



A case of *KMT2A–SEPT9* fusion–associated acute megakaryoblastic leukemia

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Abstract Acute megakaryoblastic leukemia (AMKL) constitutes ~5%–15% of cases of non-Down syndrome AML in children, and in the majority of cases, chimeric oncogenes resulting from recurrent gene rearrangements are identified. Based on these rearrangements, several molecular subsets have been characterized providing important prognostic information. One such subset includes a group of patients with translocations involving the *KMT2A* gene, which has been associated with various fusion partners in patients with AMKL. Here we report the molecular findings of a 2-yr-old girl with AMKL and t(11;17)(q23;25) found to have a *KMT2A–SEPT9* fusion identified through targeted RNA sequencing. A *KMT2A–SEPT9* fusion in this subset of patients has not previously been reported.

[Supplemental material is available for this article.]

CASE PRESENTATION

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A previously healthy 2-yr-old female was initially seen for recurrent fevers and decreased appetite. She was treated for suspected otitis media but continued to experience a poor appetite and fevers. A complete blood count showed anemia (Hb 7.2 g/dl) and thrombocytopenia (plt 60 k/μl), with a white blood cell count in the normal range (6.54 k/μl, ANC 2500/ml). A bone marrow biopsy and aspiration were performed that demonstrated a normocellular marrow with left-shifted granulopoiesis, progressive erythroid maturation, and atypical, hypolobulated megakaryocytes. Reticulin staining demonstrated variable increase in reticulin fibrosis from mild to marked (1–3+). The specimen contained an expanded blast population (30%), with blasts that were variable in size and had round nuclei, fine chromatin, variable nucleoli, and agranular cytoplasm. Some blasts showed cytoplasmic blebs with some of the larger blasts having vacuolated cytoplasm. Flow-cytometric analysis showed the blast population had expression of CD4, CD33, CD38, CD41, CD45, CD61, CD71, CD117, and CD123. The findings were consistent with a diagnosis of AMKL.

Cytogenetic analysis showed 46, XX, t(11;17)(q23;q25) in nine of 20 metaphase cells and the presence of a rearrangement involving *KMT2A* was confirmed by FISH in 8% of 300 cells. Targeted RNA sequencing identified a corresponding *KMT2A–SEPT9* fusion transcript.

The patient received induction chemotherapy consisting of daunorubicin, cytarabine, and etoposide (ADE) in combination with gemtuzumab ozogamicin. Repeat bone marrow

analysis at the end of induction demonstrated an MRD-negative complete remission. Because of the poor outcomes associated with *KMT2A* rearrangements in pediatric patients with AMKL, the decision was made to proceed to bone marrow transplantation in first complete remission. After completing two additional cycles of consolidative therapy, she received an allogeneic bone marrow transplant from an HLA-matched unrelated donor and has no evidence of disease more than 2 months after her transplant.

TECHNICAL ANALYSIS

The presence of the translocation involving Chromosomes 11 and 17 was identified by standard karyotype analysis and confirmed by a break-apart FISH probe (Abbott Molecular) (Fig. 1A,B). Targeted RNA sequencing using a customized 199-gene Archer FusionPlex panel identified the *KMT2A*–*SEPT9* transcript involving exon 7 and exon 2 of *KMT2A* and *SEPT9*, respectively (Table 1; Fig. 1C).

SUMMARY

KMT2A–*SEPT9* fusions are rare events that have been most commonly described in various myeloid leukemias exhibiting monocytic differentiation (Taki et al. 1999; Yamamoto et al. 2002; Shih et al. 2006; Strehl et al. 2006; Kurosu et al. 2008). They have infrequently been described in M0/M1/M2 AML, t-AML, and de novo myelodysplastic syndrome (Supplemental Table 1; Osaka et al. 1999; Strehl et al. 2006; Kreuziger et al. 2007; Saito et al. 2010;

