# Circular RNAs: An emerging type of RNA in cancer

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### Abstract

Circular RNAs (circRNAs), a novel type of widespread and diverse endogenous non-coding RNAs (ncRNAs), which are different from the linear RNAs, form a covalently closed continuous loop without 5' or 3' polarities. The majority of circRNAs are abundant, conserved and stable across different species, and exhibit tissue/developmental-stage-specific characteristics. They are generated primarily through a type of alternative RNA splicing called "back-splicing," in which a downstream splice donor is joined to an upstream splice acceptor through splice skipping or direct splice. Recent studies have discovered circRNAs function as microRNA sponges, binding with RNA-associated proteins to form RNA-protein complexes and then regulating gene transcription and translation into polypeptides. Emerging evidence indicates that circRNAs play important roles in the regulation of the development and progression of multiple cancers by serving as potential diagnostic and predictive biomarkers involved in tumor growth and invasion and providing new strategies for cancer diagnosis and targeted therapy. In this review, we briefly delineate the diversity and characteristics of circRNAs and discuss the highlights of the biogenesis of circRNAs and their potential functions in tumor.

### Keywords

circular RNAs, gene transcription, microRNA sponge, non-coding RNA (ncRNA)

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# Introduction

Following small RNAs (microRNAs, miRNAs) and long non-coding RNAs (LncRNAs), circular RNAs (circRNAs) are gradually recognized as a new type of endogenous non-coding RNA (ncRNA) displaying an important role in gene expression.<sup>1</sup> CircRNAs form a covalently closed loop structure, typically devoid of the terminal structures (5 'caps and 3' polyadenylated tails),<sup>2</sup> which keep themselves resistant to exonuclease R and more stable than linear RNAs. In the early 1970s, circRNAs were first found in some plant viroid, and then in Hepatitis  $\delta$  virus and some yeast mitochondrial and so on.<sup>3,4</sup> Unfortunately, although the existence of circular transcripts has been known for several decades, they are considered as molecular flukes or artifacts of aberrant RNA splicing.<sup>5</sup> Recently, novel bioinformatics and RNA deep sequencing technology have allowed comprehensive studies of circular RNA species, confirming that circRNAs are not an unexpected product of RNA transcription derived from the exon or intron of genes and are stably expressed in human, mouse, fruit fly, nematode, and other biological cells.<sup>1,6–8</sup> Unlike linear RNAs, circRNAs are generated by non-sequential back-splicing of pre-messenger RNA (mRNA) transcripts, related with reversed complementary

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Creative Commons Non Commercial CC-BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 3.0 License (http://www.creativecommons.org/licenses/by-nc/3.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage). sequences including inverted repeated Alu pairs and exon skipping and RNA-binding protein (RBP).<sup>6,9</sup> Functionally, circRNAs, as the miRNA sponges, regulate related gene expression and translating peptides.<sup>1,6,10–12</sup> Moreover, circRNAs are implicated in regulating cell proliferation and invasion in cancers, such as colon cancer, gastric cancer, and esophageal cancer, and may provide novel strategies for cancer diagnosis and therapy.<sup>13–16</sup>

# Discovery and characteristics of circRNAs

With the development of RNA sequencing and biological information technology, thousands of circRNAs across species from Archaea to human have been discovered, occupying a large proportion of the RNA family. Jeck et al.8 have identified >25,000 distinct RNA species in human fibroblasts and Memczak et al.<sup>1</sup> distinguish about 2000 circRNAs in human, 1900 in mice, and 700 in nematode. circRNAs are extraordinarily enriched in human and mouse brain, well conserved in sequence, and sometimes even detected in Drosophila brains.<sup>17</sup> In human peripheral whole blood, circRNAs have higher expression than corresponding linear mRNAs, indicating that circR-NAs may be used as biomarkers in standard clinical blood samples.<sup>18</sup>

Bioinformatic analysis reveals several shared features of circRNAs. The abundance of circR-NAs exceed that of associated linear mRNAs showing more stable property than linear mRNAs due to their resistance to exonuclease R. <sup>19</sup>They are highly conserved and predominantly cytoplasmic and possess tissue-specific characteristics.<sup>19</sup> Intronic circRNAs are primarily located in the nucleus in eukaryotes participating in regulation of gene expression at the transcription or post transcription level.<sup>20</sup> Intriguingly, some circRNAs harbor miRNA binding sites (miRNA response elements [MREs]) and attenuate endogenous miRNA-mediated transcriptional repression through target mimicry.<sup>1,8</sup>

