

# Characterization of common marmoset dysgerminoma-like tumor induced by the lentiviral expression of reprogramming factors

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## Key words

Common marmoset, FGFR, regenerating medicine, reprogramming factor, tumorigenesis

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The development of a technology to generate iPSCs from differentiated somatic cells by the transduction of a set of transcription factors, OSKM, made a significant impact in the field of basic research for regenerative medicine, in light of their potential use as a cell source for transplantation therapy for various kinds of incurable diseases.<sup>(1,2)</sup>

However, the low efficiency of iPSC generation, the need to induce their efficient differentiation into specific cell types, and the risk of tumor formation in recipients transplanted with iPSC-derived functional cells have hindered the clinical application of iPSCs.<sup>(3)</sup> The transduction of transcription factors, including the oncogene *c-MYC*, insertional mutation of the genome caused by virus vectors, and genomic instability due to the stress of long-term culture for reprogramming, might contribute to tumor development when iPSC-derived functional cells are applied to clinical practice.<sup>(4)</sup> Although the technology used to generate iPSCs has improved with the use

of OSK without M,<sup>(5)</sup> non-viral vectors,<sup>(6)</sup> other molecules such as mRNA or miRNA,<sup>(7)</sup> and chemicals,<sup>(8)</sup> tumor formation in recipients remains a major concern.<sup>(9)</sup> Despite these issues, reprogramming factor-related tumor cells have not been well-characterized to date. The CM (*Callithrix jacchus*) has several advantages as an experimental laboratory primate, including ease of handling, being inexpensive to house and feed, and a high reproductive rate.<sup>(10)</sup> Therefore, CM and CM-derived iPSCs represent useful experimental tools for testing the clinical utility of iPSC-based regenerative medicine *in vivo* and *in vitro*. In this study, we attempted to generate iPSCs from CM fibroblasts, and inadvertently produced immature malignant tumor cells. We therefore analyzed the biological characteristics of these cells *in vitro* and *in vivo*. The results may provide useful information for the development of strategies to deal with tumors unexpectedly formed in patients treated with iPSC-based therapies.

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## Materials and Methods

**Cell culture, induction of reprogramming, and proliferation assay.** Common marmoset ARCs, CM ESCs, and iPS A cells derived from fetal liver cells (provided by Erika Sasaki, KEIO-REKEN Research Center for Human Cognition, Keio University, Tokyo, Japan) were maintained in DMEM/F12 (Sigma-Aldrich, St. Louis, MO, USA) containing 20% Knock-out Serum Replacement (Gibco, Carlsbad, CA, USA), 0.1 mM non-essential amino acid (Gibco), 1 mM L-glutamine (Nacalai Tesque, Kyoto, Japan), 1% antibiotic-antimycotics (Nacalai Tesque), 0.4 mM 2-mercaptoethanol (Sigma Aldrich), and 0.12% sodium hydroxide (Nacalai Tesque). The CM DGs were maintained in DMEM/F12 containing 10% FBS at 37°C in a 5% humidified CO<sub>2</sub> atmosphere. Detailed descriptions of the cell culture, reprogramming method, and proliferation assay are provided in Figures 1 and 5.

**Plasmids and lentiviral vector production.** Human OCT3/4, SOX2, KLF4, or c-MYC was inserted into CSIV-CMV-MCS-IRES2-Venus lentiviral vectors (kindly provided by Hiroyuki Miyoshi, Riken, Tsukuba, Japan). Short hairpin RNAs targeting OCT3/4, SOX2, and c-MYC were obtained from Addgene (Cambridge, MA, USA), and shRNA targeting KLF4 was obtained from Applied Biological Materials (Richmond, BC, Canada). Lentiviruses were produced as previously described.<sup>(11)</sup>

**Microarray analysis.** Total RNA from AGM fibroblasts, ARCs, and iPS A cells were isolated using the RNeasy Mini Kit (Qiagen, Valencia, CA, USA). RNA was reverse-transcribed, biotin-labeled, and hybridized for 16 h to a marmoset genome oligonucleotide custom array Marmo2 (in preparation),<sup>12</sup> which was subsequently washed and stained in a Fluidics Station 450 (Affymetrix, Santa Clara, CA, USA) according to the manufacturer's instructions. Detailed protocols of microarray analysis are provided in Figures 2 and S5.