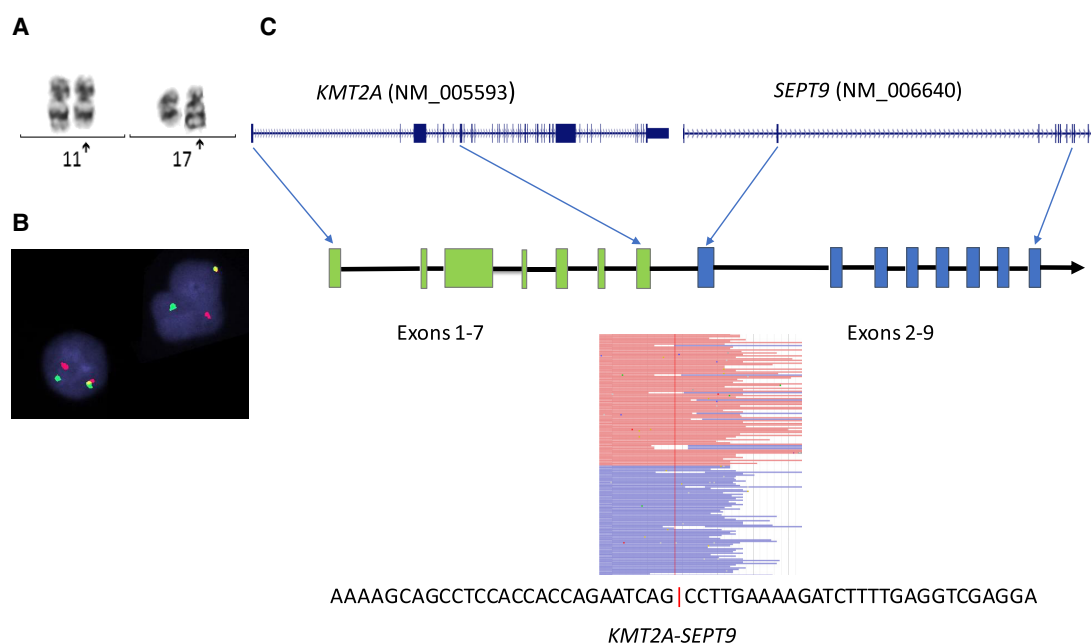


Figure 1. (A) Chromosome analysis reveals a balanced chromosome translocation between Chromosomes 11 and 17—that is, $t(11;17)(q23;q25)$ (arrows). (B) FISH analysis with a *KMT2A* break-part probe set (Abbott Molecular) shows a split *KMT2A* signal pattern—that is, $t(11;17)$. The 5' *KMT2A* and 3' *KMT2A* were labeled with green and orange, respectively. (C) Schematic illustration of the protein structure, bidirectional RNA sequencing reads, and transcript sequence of the *KMT2A* (NM_005593)–*SEPT9* (NM_006640) in-frame fusion product detected by Archer FusionPlex with exons 1–7 of *KMT2A* fused to exons 2–9 of *SEPT9*.

Table 1. KMT2A–SEPT9 fusion detected in patient

Gene 1	Gene 2	Position 1	Position 2	Transcript 1	Exon number 1	Transcript strand 1	Transcript 2	Exon number 2	Transcript strand 2	Fusion junction sequence	Frameshift class
KMT2A	SEPT9	Chr 11: 118482092	Chr 17: 77402059	NM_005933	7	+	NM_006640	2	+	AAAAGCAGCCTCCA CCACCAGAATCAG CCTTGAAAAGATCT TTTGAGGTCGAGGA	In-frame

Santos et al. 2010). To our knowledge, this is the first report of KMT2A–SEPT9 fusion–associated AMKL, as well as the first reported occurrence of any KMT2A–SEPTIN fusion occurring in AMKL (Cerveira et al. 2011). In this case the fusion is located at the intron 7 breakpoint. KMT2A–SEPT9 fusions may have a propensity to involve the intron 7 or 8 breakpoint, as the majority of reported cases involve this region. This contrasts with more common KMT2A fusion partner genes, which most frequently involve the region between exon 9 and intron 11 (Meyer et al. 2018). However, the limited number of cases prevents any definitive conclusions.

