Genetic Characterization of Human-Derived Hydatid Cysts of *Echinococcus granulosus* Sensu Lato in Heilongjiang Province and the First Report of G7 Genotype of *E. canadensis* in Humans in China



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Abstract

Cystic echinococcosis (CE) caused by the larval stage of *Echinococcus granulosus* sensu lato (s.l.) is one of the most important zoonotic parasitic diseases worldwide and 10 genotypes (G1–G10) have been reported. In China, almost all the epidemiological and genotyping studies of *E. granulosus s.l.* are from the west and northwest pasturing areas. However, in Heilongjiang Province of northeastern China, no molecular information is available on *E. granulosus s.l.* To understand and to speculate on possible transmission patterns of *E. granulosus s.l.*, we molecularly identified and genotyped 10 hydatid cysts from hepatic CE patients in Heilongjiang Province based on mitochondrial cytochrome c oxidase subunit 1 (*cox1*), cytochrome b (*cytb*) and NADH dehydrogenase subunit 1 (*nad1*) genes. Two genotypes were identified, G1 genotype (n = 6) and G7 genotype (n = 4). All the six G1 genotype isolates were identical to each other at the *cox1* locus; three and two different sequences were obtained at the *cytb* and *nad1* loci, respectively, with two *cytb* gene sequences not being described previously. G7 genotype isolates were identical to each other at the *cox1*, *cytb* and *nad1* loci; however, the *cytb* gene sequences from G1 and G7 genotypes might reflect endemic genetic characterizations. Pigs might be the main intermediate hosts of G7 genotype in our investigated area by homology analysis. The results will aid in making more effective control strategies for the prevention of transmission of *E. granulosus* s.l.

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Introduction

Cystic echinococcosis (CE, also known as hydatidosis or hydatid disease) caused by the larval stage of *Echinococcus granulosus* sensu lato (s.l.) is considered as one of the most important zoonotic parasitic diseases worldwide, particularly affecting pastoral and poor rural communities where people raise livestock in close contact with dogs [1]. The disease causes great economic loss in livestock through condemnation of infected organs (mainly liver). More importantly, CE can be responsible for a life-threatening infection in humans. Fatal cases of human CE have been reported with inoperable CE cysts of the brain and anaphylactic shock caused by rupture of liver hydatid cysts [2,3].

Humans become infected with CE by ingesting eggs released from dogs or other canids through direct contact with the animals and consumption of food and water contaminated with infected animal feces. Eggs hatch in small intestine and parasite larvae (hydatid cysts) can be found in almost any organ, with the liver being the most common. Usually, CE may develop silently over years and even decades until it surfaces with clinical signs or symptoms. Clinical symptoms are mainly related to the localization, size, and number of cysts. However, a recent study described that there seemed to be a relationship between the genotypes and the size of hydatid cysts, in which all the patients infected with G7 genotype showed smaller liver cysts than those infected with G1 genotype [3].

Molecular epidemiological data have identified 10 genotypes (G1–G10) within *E. granulosus s.l.* In China, to date, three genotypes have been found, G1, G3 and G6 genotypes in humans and G1 and G6 genotypes in animals, with G1 genotype being

Province	Human				Animal			Ref.
	No. of Samples	Genotype	đ		No. of samples	Genotype (host)		
		61	63	G6		61	G6	
Gansu	-	-			12	12 (sheep)		[4,5]
Ningxia	12	12			16	14 (sheep), 1 (goat), 1 (squirrel)		[5,6]
Qinghai	38	38			178	122 (sheep), 29 (yak), 21 (cattle), 4 (dog)	2 (goat)	[4,5,7–11]
Qinghai and Sichuan	20*	70			76*	57 (sheep), 19 (yak)		[12,13]
Sichuan	54	52	2		34	10 (sheep), 24 (yak)		[5,7,10,11,14,15]
Tibet					34	30 (sheep), 2 (yak)	2 (sheep)	[7,10,11]
Xinjiang	149	145		4	134	63 (sheep), 5 (cattle), 1 (camel), 59 (dog)	2 (cattle), 1 (camel), 3 (dog)	[4,13,16–20]

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predominant in both of them [4–20] (Table 1). Currently, G1–G3, G4, G5 and G6–10 genotypes are designated to *E. granulosus* sensu stricto (s.s.), *E. equinus*, *E. ortleppi* and *E. canadensis*, respectively [1,21].