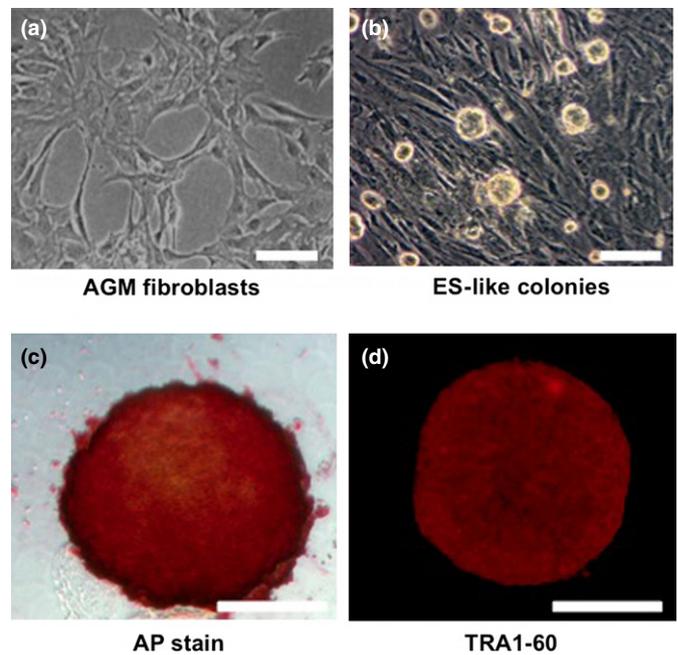
**DNA-damaging treatments.** The CM DGs were treated with 1 µg/mL MMC (Kyowa Hakko Kirin, Tokyo, Japan) or 10 µg/mL cisplatin (Sigma-Aldrich) for 1 h at 37°C. For irradiation, CM DGs were irradiated (20 Gy) using Gammacell 40 (Atomic Energy, Chalk River, Ontario, Canada). At 24 h after treatment, the cells were stained with propidium iodide (Nacalai Tesque), and the proportion of dead cells was analyzed as the sub-G<sub>1</sub> population by flow cytometry (FACSCalibur; BD Biosciences, San Jose, CA, USA).

**Statistical analysis.** Statistical analyses were carried out with the GRAPHPAD PRISM 5.0d software package (GraphPad Software, La Jolla, CA, USA). Statistical analyses were carried out using a two-tailed unpaired Student's *t*-test or one-way ANOVA followed by Tukey's multiple comparison test. *P* < 0.05 was considered statistically significant.

Additional information is provided in Supporting information.

## Results

**Characteristic of aorta-gonado-mesonephros fibroblast-derived colonies formed by transduction of reprogramming factors.** To generate CM-derived iPSCs, reprogramming factors (OSKM) were transduced into AGM fibroblasts using lentiviral vectors (Fig. 1a). Then OSKM-transduced cells were transferred to mouse embryonic fibroblast feeder cells on day 7 post-infection, and cultured in medium for CM ESCs. We found that the cells formed sphere-like structures on day 17 post-infection (Fig. 1b). Moreover, these colonies showed AP activity