AMKL is a subtype of AML with bimodal age distribution, with peaks occurring in early childhood before the age of 3 and later in adulthood (Tallman et al. 2000; Athale et al. 2001). In patients with Down syndrome (DS), AMKL is the most frequently occurring form of AML and is characterized by the presence of mutations involving GATA1 (Wechsler et al. 2002). In patients with non-DS pediatric AMKL, several molecular subsets have recently been characterized and provide valuable prognostic information (de Rooij et al. 2016, 2017; Hara et al. 2017). Commonly reoccurring rearrangements include RBM15–MKL1, CBF2T3–GLIS2, NUP98–KDM5A, and KMT2A (de Rooij et al. 2016, 2017; Hara et al. 2017). Patients with fusions involving KMT2A make up 7%–17.4% of pediatric patients with non-DS AMKL (de Rooij et al. 2016, 2017; Hara et al. 2017). Numerous KMT2A fusion partners have been identified in children with AMKL such as MLLT1, MLLT3, MLLT6, MLLT9, and MLLT10 (de Rooij et al. 2016, 2017; Hara et al. 2017). Although little is known about the prognostic implications of the various KMT2A fusion partners, collectively it appears the presence of these rearrangements is a high-risk feature associated with a greater risk of relapse and worse overall survival, indicating a role for allogeneic transplantation in first remission, which was recommended for the patient discussed in this case (de Rooij et al. 2016, 2017).

The KMT2A gene located on Chromosome 11 band q23 is a frequent target of translocation events with more than 100 recurrent rearrangements having been identified (Meyer et al. 2018). KMT2A rearrangements are commonly seen in both adult and pediatric acute leukemias but have particularly strong associations with infant ALL (Meyer et al. 2018), M4/M5 AML (Cimino et al. 1995; Schoch et al. 2003; Meyer et al. 2018), and therapy-related AML (t-AML) (Smith et al. 1994; Meyer et al. 2018), where it typically is found in patients exposed to topoisomerase II inhibitors. The KMT2A gene product is a DNA-binding protein capable of positively regulating gene expression, including the Hox family of genes, which play an important role in hematopoiesis and lymphoid cell development (Caslini et al. 2000; Milne et al. 2002). Chimeric proteins resulting from KMT2A rearrangements can efficiently transform hematopoietic precursors into leukemic stem cells (Krivtsov and Armstrong 2007). However, the fusion partner appears to play an important role in transformation because simply enhancing KMT2A promoter activity is not sufficient to induce leukemogenesis (Corral et al. 1996).

The septin family of genes is an evolutionarily conserved GTP-binding, filament-forming protein believed to be involved in polarity determination, cytoskeletal reorganization,

membrane dynamics, vesicle trafficking, and exocytosis (Kartmann and Roth 2001). Aside from *SEPT9*, several human septin genes have been identified as partners for translocation events with *KMT2A* including *SEPT5*, *SEPT6*, and *SEPT11* (Hall and Russell 2004). The role of *SEPT9* in leukemogenesis has not been clearly elucidated. However, studies have shown that variants of *SEPT9* interact with both α and γ tubulin, and cells with enhanced expression of *SEPT9* experienced defects in both cytokinesis and mitotic spindle defects, contributing to genomic instability (Peterson et al. 2011). The role of *SEPT9* in malignant transformation may not be restricted to AML/MDS, as alterations in expression or deletion of *SEPT9* are frequently observed in breast and ovarian cancer, indicating its potential role as a tumor suppressor (Kalikin et al. 2000; Burrows et al. 2003).

