

Advances in mechanism and management of bone homeostasis in osteonecrosis: a review article from basic to clinical applications

Xiao-Na Xiang, MSc^{a,b,c}, Hong-Chen He, PhD^{a,b,c}, Cheng-Qi He, PhD^{a,b,c,*}

Abstract

Osteonecrosis, characterized by bone cell death leading to impaired bone recovery, causes challenges in bone homeostasis maintenance. Bone homeostasis relies on the delicate balance between osteoclasts and osteoblasts, encompassing a series of complex and strictly regulated biological functions. Current treatments, including conservative therapies and surgeries, often fall short of expected outcomes, necessitating a reorientation towards more effective therapeutic strategies according to the pathogenesis. In this review, the authors hierarchically outlined risk factors, emerging mechanisms, and last-decade treatment approaches in osteonecrosis. By connecting mechanisms of bone homeostasis, the authors proposed future research directions should be focused on elucidating risk factors and key molecules, performing high-quality clinical trial, updating practice, and accelerating translational potential.

Keywords: bone homeostasis, osteonecrosis, pathogenesis, therapy

Introduction

Osteonecrosis, commonly referred to as avascular necrosis, atraumatic necrosis, aseptic necrosis, or ischemic necrosis, can result from trauma and has been regarded as a complication associated with autoimmune diseases^[1]. The femoral head and jaw are the most affected parts during osteonecrosis, as a pathologic process of bone cell death partly due to an interruption of blood flow, limiting the delivery of oxygen and nutrients^[2]. The mean age of presentation of osteonecrosis of the femoral head (ONFH) in the UK is 58.3 years, with a prevalence of 2 per 100 000 patients^[3]. The incidence of osteonecrosis of the jaws ranges from 0.2 per $100\ 000^{[4]}$ to $41.7\%^{[5]}$, depending on the received medication and primary diseases^[6]. Some causative conditions have been reported, such as medication use, alcohol intake, hypercoagulation, bone marrow fat embolisms, high intraosseous pressure, and vascular endothelial dysfunction^[7,8].

Copyright © 2024 The Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the terms of the [Creative Commons](http://creativecommons.org/licenses/by-nc-nd/4.0/) [Attribution-Non Commercial-No Derivatives License 4.0](http://creativecommons.org/licenses/by-nc-nd/4.0/) (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Published online 23 September 2024

HIGHLIGHTS

- Osteonecrosis is related to the various regulatory mechanisms of bone homeostasis.
- Pregnancy, immune diseases (such as SLE, ALL, SCD), trauma, alcohol intake, glucocorticoid use, and bisphosphonates use are risks for osteonecrosis.
- Current pathways (such as PI3K/AKT, WNT/β-catenin, RANK-RANKL, IFN-β, JAK/STAT3, etc.) to the mechanisms and current treatments are logically connected and summarized hierarchically.
- Future research should be directed at identifying risk factors and key molecules, performing high-quality clinical trial, updating practice, and accelerating translational potential.

The most affected organ of this disease is bone, which ultimately results in structural changes, potential collapse, and destruction[9,10]. Bone undergoes a continuous cycle of formation by osteoblasts and resorption by osteoclasts, resulting in a dynamically changing tissue, even in the face of challenging conditions such as metabolic^[11], immunological^[12], or degenerative diseases^[13]. Recent studies^[14,15] have focused on the mechanisms that couple the relation between bone resorption and formation. Osteoblasts and osteoclasts, the primary cellular components of bone, delicately balance bone homeostasis. Additionally, osteocytes are ancient bone cells, and mechanosensing stellate cells that possess a remarkable ability to detect and respond to mechanical stimuli^[16]. Osteocytes serve as crucial regulators of bone homeostasis by modulating osteoblast and osteoclast via the production of key signaling molecules in the WNT signaling and receptor activator of nuclear factor-κB ligand (RANKL) pathways^[17]. These cells not only communicate among themselves^[18], but also interact with other cells within the bone marrow, such as T and B cells, macrophages, adipocytes,

^aRehabilitation Medicine Center and Institute of Rehabilitation Medicine, West China Hospital, Sichuan University, ^bSchool of Rehabilitation Sciences, West China School of Medicine, Sichuan University and ^cKey Laboratory of Rehabilitation Medicine in Sichuan Province, Chengdu, P. R. China

Sponsorships or competing interests that may be relevant to content are disclosed at the end of this article.

^{*}Corresponding author. Address: Department of Rehabilitation Medicine and Institute of Rehabilitation Medicine, West China Hospital, Sichuan University, #37 Guoxue Alley, Wuhou strict, Chengdu, Sichuan 610041, PR China. Tel.: +86 (028) 854 238 19. E-mail: hxkfhcq2015@126.com (C-Q. He).

International Journal of Surgery (2025) 111:1101–1122

Received 4 March 2024; Accepted 15 September 2024

http://dx.doi.org/10.1097/JS9.0000000000002094

and hematopoietic pro-genitors $[19,20]$. Moreover, the processes of osteogenesis and angiogenesis are intricately connected, each relying on the other for proper function. Osteogenic cells secrete angiogenic factors to stimulate vessel growth and regulate functions, whereas endothelial cells secrete angiocrine signals that control bone remodeling^[21].

Some guidelines and comments^[22–24] about the prevention and treatment of ONFH and medication-related osteonecrosis of the jaw (MRONJ), especially by the Association Research Circulation Osseous $(ARCO)^{[22]}$, and the Multinational Association of Supportive Care in Cancer/International Society of Oral Oncology $(MASCCISOO)^{[23]}$ have published. The guidelines reported some strategies for therapy, and the clinical decision principally is made according to the disease stage and age. Among these, surgeries are important treatments for managing osteonecrosis, especially core decompression (CD). However, a meta-analysis study reported that the efficacy and effectiveness of CD alone for ONFH are no better than other joint-preserving strategies^[25]. A clear summary with a classification of current therapies is necessary. Especially, with the development of regenerative medicine, surgeries with regenerative strategies should be comprehensively presented.

In general, the pathogenesis of osteonecrosis is complex and poorly understood. Moreover, the risk factors related to osteonecrosis should be presented to avoid. With the advancement of medical technology, more available therapies have emerged for use. Some novel technologies are still in the animal research stage, while others have already undergone clinical trials. In this review, we systematically searched and summarized recent evidence about the risk factors, pathogenesis of bone homeostasis, and treatment strategies in individuals with osteonecrosis. Moreover, we discussed the potential therapies that may be used for osteonecrosis in the future.

Methods

We searched Web of Science, PubMed, and Scopus Database from 2013 to December 2023, using the Mesh terms, keywords, and combinations, such as risk factor and (osteonecrosis or ONJ or ONFH), (pathology or molecular mechanism) and (osteonecrosis or ONJ or ONFH) and (bone homeostasis or osteoblast or osteoclast or MSC), (treatment or therapy or surgery or biomaterials) and (osteonecrosis or ONJ or ONFH) and clinical trials. The reference lists of the studies selected for full-text screening were manually searched to find potentially relevant articles. We included basic and experimental studies (in vivo or in vitro) that determined the molecular pathology. We also included the clinical studies that reported the risk factors of osteonecrosis and the current treatments for osteonecrosis. Comments, conference abstracts, reviews, guidelines, and letters to editors were not included in the review. Duplicate articles were eliminated, followed by a preliminary review based on titles and abstracts. Experimental studies about the management of osteonecrosis were excluded from the systematic review. The remaining relevant literature was incorporated based on the selection criteria outlined above after reviewing the full text.

Results

The flowchart demonstrates the study selection process in our study (Fig. 1). A total of 171 studies were included for describing the risk factors (42/171), basic studies that reported the potential bone homeostasis mechanisms of the disease (66/171), while others reflected the current therapy strategies for osteonecrosis (63/171).

Risk factors

Some diseases, such as immune-mediated diseases^[26–37] and trauma[38–41] are known as usual risk factors. Moreover, the abusive consumption of alcohol^[42–44] and prolonged use of steroids^[45–50] have been recognized as promoting factors. Besides this, there are some conditions reported recently and the etiology of these risk factors and osteonecrosis need to be summarized. In this section, we focus on the relationships between the risk factors and osteonecrosis, and the risk factors and their associated incidence, prevalence, or relative risks are presented in [Table 1.](#page-2-0)

Related primary diseases or history

The risk of osteonecrosis is highly associated with systemic lupus erythematosus (SLE), acute lymphoblastic leukemia $(ALL)^{[26]}$, sickle cell disease (SCD), and human immunodeficiency virus $(HIV)^{[27,28]}$, that requires dosage of steroids^[29]. SLE is a chronic autoimmune systemic disease that requires high-dose initial corticosteroid treatment with a reported 26.9% of symptomatic osteonecrosis^[30]. The incidence of osteonecrosis during antileukemic treatment has been investigated in many studies, but results range from 1.6 to $25\%^{[31]}$. For ALL, hypertension is associated with an increased risk for symptomatic osteonecrosis[32]. Additionally, among children with ALL, increased risks were reported when receiving higher post-transplant steroid dosages and being older than ten years old at the time of transplantation^[33–35]. SCD is one of the most common hereditary blood disorders and the prevalence of ONFH ranges from 3 to 50% among individuals with SCD, which is associated

Table 1 Summary table of risk factors and their associated epidemiology.

Classification	Risk factor	Epidemiology
Related primary diseases/history	SLE	Incidence: 26.9% ^[30]
	ALL	Incidence: $1.6 - 25\%^{[31]}$
	SCD	Prevalence: 3-50% ^[36]
	HIV	Not specific
	Cancer (chemotherapy)	Incidence: 3.45% (HR for chemotherapy was 1.94) ^[51]
	Diabetes	1.16-fold increased risk vs. non-diabetes ^[52]
	Pregnancy	Prevalence: 0.375% ^[37]
Trauma/surgery	Fractures	Prevalence: 5% ^[39] (SIR 7.98/10 000 person-years ^[40])
	Tooth extraction	Incidence: $4.31\%^{[53]}$ (OR 7.6 ^[54])
	Tooth extraction with periodontitis	HR 1.61 ^[55]
Alcohol or medication use	Alcohol consumption	$>$ 3-fold vs. non-drinkers ^[43,44]
	Bisphosphonates/denosumab	Varies with drug and duration. For bisphosphonates, HR was 2.62 ^[56] . For denosumab, 1.95-3.49-fold increased risk vs. bisphosphonates ^[57]
	Glucocorticoids	Correlated with dose and duration

ALL, acute lymphoblastic leukemia; HR, hazard ratio; OR, odds ratio; SCD, sickle cell disease; SIR, standardized incidence ratios; SLE, systemic lupus erythematosus.

with the genotype and severity^[36]. For the treatment of HIV, the mean daily oral dose of glucocorticoids (GCs) was 29 mg, and the average cumulative oral dose was 30 g over 5 years^[27]. Moreover, women appear to have an increased relative risk of osteonecrosis during pregnancy^[37]. Individuals receiving chemotherapy have a higher risk for MRONJ^[51]. Additionally, patients with diabetes have an increased risk of developing $\overline{\text{c}}$ osteonecrosis^[58], which reported a 1.16-fold increased risk for $ONFH^[52].$

Trauma or surgery

Fractures of the proximal humerus, intertrochanteric fracture, and femoral neck fracture are common with a similar rate of 7% of fractures. Due to the specific vascular anatomy, these fractures predispose the humeral head to osteonecrosis after trauma^[38]. Post-traumatic ONFH is a serious consequence of femoral neck fractures, which can lead to femoral head collapse and the development of osteoarthritis^[39,40]. Spontaneous osteonecrosis of the knee referred a subchondral fracture in the medial condyle, resulting in the collapse and the formation of necrotic tissue^[41]. Tooth extraction is an additional risk factor for individuals diagnosed as $MRON[53-55]$. Moreover, extracting a periodontally compromised tooth might increase the risk of osteonecrosis that may be associated with the previous inflammation^[55,59,60]. Inflammatory response is also a key progress for diabetes associated MRONJ^[61].

Alcohol or medication use

Alcohol intake also accounts for non-traumatic osteonecrosis $[42]$ and clinical studies demonstrated that alcohol intake increased the incidence of ONFH to more than 3-fold than non-drinkers^{[43,} ^{44]}. Nonetheless, this study also illustrated that the added effect of alcohol should not superpose the overwhelming effect of steroids[44].