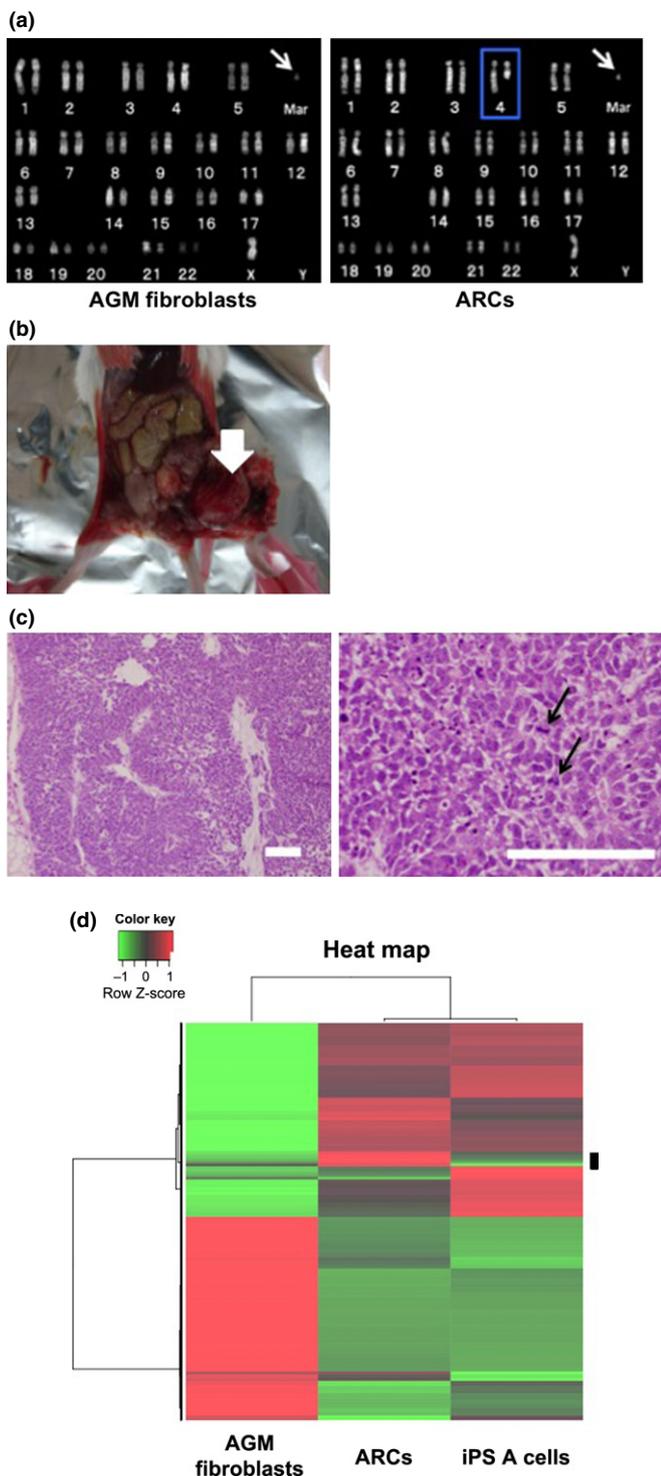
**Fig. 1.** Characterization of aorta-gonado-mesonephros (AGM) fibroblast-derived colonies formed by transduction of reprogramming factors. Representative phase-contrast images of (a) AGM fibroblasts and (b) abnormally reprogrammed cells (ARCs) forming round-shaped colonies. (c) Representative image showing expression of alkaline phosphatase (AP) activity in ARCs. (d) Immunocytochemical staining showing expression of TRA1-60 in ARCs. Bar = 100 µm.

(Fig. 1c), and expressed ESC markers such as TRA1-60, SALL1, LIN28, and DPPA4 (Figs 1d, S1). These results suggested that the reprogrammed AGM fibroblasts formed immature, round iPSC-like colonies.

**Chromosome abnormality and tumor-forming ability in abnormally reprogrammed cells.** Given that *KLF4* and *c-MYC* are well-known oncogenes,<sup>(13,14)</sup> transduction with OSKM transcription factors may cause cell transformation and chromosome instability.<sup>(15)</sup> We carried out karyotype analysis of the colony-forming cells to determine if OSKM-transduced AGM fibroblasts exhibited chromosome instability. The normal karyotype of CM cells is 44 autosomes and two sex chromosomes (46, XX or 46, XY).<sup>(16)</sup> However, the round colony-forming cells contained 44 autosomes, one X chromosome, and an abnormal marker chromosome (mar), with deletions of chromosome 4q, and were therefore denoted as 46, X, del (4q), +mar (Fig. 2a, right panel). The karyotype of the parental AGM fibroblasts was 46, X, +mar (Fig. 2a, left panel). These results suggested that the reprogramming stress induced by OSKM might have caused the deletion of 4q, although the possibility that the stress of long-term *in vitro* culture might have resulted in chromosome instability could not be excluded. These colony-forming cells were named ARCs.

To examine the ability of ARCs to differentiate into three germ layers like ESCs, we carried out an *in vitro* differentiation assay based on the protocol for human ESC differentiation.<sup>(17,18)</sup> Unlike human ESCs, ARCs did not differentiate into neural progenitors, cardiomyocytes, or hepatic cells (Fig. S2, Video S1).

We carried out *in vivo* differentiation assays by injecting ARCs into the testes of SCID mice. Approximately 6 weeks after injection, 11 of 18 mice injected with ARCs showed



**Fig. 2.** Chromosome abnormality and tumor-forming ability in abnormally reprogrammed cells (ARCs). (a) Karyotype analyses of aorta-gonadomesonephros (AGM) fibroblasts (left panel) and ARCs (right panel). Arrows indicate marker chromosome. Blue outline indicates the deletion of 4q. Mar, marker chromosome. (b) Representative photograph of dysgerminoma-like tumor formed by transplantation of ARCs into SCID mice. (c) Hematoxylin-eosin staining of dysgerminoma-like tumor tissues. Arrows in right panel indicate mitotic figures in tumor cells. Bar = 100  $\mu$ m. (d) Microarray analysis. Gene expressions in AGM fibroblasts, ARCs, and normal induced pluripotent stem (iPS) A cells were analyzed by unsupervised hierarchical clustering. A heat map using probes showing differential expression levels in each cell line is shown. Red indicates upregulation; green indicates downregulation. The black bar on the right side of the heat map shows candidate differentially expressed probes in ARCs.