The mechanism by which the *KMT2A–SEPT9* fusion drives leukemogenesis has not been firmly established. For many *KMT2A* fusion partner genes, it is believed that rearrangement events result in the fusion of transcriptional activation domains to *KMT2A* and are capable of driving leukemogenesis (So and Cleary 2003; Zeisig et al. 2003). However, as is the case with *SEPT9*, several *KMT2A* partners are localized to the cytoplasm and unlikely to have nuclear function. In a number of these partners dimerization of fusion oncoproteins has been identified as an alternative mechanism of transcriptional activation of *KMT2A* (Martin et al. 2003; So et al. 2003). Likewise, homo-oligomerization of *KMT2A–SEPT6* fusion products proved to be capable of immortalizing stem cell progenitors (Ono et al. 2005). Drawing from sequence homology across the septin family of proteins and the ability of *SEPT9* to form homodimers, it is reasonable to hypothesize that *KMT2A–SEPT9* fusion protein dimerization is a key step in leukemic transformation (Abbey et al. 2016).

In summary, this report describes the first documented case of *KMT2A–SEPT9* fusion-associated AMKL and is also the first report of any *KMT2A–SEPTIN* fusion occurring in AMKL. This rearrangement was first detected by conventional cytogenetics and confirmed by targeted RNA sequencing. Despite the numerous documented cases of the *KMT2A–SEPT9* fusion, the mechanism of its role in leukemic transformation and its prognostic impact are unclear.

ADDITIONAL INFORMATION

Data Deposition and Access

The variant described in this manuscript was deposited in ClinVar (<https://www.ncbi.nlm.nih.gov/clinvar/>) and assigned the accession number SCV000852004.

Ethics Statement

Informed and signed consent was obtained for the research performed and publication of the results. The patient was enrolled in the Memorial Sloan Kettering Cancer Center (MSKCC) targeted gene sequencing research study (Genomic profiling in cancer patients; NCT01775072) with approval from the MSKCC Institutional Review Board under protocol IRB# 12-245.

Competing Interest Statement

The authors have declared no competing interest.

Referees

Michael Roth
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Author Contributions

C.J.F., N.S., Y.Z., and M.R. conceived the study. Y.Z., J.Y., and R.B. provided figures and associated legends. All authors reviewed and drafted the manuscript.