CE is one of the major parasitic problems in humans in China. Human CE cases have been reported in 27 provinces, autonomous regions, and municipalities, with western and northwestern China being the main endemic areas [22,23]. Although Heilongjiang Province in the northeast of China is not a main endemic area of CE, the number of CE patients in hospitals has been increasing, especially in recent years since the first human CE case was reported in 1958 [24]. Dogs, pigs and sheep have been confirmed to be infected with E. granulosus s.l. [25,26]. In fact, pigs, sheep and cattle are the main economic animals, and domestic dogs are kept in large numbers by local inhabitants to guard property and livestock with the development of livestock husbandry. Thus, people have a large risk of CE infection by farming activities and home-slaughtering practices. Unhealthy life habits also increase the possibility of human infection by these pathogens via eating uncooked vegetables and drinking raw water. However, no genotyping data are available about E. granulosus s.l. isolates in this area.

Mitochondrial DNA (mtDNA) is reported to be more powerful than nuclear DNA within *E. granulosus s.l.* in constructing phylogenetic relationships among closely related species because of its rapid sequence evolution [27]. Large datasets derived from mitochondrial genomes also have the potential to resolve problematic issues in *Echinococcus* taxonomy [13,28]. The aims of the present study were to molecularly identify and genotype hydatid cysts of CE patients residing in Heilongjiang Province by sequencing and analyzing mitochondrial cytochrome c oxidase subunit I (*cox1*), cytochrome b (*cytb*) and NADH dehydrogenase subunit 1 (*nad1*) genes, to speculate on possible transmission patterns of this cestode by homology analysis, and to understand the phylogeny of genotypes of *E. granulosus s.l.* by constructing neighbor-joining trees at three loci above. The results will contribute to developing control strategies of CE in our area.

Materials and Methods

Ethics statement

This research study was approved by the Medical Ethics Review Committee of Harbin Medical University. All adult subjects gave their written informed consent for surgery. Medical Ethics Review Committee of Harbin Medical University exempted the individual informed consent for molecular identification of hydatid cysts because samples (hydatid cysts) were obtained post surgically and CE patients' names have not appeared in this study.

Collection of hydatid cyst samples

A total of 10 hydatid cysts were collected from 10 patients suffering from hepatic CE while the size of the cysts was recorded, ranging from 6.8 cm to 10.1 cm. All the patients came from Heilongjiang Province and were operated surgically at the First and the Second Affiliated Hospital of Harbin Medical University and during May 2013 to December 2013. The endocysts were preserved in 70% ethanol at 4°C.

Microscopic examination of protoscoleces

To confirm and obtain the protoscoleces of cyst samples, suspension liquid of a part of endocysts rinsed by physiological saline was centrifuged at 1500 g for five min at room temperature, and a wet preparation was made from the sediment. The slides were examined microscopically at $100 \times$ and $400 \times$ magnification.

Extraction of genome DNA

Genomic DNA was extracted from approximately 25 mg of sediment positive for protoscoleces using the DNeasy blood and tissue kit (Qiagen, Germany) according to manufacturer's instructions. DNA was eluted in 200 μL of Buffer AE and was stored at - 20°C until further use in PCR analysis.

Primers and cycle parameters of PCR amplification

The mitochondrial *cox1*, *cytb* and *nad1* genes of hydatid cysts were amplified using previously published primers. Approximately 800 bp of the *cox1* gene and 580 bp of the *cytb* gene were amplified, with the same cycle parameters, as follows: 35 cycles at 94°C for 30 s, 54°C for 30 s, and 72°C for 60 s [14]. Approximately 900 bp of the *nad1* gene was amplified using the following primers: 5'-GGGCTATTCTCAGT(C/T)TCGTA-3' and 5'-ACCCAAAACCCACATTCT(G/T)-3', with the cycle parameters as follows: an initial denaturation step at 95°C for 3 min; 30 cycles at 94°C for 1 min, 50°C for 50 s, and 72°C for 70 s, followed by a final extension step at 72°C for 10 min [4].

Each DNA preparation was analyzed at least three times by PCR. All PCR amplifications were run with negative control (non-template water control). After agarose gel electrophoresis (1.5%), all PCR products of *cox1*, *cytb* and *nad1* genes were purified and sequenced directly to identify genotypes of *E. granulosus s.l.*

DNA sequencing and molecular analysis

All PCR products were sequenced with PCR primers at each locus on an ABI PRISMTM 3730 DNA Analyzer (Applied Biosystems, Carlsbad, CA, USA), using a BigDye Terminator v3.1 Cycle Sequencing kit (Applied Biosystems, USA). Accuracy of the sequencing data was confirmed by sequencing in both directions. Nucleotide sequences obtained in the present study

Table 2. Difference in bases and amino acids at the cytb locus of E. granulosus s.l.