MRONJ is a serious disease that occurs in the jawbones and is related to anti-bone resorption drugs and anti-angiogenesis drugs, such as bisphosphonates^[62,63] and denosumab^[56]. Bisphosphonates, including zoledronic acid or zoledronate, alendronate, and pamidronate, are important pharmacological agents against osteoclast-mediated bone loss associated with various conditions, like osteoporosis, osteogenesis imperfecta, Paget's disease, hypercalcemia, and metastatic bone malignancies^[64]. These medications promote the apoptosis of osteoclasts and play an important role in maintaining osteocyte homeostasis, which is crucial for the degradation of minerals on the bone surface. However, long-term bisphosphonate treatment and old age are more likely to induce osteonecrosis of the maxillary and mandibular bones^[65]. Receiving the treatment of denosumab might have a 1.95–3.49-fold increased risk than receiving bisphosphonates[66,67]. Worse dental health is regarded as a bad influence on MRONJ[57].

The use of GCs as an anti-inflammatory and immunomodulatory agent plays a critical role in treating various conditions, including metabolic issues like osteoporosis and obesity, but multiple side effects can produce significant morbidity, with osteonecrosis being the most common[45]. GCs contribute to the development of osteonecrosis by inhibiting osteogenesis and angiogenesis, inducing endothelial damage, and thereby reducing blood supply, which results in programmed cell death and collapse. Many clinical trials have indicated that the type of steroid used, initial steroid dosage, tapering speed, concurrent medications, cumulative dosage, and overall duration are significantly linked to the risk of osteonecrosis^[46-50].

Mechanism of imbalanced bone homeostasis in osteonecrosis

Osteonecrosis is a multifactorial disease for which the pathogeny is still unknown. The main pathological feature is the progressive death of bone cells. Bone homeostasis is a dynamic and complicated physical process that involves many specific cells [\(Fig. 2\)](#page-3-0). Cellularly, bone homeostasis is mainly maintained by osteoblasts and osteoclasts, but the dynamic balance is much more complex, and many gaps remain in our knowledge. Phenotypes and molecular mechanisms of bone homeostasis related to osteonecrosis are presented in [Fig. 3.](#page-4-0)

Viability and death of bone cells

Osteoblasts programmed cell death: The proliferation of osteoblasts is inhibited, and apoptosis is induced by dexamethasone, a type of long-effect GCs. Dexamethasone potentially attenuated

Figure 2. Overview of bone homeostasis and development of lineages. The delicate balance of bone homeostasis is accomplished by the actions of two types of cells: the osteoblast lineage and osteoclast lineage cells. Osteoblast lineage cells play a role in bone mineralization and the creation of osteocytes by excreting hydroxyapatite and calcium. In contrast, osteoclast lineage cells are involved in bone resorption.

osteoblast differentiation and increased intracellular reactive oxygen species (ROS) levels^[68]. The activations of phosphatidylinositol 3-kinase (PI3K)/AKT pathway in osteoblasts were suppressed by down-regulating the expressions of p-PI3K and p-AKT, also the downstream (mTOR) was down-regulated^[69]. Knockdown of glycogen synthase kinase 3β (GSK3β) alleviated dexamethasone-induced osteoblast apoptosis by decreasing the expressions of B-cell lymphoma-2 (BCL-2) Associated X Protein (BAX), cleaved-caspase-3, cleaved-caspase 9 and increased the expression of BCL-2^[70,71]. Dexamethasone induction significantly reduced BCL-2 expression and increased the release of cytochrome c. Dexamethasone induced osteoblast apoptosis through the up-regulation of caspase-3 that mediated by signal transducer and activation of transcription (STAT) $1^{[72,73]}$. Yang et al.^[74] found that the mediator of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase family and apoptosisrelated proteins, monoacylglycerol lipase (MAGL), was significantly upregulated during GCs treatment in osteoblasts. Sirt6 is also affected in an osteonecrosis mouse model, and osteoblast/ osteocyte-specific Sirt6 knockout mice exhibited exacerbated bone loss and significant deformities^[75]. Previous studies demonstrated that GCs upregulated antagonists for WNT/βcatenin signaling pathway, including Dickkopf1 (DKK-1) and secreted frizzled-related protein-1 (SFRP1) in osteoblasts^[76]. As in mice, it displayed a three-fold rise in osteoblast apoptosis, and

28% of osteocyte apoptosis was detected in metaphyseal cortical bone^[77]. Besides apoptosis, GCs induced ferroptosis of osteoblasts via P53/SLC7A11/GPX4 pathway^[78].

Osteoclasts dysfunction: During homeostasis, osteoclasts meticulously resorb bone at specific locations, thereby ensuring the maintenance of a matrix with the appropriate levels of strength and resilience. A study indicated that the quantity, diameter, and nuclearity of osteoclasts were significantly elevated in specimens from individuals with MRONJ^[79]. Furthermore, osteoclastic expression of nuclear factor of activated T cells-c1 (NFATc1), an essential regulator of osteoclastogenesis, was significantly increased[80,81] and the plasma levels of osteoprotegerin (OPG) and RANKL were notably higher in the individuals with ONFH than the health^[82]. The RANK-RANKL axis was the most critical role in B-cell ALL-driven bone destruction^[83]. NFκB signaling also plays a critical role in the differentiation of osteoclasts. A study found that the level of IKKe was decreased in individuals with ONFH, and IKKe interacted with NF-κB-inducing kinase (NIK) to inhibit osteoclast activity by suppressing the transcription of $Nik^{[84]}$. When the use of bisphosphonates combined with dexamethasone, the block of osteoclast differentiation was significant through the expression of IFN- $\beta^{[85]}$. Furthermore, a study indicated that bromodomain-containing protein 9 (BRD9), a component of the non-canonical BAF chromatin remodeling complex, plays an important role in MRONJ. BRD9

Figure 3. Phenotypes and molecular mechanisms of bone cells and vessels related to osteonecrosis. Osteonecrosis is a complex disease. Besides immune and inflammation factors, increased cell senescence, and fibrosis, it mainly involves the apoptosis of osteoblasts and osteocytes, adipogenic differentiation of BM-MSCs, and the dysfunction of osteoclasts and BM-MSCs with the existence of risk factors (e.g., the use of glucocorticoids). Moreover, hematology impairment also contributes to the progress and inhibits the AKT signaling pathway in osteoblasts. BM-MSC, bone marrow-derived-mesenchymal stromal cell; GC, glucocorticoids.

deficiency in the myeloid lineage promoted osteoclast differentiation and increased bone resorption by suppressing interferon-β signaling and interacting with the transcription factor FOXP1, which subsequently activated *Stat1* transcription^[86].

Osteocytes apoptosis and differentiation: Osteonecrosis is characterized by excessive osteocyte death. For pathology, GCs induce osteocyte apoptosis, develop hyper-mineralization, and degeneration of the lacunocanalicular network rapidly, which resulting dysregulation of bone architecture and mineralization in human osteonecrosis^[87]. Furthermore, there was a significant buildup of low-density lipoprotein (LDL)/oxidized LDL (ox-LDL) in the osteocytes (lacunae) of necrotic areas, in contrast to healthy regions. Cell studies demonstrated that ox-LDL reduced the viability of osteocytes and enhanced their apoptosis^[88]. Both RANKL and sclerostin were markedly increased in bisphosphonates conditioned medium, moreover, Janus-activated kinase (JAK)/STAT3 pathway was subsequently activated with up-regulation of the $RANKL^[89]$. Moreover, the phosphatase and tensin homolog (PTEN)-Protein kinase B (AKT) signaling^[90,91] and PI3K/Akt/mTOR pathway^[92] also participated in osteocyte apoptosis. Moreover, a study indicated that osteocytic cell death was linked to mitochondrial pathways. Exposure to either dexamethasone or hypoxia resulted in diminished mitochondrial membrane potential, reduced levels of ATP synthase (ATP5A), and increased expression of 8-OHdG, cleaved caspases, and the X-linked inhibitor of apoptosis protein (XIAP)^[93]. Generally, the cross-talk between bone cells is complicated. A study showed that osteocytes in necrotic regions released damage-associated

molecular patterns (DAMPs) which can be sensed by osteoclasts via macrophage-inducible C-type lectin (Mincle). Mincle activation triggered osteoclastogenesis via immunoreceptor tyrosine-based activation motif-based calcium signaling pathways, triggering bone loss^[94].

BM-MSCs differentiation and apoptosis: An increase in bone marrow fat resulting from adipocyte accumulation is always regarded as a pathogenic process in osteonecrosis. Exposure to the saturated free fatty acids palmitate favored BM-MSCs (bone marrow-derived-mesenchymal stromal cells) differentiation through the adipogenic lineage at the expense of the osteoblastic phenotype. Additionally, BM-MSCs treated with steroids accumulated triglyceride, indicating that steroids increased the adipogenic differentiation of BM-MSCs^[95]. Moreover, adipogenesis was intensified and the ability to differentiate into bone was deceased in BM-MSCs of osteonecrosis^[96]. Fat mass and obesity (FTO), a key m⁶A demethylase, was found to be significantly down-regulated in individuals with osteonecrosis by targeting PPARG and was markedly upregulated during the differentiation of human MSCs into osteoblasts^[97]. One study indicated that the endogenous expression of periostin (POSTN) and SOST in BM-MSCs of ONFH was upregulated compared with developmental dysplasia of the hip group and partially inactivating the WNT/β-catenin signaling pathway^[98]. The WNT pathway is a crucial signal transduction pathway. The extracellular protein DKK1 serves as. Corticosteroids upregulated DKK1 (a negative regulator in WNT signaling) expression in BM-MSCs, which led to the delay in bone phylogeny, regeneration, and remodeling^[99,100].

The oxidative stress (OS) microenvironment of the necrotic regions leads to mitochondria damage ,stress-induced apoptosis, and senescence of BM-MSCs. The increased levels of protein 53 (P53) hindered the mitochondrial translocation of Parkin and activation of E3 ubiquitin ligase, leading to a reduction in mitophagy^[101]. Moreover, the activation of SIRT3 is also related to $\tilde{\mathrm{OS}}^{[10\tilde{2}]}$. A functional experiment demonstrated that inhibiting MAGL significantly decreased OS and partially protected BM-MSCs from apoptosis. Pathway analysis indicated that MAGL inhibition regulated oxidative stress in BM-MSCs through the Kelch-like ECH-associated protein 1 (KEAP1)/NRF2 pathway[74]. However, another study indicated BM-MSCs maintained mitochondrial function and exhibited reduced OS. Detailly, the mitochondrial membrane potentials remained stable^[93].

Hematology impairment

Osteonecrosis involves the disruption of vascular supply to the bone and hypoxia microenvironment caused by impaired blood flow could be essential pathogenic factors. The increased levels of VEGF were reported to inhibit steroid-induced osteonecrosis in a rabbit model^[46]. Additionally, VEGF, as a key mediator of angiogenesis, stimulated endothelial cell proliferation, differentiation, and survival that assessed by tube formation experiments and related to interferon-induced transmembrane protein $1^{[103]}$. It was challenging to detect HIF-1 α and VEGF in the human femoral head at end-stage ONFH, which indicated that the impairment of vascular network formation and reduced osteogenic activity ultimately led to osteonecrosis^[104]. Besides VEGF, ankyrin was a cytoskeletal protein that interacted with CD44 to promote endothelial nitric oxide synthase expression and tissue nitric oxide levels via the Akt signaling pathway^[105]. Moreover, PTEN promoted endothelial progenitor cell apoptosis under GC condition^[106]. Another reasonable mechanism was that increased PAI-1 reduced fibrinolytic function, and lastly accelerated the process of ONFH by reducing the femoral head blood flow^[67]. In juvenile mice induced by GCs, it was reported that bone angiogenesis and type H vessel (in the primary spongiosa, where trabecular bone formation begins) were disrupted, which was linked to reduced PDGF-B expression in preosteoclasts[107].

Immune and inflammation factors

The immune and skeletal system have been revealed that crossly interacted by cytokines, chemokines, receptors, signaling molecules, and transcription factors. Previous studies have suggested that activated T/Th1 cells and B cells stimulated the high expressions of RANKL and OPG in individuals with ONFH. In detail, CD4⁺ Th17 cells raised the levels of RANKL and cytokines (such as TNF- α , IL-23, IL-17^[83], and IL-15^[108]). IL-34 alone or works in coordination with macrophage-colony-stimulating factor (M-CSF) to promote osteoclastogenesis and activate extracellular regulated protein kinases (ERK), STAT3, and noncanonical NF-κB pathways, which may contribute to ONFH[109]. Moreover, immunohistochemistry results indicated that the M1/ M₂ polarization ratio increased from 3 to 10 as the condition progressed from the progressive to the end stage $[110]$. As for MRONJ a significant increase in M1 polarization also contributed to the progress^[111]. Moreover, the establishment of unique bacterial communities, along with a weakened innate

immune response, could be likely to influence disease progression^[112]. Bisphosphonates treatment of dendritic cells caused an impairment in immune functions including differentiation, maturation, migration, antigen presentation, and T-cell activation in vitro^[113]. After injecting mice with bisphosphonates and tooth extraction, immune cells from bone marrow and spleen, and purified natural killer cells from the spleen showed high induction of IFN-γ and natural killer cell-mediated cytotoxicity except for gingiva in which immune cells were in contrast^[114,115]. Moreover, exposure to bisphosphonates resulted to impaired neutrophil chemotaxis, decreased neutrophil NADPH oxidase activity, and a lower number of circulating neutrophils^[116].