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REFERENCES

- Abbey M, Hakim C, Anand R, Lafera J, Schambach A, Kispert A, Taft MH, Kaefer V, Kotlyarov A, Gaestel M, et al. 2016. GTPase domain driven dimerization of SEPT7 is dispensable for the critical role of septins in fibroblast cytokinesis. *Sci Rep* **6**: 20007.
- Athale UH, Razzouk BI, Raimondi SC, Tong X, Behm FG, Head DR, Srivastava DK, Rubnitz JE, Bowman L, Pui CH, et al. 2001. Biology and outcome of childhood acute megakaryoblastic leukemia: a single institution's experience. *Blood* **97**: 3727–3732.
- Burrows JF, Chanduloy S, McIlhatton MA, Nagar H, Yeates K, Donaghy P, Price J, Godwin AK, Johnston PG, Russell SE. 2003. Altered expression of the septin gene, *SEPT9*, in ovarian neoplasia. *J Pathol* **201**: 581–588.
- Caslini C, Shilatifard A, Yang L, Hess JL. 2000. The amino terminus of the mixed lineage leukemia protein (MLL) promotes cell cycle arrest and monocytic differentiation. *Proc Natl Acad Sci* **97**: 2797–2802.
- Cerveira N, Bizarro S, Teixeira MR. 2011. *MLL-SEPTIN* gene fusions in hematological malignancies. *Biol Chem* **392**: 713–724.
- Cimino G, Rapanotti MC, Elia L, Biondi A, Fizzotti M, Testi AM, Tosti S, Croce CM, Canaani E, Mandelli F, et al. 1995. *ALL-1* gene rearrangements in acute myeloid leukemia: association with M4–M5 French–American–British classification subtypes and young age. *Cancer Res* **55**: 1625–1628.
- Corral J, Lavenir I, Impey H, Warren AJ, Forster A, Larson TA, Bell S, McKenzie AN, King G, Rabbitts TH. 1996. An *MLL-AP9* fusion gene made by homologous recombination causes acute leukemia in chimeric mice: a method to create fusion oncogenes. *Cell* **85**: 853–861.
- de Rooij JD, Masetti R, van den Heuvel-Eibrink MM, Cayuela JM, Trka J, Reinhardt D, Rasche M, Sonneveld E, Alonzo TA, Fornerod M, et al. 2016. Recurrent abnormalities can be used for risk group stratification in pediatric AMKL: a retrospective intergroup study. *Blood* **127**: 3424–3430.
- de Rooij JDE, Branstetter C, Ma J, Li YJ, Walsh MP, Cheng JJ, Obulkasim A, Dang JJ, Easton J, Verboon LJ, et al. 2017. Pediatric non–Down syndrome acute megakaryoblastic leukemia is characterized by distinct genomic subsets with varying outcomes. *Nat Genet* **49**: 451–456.
- Hall PA, Russell SEH. 2004. The pathobiology of the septin gene family. *J Pathol* **204**: 489–505.
- Hara Y, Shiba N, Ohki K, Tabuchi K, Yamato G, Park MJ, Tomizawa D, Kinoshita A, Shimada A, Arakawa H, et al. 2017. Prognostic impact of specific molecular profiles in pediatric acute megakaryoblastic leukemia in non–Down syndrome. *Genes Chromosomes Cancer* **56**: 394–404.
- Kalikin LM, Sims HL, Petty EM. 2000. Genomic and expression analyses of alternatively spliced transcripts of the *MLL* septin-like fusion gene (*MSF*) that map to a 17q25 region of loss in breast and ovarian tumors. *Genomics* **63**: 165–172.
- Kartmann B, Roth D. 2001. Novel roles for mammalian septins: from vesicle trafficking to oncogenesis. *J Cell Sci* **114**(Pt 5): 839–844.
- Kreuziger LM, Porcher JC, Ketterling RP, Steensma DP. 2007. An *MLL-SEPT9* fusion and t(11;17)(q23;q25) associated with de novo myelodysplastic syndrome. *Leuk Res* **31**: 1145–1148.
- Krivtsov AV, Armstrong SA. 2007. *MLL* translocations, histone modifications and leukaemia stem-cell development. *Nat Rev Cancer* **7**: 823–833.
- Kurosu T, Tsuji K, Ohki M, Miki T, Yamamoto M, Kakihana K, Koyama T, Taniguchi S, Miura O. 2008. A variant-type *MLL/SEPT9* fusion transcript in adult de novo acute monocytic leukemia (M5b) with t(11;17)(q23;q25). *Int J Hematol* **88**: 192–196.
- Martin ME, Milne TA, Bloyer S, Galoian K, Shen W, Gibbs D, Brock HW, Slany R, Hess JL. 2003. Dimerization of *MLL* fusion proteins immortalizes hematopoietic cells. *Cancer Cell* **4**: 197–207.
- Meyer C, Burmeister T, Groger D, Tsaur G, Fechina L, Renneville A, Sutton R, Venn NC, Emerenciano M, Pombo-de-Oliveira MS, et al. 2018. The *MLL* recombinome of acute leukemias in 2017. *Leukemia* **32**: 273–284.
- Milne TA, Briggs SD, Brock HW, Martin ME, Gibbs D, Allis CD, Hess JL. 2002. *MLL* targets SET domain methyltransferase activity to *Hox* gene promoters. *Mol Cell* **10**: 1107–1117.
- Ono R, Nakajima H, Ozaki K, Kumagai H, Kawashima T, Taki T, Kitamura T, Hayashi Y, Nosaka T. 2005. Dimerization of *MLL* fusion proteins and FLT3 activation synergize to induce multiple-lineage leukemogenesis. *J Clin Invest* **115**: 919–929.