Genotype/species	Accession no.	Homology (%) Ref sequence	Codon (amino acid)/Nucleotide position ^a		
G1/E. granulosus s.s.	AB622276		ATT (I)/520	TAC (Y)/612	TTG (L)/721
	KJ556990	100	ATT (I)/520	TAC (Y)/612	TTG (L)/721
	KJ556991	99.83	ATT (I)/520	TAT (Y)/612	TTG (L)/721
	KJ556992	99.65	GTT (V)/520	TAC (Y)/612	CTG (L)/721
G7/E. canadensis	AB235847	Ref sequence	ATT (I)/451	TTA (L)/855	
	KJ556989	99.65	GTT (V)/451	TTG (L)/855	

^aNucleotide position numbers according to AB235847, with the beginning of the coding region of the *cytb* gene as position no. 1. doi:10.1371/journal.pone.0109059.t002

Table 3. Distribution of G7 genotype of E. canadensis in humans and animals worldwide.

Country	No. of humans	No. of animals (host)	Amplified gene(s)	Ref.
Argentina		76 (pig)	cox1, nad1, ITS1	[35–40]
Austria	23		nad1	[3]
Brazil		3 (pig), 1 (cattle)	cox1, mdh, EgAgB4	[41,42]
Former Yugoslavia	9		nad1	[3]
Greece		20 (goat)	cox1, nad1	[43]
Hungary	1		nad1	[3]
Italy		2 (pig)	cox1, nad1, 12 S rRNA	[44]
Mexico		8 (pig)	<i>cox1, nad1,</i> ITS1, Eg9, Eg16	[45,46]
Peru		12 (pig)	cox1, nad1, ef1a	[47,48]
Poland	30	2 (pig)	<i>cox1, nad1,</i> ITS1	[31,49]
Portugal		1 (cattle)	cox1	[50]
Romania		18 (pig)	cox1, nad1, mdh, EgAgB4	[41,51]
Slovakia	2	18 (pig), 2 (cattle)	nad1, ITS1, 12S rRNA	[32,52]
Spain		34 (pig), 4 (wild boar), 6 (goat)	cox1, nad1	[53,54]
South Africa	2		nad1, 12S rRNA	[33]
Turkey	1	2 (sheep)	cox1, nad1, 12S rRNA	[34]

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were subjected to BLAST searches (http://www.ncbi.nlm.nih. gov/blast/), and were then aligned and analyzed with each other and *E. granulosus* reference sequences downloaded from GenBank using Clustal X 1.83.

Phylogenetic analysis

To understand phylogeny of genotypes of *E. granulosus s.l.*, three neighboring-joining trees were constructed at *cox1*, *cytb* and *nad1* loci, respectively, using the software Mega 5 (http://www.megasoftware.net/) based on the evolutionary distances calculated by Kimura-2-parameter model. The reliability of these trees was assessed using the bootstrap analysis with 1000 replicates.

Nucleotide sequence accession numbers

Representative nucleotide sequences obtained in this study were deposited in the GenBank database under the accession numbers KJ556989 to KJ556997.

Results

All the cysts were confirmed to be fertile (with protoscoleces) by microscopy. Two genotypes were identified out of 10 hydatid cysts based on the *cox1*, *cytb* and *nad1* genes: G1 (n = 6) and G7 (n = 4). However, one cyst identified as G7 genotype was not amplified successfully at the *cytb* locus.

Genetic characterizations of G1 genotype of *E. granulosus s.s.*

Six cox1 gene sequences identified as G1 genotype were identical to each other (KJ556996) and showed 100% similarity

with those from humans in Mongolia (AB787540–45, AB787547), Russia (AB777904, AB777907 and AB777908), and China (AB688608, AB688616, AB688617, AF458874 and JF906154); from sheep in Jordan (AB688591, AB688594), Iran (JQ250806), Peru (AB688621) and China (EU072107, AY377836 and AB491414); from cattle (JQ318001 and AY278068) and yaks (JF906154) in China; from a cat in Russia (AB622277).

Three different *cytb* gene sequences were obtained from six hydatid cysts identified as G1 genotype, with one to three base differences between one another. One sequence (KJ556990) (n = 4) had 100% similarity with the human-derived (AB786664) and yakderived (AY278067) sequences in China, and a cat-derived sequence (AB622276) in Russia. The other two (KJ556991 and KJ556992) were not described previously, having 99.83% and 99.65% similarity with that from a cat (AB622276), respectively. However, only the base difference (A to G) led to change of one amino acid (isoleucine to valine) (Table 2).