## **Biogenesis of circRNAs**

Recent studies have shown the biogenesis of circR-NAs through back-splicing differs from the canonical splicing of linear RNAs. They are divided into exonic circRNAs, intronic circRNAs, and exonintron circRNAs. Jeck et al.<sup>8</sup> put forth two models about the origination of circRNAs, termed as lariat-driven circularization and intron-pairing-driven circularization. It is demonstrated that exon circularization depends on flanking intronic complementary sequences and alternative formation of inverted repeated Alu pairs causing alternative circularization, and resulting in multiple circular RNA transcripts produced from a single gene.<sup>20</sup>

In addition, RBPs serve as activators or inhibitors of the processing of circRNAs in some conditions. It is shown that the production of over one-third of abundant circRNAs are dynamically regulated by the alternative splicing factor, Quaking (QKI), and the addition of QKI motifs are sufficient to induce de novo circRNA formation from transcripts that are normally and linearly spliced.<sup>21</sup> However, double-strand RNA-editing enzyme ADAR1 antagonizes circRNA expression by melting stems structure.<sup>22</sup> The detailed mechanism of the formation of circRNAs remains elusive.

# **Functions of circRNAs**

CircRNAs have important functions in gene regulation from the transcription or post-transcription level, then affecting the level of gene expression.

### CircRNAs as miRNA sponges

Emerging evidence shows that a large number of circRNAs as miRNA sponges negatively regulate miRNAs. Human circRNA cerebellar degeneration-related protein 1 transcript (CDR1) acts as a miR-7 sponge by binding with miRNA effector complexes and positively associates with miRNA target site Argonaute in a miR-7-dependent manner, involved in suppressing miR-7 activity and increasing miR-7 target gene expression level.<sup>1</sup> Similarly, the testis-specific circRNA, sex-determining region Y (Sry), serves as the miR-138 sponge,<sup>23</sup> suggesting that circRNAs as the miRNA

# CircRNAs as gene transcription and expression regulators

Besides regulating miRNAs, some circRNAs are predicted to function as robust post-transcriptional regulators of gene expression, and bind and regulate linear RNA transcription and protein expression.<sup>25</sup> Some circRNAs are circularized with introns "retained" between exons; we term them exon-intron circRNAs (ElciRNAs). ElciRNAs are predominantly localized in the nucleus, interact with U1 snRNP, and enhance transcription of their parental genes.<sup>11</sup> The circRNA circMbl and its flanking introns contain conserved muscleblind (MBL) binding sites, which are specifically bound by MBL. Meanwhile, the MBL levels strongly affects circMbl biosynthesis via the MBL-binding sites, suggesting that circRNAs function in gene regulation by competing with linear splicing.<sup>6</sup> In addition, circular intronic RNA (ciRNA) ci-ankrd52, largely accumulating to its sites of transcription, associates with elongation Pol II machinery and acts as a positive regulator of Pol II transcription. Knockdown of ci-ankrd52 leads to the reduced expression of their parent genes.<sup>10</sup> Above all, we come to a conclusion that exonic circRNAs (ecircRNAs) may have regulatory functions as miRNA sponges in the cytoplasm, while ciRNAs and ElciRNAs seem to exert transcriptional regulation in the nucleus.

# **CircRNAs** and cancer

CircRNAs are viewed as large species of transcripts in eukaryotic cells. The evolutionary conservation of circularization conceals certain important functions and recent works have suggested that circRNAs play crucial roles in multiple cellular processes in the initiation and development of diseases such as atherosclerosis,<sup>26</sup> Alzheimer's disease,<sup>27</sup> and many other ones. Growing evidence shows that circRNAs have critical functions in tumor (Table 1) and serve as the huge diagnostic and therapeutic potentials for cancer.