The production of inflammatory cytokines plays an important role in the pathogenesis. Nod-like receptor pyrin domain-containing protein 3 (NLRP3) inflammasome, caspase-1, and IL-1β in macrophages contributed to the persistent inflammation^{[61,117}] . There was a debate about the existence of TNF-α. Zheng and their colleague figured that TNF- α induces the apoptosis of osteoblasts with a high expression in necrotic regions^[118], while Fang et al.^[119] demonstrated that the content of TNF- α in necrotic tissue is much lower than that of normal tissue. TNF-α is also related to autophagy, which is beneficial for interrupting the process of osteonecrosis, therefore TNF-α might should not be considered as a therapeutic target. IL-9 was also highly expressed in osteonecrosis, leading to an elevated level of inflammationrelated cytokines and cartilage matrix-degrading enzymes, alongside heightened activation of JAK-STAT signaling^[120].

Fibrosis and other novel findings

A recent study figured that the extension of fibrosis on specimens from biopsies in ONFH is a predictor of the outcome of the core decompression^[121]. In Legg-Calvé-Perthes disease (LCPD), an increased fibrous tissue and adipose tissue was detected in the bone marrow space and a decreased osteogenesis was reported in a piglet model^[122]. However, the mechanisms of fibrosis are merely explored.

Chen et al. found that hsa-miR-200b-3p and hsa-miR-206 in urinary exosomes might serve as non-invasive biomarkers for GCs-induced ONFH and the Hippo, PI3K-AKT, TGF-β and Wnt signaling were enriched in bioinformation analysis^[123]. Shen and their colleague indicated that zoledronic acid activated the NF-κB signaling pathway and macrophages derived miR-149-5p regulate biological functions of endothelial cells via the Rap1a/ Rap1b/VEGFR2 pathway^[124]. MiR-708 was also found to be upregulated in individuals with ONFH and in cell experiments. It targeted SMAD3 and inhibited osteogenic differentiation and adipogenesis of MSCs^[125]. An increase in miR-141^[126] and reductions in miR-17-5p^[127], miR-21-5p^[128], miR-4523^[129], miR-122-5 $p^{[130]}$, and miR-26a^[131] are potential mechanisms for ONFH. Furthermore, Fang et al.^[132] found that tsRNA-10277 was significantly down-regulated in plasma exosomes of individuals with GCs-induce ONFH compared to that in healthy individuals and tsRNA-10277 showed positive effects on osteogenic differentiation ability of BM-MSCs in vitro.

Advancements in decade for prevention and treatment of osteonecrosis

The studies published in the past decade related to clinical use and effectiveness included in this review are summarized in [Table 2](#page-6-0). A

Characteristics of included studies about clinical use and effectiveness.

1107

(Continued)

1108

(Continued)

ARCO, Association Research Circulation Osseous: ATA, atmospheres absolute; BMAC, bone marrow aspirate concentrates; BM-MNCs, bone marrow-derived mononuclear cells; BM-MSCs, bone marrow-derived MSCs; CD, core decompression; CT, computed tomography; EPC, endothelial progenitor cells; EQ-5D, EuroQol-5 Dimension; ESWT, extracorporeal shock wave therapy; GCSF, granulocyte colony-stimulating factor; HAM, human amniotic membrane; HBOT, Hyperbaric o Association Hip-Disease Evaluation Questionnaire; HHS, Harris Hip Score; hUC-MSCs, human umbilical cord-derived MSCs; IKDC, Knee Documentation Committee subjective knee form; JHEQ, Japanese Orthopaedic Association Hip-Dise Japanese Orthopaedic Association; KSS, knee society score; L-PRF, L-Platelet-Rich Fibrin; MRONJ, medication-related osteonecrosis of the jaw; MSCs, mesenchymal stromal cells; NA, not applied; NT, not mentioned; OHS, Oxford head; OPG, osteoprotegerin; PBSCs, peripheral blood stem cells; PEMFs, pulsed electromagnetic fields therapy; PMA, Postel and Merle d'Aubigné score; RCT, randomized controlled trial; rhBMP-2, recombinant human bone morphog fibroblast growth factor; ROM, range-of-motion; SCD, sickle cell disease; SF-36, Medical Outcomes Study 36-Item Short-Form Health Survey; SONK, spontaneous osteonecrosis of the knee; SPECT, single photon emission computed Vascular Fraction; THA, total hip arthroplasty; TKA, total knee arthroplasty; VAC, vacuum-assisted closure; VAS, Visual Analog Scale; WOMAC, The Western Ontario and McMaster Universities Osteoarthritis.

Figure 4. Current prevention and treatment of osteonecrosis. BMAC, bone marrow aspirate concentrates; BM-MNCs, bone marrow-derived mononuclear cells; CD, core decompression; HBOT, hyperbaric oxygen therapy; PRP, platelet-rich plasma; THA, total hip arthroplasty.

well-established staging system should be used, in general, these treatments can be divided into prevention, conservative therapies, surgeries, and surgeries with additional therapies (Fig. 4). The advantages and disadvantages of various treatments are listed in [Table 3.](#page-12-0)

Prevention of osteonecrosis

For osteonecrosis, early diagnosis, coordination of care, and regular visits play important roles in the management. For traumatic ONFH, the blood supply for the femoral head should be evaluated regularly and should be protected during internal fixation^[194]. For the long-term use of steroids, vasodilator drugs may be considered. In general, individuals with risks of osteonecrosis should be aware of these risk factors during patient education and try to avoid the mentioned risks.

Conservative therapy

For ONFH, the consensus has reported that orthotics (canes and crutches) could promote pain relief and walking ability by reducing weight-bearing loading. Hyperbaric oxygen therapy (HBOT) has beneficial effects, and its mechanism might include upregulating serum OPG and/or inhibiting osteoclast activation. HBOT significantly reduced lesion size in early-stage patients^[133] . Significant changes in serum biomarkers for angiogenesis, osteogenesis, anti-inflammation, pain threshold, and tissue regeneration were also reported after extracorporeal shock waves^[134]. Ozone therapy also has the potential to improve symptoms and delay the need for total hip arthroplasty $(THA)^{[135]}$. Pulsed electromagnetic fields therapy with a dosage of 6 h daily for 90 days reduced knee pain and necrosis area of $knee^{[136]}$. For spontaneous osteonecrosis of the knee, Atoun and colleagues reported that a biomechanical therapy (an orthosis for the foot) could reduce pain and increase activity^[137]. For MSCs, BM-MSCs alone improved the quality of life of leukemia survivors with ankle osteonecrosis $[138]$, and ONFH in females after pregnancy^[37]. Hernigou et al.^[39] reported that BM-MSCs obtained better survival time before revision compared to THA. De Rojas et al.^[139] reported that BM-MSCs therapy was a novel treatment for children and young adults with ONFH after overcoming ALL, although the X-ray and MRI imaging of the femoral head deteriorated. Hernigou et al .^[140] also indicated that BM-MSCs gained similar improvement with a lower rate of complications compared with total knee arthroplasty. Autologous peripheral blood stem cells^[141] and umbilical cord-derived $\text{MSCs}^{[142]}$ were proven to be another origin of MSCs. Besides MSCs, bone marrow-derived mononuclear cells (BM-MNCs) also showed similar effects^[143]. Moreover, bisphosphonate treatment should not be considered as a potential therapy for preventing osteonecrosis^[144].

For MRONJ, conservative measures may include the use of antimicrobial mouth rinses and antibiotics^[145], effective oral

BM-MSCs, bone marrow-derived MSCs; CD, core decompression; ESWT, extracorporeal shock wave therapy; HBOT, hyperbaric oxygen therapy; L-PRF, L-platelet-rich fibrin; MNC, mononuclear cells; MSCs, mesenchymal stromal cells; N blood stem cells; PEMFs, pulsed electromagnetic fields therapy; PRP, platelet-rich plasma; rhBMP-2, recombinant human bone morphogenetic protein-2; rhFGF, recombinant human fibroblast growth factor; THA, total hip arthropl

1113

hygiene, and conservative surgical interventions. Moreover, Sim $et al.^[146]$ reported that teriparatide (an osteoanabolic medication) provided another choice for treating lesions in MRONJ, which is consistent with the study of Jung and colleagues^[147]. Laimer et al .^[148] reported that intraoral vacuum-assisted closure therapy can reduce pain and increase the degree of cleanliness that provides a good condition for healing the closure.

Invasive operation

Surgery: The intramedullary pressure has been reported to be elevated in individuals with ONFH, and the high bone marrow pressure was regarded as one of the most common reasons for the development of ONFH. Regarding this, CD was considered as a useful technique for intramedullary decompression of the osteonecrosis regions. In general, Kirschner wire was placed to locate the necrotic area and direction of the wire, then a suitable support rod was inserted toward the anterolateral necrotic area of the femoral head until it was safe beneath the subchondral bone. Moreover, Yue et al.^[149] designed a novel method, called a single approach to double-channel CD and bone grafting with structural bone support. After CD, one of the two channels (top one) was filled with a support rod and grafting bone, while the bottle one only filled with bone matrix. Researchers often designed the CD as an intervention in a control group according to the publishments in the last 10 years. Wang and colleagues reported that the light bulb technique (one of the non-vascularized bone grafting techniques) had similar outcomes to $CD^{[150]}$. Civinini and colleagues reported that CD with a calcium sulfate/calcium phosphate bioceramic could make progress in a quantitative computed tomography analysis $^{[151]}$. Another study combined autologous bone and CD, which was also reported positively $[152]$.

Bone grafting transfer is a common surgical strategy for ONFH^[153]. Wang et al.^[150] reported the light bulb technique through a direct anterior approach, which allowed the removal of more necrotic bone by making a round bone window 1.0 cm in diameter using osteotomes and then a high-speed drill and a curette. Compared to non-vascularized bone grafting, vascularized bone grafting seems to be more beneficial for large joints, such as the hip. In general, a bone (usually fibula) was cut, a bone groove was made as before, the distal end of the fibula was grafted, and then the free peroneal vessels and the lateral femoral circumflex artery and vein were sutured^[154,155]. Cao and their colleague performed a study to compare the effect of vascularized fibular grafting and CD, and the results supported the application of free vascularized fibular grafting^[154]. Sun et al.^[156] reported pedicled vascularized iliac bone grafting transfer improved function 6 months after surgery and Zhang et al ^[157] reported similar outcomes one year after the operation. Bone grafts combined with $BMSCs^{[158]}$ and biomaterial bone grafts seem to be effective treatment methods^[159]. Kang and the colleague figured that bone grafts combined with BMSCs might be more effective for individuals with medium-sized lesions $[160]$. Nevertheless, Chen and the colleague reported that the metal trabecular bone with bone dust of the femoral trochanteric region and femur neck was superior to free vascularized fibular graft^[155]. Electromagnetic field stimulation as an additional treatment to autologous bone grafting might not have some extra beneficial effects^[161]

Femoral transtrochanteric rotational osteotomy, a wildly known technique to alleviate symptoms and prevent the

deterioration of ONFH, required sufficient intact area at the lateral femoral head. The bone cut was through the trochanter, and the femoral head was anterior or posterior rotated properly to improve its position within the hip socket^[162]. Utsunomiya and their colleagues reviewed previous patients who underwent THA after Sugioka transtrochanteric anterior rotational osteotomy, and considered the effects to be comparable to those of THA^[163]. Curved varus osteotomy was conducted on patients who exhibited more than one-third coverage of the weightbearing region with an intact articular surface. A curved osteotomy and a varus position were needed for performing this surgery. For patients younger than 50 years, curved varus osteotomy was comparable with THA for functional outcomes, survival rate, and sporting activities after a mean follow-up period of 10 years^[164]. Curved intertrochanteric varus osteotomy combined with bone impaction grafting yielded more positive outcomes when participants were carefully selected, and the procedure was executed accurately^[165].