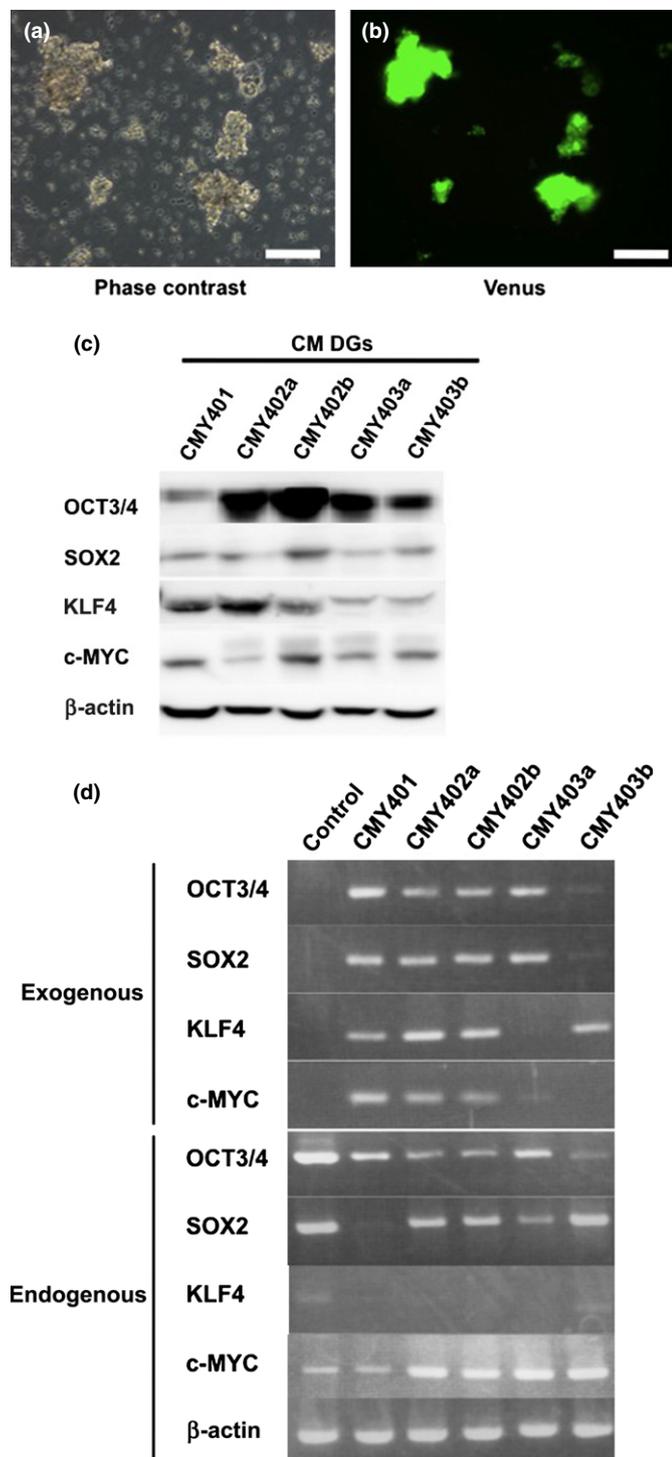
tumor formation (Fig. 2b), whereas no mice injected with AGM fibroblasts showed tumor formation (0/3, data not shown). Staining with H&E revealed that the tumors were relatively homogenous, with high cellular density, necrosis, and pleomorphism, indicating their malignant phenotype (Fig. 2c). In addition, tumors were composed of nests and sheets of uniform round or polygonal cells with abundant, clear to faintly eosinophilic cytoplasm with well-demarcated cytoplasmic borders and a delicate network of thin-walled blood vessels in the tumor nests (Fig. 2c, right panel). Furthermore, immunohistochemical analyses revealed that the tumor cells were focally and weakly immunopositive for vimentin, and immunonegative for the differentiation markers cytokeratin, S100, desmin,  $\alpha$ -smooth muscle actin, and neuron-specific enolase (data not shown). Tumor tissues also expressed c-KIT, but not CD30 or CD45 (Fig. S3). These molecular expression profiles implied that the tumor was equivalent to human malignant dysgerminoma, rather than other types of immature tumors such as embryonal carcinoma, yolk sac tumor, or teratoma.<sup>(19,20)</sup> The tumor was named CM DG.