Two different *nad1* gene sequences were obtained from six hydatid cysts identified as G1 genotype, and there was one base difference between them. One sequence (KJ556993) (n = 5) had 100% similarity with the sequences from humans in China (JX217926); from sheep in China (JX217856, JX217868, JX217883 and JX217895), Morocco (EF367295–EF367301) and Iran (FJ010971–FJ010973 and GQ358009–CQ358013); from goats in Iran (HM055621–HM055624 and GQ358008); from cattle in Iran (HM055617 and GQ358005); from a camel in Iran (GQ358001); from a yak in China (JX217911). The other one (KJ556994) had 100% homology with the sequence from a human in China (JX217920).

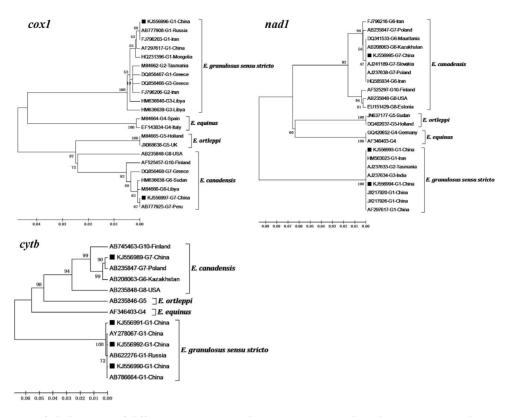


Figure 1. Comparison of Phylogenies of different genotypes within *E. granulosus s. l.* **based on** *cox1, cytb* **and** *nad1* **sequences.** Three phylogenetic trees were constructed using the Neighbor-joining distance method analysis with a Kimura-2-parameter model. The reliability of these trees was assessed using the bootstrap analysis with 1000 replicates. The GenBank accession number, the genotype and the country of origin were given for each isolate of *E. granulosus s.l.* when available. The squares indicate the sequences of *E. granulosus s.l.* from the present study. doi:10.1371/journal.pone.0109059.g001

Genetic characterizations of G7 genotype of *E. canadensis*

Four *cox1* gene sequences identified as G7 genotype were identical to each other (KJ556997) and showed 100% similarity with those from a human in Argentina (JN176935) and from pigs in Peru (AB777924, AB777925 and AB458678), Argentina (GU980914) and Romania (EU048820).

Three *cytb* gene sequences were obtained in the present study. All of them were identical to each other and were not described previously. The representative *cytb* gene sequence (KJ556989) had 99.65% similarity with that from a pig (AB235847), having two base variations. However, only one base variation (A to G) led to change of one amino acid (isoleucine to valine) (Table 2).

Four *nad1* gene sequences of G7 genotype were identical to each other (KJ556995) and were completely homologous to pigderived sequences in Slovakia (AJ241189) and Poland (AJ241196 and AJ241212).

Discussion

Human CE is a cause of serious concern due to increase in morbidity in humans and economic impact of diagnosis and treatment in China; however, the degree of endemicity of echinococcosis differs with regions in China [22,23]. The fact of increasing number of sporadic CE patients in Heilongjiang Province arouses our interest in understanding genetic characterization of *E. granulosus* isolates, which will be helpful to speculate on possible transmission patterns.

In the present study, six of ten hydatid cysts were identified as G1 genotype of *E. granulosus s.s.* G1 genotype is reported to have

a predominance in humans and animals in most studies [23]. Molecular epidemiological data of CE have revealed that G1 genotype is responsible for 88.5% of human CE worldwide [1]. In China, G1 genotype can be seen in CE cases, 98.1% (318/324) for humans versus 97.9% (474/484) for animals (Table 1). Meanwhile, differences in the genetic characterizations within G1 genotype strains were noticed in the present study. Six cox1 gene sequences were identical to each other while three and two different gene sequences were obtained at the cytb and nad1 loci, respectively, showing a conservative nature of G1 genotype at the cox1 locus. However, in Algeria, eight and three different gene sequences of G1 genotype were obtained among 30 hydatid cysts from humans, cattle and sheep at the cox1 and nad1 loci, respectively [29]. In Kenya, 49 hydatid cysts of G1 genotype from humans had 100% homology with each other at the cox1 and nad1 loci, respectively [30]. In addition, the findings of the two new cytb gene sequences of G1 genotype (KJ556991 and KJ556992) also possibly reflected endemic genetic characterizations of E. granulosus s.l. However, the base variation of the cytb sequence (KJ556991) was found to be insignificant due to the degeneracy of amino acids, for the two codons (TAC and TAT) can be translated to the same amino acid (tyrosine) (Table 2).