The last two choices of surgery are hemiarthroplasty and THA. A recent decade of studies merely reported the outcomes of bipolar hemiarthroplasty for ONFH. The reason might be the mid-to-long-term results in the early stage might be generally good, nonetheless, individuals may experience postoperative pain in the buttocks and groin, along with the potential migration of the outer head. The long-term results indicated that cemented THA using modern cementing techniques might be an effective and beneficial treatment option. However, using a cemented conventional stem as the femoral component may lead to a subset of requires for hip revision^[166]. Hence, cementless THA was considered, although the risk of stress shielding along with some thigh pain increased. The long-term results from a large cohort multicenter survey indicated that contemporary cementless THA in young individuals with hematological disease following bone marrow transplantation did not increase the surgical complications, revision, reoperation, readmission, or mortality $[167]$.

For MRONJ, recent American Association of Oral and Maxillofacial Surgeons guidance recommended debridement to relieve soft-tissue irritation and infection control at stage 2 (softtissue inflammatory swelling or infection), with resection considered at stage 3 (radiographic evidence of osteolysis extending to the inferior border of mandible). Voss et al ^[168] figured that surgical management with MSCs further promoted the healing of MRONJ. A comprehensive strategy for achieving healing in MRONJ may involve thoroughly removing necrotic bone, smoothing sharp bony edges, and carefully closing the wound, all supported by perioperative antibiotic treatment. Calvani et al.^[169] reported bovine lactoferrin could promote wound repair in individuals with suffering from MRONJ with the progressive destruction of bone.

Additional therapy: Last decade, cell-based therapy was rapidly developed and usually combined with orthopedic surgery. However, the necessity for the application of cell therapy is still controversial^[24]. Among the substantial regenerative options of cell-based therapy, bone marrow aspirate concentrates (BMAC) was frequently used mainly in Europe and the United States, while BM-MNCs, BM-MSCs, PBSCs^[170], and plateletrich plasma $(PRP)^{[195]}$ were also applied. Many clinical studies^[171,172] have investigated the effect of BMAC combined with CD, which indicated the elimination of cell proliferation but did not produce any improvement at the later stage. Moreover, BM-MSCs therapy with CD improved the outcome of the

humeral head osteonecrosis^[38,173] and ONFH and bone regeneration in the early stages of ONFH^[174–178]. For talus osteonecrosis, Hernigou et al. found similar results^[179]. Wu et al.^[180] indicated that the clinical effect was related to the quality of BM-MSCs, in detail, the repair of the necrosis area was a positive correlation to the osteogenic and chondrogenic differentiation ability. Emadedin et $dL^{[181]}$ injected bone marrow-derived CD133⁺ cells (an early hematopoietic stem cell marker) during CD, which significantly and clinically improved pain relief and function. The rehabilitation program after BM-MNCs injection significantly improved the external rotation range of motion and strength of extensor and abductor muscle compared with that before treatment^[182]. Nonetheless, Jayankura and colleagues^[183] indicated that autologous osteoblastic cells addicted to CD did not take extra benefits in early-stage ONFH patients. Moreover, Hauzeur reported that osteoblast therapy obtained similar effects with BMAC^[184]. Hernandez et al.^[185] also reported that CD combined autologous BMAC and tricalcium phosphate therapy show no improvement in preventing progression to collapse.

Growth factors have been suggested, but their efficacy remained uncertain. Regarding growth factors, recombinant human BMP-2 and BMP-7 were combined with bone grafting. Additionally, clinical trials using gelatin hydrogel impregnated with recombinant human fibroblast growth factor (rhFGF)-2 were performed for individuals with pre-collapse ONFH^[186]. Kuroda and colleague^[187] also figured that rhFGF-2 treatment alone also had positive effects on activity and joint preservation time improvement. Gao and colleagues combined BMP-2 and bone marrow-derived cells during CD, which also prevented the progression of osteonecrosis in young individuals^[188].

For MRONJ, the human amniotic membrane is used as a low immunogenicity regenerative strategy and is reported to help reduce pain^[189] and promote wound healing^[190]. Bouland and their colleagues[191] designed a leukocyte-rich platelet-rich fibrin seeded with stromal vascular fraction, which was available and promoted the healing of buccal mucosa. Besides this, Park et al ^[192] combined leukocyte-rich platelet-rich fibrin with rhBMP-2, which also promoted MRONJ healing. Concentrated growth factor may provide limited advantages in promoting the healing of surgical sites^[193].

Discussion

Scope

This review aims to comprehensively sum up the risk factors for osteonecrosis. For individuals with high-risk primary diseases, they should monitor the susceptible structure of osteonecrosis, and prevent the development of disease. For traumatic damaged vessels, surgeons should repair the vessels as soon as they can and monitor the blood supply on time. For the overused steroids, proper prescription and prevention treatment can be used. Secondly, this review aims to outline various mechanisms of osteonecrosis, especially for ONFH and MRONJ. Besides a deeper understanding of the disease, comprehensively demonstrating the mechanisms could promote the development and application of potential biotherapies. Lastly, we aim to classify the current therapeutic strategies for osteonecrosis, and hope could help make the standard process. Some of the treatment strategies focused on the prevention of disease progression, recovery of blood supply, and the surrounding environment. Others reported the application of different surgeries. It is worth noticing that additional cell-based injections or implants with orthopedic surgery, such as CD have been applied more and more in clinics^[196]. The effectiveness of additional cell-based therapy requires more high-quality clinical trials with consistent cell preparation and intervention methods.

Limitations of osteonecrosis research

Although there already exist many therapeutic strategies for osteonecrosis, efficacy and effectiveness are still controversial for medical decisions. Revision surgery may be required and symptoms (such as pain) remain^[167]. Developing novel, valuable, and highly efficient treatments is still an urgent problem and a big challenge. We figured out the current limitations of studies of osteonecrosis and tried to identify future solution ideas as below and presented in [Fig. 5](#page-15-0).

Undetermined risk factors and complex mechanisms with primary diseases

With the development of epidemiology, the risk factors of osteonecrosis have gradually become clear. However, the results were various among different areas and populations. Additionally, although we summarized several mechanisms about the imbalanced bone hemostasis during osteonecrosis, merely basic studies focused on the primary diseases that might increase the occurrence of osteonecrosis. Moreover, current research often reported the phenotypes, rather than molecular mechanisms.

Vacant position for superior treatment and insufficient evidence for regenerative medicine

Despite we classified the current therapeutic strategies for osteonecrosis, there are still limitations in choosing the most effective treatment. Although the study of Hernigou and colleague reported that MSCs transplantation was better than $arthroplasty^{[140]}$, the conclusion may be various among other locations of osteonecrosis. According to the publishments in decade, the combination of regenerative medicine and surgeries gained more focus. Nonetheless, the study design of these studies mostly was case series, and lack of comparisons and results of long-term follow-up. The low-grade evidence for additional cellbased therapy might limit the applications.

Roadblocks in translational medicine

Although studies over the last decades have provided much progression, the integration of these findings into a reproducible treatment strategy applicable to the entire spectrum of osteonecrosis remains a challenge. Additionally, we still need to consider the common gaps for translational medicine, such as resolution of drugs, immune aggravation, low proportion of cells surviving after transplant, inflammation, fibrosis, and how to control the "growth-stop signals" and modulation of various signaling pathways^[197].

Future directions of osteonecrosis research

Identify risk factors and key molecules

A long follow-up and worldwide cohort study may be urgent and necessary for identifying more risk factors. Avoid the risk factors

Figure 5. Limitations and future directions of osteonecrosis. EVs, extracellular vehicles; MSCs, mesenchymal stromal cells; RCT, randomized controlled trials; tFNAs, tetrahedral framework nucleic acids.

and get prevention as soon as possible could slow the process of disease. For basic study, gene knockout or gene targeting techniques may result in a deeper understanding of the pathology of osteonecrosis. Additionally, upcoming research could aim to clarify the molecular mechanisms that contribute to the protective effects of gene therapy, such as PGK1. Functional studies in vitro demonstrated that phosphoglycerate kinase 1 (PGK1) shRNA largely attenuated dexamethasone-induced ROS following the death of osteoblasts^[198]. Investigating the regulation of these key targets with signaling pathways involved in bone homeostasis could provide valuable insights for novel therapies.

Perform high-quality clinical trial and update practice

Owing to the study design and different sites of osteonecrosis, popularize of findings from clinical studies is challenging. Welldesigned multicenter RCTs could solve the clinical and methodological heterogeneity, which are essential for the current treatments, especially for conservative treatments and additional therapies with CD or bone grafting. Moreover, a long-term follow-up (such as 3–10 years) should be designed for clinical studies involving MSCs. Moreover, consistent cell preparation (such as cell origin, cell viability, and differentiation potency) and intervention methods (volume and the same combinational strategy)

should be considered since the report of Wu et al.^[180] indicated that the quality of BM-MSCs affected the clinical effectiveness.

For clinical practice, cell-based therapies or drugs with surgeries has becoming a trend. Besides current additional therapies, exercise and physical therapies before and after operation may also help return to life^[182]. An integrative treatment that contains conservative treatment, surgery, regenerative medicine should be performed in future research. Additionally, translational medicine implements with CD, which as the most classical technique should be considered by investigators moving forwards.

Accelerate translational potential

An ideal pharmacological therapy for osteonecrosis should help reduce the inflammatory and promote bone repair by stimulating blood supply aiming at inducing progenitor growth and improving the microenvironment. Epigenetic modification (such as miR-214^[199] and miR-26a^[131]) and modulation of various signaling pathways (such as a local delivery system with pharmacological mediator of BRD9 and flexible injectable silk fibroin hydrogel^[86], BMP-2^[188], and FGF-2^[186,187]) are potential therapeutic avenues and proved in pre-clinical trials.

For tissue engineering, scaffolds can serve as mechanical support or carriers. Researchers aim to develop innovative biomaterial strategies that enhance bone regeneration and provide low-grade evidence (animal and cell experiments), such as poly (lactic-co-glycolic acid)/β-calcium phosphate/icariin (PLGA/ $TCP^{[200-203]})$, synthetic calcium phosphate ceramics^[204], injectable hydrogel with microsphere bone cement^[205], magnesiumbased layered double hydroxide nanosheets[206], strontium-doped calcium polyphosphate (SCPP)^[207], and biodegradable poly (ethylene glycol) maleate citrate (PEGMC). MSCs, extracellular vehicles (EVs), miRNAs (such as MiR@TDNs/Li-hep-gel^[100], which upregulated the Wnt signaling pathway), and proteins (such as VEGF-loaded gelatin microspheres^[208] and BMP-VEGF-PLGA-calcium polyphosphate composite^[209]) are also used and could as common seeds combined with scaffolds.

Lastly, the current study emphasized that nanoparticles may serve as potential carriers for gene therapy and drug delivery. Nano-hydroxyapatite-copper-lithium (Cu-Li-nHA) composite^[104] and tetrahedral framework nucleic acids (tFNAs) were innovative nanomaterials showing potential for drug delivery, and recent studies indicated that tFNAs restored osteogenesis dysfunction and attenuated BM-MSCs apoptosis in treating $\text{ONFH}^{[210]}$, and promoted angiogenesis in treating MRONJ^[211].

Conclusion

Osteonecrosis is a refractory disease marked by the death of bone cells, and maintaining bone homeostasis is a challenge for this disease. In this review, we have discussed the roles of osteoblasts, osteoclasts, osteocytes, BM-MSCs, and bone marrow niche and angiogenesis. Pregnancy, diseases (such as SLE, ALL, SCD, HIV), trauma, alcohol intake, GCs use, and bisphosphonates use are risks for osteonecrosis. There are several proven mechanisms of impairment in bone homeostasis for osteonecrosis, including apoptosis of osteoblasts, osteocytes and BM-MSCs, dysfunction of osteoclasts, immune and inflammation, adipogenic differentiation, cell senescence, hematology impairment, and fibrosis. We have attempted to logically connect current pathways to these mechanisms and summarize the current treatments hierarchically. However, some limitations still exist, including undetermined risk factors, complex mechanisms with primary diseases, vacant positions for superior treatment, insufficient evidence for regenerative medicine, and roadblocks in translational medicine. Ongoing and future research should be directed at elucidating risk factors and identifying new targets, performing high-quality clinical trial, clarifying the standard quality of cell therapies, updating clinical practice (combination of conservative treatments, surgeries and regenerative medicine), accelerating translational medicine by developing pharmacological therapies, tissue engineering, and nanotechnology.

Ethical approval

Not applicable.

Consent

Not applicable.

