-eight-day oral toxicity, genotoxicity, and gender-related tissue distribution of silver nanoparticles in Sprague-Dawley rats. *Inhal. Toxicol.*, **20**, 575-583.
- Kulvietis, V., Zalgevičienė, V., Didziapetrienė, J. and Rotomskis, R. (2011). Transport of nanoparticles through the placental barrier. *Tohoku J. Exp. Med.*, **225**, 225-234.
- Menezes, V., Malek, A. and Keelan, J.A. (2011). Nanoparticulate drug delivery in pregnancy: placental passage and fetal exposure. *Curr. Pharm. Biotechnol.*, **12**, 731-742.
- Park, K., Park, E.J., Chun, I.K., Choi, K., Lee, S.H., Yoon, J. and Lee, B.C. (2011). Bioavailability and Toxicokinetics of Citrate-coated Silver Nanoparticles in Rats. *Arch. Pharm. Res.*, **34**, 153-158.
- Takeda, K., Suzuki, K.I., Ishihara, A., Kubo-Irie, M., Fujimoto, R., Tabata, M., Oshio, S., Nihei, Y., Ihara, T. and Sugamata, M. (2009). Nanoparticles transferred from pregnant mice to their offspring can damage the genital and cranial nerve systems. *J. Health Sci.*, **55**, 95-102.
- Tang, J., Xiong, L., Wang, S., Wang, J., Liu, L., Li, J., Yuan, F. and Xi, T. (2009). Distribution, translocation and accumulation of silver nanoparticles in rats. *J. Nanosci. Nanotechnol.*, **9**, 4924-4932.