In the present study, four hydatid cysts were identified as G7 genotype, which currently has been assigned to *E. canadensis*, together with G6, G8 and G10 genotypes. To date, G7 genotype has been reported in humans from seven countries [3,31-34] and in five animal species as intermediate hosts from 12 countries [32,34-54] (Table 3). In general, G7 genotype is not as common as G1 genotype either in humans or in animals worldwide. However, it has been reported that 100% (30/30), 92.0% (23/25)

and, 33.3% (9/27) of human-derived hydatid cysts were indentified as G7 genotype in Poland, Austria and the former Yugoslavia, respectively [3,31]. Differences in population of E. granulosus s.l. might be related to geographical locations. Homology analysis revealed that four hydatid cysts had 100% similarity with those from pigs [39,47] at the cox1 and nad1 loci (AJ241189, AJ241196 and AJ241212). The result showed that pigs might be intermediate hosts of G7 genotype of E. canadensis in Heilongjiang Province. In fact, early in 2000, 4.5% of pigs were reported to be infected with hydatid cysts in our province [25]. Based on our summary of previous molecular data of E. granulosus s.l., pigs appear to be a principal animal intermediate host for G7 genotype, accounting for the highest prevalence of 82.8% (173/209) in pigs (Table 3). In Heilongjiang Province, the pig industry is a major economic component. Once pigs are infected with E. granulosus s.l., the habit of feeding raw offal of animals to dogs will easily lead to E. granulosus s.l. infection in dogs. Thus, this parasite can complete its life cycle in our area.

This is the first report of G7 genotype in humans in China. Identification of G7 genotype might also be attributable to the fact of the increasing international travels and livestock trades as well as the increasing number of dogs imported in recent decades. Of course, we could not rule out the possibility that humans affect CE by contacting local dogs. Early in 2002, 4% dogs were reported to be infected with *E. granulosus s.l.* in Heilongjiang Province [25]. However, they were only identified by microscopy. Thus, true transmission patterns of CE in investigated area still need to be elucidated by molecularly analyzing more samples from animals in the future.

A single locus has been used in most of epidemiological studies of *E. granulosus s.l.* However, genetic data of several genes provide more information, which will be helpful to understand the taxonomic status and phylogeny between inter- and intragenotypes of *E. granulosus s.l.*, and are especially valuable to designate a new genotype/species of *E. granulosus s.l.* McManus questioned the validity of G9 genotype, since the three same *E. granulosus s.l.* isolates from humans were identified as G9 and G7 genotypes based on ITS1-RFLP and based on sequencing of the *nad1* gene [55,56]. In the present study, by a phylogenetic analysis

References

- Alvarez Rojas CA, Romig T, Lightowlers MW (2014) *Echinococcus granulosus* sensu lato genotypes infecting humans - review of current knowledge. Int J Parasitol 44: 9–18.
- Eren B, Turkmen N, Fedakar R (2010) Allergic myocardial infarction due to hydatid cyst: an autopsy case. Bratisl Lek Listy 111: 351–352.
- Schneider R, Gollackner B, Schindl M, Tucek G, Auer H (2010) *Echinococcus canadensis* G7 (pig strain): an underestimated cause of cystic echinococcosis in Austria. Am J Trop Med Hyg 82: 871–874.
- Yang J, Jia W, Jing T, Tian G, Cheng X (2004) Analysis of genetic variation of *Echimococcus granulosus* collected from three provinces in China. Chin J Vet Sci Tech 34: 12–16. (in Chinese).
- Yang YR, Rosenzvit MC, Zhang LH, Zhang JZ, McManus DP (2005) Molecular study of *Echinococcus* in west-central China. Parasitology 131: 547– 555.
- Yang YR, Liu T, Bai X, Boufana B, Craig PS, et al. (2009) Natural infection of the ground squirrel (*Spermophilus* spp.) with *Echinococcus granulosus* in China. PLoS Negl Trop Dis 3: e518.
- Hu D, Wang N, Zhong X, Wang J, Yan N, et al. (2013) Molecular Identification and Genetic Variation of *Echinococcus granulosus* in the Tibetan Plateau. Acta Vet 44: 14. (in Chinese).
- Liu Q, Cao L, Zhang Y, Xu D, Shang L, et al. (2013) Genotypes of *Echinococcus granulosus* in animals from Yushu, Northeastern China. Vector Borne Zoonotic Dis 13: 134–137.
- Ma SM, Maillard S, Zhao HL, Huang X, Wang H, et al. (2008) Assessment of *Echinococcus granulosus* polymorphism in Qinghai province, People's Republic of China. Parasitol Res 102: 1201–1206.
- Wang N, Wang J, Hu D, Zhong X, Jiang Z, et al. (2013) Genetic variability of *Echinococcus granulosus* based on the mitochondrial 16S ribosomal RNA gene. Mitochondr DNA. Available: http://www.ncbi.nlm.nih.gov/pubmed/