Source of funding

The National Natural Science Foundation of China (81972146, 82272599), the Department of Science and Technology of Sichuan Province (2021YFS0004), and 1.3.5 project for disciplines of excellence of West China Hospital, Sichuan University (ZYGD23014).

Author contribution

X.N.X.: conceptualization, investigation, visualization, writing —original draft. H.C.H.: writing—review and editing, supervision. C.Q.H.: conceptualization, writing—review, supervision, funding acquisition. All the authors approve and agree with the content of the review.

Conflicts of interest disclosure

The authors declare no conflicts of interest.

Research registration unique identifying number (UIN)

Not applicable.

Guarantor

Cheng-Qi He.

Data availability statement

This manuscript is a review and makes use of publicly available data from published studies, therefore, no original data are available for sharing.

Provenance and peer review

Not commissioned, externally peer-reviewed.

Acknowledgement

The authors thank Si-Yi Zhu, Hao-Nan Wang, and Ping Mou in West China Hospital and Jie Deng in Department of Orthopedics and Sports Medicine, Erasmus MC University Medical Center for their assistance in the outline, and the authors acknowledge funding from the National Natural Science Foundation of China (81972146, 82272599), the Department of Science and Technology of Sichuan Province (2021YFS0004), 1.3.5 project for disciplines of excellence of West China Hospital, Sichuan University (ZYGD23014).

References

- [1] Yan R, Jiang R, Hu L, et al. Establishment and assessment of rodent models of medication-related osteonecrosis of the jaw (MRONJ). Int J Oral Sci 2022;14:41.
- [2] Quan H, Ren C, He Y, et al. Application of biomaterials in treating early osteonecrosis of the femoral head: research progress and future perspectives. Acta Biomater 2023;164:15–73.
- [3] Lamb JN, Holton C, O'Connor P, et al. Avascular necrosis of the hip. BMJ (Clinical Research ed) 2019;365:12178.
- [4] Perry DC, Machin DM, Pope D, et al. Racial and geographic factors in

the incidence of Legg-Calve-Perthes' disease: a systematic review. Am J Epidemiol 2012;175:159–66.