# The correlation between circRNAs and miRNAs

CircRNAs are involved in fine-tuning the level of miRNA-mediated regulation of gene expression. In particular, their interaction with tumor-related miRNAs indicates a great significance in tumor. Ghosal et al.<sup>45</sup> have analyzed human disease-associated miRNA data collected from 174 diseases including cancer, found that circRNAs interact with these disease-associated miRNAs via the Gene ontology (GO) enrichment analysis, and identified enriched genes associated with particular biological processes including 22 stimulus

response genes and 43 cell cycle-related genes in breast cancer. Furthermore, Circ2Trait, a first comprehensive database of potential association of circRNAs with human diseases is built up.

### CircRNAs as miRNA sponges in cancer

It has been shown that miR-7 have tremendous effects on the development of a variety of cancers, such as breast cancer, hepatocellular carcinoma, cervical cancer, Schwannoma tumor, tongue cancer, lung neoplasm, gastric cancer, and colorectal cancer.<sup>28</sup> ciRS-7 as a miR-7 sponge strongly quenches the activity of mir-7 via increase of the expression of miR-7 target oncogenes and decrease of the tumor suppression genes involving in lung cancer-associated biological processes and prognosis.<sup>46</sup> In addition, viral oncogene E6/E7 is associated with miR-7 overexpression in HPV-positive HeLa cell line.47 Another well-known miRNA sponge is circSry, which contains miR-138 binding sites and functions as a miR-138 sponge.<sup>23</sup> Cir-ITCH act as a miRNA sponge of miR-7, miR-17, and miR-214 in esophageal squamous cell carcinoma and as sponges of miR-7 and miR-20a in colorectal cancer.<sup>16,36</sup> CircZEB1.5, circ-ZEB1.19, circZEB-1.17, and circZEB1.33 as the miR-200 sponge are implicated in the suppression of the lung cancer progression.<sup>37</sup>

# CircRNAs regulation of cancer-related signaling pathways

The ciRS-7/miR-7 axis is probably involved in cancer-associated biological processes via downregulating oncogenic gene expression, such as EGFR and XIAP, as well as inhibiting the tumor-suppressor gene expression, such as KLF4 in cancer.<sup>28</sup> Through sequestering and inhibiting miR-7 activity, ciRS-7 promotes the initiation and development of cancer, such as cervical cancer and hepatocellular carcinoma.<sup>23,28,32</sup> Additionally, miR-7 indirectly upregulates E-cadherin and results in reduced epithelial to mesenchyme transition (EMT), participating in tumorigenesis and cancer progression.<sup>48</sup>

Aberrant regulation of the Wnt signaling pathway is considered as a prevalent theme in cancer biology. It has been reported that cir-ITCH is involved in tumor formation and chemosensitivity through regulation of miRNA mediated Wnt/ $\beta$ -catenin signaling pathway via ubiquitination and degradation of phosphorylated Dvl2.<sup>16,36</sup>

CircRNA	Gene symbol	Gene position	Function	Disease
CDR I as/ciRS-7	CDRI	chrX:139865339–139866824	miR-7 sponges <sup>1</sup>	Neuroblastomas <sup>28</sup> Astrocytoma <sup>28</sup> Lung carcinomas <sup>28,29</sup> Cervical cancer <sup>23</sup> Breast cancer <sup>28, 30</sup> Schwannoma tumor <sup>31</sup> Hepatocellular carcinoma <sup>32</sup> Gastric carcinoma <sup>33</sup> Tongue cancer <sup>34</sup> Colorectal cancer <sup>35</sup> Esophageal carcinoma <sup>16</sup>
Cir-ITCH	ITCH	chr20:33001547-33037285	miR-7,miR-17,miR-214 sponges miR-7,miR-20a sponges	Esophageal carcinoma <sup>16</sup> Colorectal cancer <sup>36</sup>
circ-ZEB1.5	ZEBI	chr10:31749965-31791437	miR-200 sponges	Lung cancer <sup>37</sup>
circ-ZEB1.19		chr10:31749965-31750166		
circ-ZEB1.17		chr10:31661946-31750166		
circ-ZEB1.33		chr10:31749965-31791437		
circMYLK	MYLK	chr3:123366070-123368041	Bind competitively with miRNA-29a-3p	Bladder carcinoma <sup>38</sup>
circ-Foxo3	FOXO3	chr6:108984657-108986092	Enhance Foxo3 expression	Breast cancer <sup>39</sup>
circ-KLDHC10	KLDHC10	chr7:129760588-129762042	Upregulated	Colorectal cancer <sup>40</sup>
hsa_circ_0007374	AZINI	chr8:103846416-103852051	Upregulated	Colorectal cancer <sup>41</sup>
hsa_circ_0006229	TNS3	chr7:47384352-47385954	Downregulated	Colorectal cancer <sup>41</sup>
hsa_circ_002059	KIAA0907	chr1:155891165-155895634	Potential biomarker	Gastric carcinoma <sup>15</sup>
hsa_circ_0001649	SHPRH	chr6:146209155-146216113	Potential biomarker	Hepatocellular carcinoma <sup>42</sup>
hsa_circ_0005075	EIF4G3	chr1:21377358-21415706	Potential biomarker	Hepatocellular carcinoma <sup>43</sup>
has_circ_001988	FBXW7	chr4:153332454-153333681	Potential biomarker	Colorectal cancer <sup>14</sup>
hsa_circ_100855	-	-	Potential biomarker	Laryngeal carcinoma <sup>44</sup>
hsa_circ_104912	-	_	Potential biomarker	Laryngeal carcinoma <sup>44</sup>