We next carried out soft agar assays to determine if ARCs were transformed and showed anchorage-independent growth as a result of ectopic expression of reprogramming factors. The ARCs were cultured in 0.5% agarose-containing medium for 20 days, and the number of colonies was counted. The ARCs formed many colonies, compared with parental AGM fibroblasts (Fig. S4a and data not shown). These results strongly suggested that ARCs were transformed during reprogramming, and acquired the capacity for anchorage-independent growth. To clarify the contribution of reprogramming factors that could transform AGM fibroblasts, we transduced various combinations of these factors into AGM fibroblasts, and examined if the transduced cells were transformed by the colony formation assay on mouse embryonic fibroblasts, AP staining assay, and soft agar assay. The iPSC-like colonies were found in OSKM- and OSM-transduced cells (OSKM,  $30 \pm 3/5000$ ; OSM,  $6 \pm 0/5000$ ), but they were not found at all when OSK, OS, OM, SM, O, S, K, or M were transduced (Fig. S4b). AP activity was found in both OSKM- and OSM-transduced cells, although OSM-transduced cells showed weaker AP activity than OSKM-transduced cells (Fig. S4c). In soft agar assay, the anchorage-independent growth was found in both OSKM- and OSM-transduced cells (Fig. S4d; OSKM,  $160 \pm 23/1000$ ; OSM,  $163 \pm 10/1000$ ). These results indicated that the simultaneous expression of OCT3/4, SOX2, and c-MYC was at least required for the transformation of AGM fibroblasts, while KLF4 did not play a major role in the transformation of AGM fibroblasts.

Tomioka *et al.* reported the establishment of CM iPSCs (iPS A cells) showing normal karyotype.<sup>(12)</sup> To characterize the gene expression in ARCs, we carried out microarray analyses using mRNA from ARCs, iPS A cells, and AGM fibroblasts. According to the clustering pattern and the heat map, 171 probes that showed higher expression levels in ARCs as compared to other cells were selected as candidate differentially expressed genes in ARCs (Fig. 2d). Moreover, we focused on the genes specifically highly expressed in ARCs compared to those in iPS A cells, and the top seven genes highly expressed in ARCs were selected (*ZFHX4*, *PCDH19*, *NFIX*, *HOXC8*, *STMN2*, *SERPINA3*, and *CXORF67*). Then we validated these data by semiquantitative RT-PCR analyses, and five genes (*ZFHX4*, *NFIX*, *HOXC8*, *STMN2*, and *CXORF67*) were confirmed to be more expressed in ARCs than those in controls (Fig. S5). The high expression of these five genes might be characteristics of ARCs.

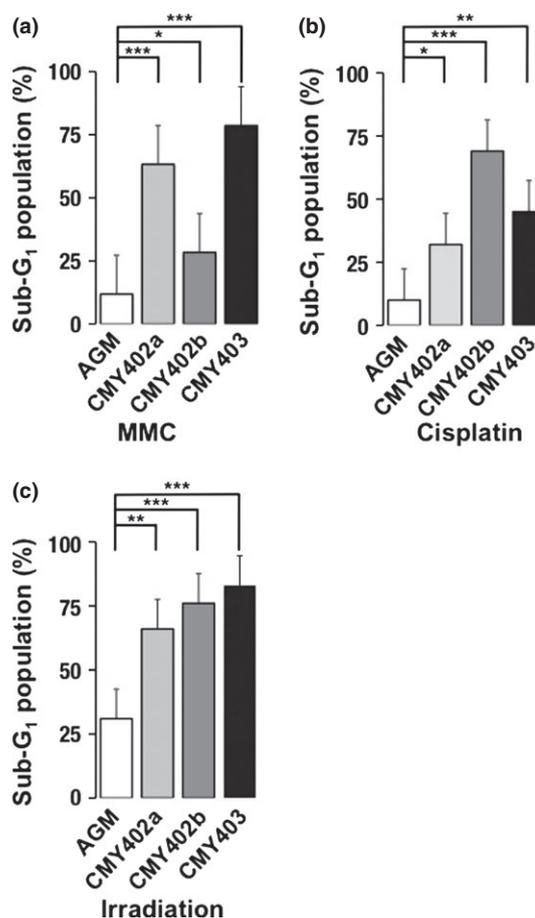
**Characteristic of CM DGs.** We then surgically removed CM DGs and cultured them *in vitro* to examine their biological characteristics. The CM DGs could grow infinitely in a semi-

floating state in the culture dish, and showed continuous expression of Venus fluorescent protein (Fig. 3a,b). We generated five CM DG cell lines (CMY401, CMY402a, CMY402b, CMY403a, and CMY403b) from five independent tumors formed by the injection of ARCs into SCID mice, and found that all four transduced reprogramming factors were integrated into their genomes (Fig. S6a). Both endogenous and exogenous reprogramming factors were expressed in these cell lines (Fig. 3c,d).