- [5] Littooij AS, Kwee TC, Enriquez G, et al. Whole-body MRI reveals high incidence of osteonecrosis in children treated for Hodgkin lymphoma. Br J Haematol 2017;176:637–42.
- [6] Saad F, Brown JE, Van Poznak C, et al. Incidence, risk factors, and outcomes of osteonecrosis of the jaw: integrated analysis from three blinded active-controlled phase III trials in cancer patients with bone metastases. Ann Oncol 2012;23:1341–7.
- [7] Chang C, Greenspan A, Gershwin ME. The pathogenesis, diagnosis and clinical manifestations of steroid-induced osteonecrosis. J Autoimmun 2020;110:102460.
- [8] Reid IR, Cornish J. Epidemiology and pathogenesis of osteonecrosis of the jaw. Nat Rev Rheumatol 2011;8:90-6.
- [9] Zheng L-Z, Wang J-L, Kong L, et al. Steroid-associated osteonecrosis animal model in rats. J Orthop Transl 2018;13:13–24.
- [10] Paparella ML, Brandizzi D, Santini-Araujo E, et al. Histopathological features of osteonecrosis of the jaw associated with bisphosphonates. Histopathology 2012;60:514–6.
- [11] Zhu Q, Fu Y, Cui C-P, et al. OTUB1 promotes osteoblastic bone formation through stabilizing FGFR2. Signal Transduct Target Ther 2023;8:142.
- [12] Redlich K, Ziegler S, Kiener HP, et al. Bone mineral density and biochemical parameters of bone metabolism in female patients with systemic lupus erythematosus. Ann Rheumat Dis 2000;59:308–10.
- [13] van Gastel N, Carmeliet G. Metabolic regulation of skeletal cell fate and function in physiology and disease. Nat Metab 2021;3:11–20.
- [14] Uenaka M, Yamashita E, Kikuta J, et al. Osteoblast-derived vesicles induce a switch from bone-formation to bone-resorption in vivo. Nat Commun 2022;13:1066.
- [15] Sims NA, Martin TJ. Osteoclasts provide coupling signals to osteoblast lineage cells through multiple mechanisms. Annu Rev Physiol 2020;82: 507–29.
- [16] Robling AG, Bonewald LF. The osteocyte: new insights. Annu Rev Physiol 2020;82:485–506.
- [17] Hofbauer LC, Lademann F, Rauner M. Deconstructing cellular senescence in bone and beyond. J Clin Invest 2023;133:e169069.
- [18] Li D, Liu J, Guo B, et al. Osteoclast-derived exosomal miR-214-3p inhibits osteoblastic bone formation. Nat Commun 2016;7:10872.
- Zaidi M, Yuen T, Sun L, et al. Regulation of skeletal homeostasis. Endocr Rev 2018;39:701–18.
- [20] Bae S, Kim K, Kang K, et al. RANKL-responsive epigenetic mechanism reprograms macrophages into bone-resorbing osteoclasts. Cell Mol Immunol 2023;20:94–109.
- [21] Di Maggio N, Banfi A. The osteo-angiogenic signaling crosstalk for bone regeneration: harmony out of complexity. Curr Opin Biotechnol 2022;76:102750.
- [22] Zhao D, Zhang F, Wang B, et al. Guidelines for clinical diagnosis and treatment of osteonecrosis of the femoral head in adults (2019 version). J Orthop Translat 2020;21:100–10.
- [23] Yarom N, Shapiro CL, Peterson DE, et al. Medication-Related Osteonecrosis of the Jaw: MASCC/ISOO/ASCO Clinical Practice Guideline. J Clin Oncol 2019;37:2270–90.
- [24] Meermans G. CORR Insights®: Did Osteoblastic Cell Therapy Improve the Prognosis of Pre-fracture Osteonecrosis of the Femoral Head? A Randomized Controlled Trial. Clin Orthop Relat Res 2020;478: 1316–8.
- [25] Sadile F, Bernasconi A, Russo S, et al. Core decompression versus other joint preserving treatments for osteonecrosis of the femoral head: a meta-analysis. Br Med Bull 2016;118:33–49.
- [26] Padhye B, Dalla-Pozza L, Little D, et al. Incidence and outcome of osteonecrosis in children and adolescents after intensive therapy for acute lymphoblastic leukemia (ALL). Cancer Med 2016;5:960–7.
- [27] Krez A, Lane J, Heilbronner A, et al. Risk factors for multi-joint disease in patients with glucocorticoid-induced osteonecrosis. Osteoporos Int 2021;32:2095–103.
- [28] Borges $\hat{A}H$, Hoy J, Florence E, *et al.* Antiretrovirals, fractures, and osteonecrosis in a large international HIV cohort. Clin Infect Dis 2017; 64:1413–21.
- [29] Mimura N, Iwamoto T, Furuta S, et al. Prevalence and risk factors of osteonecrosis of the femoral head in patients with ANCA-associated vasculitis: a multicentre cohort study. RMD Open 2023;9:e002787.
- [30] Kuroda T, Tanabe N, Wakamatsu A, et al. High triglyceride is a risk factor for silent osteonecrosis of the femoral head in systemic lupus erythematosus. Clin Rheumatol 2015;34:2071–7.
- [31] Brivio E, Cossio A, Borra D, et al. Osteonecrosis in paediatric acute lymphoblastic leukaemia: Incidence, risk factors, radiological patterns and evolution in a single-centre cohort. Br J Haematol 2022;197:602–8.
- [32] Janke LJ, Van Driest SL, Portera MV, et al. Hypertension is a modifiable risk factor for osteonecrosis in acute lymphoblastic leukemia. Blood 2019;134:983–6.
- [33] Girard P, Auquier P, Barlogis V, et al. Symptomatic osteonecrosis in childhood leukemia survivors: prevalence, risk factors and impact on quality of life in adulthood. Haematologica 2013;98:1089–97.
- [34] Sakamoto K, Imamura T, Kihira K, et al. Low incidence of osteonecrosis in childhood acute lymphoblastic leukemia treated with ALL-97 and ALL-02 study of Japan Association of Childhood Leukemia Study Group. J Clin Oncol 2018;36:900–7.
- [35] Mogensen SS, Harila-Saari A, Mäkitie O, et al. Comparing osteonecrosis clinical phenotype, timing, and risk factors in children and young adults treated for acute lymphoblastic leukemia. Pediatr Blood Cancer 2018;65:e27300.
- [36] Brandalise SR, Assis R, Laranjeira ABA, et al. Low-dose methotrexate in sickle-cell disease: a pilot study with rationale borrowed from rheumatoid arthritis. Exp Hematol Oncol 2017;6:18.
- [37] Hernigou P, Rigoulot G, Auregan JC, et al. Unusual indication of Cell therapy for hip osteonecrosis after pregnancy. SICOT J 2018;4:46.
- [38] Hernigou J, Bastard C, Dubory A, et al. Cell therapy for posttraumatic shoulder osteonecrosis. Morphologie 2021;105:162–9.
- [39] Hernigou J, Housset V, Dubory A, et al. Cell therapy for post-traumatic hip osteonecrosis in young patients. Morphologie 2021;105:127–33.
- [40] Bergman J, Nordström A, Nordström P. Epidemiology of osteonecrosis among older adults in Sweden. Osteoporos Int 2019;30:965–73.
- [41] Bittner J, Hartstein A. Spontaneous osteonecrosis of the knee. J Orthop Sports Phys Ther 2018;48:824.
- [42] Ismail T, Osinga R, Todorov A Jr, et al. Engineered, axially-vascularized osteogenic grafts from human adipose-derived cells to treat avascular necrosis of bone in a rat model. Acta Biomater 2017;63:236–45.
- [43] Yoon BH, Kim TY, Shin IS, et al. Alcohol intake and the risk of osteonecrosis of the femoral head in Japanese populations: a doseresponse meta-analysis of case-control studies. Clin Rheumatol 2017; 36:2517–24.
- [44] Fukushima W, Yamamoto T, Takahashi S, et al. The effect of alcohol intake and the use of oral corticosteroids on the risk of idiopathic osteonecrosis of the femoral head: a case-control study in Japan. Bone Joint J 2013;95-b:320–5.
- [45] Chen CY, Du W, Rao SS, et al. Extracellular vesicles from human urinederived stem cells inhibit glucocorticoid-induced osteonecrosis of the femoral head by transporting and releasing pro-angiogenic DMBT1 and anti-apoptotic TIMP1. Acta Biomater 2020;111:208–20.
- [46] Zheng Y, Zheng Z, Zhang K, et al. Osteonecrosis in systemic lupus erythematosus: Systematic insight from the epidemiology, pathogenesis, diagnosis and management. Autoimmun Rev 2022;21:102992.
- [47] Bakhshi Z, Yadav S, Harmsen WS, et al. Osteonecrosis in inflammatory bowel disease: clinical features, risk factor analysis, and outcomes. Inflamm Bowel Dis 2023;29:1223–30.
- [48] Borchmann S, Müller H, Haverkamp H, et al. Symptomatic osteonecrosis as a treatment complication in Hodgkin lymphoma: an analysis of the German Hodgkin Study Group (GHSG). Leukemia 2019;33:439–46.
- [49] Messer JG, Jiron JM, Mendieta Calle JL, et al. Zoledronate treatment duration is linked to bisphosphonate-related osteonecrosis of the jaw prevalence in rice rats with generalized periodontitis. Oral Dis 2019;25: 1116–35.
- [50] Felten R, Perrin P, Caillard S, et al. Avascular osteonecrosis in kidney transplant recipients: Risk factors in a recent cohort study and evaluation of the role of secondary hyperparathyroidism. PLoS One 2019;14: e0212931.
- [51] Lai TY, Wang TH, Liu CJ, et al. Risk factors for osteonecrosis of the jaw in oral cancer patients after surgery and eventual adjuvant treatment: the potential role of chemotherapy. Radiother Oncol 2017;123:406–11.
- [52] Lai S-W, Lin C-L, Liao K-F. Real-world database examining the association between avascular necrosis of the femoral head and diabetes in Taiwan. Diabetes Care 2019;42:39–43.
- [53] Fujieda Y, Doi M, Asaka T, et al. Incidence and risk of antiresorptive agent-related osteonecrosis of the jaw (ARONJ) after tooth extraction in patients with autoimmune disease. J Bone Miner Metab 2020;38:581–8.
- [54] Barasch A, Cunha-Cruz J, Curro F, et al. Dental risk factors for osteonecrosis of the jaws: a CONDOR case-control study. Clin Oral Investig 2013;17:1839–45.
- [55] Kwoen MJ, Park JH, Kim KS, et al. Association between periodontal disease, tooth extraction, and medication-related osteonecrosis of the jaw in women receiving bisphosphonates: a national cohort-based study. J Periodontol 2023;94:98–107.
- [56] Bracchi P, Zecca E, Brunelli C, et al. A real-world study on the prevalence and risk factors of medication related osteonecrosis of the jaw in cancer patients with bone metastases treated with Denosumab. Cancer Med 2023;12:18317-26.
- [57] Kizub DA, Miao J, Schubert MM, et al. Risk factors for bisphosphonate-associated osteonecrosis of the jaw in the prospective randomized trial of adjuvant bisphosphonates for early-stage breast cancer (SWOG 0307). Support Care Cancer 2021;29:2509–17.
- [58] Lim SJ, Yeo I, Park CW, et al. Risk factors for osteonecrosis of the femoral head in brain tumor patients receiving corticosteroid after surgery. PLoS One 2020;15:e0238368.
- [59] Williams DW, Ho K, Lenon A, et al. Long-term ligature-induced periodontitis exacerbates development of bisphosphonate-related osteonecrosis of the jaw in mice. J Bone Miner Res 2022;37:1400–10.
- [60] Soutome S, Otsuru M, Hayashida S, et al. Relationship between tooth extraction and development of medication-related osteonecrosis of the jaw in cancer patients. Sci Rep 2021;11:17226.
- [61] Zhang Q, Yu W, Lee S, et al. Bisphosphonate induces osteonecrosis of the jaw in diabetic mice via NLRP3/caspase-1-dependent IL-1β mechanism. J Bone Miner Res 2015;30:2300–12.
- [62] Kim SH, Lee YK, Kim TY, et al. Incidence of and risk for osteonecrosis of the jaw in Korean osteoporosis patients treated with bisphosphonates: a nationwide cohort-study. Bone 2021;143:115650.
- [63] Van Poznak C, Reynolds EL, Estilo CL, et al. Osteonecrosis of the jaw risk factors in bisphosphonate-treated patients with metastatic cancer. Oral Dis 2022;28:193–201.
- [64] Kuroshima S, Sasaki M, Nakajima K, et al. Transplantation of noncultured stromal vascular fraction cells of adipose tissue ameliorates osteonecrosis of the jaw-like lesions in mice. J Bone Miner Res 2018;33: 154–66.
- [65] Di Fede O, Fusco V, Matranga D, et al. Osteonecrosis of the jaws in patients assuming oral bisphosphonates for osteoporosis: a retrospective multi-hospital-based study of 87 Italian cases. Eur J Intern Med 2013;24:784–90.
- [66] Everts-Graber J, Lehmann D, Burkard JP, et al. Risk of osteonecrosis of the jaw under denosumab compared to bisphosphonates in patients with osteoporosis. J Bone Miner Res 2022;37:340–8.
- [67] Nordström P, Bergman J, Ballin M, et al. Bone-specific drugs and osteonecrosis of sites other than the jaw: a nationwide cohort study. J Bone Miner Res 2020;35:1703–10.
- [68] Liu W, Zhao Z, Na Y, et al. Dexamethasone-induced production of reactive oxygen species promotes apoptosis via endoplasmic reticulum stress and autophagy in MC3T3-E1 cells. Int J Mol Med 2018;41: 2028–36.
- [69] Xu WN, Zheng HL, Yang RZ, et al. HIF-1α regulates glucocorticoidinduced osteoporosis through PDK1/AKT/mTOR signaling pathway. Front Endocrinol (Lausanne) 2019;10:922.
- [70] Deng S, Dai G, Chen S, et al. Dexamethasone induces osteoblast apoptosis through ROS-PI3K/AKT/GSK3β signaling pathway. Biomed Pharmacother 2019;110:602–8.
- [71] Nie Z, Chen S, Deng S, et al. Gene expression profiling of osteoblasts subjected to dexamethasone-induced apoptosis with/without GSK3βshRNA. Biochem Biophys Res Commun 2018;506:41–7.
- [72] Feng Z, Zheng W, Tang Q, et al. Fludarabine inhibits STAT1-mediated up-regulation of caspase-3 expression in dexamethasone-induced osteoblasts apoptosis and slows the progression of steroid-induced avascular necrosis of the femoral head in rats. Apoptosis 2017;22:1001–12.
- [73] Cui Y, Zhang W, Yang P, et al. Menaquinone-4 prevents medicationrelated osteonecrosis of the jaw through the SIRT1 signaling-mediated inhibition of cellular metabolic stresses-induced osteoblast apoptosis. Free Radic Biol Med 2023;206:33–49.
- [74] Yang N, Sun H, Xue Y, et al. Inhibition of MAGL activates the Keap1/ Nrf2 pathway to attenuate glucocorticoid-induced osteonecrosis of the femoral head. Clin Transl Med 2021;11:e447.
- [75] Zhang Z, Song Y, Wang SI, et al. Osteoblasts/osteocytes sirtuin6 is vital to preventing ischemic osteonecrosis through targeting VDR-RANKL signaling. J Bone Miner Res 2021;36:579–90.
- [76] Kato T, Khanh VC, Sato K, et al. Elevated expression of Dkk-1 by glucocorticoid treatment impairs bone regenerative capacity of adipose tissue-derived mesenchymal stem cells. Stem Cells Dev 2018;27:85–99.
- [77] Weinstein RS, Jilka RL, Parfitt AM, et al. Inhibition of osteoblastogenesis and promotion of apoptosis of osteoblasts and osteocytes by glucocorticoids. Potential mechanisms of their deleterious effects on bone. J Clin Invest 1998;102:274–82.
- [78] Sun F, Zhou JL, Liu ZL, et al. Dexamethasone induces ferroptosis via P53/SLC7A11/GPX4 pathway in glucocorticoid-induced osteonecrosis of the femoral head. Biochem Biophys Res Commun 2022;602:149–55.
- [79] Gross C, Weber M, Creutzburg K, et al. Osteoclast profile of medication-related osteonecrosis of the jaw secondary to bisphosphonate therapy: a comparison with osteoradionecrosis and osteomyelitis. J Transl Med 2017;15:128.
- [80] Wehrhan F, Gross C, Creutzburg K, et al. Osteoclastic expression of higher-level regulators NFATc1 and BCL6 in medication-related osteonecrosis of the jaw secondary to bisphosphonate therapy: a comparison with osteoradionecrosis and osteomyelitis. J Transl Med 2019; 17:69.
- [81] Zhi X, Fang C, Gu Y, et al. Guaiacol suppresses osteoclastogenesis by blocking interactions of RANK with TRAF6 and C-Src and inhibiting NF-κB, MAPK and AKT pathways. J Cell Mol Med 2020;24:5122–34.
- [82] He MC, Zhang J, Chen XJ, et al. Osteoclastic activity was associated with the development of steroid-induced osteonecrosis of femoral head. Artif Cells Nanomed Biotechnol 2020;48:1036–46.
- [83] Rajakumar SA, Papp E, Lee KK, et al. B cell acute lymphoblastic leukemia cells mediate RANK-RANKL-dependent bone destruction. Sci Transl Med 2020;122020/09/18. doi:10.1126/scitranslmed.aba5942
- [84] Liu Y, Shan H, Zong Y, et al. IKKe in osteoclast inhibits the progression of methylprednisolone-induced osteonecrosis. Int J Biol Sci 2021;17: 1353–60.
- [85] Kalkar P, Cohen G, Tamari T, et al. IFN-β mediates the anti-osteoclastic effect of bisphosphonates and dexamethasone. Front Pharmacol 2022; 13:1002550.
- [86] Du J, Liu Y, Wu X, et al. BRD9-mediated chromatin remodeling suppresses osteoclastogenesis through negative feedback mechanism. Nat Commun 2023;14:1413.
- [87] Fowler TW, Acevedo C, Mazur CM, et al. Glucocorticoid suppression of osteocyte perilacunar remodeling is associated with subchondral bone degeneration in osteonecrosis. Sci Rep 2017;7:44618.
- [88] Wang XY, Ma TL, Chen KN, et al. Accumulation of LDL/ox-LDL in the necrotic region participates in osteonecrosis of the femoral head: a pathological and in vitro study. Lipids Health Dis 2021;20:167.
- [89] Kim HJ, Kim HJ, Choi Y, et al. Zoledronate enhances osteocyte-mediated osteoclast differentiation by IL-6/RANKL axis. Int J Mol Sci 2019; 20:1467.
- [90] Kuang MJ, Huang Y, Zhao XG, et al. Exosomes derived from Wharton's jelly of human umbilical cord mesenchymal stem cells reduce osteocyte apoptosis in glucocorticoid-induced osteonecrosis of the femoral head in rats via the miR-21-PTEN-AKT signalling pathway. Int J Biol Sci 2019;15:1861–71.
- [91] Zhao J, Ma XL, Ma JX, et al. TET3 mediates alterations in the epigenetic marker 5hmC and Akt pathway in steroid-associated osteonecrosis. J Bone Miner Res 2017;32:319–32.
- [92] Wang XY, Gong LJ, Huang JM, et al. Pinocembrin alleviates glucocorticoid-induced apoptosis by activating autophagy via suppressing the PI3K/Akt/mTOR pathway in osteocytes. Eur J Pharmacol 2020;880: 173212.
- [93] Shimasaki M, Ueda S, Ichiseki T, et al. Resistance of bone marrow mesenchymal stem cells in a stressed environment—comparison with osteocyte cells. Int J Med Sci 2021;18:1375–81.
- [94] Andreev D, Liu M, Weidner D, et al. Osteocyte necrosis triggers osteoclast-mediated bone loss through macrophage-inducible C-type lectin. J Clin Invest 2020;130:4811–30.
- [95] Fang S, Li Y, Chen P. Osteogenic effect of bone marrow mesenchymal stem cell-derived exosomes on steroid-induced osteonecrosis of the femoral head. Drug Des Devel Ther 2019;13:45–55.
- [96] Houdek MT, Wyles CC, Packard BD, et al. Decreased osteogenic activity of mesenchymal stem cells in patients with corticosteroidinduced osteonecrosis of the femoral head. J Arthroplasty 2016;31: 893–8.
- [97] Chen L-S, Zhang M, Chen P, et al. The m6A demethylase FTO promotes the osteogenesis of mesenchymal stem cells by downregulating PPARG. Acta Pharmacol Sin 2022;43:1311–23.
- [98] Han L, Gong S, Wang R, et al. Knockdown of POSTN inhibits osteogenic differentiation of mesenchymal stem cells from patients with steroid-induced osteonecrosis. Front Cell Dev Biol 2020;8:606289.