based on three neighbor-joining trees at the cox1, cytb and nad1 loci, respectively (Fig. 1), it could be seen that at the cox1 and nad1 loci, it was impossible to separate G6 and G7 genotypes within *E. canadensis* as well as G1, G2 and G3 genotypes within *E. granulosus s.s.* However, the phenomenon that G6 and G7 genotypes of *E. canadensis* seemed to be different from each other at the cytb locus might be related to fewer sequences available currently. In spite of this, it is generally scientific and reasonable to regard G6 and G7 genotypes as one unit within *E. canadensis*.

Hydatid cysts of G1 genotype were observed to be a little larger than those of G7 genotype in the present study, with 9.7 cm versus 7.1 cm in diameter on average. A similar result was seen by comparing CE cysts from 68 patients infected with G1 genotype to those from 33 patients infected with G7 genotype conducted in Austria (10.7 cm for G1 genotype cysts and 5.9 cm for G7 genotype cysts) [3]. However, due to few data involving genotypes and sizes of hydatid cysts, it is unclear about the relationship between them.

To the best of our knowledge, the finding of G7 genotype (E. canadensis G6/7) in China shows that this genotype possibly has a wider geographical distribution than previously considered [57]. The previous reports of adults of E. granulosus s.l. in dogs, and hydatid cysts in sheep and pigs in Heilongjiang Province suggest the presence of two transmission patterns in our investigated areas: dog-sheep and dog-pig life cycles. However, the relationship between transmission patterns and species/genotype of E. granulosus s.l. needs to be elucidated by molecular epidemiological investigations from animals. We will try to seek the cooperation of CE patients to carry out a study of risk factors by questionnaire of their life style habits. The data will also be valuable to make control strategies for the prevention of transmission of this parasite.

Author Contributions

Conceived and designed the experiments: AL W. Zhang TZ. Performed the experiments: DY TZ. Analyzed the data: DY HL W. Zhao. Contributed reagents/materials/analysis tools: ZZ DP TJ JC YS. Contributed to the writing of the manuscript: W. Zhang AL DY.

?term=Genetic+variability+of+Echinococcus+granulosus+based+on+the+ mitochondrial+16S+ribosomal+RNA+gene. Accessed 14 October 2013.

- Yan N, Nie HM, Jiang ZR, Yang AG, Deng SJ, et al. (2013) Genetic variability of *Echinococcus granulosus* from the Tibetan plateau inferred by mitochondrial DNA sequences. Vet Parasitol 196: 179–183.
- Li T, Ito A, Nakaya K, Qiu J, Nakao M, et al. (2008) Species identification of human echinococcosis using histopathology and genotyping in northwestern China. Trans R Soc Trop Med Hyg 102: 585–590.
- Nakao M, Li T, Han X, Ma X, Xiao N, et al. (2010) Genetic polymorphisms of *Echinococcus* tapeworms in China as determined by mitochondrial and nuclear DNA sequences. Int J Parasitol 40: 379–385.
- Xiao N, Qiu J, Nakao M, Nakaya K, Yamasaki H, et al. (2003) Short report: Identification of *Echinococcus* species from a yak in the Qinghai-Tibet plateau region of China. Am J Trop Med Hyg 69: 445–446.
- Yang YR, McManus DP, Huang Y, Heath DD (2009) *Echinococcus granulosus* infection and options for control of cystic echinococcosis in Tibetan communities of Western Sichuan Province, China. PLoS Negl Trop Dis 3: e426.
- Bart JM, Abdukader M, Zhang YL, Lin RY, Wang YH, et al. (2006) Genotyping of human cystic echinococcosis in Xinjiang, PR China. Parasitology 133: 571– 579.
- Liu C, Ma X, Ding J, Chen H, Zhang N (2010) Genotyping Echinococcus granulosus isolates from sheep in Xinjiang. Med inform 23: 341. (in Chinese).
- Ma X, Mamutil W, Ding J, Lin R, Zhang Y, et al. (2008) Application of PCR-RFLP method for identification of *Echinococcus granulosus* genotypes. J Pathog Biol 3: 281–283. (in Chinese).
- Zhang LH, Chai JJ, Jiao W, Osman Y, McManus DP (1998) Mitochondrial genomic markers confirm the presence of the camel strain (G6 genotype) of *Echinococcus granulosus* in north-western China. Parasitology 116: 29–33.