<b>Table I.</b> CircRNAs associated with hur	man tumor.
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#### CircRNAs as potential biomarkers in cancer

Most circRNAs are abundant and conserved across species and exhibit tissue/developmental-stagespecific property. Importantly, circRNAs are stably expressed in saliva, blood, and exosomes as the promising biomarkers for diagnosis, prognosis, and therapeutic response for cancer patients. circRNA 100855 is upregulated and circRNA 104912 is downregulated in laryngeal squamous cell cancer tissues (LSCC) tissues, and their expressions are significantly related with tumor stage and neck nodal metastasis, suggesting an important role in the tumorigenesis of LSCC.44 Similarly, circ 002059, significantly downregulated in gastric cancer, is correlated with distal metastasis, TNM stage, gender, and age, implying a potential novel biomarker for the diagnosis of gastric carcinoma.<sup>15</sup> Other circRNAs have been reported aberrantly expressed in human cancers such as hepatocellular carcinoma and colorectal

carcinoma,<sup>14,42–43</sup> indicating the critical role of circRNAs in physiological and pathological processes of cancer.

### Conclusion

Previously, circRNAs, as a recently discovered RNA type, were thought to be "scrambled" exons without known functions. With the potential functions of circRNAs as miRNA "sponges" in human diseases, they are a hotspot in the field of RNA research. CircRNAs are mainly focused on their expressing profiles, characteristics, biogenesis, and functional mechanisms in various diseases including cancer. These findings will disclose the physiological and pathological processes of the vast majority of circRNAs as new biomarkers and potential therapeutic targets in cancer. Developing circRNA-based therapeutic strategies is expected to be applied to clinical practice in cancer as early as possible.

#### **Declaration of conflicting interests**

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

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#### Reference

- 1. Memczak S, Jens M, Elefsinioti A, et al. (2013) Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* 495: 333–338.
- Chen LL and Yang L. (2015) Regulation of circRNA biogenesis. *RNA Biology* 12: 381–388.
- 3. Sanger HL, Klotz G, Riesner D, et al. (1976) Viroids are single-stranded covalently closed circular RNA molecules existing as highly base-paired rod-like structures. *Proceedings of the National Academy of Sciences of the United States of America* 73: 3852– 3856.
- Arnberg AC, Van Ommen GJ, Grivell LA, et al. (1980) Some yeast mitochondrial RNAs are circular. *Cell* 19: 313–319.
- Cocquerelle C, Mascrez B, Hetuin D, et al. (1993) Mis-splicing yields circular RNA molecules. *FASEB Journal* 7: 155–160.
- Ashwal-Fluss R, Meyer M, Pamudurti NR, et al. (2014) circRNA biogenesis competes with pre-mRNA splicing. *Molecular Cell* 56: 55–66.
- Wang PL, Bao Y, Yee MC, et al. (2014) Circular RNA is expressed across the eukaryotic tree of life. *PLoS One* 9: e90859.
- 8. Jeck WR, Sorrentino JA, Wang K, et al. (2013) Circular RNAs are abundant, conserved, and associated with ALU repeats. *RNA* 19: 141–157.
- 9. Zheng Q, Bao C, Guo W, et al. (2016) Circular RNA profiling reveals an abundant circHIPK3 that regulates cell growth by sponging multiple miRNAs. *Nature Communications* 7: 11215.
- Zhang Y, Zhang XO, Chen T, et al. (2013) Circular intronic long noncoding RNAs. *Molecular Cell* 51: 792–806.
- 11. Li Z, Huang C, Bao C, et al. (2015) Exon-intron circular RNAs regulate transcription in the nucleus. *Nature Structural & Molecular Biology* 22: 256–264.
- 12. Granados-Riveron JT and Aquino-Jarquin G. (2016) The complexity of the translation ability of circRNAs. *Biochimica et Biophysica Acta* 1859: 1245–1251.
- Xie H, Ren X, Xin S, et al. (2016) Emerging roles of circRNA\_001569 targeting miR-145 in the proliferation and invasion of colorectal cancer. *Oncotarget* 7: 26680–26691.