**Fig. 3.** Characterization of common marmoset dysgerminoma-like (CM DG) cells in culture. (a) Representative phase-contrast image of CM DGs. (b) Immunofluorescent image of Venus expression in CM DGs. Bar = 100 μm. (c) Western blot analysis showing expression of reprogramming factors in CM DG cell lines. (d) RT-PCR analysis showing the expression of endogenous or exogenous reprogramming factors in CM DGs. Cj11 (CM embryonic stem cell line) was used as control.

**Effects of DNA-damaging agents and irradiation on CM DGs.** Dysgerminomas are generally sensitive to cisplatin and irradiation.<sup>(21,22)</sup> We therefore examined the effects of DNA-damaging agents such as MMC and cisplatin on CM DGs. Three CM DG cell lines (CMY402a, CMY402b, and CMY403a) were treated with MMC for 1 h, and the proportion of dead cells was analyzed by flow cytometry at 24 h after treatment. The percentage of cells with a sub-G<sub>1</sub> DNA content was taken as a measure of dead cells in the population. The proportion of dead cells in MMC-treated CM DG cultures was significantly higher than that in controls (MMC-treated AGM fibroblasts) (Figs 4a, S7). Similar results were obtained when these three cell lines were treated with cisplatin or irradiation (Figs 4b,c, S8, S9). These results suggested that CM DGs were



**Fig. 4.** Effects of DNA-damaging agents and irradiation on common marmoset dysgerminoma-like cells (CM DGs). The cells were treated with (a) mitomycin C (MMC), (b) cisplatin, or (c) irradiation, and the proportions of sub-G<sub>1</sub> populations in aorta-gonado-mesonephros (AGM) fibroblasts or CM DG cell lines were analyzed by FACS. Results are shown as means ± SD. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001.

more sensitive to DNA damage than their parental AGM fibroblasts.

We carried out inverse PCR analyses to identify the integration sites of the lentiviral vectors expressing reprogramming factors in CM DGs. OCT 3/4-, SOX2-, KLF4-, and c-MYC-expressing lentiviral vectors were integrated into 5, 12, 5, and 9 genomic sites, respectively (Fig. S6b, Table S1). The possibility that multiple integrations of lentiviral vectors into the genome caused chromosome instability, leading to the formation of CM DGs, could therefore not be excluded.

**Dependence of CM DGs growth on c-MYC and bFGF signalings.**  
 To address the question of whether proliferation of CM DGs was dependent on reprogramming factors, we observed the proliferation rate after suppression of each reprogramming factor by shRNA (Fig. S10). Suppression of c-MYC or all four reprogramming factors greatly inhibited the proliferation of CM DGs, indicating that the growth of CM DGs was highly dependent on c-MYC (Fig. 5a).

The proliferation of human ESCs is known to be promoted by bFGF signaling.<sup>(23)</sup> We examined the possibility that the growth of CM DGs might also be enhanced by bFGF signaling by analyzing the proliferation of CM DGs cultured in medium with or without bFGF. The growth of CM DGs was highly dependent on bFGF (Fig. 5b). Consistent with these results, BGJ398, an inhibitor for FGFR 1 to 4, remarkably inhibited the growth of CM DGs in a dose-dependent manner (Fig. 5c). Moreover, FACS analyses revealed that the sub-G<sub>1</sub> population, representing dead cells, was increased in the presence of BGJ398 (Fig. S11). It should be emphasized that the IC<sub>50</sub> of BGJ398 (59 nM) was lower for CM DGs than for their parental AGM fibroblasts and control CM skin fibroblasts (Fig. 5d), indicating higher sensitivity of CM DGs. These results suggested that the growth of CM DGs was dependent on bFGF signaling, and therefore FGFR inhibitor could be used to control the growth of the reprogramming factor-induced tumor.