- Zhang Y, Bart JM, Giraudoux P, Craig P, Vuitton D, et al. (2006) Morphological and molecular characteristics of *Echinococcus multilocularis* and *Echinococcus granulosus* mixed infection in a dog from Xinjiang, China. Vet Parasitol 139: 244–248.
- Nakao M, Lavikainen A, Yanagida T, Ito A (2013) Phylogenetic systematics of the genus *Echinococcus* (Cestoda: Taeniidae). Int J Parasitol 43: 1017–1029.
- Mahmud AL, Yan W (2011) Geographic distribution, hazard and prevention strategies of echinococcosis. Xinjiang Med J 41: 96–100. (in Chinese).
- Grosso G, Gruttadauria S, Biondi A, Marventano S, Mistretta A (2012) Worldwide epidemiology of liver hydatidosis including the Mediterranean area. World J Gastroenterol 18: 1425–1437.
- Zhang J (1958) A case of giant hepatic hydatid cyst misdiagnosed as ovarian cyst. Chin J Obstet Gynecol 6: 93–94. (in Chinese).
- 25. Qu H, Sun Y, Li B, Liu W, Sun Y, et al. (2000) Investigation on the larvae of tapeworm in pigs and the tapeworms in dogs in part of cities and counties of Heilongjiang Province, China. Chin J Vet Sci Tech 30: 13–14. (in Chinese).
- Zhang H, Ding Y, Qiao Y (2002) Investigation of parasites in sheep from Shuangcheng Town in Heilongjiang Province, China. Chin J Vet Sci Tech 32: 15–16. (in Chinese).
- Rostami Nejad M, Taghipour N, Nochi Z, Mojarad EN, Mohebbi SR, et al. (2012) Molecular identification of animal isolates of *Echinococcus granulosus* from Iran using four mitochondrial genes. J Helminthol 86: 485–92.
- Rostami Nejad, M, Nazemalhosseini Mojarad E, Fasihi Harandi M (2010) Echinococcosis: based on molecular studies in Iran. Gastroenterolology and Hepatology from Bed to Bench 3: 169–176.
- Bardonnet K, Benchikh-Elfegoun MC, Bart JM, Harraga S, Hannache N, et al. (2003) Cystic echinococcosis in Algeria: cattle act as reservoirs of a sheep strain and may contribute to human contamination. Vet Parasitol 116: 35–44.
- Casulli A, Zeyhle E, Brunetti E, Pozio E, Meroni V, et al. (2010) Molecular evidence of the camel strain (G6 genotype) of *Echinococcus granulosus* in humans from Turkana, Kenya. Trans R Soc Trop Med Hyg 104: 29–32.
- Dybicz M, Gierczak A, Dabrowska J, Rdzanek Ł, Michałowicz B (2013) Molecular diagnosis of cystic echinococcosis in humans from central Poland. Parasitol Int 62: 364–367.
- Turceková L, Snábel V, D'Amelio S, Busi M, Dubinský P (2003) Morphological and genetic characterization of *Echinococcus granulosus* in the Slovak Republic. Acta Trop 85: 223–229.
- Mogoye BK, Menezes CN, Wong ML, Stacey S, von Delft D, et al. (2013) First insights into species and genotypes of *Echinococcus* in South Africa. Vet Parasitol 196: 427–432.
- 34. Snábel V, Altintas N, D'Amelio S, Nakao M, Romig T, et al. (2009) Cystic echinococcosis in Turkey: genetic variability and first record of the pig strain (G7) in the country. Parasitol Res 105: 145–154.
- Kamenetzky L, Gutierrez AM, Canova SG, Haag KL, Guarnera EA, et al. (2002) Several strains of *Echinococcus granulosus* infect livestock and humans in Argentina. Infect Genet Evol 2: 129–136.
- Rosenzvit MC, Zhang LH, Kamenetzky L, Canova SG, Guarnera EA, et al. (1999) Genetic variation and epidemiology of *Echinococcus granulosus* in Argentina. Parasitology 118: 523–530.
- Kamenetzky L, Canova SG, Guarnera EA, Rosenzvit MC (2000) *Echinococcus granulosus*: DNA extraction from germinal layers allows strain determination in fertile and nonfertile hydatid cysts. Exp Parasitol 95: 122–127.
- Haag KL, Ayala FJ, Kamenetzky L, Gutierrez AM, Rosenzvit M (2004) Livestock trade history, geography, and parasite strains: the mitochondrial genetic structure of *Echinococcus granulosus* in Argentina. J Parasitol 90: 234– 239.