- Wang X, Zhang Y, Huang L, et al. (2015) Decreased expression of hsa\_circ\_001988 in colorectal cancer and its clinical significances. *International Journal* of Clinical and Experimental Pathology 8: 16020– 16025.
- Li P, Chen S, Chen H, et al. (2015) Using circular RNA as a novel type of biomarker in the screening of gastric cancer. *Clinica Chimica Acta* 444: 132–136.
- Li F, Zhang L, Li W, et al. (2015) Circular RNA ITCH has inhibitory effect on ESCC by suppressing the Wnt/beta-catenin pathway. *Oncotarget* 6: 6001– 6013.
- 17. Rybak-Wolf A, Stottmeister C, Glazar P, et al. (2015) Circular RNAs in the mammalian brain are highly abundant, conserved, and dynamically expressed. *Molecular Cell* 58: 870–885.
- Memczak S, Papavasileiou P, Peters O, et al. (2015) Identification and characterization of circular RNAs as a new class of putative biomarkers in human blood. *PLoS One* 10: e0141214.
- Salzman J, Gawad C, Wang PL, et al. (2012) Circular RNAs are the predominant transcript isoform from hundreds of human genes in diverse cell types. *PLoS One* 7: e30733.
- 20. Zhang XO, Wang HB, Zhang Y, et al. (2014) Complementary sequence-mediated exon circularization. *Cell* 159: 134–147.
- 21. Conn SJ, Pillman KA, Toubia J, et al. (2015) The RNA binding protein quaking regulates formation of circRNAs. *Cell* 160: 1125–1134.
- 22. Ivanov A, Memczak S, Wyler E, et al. (2015) Analysis of intron sequences reveals hallmarks of circular RNA biogenesis in animals. *Cell Reports* 10: 170–177.
- Hansen TB, Jensen TI, Clausen BH, et al. (2013) Natural RNA circles function as efficient microRNA sponges. *Nature* 495: 384–388.
- 24. Thomas LF and Saetrom P. (2014) Circular RNAs are depleted of polymorphisms at microRNA binding sites. *Bioinformatics* 30: 2243–2246.
- Dudekula DB, Panda AC, Grammatikakis I, et al. (2016) CircInteractome: A web tool for exploring circular RNAs and their interacting proteins and micro-RNAs. *RNA Biology* 13: 34–42.
- Burd CE, Jeck WR, Liu Y, et al. (2010) Expression of linear and novel circular forms of an INK4/ARFassociated non-coding RNA correlates with atherosclerosis risk. *PLoS Genetics* 6: e1001233.
- Lukiw WJ. (2013) Circular RNA (circRNA) in Alzheimer's disease (AD). Frontiers in Genetics 4: 307.
- Hansen TB, Kjems J and Damgaard CK. (2013) Circular RNA and miR-7 in cancer. *Cancer Research* 73: 5609–5612.
- 29. Liu H, Wu X, Huang J, et al. (2015) miR-7 modulates chemoresistance of small cell lung cancer by

repressing MRP1/ABCC1. International Journal of Experimental Pathology 96: 240–247.