- [99] Zhun W, Donghai L, Zhouyuan Y, et al. Efficiency of cell therapy to GC-induced ONFH: BMSCs with Dkk-1 interference is not superior to unmodified BMSCs. Stem Cells Int 2018;2018:1340252.
- [100] Li D, Yang Z, Luo Y, et al. Delivery of MiR335-5p-pendant tetrahedron DNA nanostructures using an injectable heparin lithium hydrogel for challenging bone defects in steroid-associated osteonecrosis. Adv Healthc Mater 2022;11:e2101412.
- [101] Zhang F, Peng W, Zhang J, et al. P53 and Parkin co-regulate mitophagy in bone marrow mesenchymal stem cells to promote the repair of early steroid-induced osteonecrosis of the femoral head. Cell Death Dis 2020; 11.42
- [102] Chen L, Wang BZ, Xie J, et al. Therapeutic effect of SIRT3 on glucocorticoid-induced osteonecrosis of the femoral head via intracellular oxidative suppression. Free Radic Biol Med 2021;176:228–40.
- [103] Kim BS, Yang SS, Kim CS, et al. Zoledronate suppresses VEGF-induced capillary tube formation and inhibits expression of interferon‑induced transmembrane protein-1 in human umbilical vein endothelial cells. Int J Mol Med 2018;41:2879–84.
- [104] Li B, Lei Y, Hu Q, et al. Porous copper- and lithium-doped nanohydroxyapatite composite scaffold promotes angiogenesis and bone regeneration in the repair of glucocorticoids-induced osteonecrosis of the femoral head. Biomed Mater 2021;16. doi:10.1088/1748-605X/ ac246e
- [105] Lin RLC, Sung PH, Wu CT, et al. Decreased ankyrin expression is associated with repressed eNOS Signaling, cell proliferation, and osteogenic differentiation in osteonecrosis of the femoral head. J Bone Joint Surg Am 2022;104:2–12.
- [106] Yao X, Yu S, Jing X, et al. PTEN inhibitor VO-OHpic attenuates GCassociated endothelial progenitor cell dysfunction and osteonecrosis of the femoral head via activating Nrf2 signaling and inhibiting mitochondrial apoptosis pathway. Stem Cell Res Ther 2020;11:140.
- [107] Tuckermann J, Adams RH. The endothelium-bone axis in development, homeostasis and bone and joint disease. Nat Rev Rheumatol 2021;17: 608–20.
- [108] Zhou Z, Lin Y, Pan C, et al. IL-15 deficiency alleviates steroid-induced osteonecrosis of the femoral head by impact osteoclasts via RANKL-RANK-OPG system. Immun Ageing 2020;17:19.
- [109] Wang F, Min HS, Shan H, et al. IL-34 aggravates steroid-induced osteonecrosis of the femoral head via promoting osteoclast differentiation. Immune Netw 2022;22:e25.
- [110] Tan Z, Wang Y, Chen Y, et al. The dynamic feature of macrophage M1/ M2 imbalance facilitates the progression of non-traumatic osteonecrosis of the femoral head. Front Bioeng Biotechnol 2022;10:912133.
- [111] Weber M, Homm A, Müller S, et al. Zoledronate causes a systemic shift of macrophage polarization towards M1 in vivo. Int J Mol Sci 2021;22: 1323.
- [112] Pushalkar S, Li X, Kurago Z, et al. Oral microbiota and host innate immune response in bisphosphonate-related osteonecrosis of the jaw. Int J Oral Sci 2014;6:219–26.
- [113] Elsayed R, Kurago Z, Cutler CW, et al. Role of dendritic cell-mediated immune response in oral homeostasis: a new mechanism of osteonecrosis of the jaw. Faseb j 2020;34:2595–608.
- [114] Kaur K, Kanayama K, Wu QQ, et al. Zoledronic acid mediated differential activation of NK cells in different organs of WT and Rag2(-/-) mice; stark differences between the bone marrow and gingivae. Cell Immunol 2022;375:104526.
- [115] Kaur K, Sun Y, Kanayama K, et al. Augmentation of IFN- γ by bone marrow derived immune cells in the presence of severe suppression of IFN-γ in gingivae induced by zoledronic acid and denosumab in Hu-BLT mice model of ONJ. Front Endocrinol (Lausanne) 2023;14: 1111627.
- [116] Kuiper JW, Forster C, Sun C, et al. Zoledronate and pamidronate depress neutrophil functions and survival in mice. Br J Pharmacol 2012; 165:532–9.
- [117] Ying J, Wang P, Ding Q, et al. Peripheral blood stem cell therapy does not improve outcomes of femoral head osteonecrosis with cap-shaped separated cartilage defect. J Orthop Res 2020;38:269–76.
- [118] Zheng LW, Wang WC, Mao XZ, et al. TNF-α regulates the early development of avascular necrosis of the femoral head by mediating osteoblast autophagy and apoptosis via the p38 MAPK/NF-κB signaling pathway. Cell Biol Int 2020;44:1881–9.
- [119] Fang B, Wang D, Zheng J, et al. Involvement of tumor necrosis factor alpha in steroid-associated osteonecrosis of the femoral head: friend or foe? Stem Cell Res Ther 2019;10:5.
- [120] Geng W, Zhang W, Ma J. IL-9 exhibits elevated expression in osteonecrosis of femoral head patients and promotes cartilage degradation through activation of JAK-STAT signaling in vitro. Int Immunopharmacol 2018;60:228–34.
- [121] Sadile F, Bernasconi A, Carbone F, et al. Histological fibrosis may predict the failure of core decompression in the treatment of osteonecrosis of the femoral head. Int J Surg 2017;44:303–8.
- [122] Deng Z, Ren Y, Park MS, et al. Damage associated molecular patterns in necrotic femoral head inhibit osteogenesis and promote fibrogenesis of mesenchymal stem cells. Bone 2022;154:116215.
- [123] Chen D, Zhang G, Li Y, et al. Up-regulation of urinary exosomal hsamicroRNA-200b-3p and hsa-microRNA-206 in patients of steroidinduced osteonecrosis of femoral head. Am J Transl Res 2021;13: 7574–90.
- [124] Shen X, Zhu W, Zhang P, et al. Macrophage miR-149-5p induction is a key driver and therapeutic target for BRONJ. JCI Insight 2022;7: e159865.
- [125] Hao C, Yang S, Xu W, et al. MiR-708 promotes steroid-induced osteonecrosis of femoral head, suppresses osteogenic differentiation by targeting SMAD3. Sci Rep 2016;6:22599.
- [126] Tian L, Sun S, Li W, et al. Down-regulated microRNA-141 facilitates osteoblast activity and inhibits osteoclast activity to ameliorate osteonecrosis of the femoral head via up-regulating TGF-β2. Cell Cycle 2020; 19:772–86.
- [127] Jia J, Feng X, Xu W, et al. MiR-17-5p modulates osteoblastic differentiation and cell proliferation by targeting SMAD7 in non-traumatic osteonecrosis. Exp Mol Med 2014;46:e107.
- [128] Fang S, Liu Z, Wu S, et al. Pro-angiognetic and pro-osteogenic effects of human umbilical cord mesenchymal stem cell-derived exosomal miR-21-5p in osteonecrosis of the femoral head. Cell Death Discov 2022;8: 226.
- [129] Liang JQ, Zhou ZT, Bo L, et al. Phosphoglycerate kinase 1 silencing by a novel microRNA microRNA-4523 protects human osteoblasts from dexamethasone through activation of Nrf2 signaling cascade. Cell Death Dis 2021;12:964.
- [130] Liao W, Ning Y, Xu HJ, et al. BMSC-derived exosomes carrying microRNA-122-5p promote proliferation of osteoblasts in osteonecrosis of the femoral head. Clin Sci (Lond) 2019;133:1955–75.
- [131] Li G, Liu H, Zhang X, et al. The protective effects of microRNA-26a in steroid-induced osteonecrosis of the femoral head by repressing EZH2. Cell Cycle 2020;19:551–66.
- [132] Fang S, He T, Jiang J, et al. Osteogenic effect of tsRNA-10277-loaded exosome derived from bone mesenchymal stem cells on steroid-induced osteonecrosis of the femoral head. Drug Des Devel Ther 2020;14: 4579–91.
- [133] Vezzani G, Quartesan S, Cancellara P, et al. Hyperbaric oxygen therapy modulates serum OPG/RANKL in femoral head necrosis patients. J Enzyme Inhib Med Chem 2017;32:707–11.
- [134] Wang CJ, Huang CC, Yip HK, et al. Dosage effects of extracorporeal shockwave therapy in early hip necrosis. Int J Surg 2016;35:179–86.
- [135] An IX, Wu GP, Niu K, *et al*. Treatment of femoral head osteonecrosis with ozone therapy: pilot trial of a new therapeutic approach. Pain Physician 2022;25:E43–e54.
- [136] Marcheggiani Muccioli GM, Grassi A, Setti S, et al. Conservative treatment of spontaneous osteonecrosis of the knee in the early stage: pulsed electromagnetic fields therapy. Eur J Radiol 2013;82:530–7.
- [137] Atoun E, Mor A, Segal G, et al. A non-invasive, home-based biomechanical therapy for patients with spontaneous osteonecrosis of the knee. J Orthop Surg Res 2016;11:139.
- [138] Hernigou P, Auregan JC, Dubory A, et al. Ankle osteonecrosis in fiftyone children and adolescent's leukemia survivors: a prospective randomized study on percutaneous mesenchymal stem cells treatment. Int Orthop 2021;45:2383–93.
- [139] de Rojas T, Martinez-Alvarez S, Lerma-Lara S, et al. Outcome of childhood leukaemia survivors and necrosis of the femoral head treated with autologous mesenchymal stem cells. Clin Transl Oncol 2018;20: 584–90.
- [140] Hernigou P, Auregan JC, Dubory A, et al. Subchondral stem cell therapy versus contralateral total knee arthroplasty for osteoarthritis following secondary osteonecrosis of the knee. Int Orthop 2018;42:2563–71.
- [141] Pan J, Ding Q, Lv S, et al. Prognosis after autologous peripheral blood stem cell transplantation for osteonecrosis of the femoral head in the pre-collapse stage: a retrospective cohort study. Stem Cell Res Ther 2020;11:83.
- [142] Chen C, Qu Z, Yin X, et al. Efficacy of umbilical cord-derived mesenchymal stem cell-based therapy for osteonecrosis of the femoral head: a three-year follow-up study. Mol Med Rep 2016;14:4209–15.
- [143] Mao Q, Jin H, Liao F, et al. The efficacy of targeted intraarterial delivery of concentrated autologous bone marrow containing mononuclear cells in the treatment of osteonecrosis of the femoral head: a five year followup study. Bone 2013;57:509–16.
- [144] Meier C, Kraenzlin C, Friederich NF, et al. Effect of ibandronate on spontaneous osteonecrosis of the knee: a randomized, double-blind, placebo-controlled trial. Osteoporos Int 2014;25:359–66.
- [145] Pichardo SEC, Ten Broek FW, Richard van Merkesteyn JP. Treatment of pathologic fractures of the mandible in stage III medication-related osteonecrosis of the jaw-an observational study. J Craniomaxillofac Surg 2018;46:1241–6.
- [146] Sim IW, Borromeo GL, Tsao C, et al. Teriparatide promotes bone healing in medication-related osteonecrosis of the jaw: a placebo-controlled, randomized trial. J Clin Oncol 2020;38:2971–80.
- [147] Jung J, Yoo HY, Kim GT, et al. Short-term teriparatide and recombinant human bone morphogenetic protein-2 for regenerative approach to medication-related osteonecrosis of the jaw: a preliminary study. J Bone Miner Res 2017;32:2445–52.
- [148] Laimer J, Steinmassl O, Hechenberger M, et al. Intraoral vacuumassisted closure therapy—a pilot study in medication-related osteonecrosis of the jaw. J Oral Maxillofac Surg 2017;75:2154–61.
- [149] Yue Ja, Guo X, Wang R, et al. Single approach to double-channel core decompression and bone grafting with structural bone support for treating osteonecrosis of the femoral head in different stages. J Orthop Surg Res 2020;15:198.
- [150] Wang Q, Li D, Yang Z, et al. Femoral head and neck fenestration through a direct anterior approach combined with compacted autograft for the treatment of early stage nontraumatic osteonecrosis of the femoral head: a retrospective study. J Arthroplasty 2020;35:652–60.
- [151] Civinini R, Capone A, Carulli C, et al. The kinetics of remodeling of a calcium sulfate/calcium phosphate bioceramic. J Mater Sci Mater Med 2017;28:137.
- [152] Landgraeber S, Warwas S, Claßen T, et al. Modifications to advanced Core decompression for treatment of Avascular necrosis of the femoral head. BMC Musculoskelet Disord 2017;18:479.
- [153] Fontecha CG, Roca I, Barber I, et al. Femoral head bone viability after free vascularized fibular grafting for osteonecrosis: SPECT/CT study. Microsurgery 2016;36:573–7.
- [154] Cao L, Guo C, Chen J, et al. Free vascularized fibular grafting improves vascularity compared with core decompression in femoral head osteonecrosis: a randomized clinical trial. Clin Orthop Relat Res 2017;475: 2230–40.
- [155] Chen XT, Zhu YJ, Liu YW, et al. Metal trabecular bone reconstruction system better improves clinical efficacy and biomechanical repair of osteonecrosis of the femoral head than free vascularized fibular graft: a case-control study. J Cell Physiol 2019;234:20957–68.
- [156] Sun J, Li Z, Liu S, et al. Biodegradable magnesium screw, titanium screw and direct embedding fixation in pedicled vascularized iliac bone graft transfer for osteonecrosis of the femoral head: a randomized controlled study. J Orthop Surg Res 2023;18:523.
- [157] Zhang Y, Wang X, Jiang C, et al. Biomechanical research of medial femoral circumflex vascularized bone-grafting in the treatment of earlyto-mid osteonecrosis of the femoral head: a finite element analysis. J Orthop Surg Res 2022;17:441.
- [158] Aoyama T, Goto K, Kakinoki R, et al. An exploratory clinical trial for idiopathic osteonecrosis of femoral head by cultured autologous multipotent mesenchymal stromal cells augmented with vascularized bone grafts. Tissue Eng Part B Rev 2014;20:233–42.
- [159] Zhao D, Huang S, Lu F, et al. Vascularized bone grafting fixed by biodegradable magnesium screw for treating osteonecrosis of the femoral head. Biomaterials 2016;81:84–92.
- [160] Kang JS, Moon KH, Kim BS, et al. Clinical results of auto-iliac cancellous bone grafts combined with implantation of autologous bone marrow cells for osteonecrosis of the femoral head: a minimum 5-year follow-up. Yonsei Med J 2013;54:510–5.
- [161] Windisch C, Kolb W, Röhner E, et al. Invasive electromagnetic field treatment in osteonecrosis of the femoral head: a prospective cohort study. Open Orthop J 2014;8:125–9.
- [162] Morita D, Hasegawa Y, Okura T, et al. Long-term outcomes of transtrochanteric rotational osteotomy for non-traumatic osteonecrosis of the femoral head. Bone Joint J 2017;99-B:175–83.
- [163] Utsunomiya T, Motomura G, Ikemura S, et al. The results of total hip arthroplasty after sugioka transtrochanteric anterior rotational osteotomy for osteonecrosis. J Arthroplasty 2017;32:2768–73.
- [164] Osawa Y, Seki T, Okura T, et al. Curved intertrochanteric varus osteotomy vs total hip arthroplasty for osteonecrosis of the femoral head in patients under 50 years old. J Arthroplasty 2020;35:1600–5.
- [165] Osawa Y, Seki T, Okura T, et al. Long-term outcomes of curved intertrochanteric varus osteotomy combined with bone impaction grafting for non-traumatic osteonecrosis of the femoral head. Bone Joint J 2021;103-B:665–71.
- [166] Miladi M, Villain B, Mebtouche N, et al. Interest of short implants in hip arthroplasty for osteonecrosis of the femoral head: comparative study "uncemented short" vs "cemented conventional" femoral stems. Int Orthop 2018;42:1669–74.
- [167] Kim SC, Lim YW, Jo WL, et al. Long-term results of total hip arthroplasty in young patients with osteonecrosis after allogeneic bone marrow transplantation for hematological disease: a multicenter, propensity-matched cohort study with a mean 11-year follow-up. J Arthroplasty 2021;36:1049–54.
- [168] Voss PJ, Matsumoto A, Alvarado E, et al. Treatment of stage II medication-related osteonecrosis of the jaw with necrosectomy and autologous bone marrow mesenchymal stem cells. Odontology 2017;105: 484–93.
- [169] Calvani F, Cutone A, Lepanto MS, et al. Efficacy of bovine lactoferrin in the post-surgical treatment of patients suffering from bisphosphonaterelated osteonecrosis of the jaws: an open-label study. Biometals 2018; 31:445–55.
- [170] Mao Q, Wang W, Xu T, et al. Combination treatment of biomechanical support and targeted intra-arterial infusion of peripheral blood stem cells mobilized by granulocyte-colony stimulating factor for the osteonecrosis of the femoral head: a randomized controlled clinical trial. J Bone Miner Res 2015;30:647–56.
- [171] Hauzeur JP, De Maertelaer V, Baudoux E, et al. Inefficacy of autologous bone marrow concentrate in stage three osteonecrosis: a randomized controlled double-blind trial. Int Orthop 2018;42:1429–35.
- [172] Boontanapibul K, Huddleston JI, Amanatullah DF, et al. Modified Kerboul angle predicts outcome of core decompression with or without additional cell therapy. J Arthroplasty 2021;36:1879–86.
- [173] Hernigou P, Hernigou J, Scarlat M. Mesenchymal stem cell therapy improved outcome of early post-traumatic shoulder osteonecrosis: a prospective randomized clinical study of fifty patients with over ten year follow-up. Int Orthop 2021;45:2643–52.
- [174] Daltro GC, Fortuna V, de Souza ES, et al. Efficacy of autologous stem cell-based therapy for osteonecrosis of the femoral head in sickle cell disease: a five-year follow-up study. Stem Cell Res Ther 2015;6:110.
- [175] Lim YW, Kim YS, Lee JW, et al. Stem cell implantation for osteonecrosis of the femoral head. Exp Mol Med 2013;45:e61.
- [176] Blanco JF, Garcia-Garcia FJ, Villaron EM, et al. Long-term results of a phase I/II clinical trial of autologous mesenchymal stem cell therapy for femoral head osteonecrosis. J Clin Med 2023;12:2117.
- [177] Mardones R, Camacho D, Monsalvo F, et al. Treatment of osteonecrosis of the femoral head by core decompression and implantation of fully functional ex vivo-expanded bone marrow-derived mesenchymal stem cells: a proof-of-concept study. Stem Cells Cloning 2019;12:11–6.
- [178] Hernigou P, Dubory A, Homma Y, et al. Cell therapy versus simultaneous contralateral decompression in symptomatic corticosteroid osteonecrosis: a thirty year follow-up prospective randomized study of one hundred and twenty five adult patients. Int Orthop 2018;42: 1639–49.
- [179] Hernigou P, Dubory A, Flouzat Lachaniette CH, et al. Stem cell therapy in early post-traumatic talus osteonecrosis. Int Orthop 2018;42: $2949 - 56$
- [180] Wu ZY, Sun Q, Liu M, et al. Correlation between the efficacy of stem cell therapy for osteonecrosis of the femoral head and cell viability. BMC Musculoskelet Disord 2020;21:55.
- [181] Emadedin M, Karimi S, Karimi A, et al. Autologous bone marrowderived CD133 cells with core decompression as a novel treatment method for femoral head osteonecrosis: a pilot study. Cytotherapy 2019;21:107–12.
- [182] Aoyama T, Fujita Y, Madoba K, et al. Rehabilitation program after mesenchymal stromal cell transplantation augmented by vascularized bone grafts for idiopathic osteonecrosis of the femoral head: a preliminary study. Arch Phys Med Rehabil 2015;96:532–9.
- [183] Jayankura M, Thomas T, Seefried L, et al. Does adjunction of