- Osaka M, Rowley JD, Zeleznik-Le NJ. 1999. MSF (MLL septin-like fusion), a fusion partner gene of MLL, in a therapy-related acute myeloid leukemia with a t(11;17)(q23;q25). *Proc Natl Acad Sci* **96**: 6428–6433.
- Peterson EA, Stanbery L, Li C, Kocak H, Makarova O, Petty EM. 2011. SEPT9_i1 and genomic instability: mechanistic insights and relevance to tumorigenesis. *Genes Chromosomes Cancer* **50**: 940–949.
- Saito H, Otsubo K, Kakimoto A, Komatsu N, Ohsaka A. 2010. Emergence of two unrelated clones in acute myeloid leukemia with MLL–SEPT9 fusion transcript. *Cancer Genet Cytogenet* **201**: 111–115.
- Santos J, Cerveira N, Correia C, Lisboa S, Pinheiro M, Torres L, Bizarro S, Vieira J, Viterbo L, Mariz JM, et al. 2010. Coexistence of alternative MLL–SEPT9 fusion transcripts in an acute myeloid leukemia with t(11;17)(q23;q25). *Cancer Genet Cytogenet* **197**: 60–64.
- Schoch C, Schnittger S, Klaus M, Kern W, Hiddemann W, Haferlach T. 2003. AML with 11q23/MLL abnormalities as defined by the WHO classification: incidence, partner chromosomes, FAB subtype, age distribution, and prognostic impact in an unselected series of 1897 cytogenetically analyzed AML cases. *Blood* **102**: 2395–2402.
- Shih LY, Liang DC, Fu JF, Wu JH, Wang PN, Lin TL, Dunn P, Kuo MC, Tang TC, Lin TH, et al. 2006. Characterization of fusion partner genes in 114 patients with de novo acute myeloid leukemia and MLL rearrangement. *Leukemia* **20**: 218–223.
- Smith MA, Rubinstein L, Ungerleider RS. 1994. Therapy-related acute myeloid leukemia following treatment with epipodophyllotoxins: estimating the risks. *Med Pediatr Oncol* **23**: 86–98.
- So CW, Cleary ML. 2003. Common mechanism for oncogenic activation of MLL by forkhead family proteins. *Blood* **101**: 633–639.
- So CW, Lin M, Ayton PM, Chen EH, Cleary ML. 2003. Dimerization contributes to oncogenic activation of MLL chimeras in acute leukemias. *Cancer Cell* **4**: 99–110.
- Strehl S, Konig M, Meyer C, Schneider B, Harbott J, Jager U, von Bergh AR, Loncarevic IF, Jarosova M, Schmidt HH, et al. 2006. Molecular dissection of t(11;17) in acute myeloid leukemia reveals a variety of gene fusions with heterogeneous fusion transcripts and multiple splice variants. *Genes Chromosomes Cancer* **45**: 1041–1049.
- Taki T, Ohnishi H, Shinohara K, Sako M, Bessho F, Yanagisawa M, Hayashi Y. 1999. AF17q25, a putative septin family gene, fuses the MLL gene in acute myeloid leukemia with t(11;17)(q23;q25). *Cancer Res* **59**: 4261–4265.
- Tallman MS, Neuberger D, Bennett JM, Francois CJ, Paietta E, Wiernik PH, Dewald G, Cassileth PA, Oken MM, Rowe JM. 2000. Acute megakaryocytic leukemia: the Eastern Cooperative Oncology Group experience. *Blood* **96**: 2405–2411.
- Wechsler J, Greene M, McDevitt MA, Anastasi J, Karp JE, Le Beau MM, Crispino JD. 2002. Acquired mutations in GATA1 in the megakaryoblastic leukemia of Down syndrome. *Nat Genet* **32**: 148–152.
- Yamamoto K, Shibata F, Yamaguchi M, Miura O. 2002. Fusion of MLL and MSF in adult de novo myelomonocytic leukemia (M4) with t(11;17)(q23;q25). *Int J Hematol* **75**: 503–507.
- Zeisig BB, Schreiner S, García-Cuellar MP, Slany RK. 2003. Transcriptional activation is a key function encoded by MLL fusion partners. *Leukemia* **17**: 359–365.