- Kong X, Li G, Yuan Y, et al. (2012) MicroRNA-7 inhibits epithelial-to-mesenchymal transition and metastasis of breast cancer cells via targeting FAK expression. *PLoS One* 7: e41523.
- Saydam O, Senol O, Wurdinger T, et al. (2011) miRNA-7 attenuation in Schwannoma tumors stimulates growth by upregulating three oncogenic signaling pathways. *Cancer Research* 71: 852–861.
- 32. Fang Y, Xue JL, Shen Q, et al. (2012) MicroRNA-7 inhibits tumor growth and metastasis by targeting the phosphoinositide 3-kinase/Akt pathway in hepatocellular carcinoma. *Hepatology* 55: 1852–1862.
- Zhao X, Dou W, He L, et al. (2013) MicroRNA-7 functions as an anti-metastatic microRNA in gastric cancer by targeting insulin-like growth factor-1 receptor. *Oncogene* 32: 1363–1372.
- Jiang L, Liu X, Chen Z, et al. (2010) MicroRNA-7 targets IGF1R (insulin-like growth factor 1 receptor) in tongue squamous cell carcinoma cells. *Biochemical Journal* 432: 199–205.
- Xu K, Chen Z, Qin C, et al. (2014) miR-7 inhibits colorectal cancer cell proliferation and induces apoptosis by targeting XRCC2. *Onco Targets and Therapy* 7: 325–332.
- 36. Huang G, Zhu H, Shi Y, et al. (2015) cir-ITCH plays an inhibitory role in colorectal cancer by regulating the Wnt/beta-catenin pathway. *PLoS One* 10: e0131225.
- Liu YC, Li JR, Sun CH, et al. (2016) CircNet: A database of circular RNAs derived from transcriptome sequencing data. *Nucleic Acids Research* 44: D209–215.
- Huang M, Zhong Z, Lv M, et al. (2016) Comprehensive analysis of differentially expressed profiles of lncR-NAs and circRNAs with associated co-expression and ceRNA networks in bladder carcinoma. *Oncotarget* 7: 47186–47200.
- 39. Yang W, Du WW, Li X, et al. (2016) Foxo3 activity promoted by non-coding effects of circular RNA and

Foxo3 pseudogene in the inhibition of tumor growth and angiogenesis. *Oncogene* 35: 3919–3931.

- 40. Li Y, Zheng Q, Bao C, et al. (2015) Circular RNA is enriched and stable in exosomes: A promising biomarker for cancer diagnosis. *Cell Research* 25: 981–984.
- Bachmayr-Heyda A, Reiner AT, Auer K, et al. (2015) Correlation of circular RNA abundance with proliferation–exemplified with colorectal and ovarian cancer, idiopathic lung fibrosis, and normal human tissues. *Scientific Reports* 5: 8057.
- 42. Qin M, Liu G, Huo X, et al. (2015) Hsa\_circ\_0001649: A circular RNA and potential novel biomarker for hepatocellular carcinoma. Cancer Biomarkers: Section A of Disease Markers 16: 161–169.
- Shang X, Li G, Liu H, et al. (2016) Comprehensive circular RNA profiling reveals that hsa\_circ\_0005075, a new circular RNA biomarker, is involved in hepatocellular carcinoma development. *Medicine (Baltimore)* 95: e3811.
- Xuan L, Qu L, Zhou H, et al. (2016) Circular RNA: A novel biomarker for progressive laryngeal cancer. *American Journal of Translational Research* 8: 932–939.
- 45. Ghosal S, Das S, Sen R, et al. (2013) Circ2Traits: A comprehensive database for circular RNA potentially associated with disease and traits. *Frontiers in Genetics* 4: 283.
- 46. Chou YT, Lin HH, Lien YC, et al. (2010) EGFR promotes lung tumorigenesis by activating miR-7 through a Ras/ERK/Myc pathway that targets the Ets2 transcriptional repressor ERF. *Cancer Research* 70: 8822–8831.
- Honegger A, Schilling D, Bastian S, et al. (2015) Dependence of intracellular and exosomal microR-NAs on viral E6/E7 oncogene expression in HPVpositive tumor cells. *PLoS Pathogens* 11: e1004712.
- 48. Kong X, Li G, Yuan Y, et al. (2012) MicroRNA-7 inhibits epithelial-to-mesenchymal transition and metastasis of breast cancer cells via targeting FAK expression. *PLoS One* 7: e41523.