autologous osteoblastic cells improve the results of core decompression in early-stage femoral head osteonecrosis? A double-blind, randomized trial. Clin Orthop Relat Res 2023;481:1527–40.

- [184] Hauzeur JP, Lechanteur C, Baudoux E, et al. Did osteoblastic cell therapy improve the prognosis of pre-fracture osteonecrosis of the femoral head? A randomized, controlled trial. Clin Orthop Relat Res 2020;478:1307–15.
- [185] Hernandez A, Nunez JH, Sallent A, et al. Core decompression combined with implantation of autologous bone marrow concentrate with tricalcium phosphate does not prevent radiographic progression in early stage osteonecrosis of the hip. Clin Orthop Surg 2020;12:151–7.
- [186] Kuroda Y, Asada R, So K, et al. A pilot study of regenerative therapy using controlled release of recombinant human fibroblast growth factor for patients with pre-collapse osteonecrosis of the femoral head. Int Orthop 2016;40:1747–54.
- [187] Kuroda Y, Tanaka T, Miyagawa T, et al. Recombinant human FGF-2 for the treatment of early-stage osteonecrosis of the femoral head: TRION, a single-arm, multicenter, Phase II trial. Regen Med 2021;16:535–48.
- [188] Gao F, Sun W, Guo W, et al. Combined with bone marrow-derived cells and rhBMP-2 for osteonecrosis after femoral neck fractures in children and adolescents: a case series. Sci Rep 2016;6:30730.
- [189] Ragazzo M, Val M, Montagner G, et al. Human amniotic membrane: an improvement in the treatment of Medication-related osteonecrosis of the jaw (MRONJ)? A case-control study. Cell Tissue Bank 2022;23:129–41.
- [190] Odet S, Meyer C, Gaudet C, et al. Tips and tricks and clinical outcome of cryopreserved human amniotic membrane application for the management of medication-related osteonecrosis of the jaw (MRONJ): a pilot study. Front Bioeng Biotechnol 2022;10:936074.
- [191] Bouland C, Meuleman N, Widelec J, et al. Case reports of medication-related osteonecrosis of the jaw (MRONJ) treated with uncultured stromal vascular fraction and L-PRF. J Stomatol Oral Maxillofac Surg 2021;122:212–8.
- [192] Park JH, Kim JW, Kim SJ. Does the addition of bone morphogenetic protein 2 to platelet-rich fibrin improve healing after treatment for medication-related osteonecrosis of the jaw? J Oral Maxillofac Surg 2017;75:1176–84.
- [193] Yüce MO, Adalı E, Işık G. The effect of concentrated growth factor (CGF) in the surgical treatment of medication-related osteonecrosis of the jaw (MRONJ) in osteoporosis patients: a randomized controlled study. Clin Oral Investig 2021;25:4529–41.
- [194] Li Z, Wang Y, Xiao K, et al. Emerging role of exosomes in the joint diseases. Cell Physiol Biochem 2018;47:2008–17.
- [195] Li Y, Tang J, Hu Y, et al. A study of autologous stem cells therapy assisted regeneration of cartilage in avascular bone necrosis. Eur Rev Med Pharmacol Sci 2015;19:3833–7.
- [196] Weldy JM, Taghavi S. Stem cell therapy as an adjunct to surgery. Int J Surg (London, England) 2019;68:182.
- [197] McKinley KL, Longaker MT, Naik S. Emerging frontiers in regenerative medicine. Science 2023;380:796–8.
- [198] Liang J, Zhang XY, Zhen YF, et al. PGK1 depletion activates Nrf2 signaling to protect human osteoblasts from dexamethasone. Cell Death Dis 2019;10:888.
- [199] Wang C, Sun W, Ling S, et al. AAV-anti-miR-214 prevents collapse of the femoral head in osteonecrosis by regulating osteoblast and osteoclast activities. Mol Ther Nucleic Acids 2019;18:841–50.
- [200] Lai Y, Li Y, Cao H, et al. Osteogenic magnesium incorporated into PLGA/TCP porous scaffold by 3D printing for repairing challenging bone defect. Biomaterials 2019;197:207–19.
- [201] Rodríguez-Lozano FJ, Oñate-Sánchez R, Gonzálvez-García M, et al. Allogeneic bone marrow mesenchymal stem cell transplantation in tooth extractions sites ameliorates the incidence of osteonecrotic jaw-like lesions in zoledronic acid-treated rats. J Clin Med 2020;9: 1649.
- [202] Lai Y, Cao H, Wang X, et al. Porous composite scaffold incorporating osteogenic phytomolecule icariin for promoting skeletal regeneration in challenging osteonecrotic bone in rabbits. Biomaterials 2018;153:1–13.
- [203] Wang G, Li Y, Sun T, et al. BMSC affinity peptide-functionalized βtricalcium phosphate scaffolds promoting repair of osteonecrosis of the femoral head. J Orthop Surg Res 2019;14:204.
- [204] Paulo S, Laranjo M, Abrantes AM, et al. Synthetic calcium phosphate ceramics as a potential treatment for bisphosphonate-related osteonecrosis of the jaw. Materials (Basel) 2019;12:1840.
- [205] Ma C, Andre G, Edwards D, et al. A rat model of ischemic osteonecrosis for investigating local therapeutics using biomaterials. Acta Biomater 2021;132:260–71.
- [206] Wang Y, Mei X, Bian Y, et al. Magnesium-based layered double hydroxide nanosheets: a new bone repair material with unprecedented osteogenic differentiation performance. Nanoscale 2020;12:19075–82.
- [207] Kang P, Xie X, Tan Z, et al. Repairing defect and preventing collapse of femoral head in a steroid-induced osteonecrotic of femoral head animal model using strontium-doped calcium polyphosphate combined BM-MNCs. J Mater Sci Mater Med 2015;26:80.
- [208] Luo Y, Li D, Xie X, et al. Porous, lithium-doped calcium polyphosphate composite scaffolds containing vascular endothelial growth factor (VEGF)-loaded gelatin microspheres for treating glucocorticoid-induced osteonecrosis of the femoral head. Biomed Mater 2019;14:035013.
- [209] Zhang HX, Zhang XP, Xiao GY, et al. In vitro and in vivo evaluation of calcium phosphate composite scaffolds containing BMP-VEGF loaded PLGA microspheres for the treatment of avascular necrosis of the femoral head. Mater Sci Eng C Mater Biol Appl 2016;60:298–307.
- [210] Zhao Y, Li S, Feng M, et al. Effects of puerarin-loaded tetrahedral framework nucleic acids on osteonecrosis of the femoral head. Small (Weinheim an Der Bergstrasse, Germany) 2023;19:e2302326.
- [211] Zhao D, Xiao D, Liu M, et al. Tetrahedral framework nucleic acid carrying angiogenic peptide prevents bisphosphonate-related osteonecrosis of the jaw by promoting angiogenesis. Int J Oral Sci 2022;14:23.