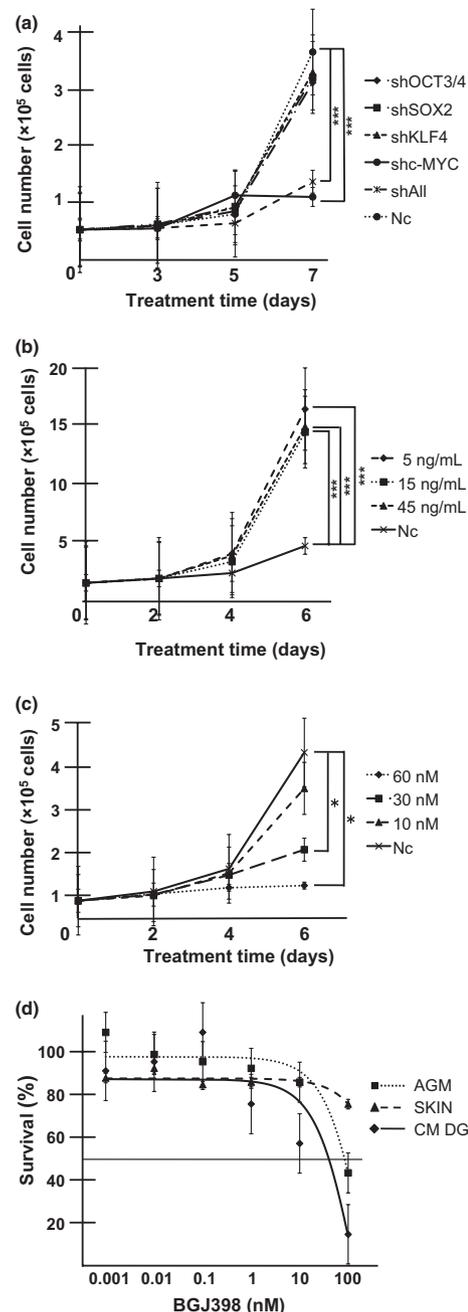
## Discussion

In this study we investigated the characteristics of ARCs and CM DGs generated in the reprogramming process of CM AGM fibroblasts by Yamanaka factors.

A normal iPSC line of iPS A cells, showed the expression of ES markers, pluripotency, and flattened morphology, like human iPSCs.<sup>(20)</sup> In contrast, ARCs showed sphere-like structures, like mouse iPSCs.<sup>(1)</sup> This morphological difference between iPS A cells and ARCs might be useful to select “true” iPSCs derived from CM, although the underlying molecular mechanisms responsible for this morphological difference remain unknown.

We found, by microarray analyses, that the gene expression pattern in ARCs was more similar to that in iPS A cells than that in AGM fibroblasts, suggesting that reprogramming processes have been done in ARCs by the transduction of reprogramming factors. We also found that genes such as *ZFH4*, *NFIX*, *HOXC8*, *STMN2*, and *CXORF67* were highly expressed in ARCs. It should be noted that, among these, *HOXC8* is known to be a transcriptional factor related to tumorigenesis.<sup>(24)</sup> Therefore, these candidates of markers might be useful to predict the tumorigenic potential of iPSCs. Further evaluation is required to confirm our hypothesis.

The original AGM fibroblasts had an abnormal marker chromosome (mar; Fig. 2a, left panel). Although tumor formation was not evident caused in SCID mice (data not shown), this chromosome instability might also be one of the inducers of



**Fig. 5.** Dependence of common marmoset dysgerminoma-like (CM DG) cell growth on c-MYC and basic fibroblast growth factor (bFGF) signaling. (a) Inhibition of CM DG growth by knockdown of c-MYC. Cells ( $3 \times 10^4$ ) were seeded on 24-well plates and transduced with shRNA targeting OCT3/4, SOX2, KLF4, c-MYC, or all reprogramming factors (shAll). Cell growth curves were analyzed by cell counts at the indicated time points. Results are shown as means  $\pm$  SD. \*\*\* $P < 0.001$ . Nc, negative control (mock vector). (b) Growth rate of CM DGs was promoted by the addition of bFGF. Cells were cultured in the presence or absence (Nc) of bFGF. Cell numbers were counted at the indicated time points. Results are shown as means  $\pm$  SD. \*\*\* $P < 0.001$ . (c) FGFR inhibitor suppressed CM DG growth. Cells were cultured in the presence or absence (Nc) of the FGFR1-4 inhibitor BGJ398; bFGF was added at 5 ng/mL. Cell numbers were counted at the indicated time points. Results are shown as means  $\pm$  SD. \* $P < 0.05$ . (d) CM DGs, aorta-gonado-mesonephros fibroblasts (AGM), and CM skin fibroblasts (SKIN) were treated with different concentrations of BGJ398 for 3 days, and the growth-inhibitory effects were analyzed by MTS assay. The IC<sub>50</sub> for CM DGs was lower than those for parental AGM fibroblasts and control CM skin fibroblasts. Results are shown as means  $\pm$  SD.

carcinogenesis during the reprogramming process. Thus, needless to say, to generate “safe” iPSCs, validation of the karyotype of the original cells is needed. Moreover, ARCs lost chromosome 4q and X or Y, and possessed an abnormal marker chromosome (mar). Various tumor suppressors including human tumor suppressor gene 1, large tumor suppressor 1 and P36 transformed follicular lymphoma gene have been identified on chromosome 4q in CM cells (Table S2), suggesting that loss of these tumor suppressors might have induced the transformation of CM AGM fibroblasts during reprogramming, although the possibility that translocation of chromosome 4q occurred during the reprogramming process caused the transformation of cells could not be excluded.