- Soriano SV, Pierangeli NB, Pianciola L, Mazzeo M, Lazzarini LE, et al. (2010) Molecular characterization of *Echinococcus* isolates indicates goats as reservoir for *Echinococcus canadensis* G6 genotype in Neuquén, Patagonia Argentina. Parasitol Int 59: 626–628.
- Soriano SV, Pierangeli NB, Pianciola LA, Mazzeo M, Lazzarini LE, et al. (2013) The optimum cut-off value to differentiate *Echinococcus granulosus sensu stricto* from other species of *E. granulosus sensu lato* using larval rostellar hook morphometry. J Helminthol 10: 1–8.
- Badaraco JL, Ayala FJ, Bart JM, Gottstein B, Haag KL (2008) Using mitochondrial and nuclear markers to evaluate the degree of genetic cohesion among *Echinococcus* populations. Exp Parasitol 119: 453–459.
- Monteiro DU, Botton SA, Tonin AA, Azevedo MI, Graichen DA, et al. (2014) *Echinococcus canadensis* (G7) and *Echinococcus granulosus sensu stricto* (G1) in swine of southern Brazil. Vet Parasitol 202: 335–338.
- Varcasia A, Canu S, Kogkos A, Pipia AP, Scala A, et al. (2007) Molecular characterization of *Echinococcus granulosus* in sheep and goats of Peloponnesus, Greece. Parasitol Res 101: 1135–1139.
- Varcasia A, Canu S, Lightowlers MW, Scala A, Garippa G (2006) Molecular characterization of *Echinococcus granulosus* strains in Sardinia. Parasitol Res 98: 273–277.
- Villalobos N, González LM, Morales J, de Aluja AS, Jiménez MI, et al. (2007) Molecular identification of *Echinococcus granulosus* genotypes (G1 and G7) isolated from pigs in Mexico. Vet Parasitol 147: 185–189.
- Cruz-Reyes A, Constantine CC, Boxell AC, Hobbs RP, Thompson RC (2007) *Echinococcus granulosus* from Mexican pigs is the same strain as that in Polish pigs. J Helminthol 81: 287–292.
- Moro PL, Nakao M, Ito A, Schantz PM, Cavero C, et al. (2009) Molecular identification of *Echinococcus* isolates from Peru. Parasitol Int 58: 184–186.
- Sánchez E, Cáceres O, Náquira C, Miranda E, Samudio F, et al. (2012) *Echinococcus granulosus* genotypes circulating in alpacas (*Lama pacos*) and pigs (*Sus scrofa*) from an endemic region in Peru. Mem Inst Oswaldo Cruz 107: 275– 278.
- Bowles J, Blair D, McManus DP (1992) Genetic variants within the genus *Echinococcus* identified by mitochondrial DNA sequencing. Mol Biochem Parasitol 54: 165–173.
- Beato S, Parreira R, Roque C, Gonçalves M, Silva L, et al. (2013) *Echinococcus granulosus* in Portugal: the first report of the G7 genotype in cattle. Vet Parasitol 198: 235–239.
- Bart JM, Morariu S, Knapp J, Ilie MS, Pitulescu M, et al. (2006) Genetic typing of *Echinococcus granulosus* in Romania. Parasitol Res 98: 130–137.
- Snábel V, D'Amelio S, Mathiopoulos K, Turceková L, Dubinský P (2000) Molecular evidence for the presence of a G7 genotype of *Echinococcus* granulosus in Slovakia. J Helminthol 74: 177–181.
- González LM, Daniel-Mwambete K, Montero E, Rosenzvit MC, McManus DP, et al. (2002) Further molecular discrimination of Spanish strains of *Echinococcus* granulosus. Exp Parasitol 102: 46–56.
- Daniel Mwambete K, Ponce-Gordo F, Cuesta-Bandera C (2004) Genetic identification and host range of the Spanish strains of *Echinococcus granulosus*. Acta Trop 91: 87–93.
- Scott JC, Stefaniak J, Pawlowski ZS, McManus DP (1997) Molecular genetic analysis of human cystic hydatid cases from Poland: identification of a new genotypic group (G9) of *Echinococcus granulosus*. Parasitology 114: 37–43.
- McManus DP (2006) Molecular discrimination of taeniid cestodes. Parasitol Int 55 Suppl: S31–37.
- Cardona GA, Carmena D (2013) A review of the global prevalence, molecular epidemiology and economics of cystic echinococcosis in production animals. Vet Parasitol 192: 10–32.