It is also possible that the continuous activation of ectopically-transduced transcription factors, including the oncogene *c-MYC*, might have contributed to cell transformation, as described previously.<sup>(25,26)</sup> Indeed, CM DGs overexpressed *c-MYC*, and their growth was highly dependent on *c-MYC* expression, suggesting that downregulation of *c-MYC* might represent a possible strategy for inhibiting the growth of reprogramming factor-related tumors.

Insertional mutation caused by the integration of lentiviral vectors into the genome might also have promoted cell transformation. Lentiviral vectors expressing reprogramming factors were integrated into at least 31 different genomic sites in CM DGs, some of which were in the vicinity of protein-encoding genes. Moreover, the expression of reprogramming factors transduced by lentiviral vectors continued for over a year in ARCs (data not shown). A safer method, without genome integration, is therefore required for the delivery of reprogramming factors to somatic cells to generate iPSCs applicable for transplantation therapies. Although Sendai virus vectors or transfection of DNA or mRNA may be safer methods,<sup>(27)</sup> these are lengthy processes that can take more than 1 month to obtain iPSCs,<sup>(28)</sup> which could also cause stress and lead to genomic instability and subsequent tumor formation. More sophisticated, safer, and more rapid methods of reprogramming might be desirable.

Common marmoset DGs resembled human dysgerminomas in terms of both their pathology and sensitivity to irradiation and

DNA-damaging agents.<sup>(21,22)</sup> In addition, the growth of CM DGs was significantly inhibited by an FGFR1-4 inhibitor. Therefore irradiation, chemotherapy, and FGFR1-4 inhibitors might be effective strategies for controlling human dysgerminomas, and also for tumors that develop in patients treated with iPSC-based therapies.

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## Disclosure Statement

The authors have no conflicts of interest.

## Abbreviations

AGM	aorta-gonado-mesonephros
AP	alkaline phosphatase
ARC	abnormally reprogrammed cell
bFGF	basic fibroblast growth factor
CM	common marmoset
DG	human dysgerminoma-like cell
ESC	embryonic stem cell
FGFR	fibroblast growth factor receptor
iPSC	induced pluripotent stem cell
K	KLF4
M	<i>c-MYC</i>
MMC	mitomycin C
O	OCT3/4
S	SOX2

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## Supporting Information

Additional supporting information may be found in the online version of this article:

**Fig. S1.** Expression of embryonic stem cell (ESC) markers in abnormally reprogrammed cells (ARCs).

**Fig. S2.** Impaired differentiation of abnormally reprogrammed cells (ARCs).

**Fig. S3.** Expression of c-KIT, CD30, and CD45 in common marmoset dysgerminoma-like cells.

**Fig. S4.** Colony formation of aorta-gonado-mesonephros fibroblasts by the transduction of OCT3/4, SOX2, KLF4, and c-MYC (OSKM) and OSM.

**Fig. S5.** Validation of genes showing upregulation in abnormally reprogrammed cells (ARCs) compared to normal induced pluripotent stem (iPS) A cells in microarray analysis.

**Fig. S6.** Integration of reprogramming genes into the genome of common marmoset dysgerminoma-like cells (CM DGs).

**Fig. S7.** Fluorescence-activated cell sorter analyses to reveal effects of mitomycin C treatment on common marmoset dysgerminoma-like cell lines.

**Fig. S8.** Fluorescence-activated cell sorter analyses to reveal effects of cisplatin treatment on common marmoset dysgerminoma-like cell lines.

**Fig. S9.** Fluorescence-activated cell sorter analyses to reveal effects of irradiation on common marmoset dysgerminoma-like cell lines.

**Fig. S10.** Knockdown of OCT3/4, SOX2, KLF4, or c-MYC by shRNA in common marmoset dysgerminoma-like cell lines.

**Fig. S11.** Induction of cell death in common marmoset dysgerminoma-like cells by BGJ398.

**Table S1.** Lentiviral vector integration sites in common marmoset (CM) dysgerminoma-like cells.

**Table S2.** Human homologs of candidate tumor suppressors located on chromosome 4q in common marmoset (CM).

**Video S1.** *In vitro* differentiation assay to assess the ability of abnormally reprogrammed cells to differentiate into cardiomyocytes.

**Data S1.** Materials and Methods.