

Review



# Neuroinflammation Modulation via $\alpha$ 7 Nicotinic Acetylcholine Receptor and Its Chaperone, RIC-3

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**Abstract:** Nicotinic acetylcholine receptors (nAChRs) are widely expressed in or on various cell types and have diverse functions. In immune cells nAChRs regulate proliferation, differentiation and cytokine release. Specifically, activation of the  $\alpha$ 7 nAChR reduces inflammation as part of the cholinergic anti-inflammatory pathway. Here we review numerous effects of  $\alpha$ 7 nAChR activation on immune cell function and differentiation. Further, we also describe evidence implicating this receptor and its chaperone RIC-3 in diseases of the central nervous system and in neuroinflammation, focusing on multiple sclerosis (MS) and its animal model, experimental autoimmune encephalomyelitis (EAE). Deregulated neuroinflammation due to dysfunction of  $\alpha$ 7 nAChR provides one explanation for involvement of this receptor and of RIC-3 in neurodegenerative diseases. In this review, we also provide evidence implicating  $\alpha$ 7 nAChRs and RIC-3 in neurodegenerative diseases such as Alzheimer's disease (AD) and Parkinson's disease (PD) involving neuroinflammation. Besides, we will describe the therapeutic implications of activating the cholinergic anti-inflammatory pathway for diseases involving neuroinflammation.

**Keywords:** cholinergic anti-inflammatory pathway; neuroinflammation; α7 nicotinic acetylcholine receptor; RIC-3; multiple sclerosis; experimental autoimmune encephalomyelitis; Parkinson's disease; Alzheimer's disease

# 1. Introduction

Nicotinic acetylcholine receptors (nAChRs) belong to a large and diverse family of acetylcholine-gated cationic ion channels. In mammals this family is composed of nine alpha subunits, whose N-terminus contains two disulfide linked vicinal cysteines enabling ligand binding, and seven non-alpha subunits. Most nAChR subunits assemble to form heteromeric receptors whereas  $\alpha$ 7 and  $\alpha$ 9 nAChR subunits can form homomeric receptors. Folding assembly and trafficking of mature nAChRs from the endoplasmic reticulum to the plasma membrane is a complex process which requires assistance from multiple cellular factors. Several proteins were shown to affect functional expression and properties of nAChRs including Resistance to Inhibitors of Cholinesterase-3 (RIC-3) and NACHO [1,2].

The nAChRs express widely and have multiple functions. In skeletal muscle nAChRs mediate excitation in the neuromuscular junction. Similarly, in the autonomic nervous system nAChRs are needed for excitatory synaptic transmission [3,4]. Functions of nAChRs, however, are not limited to synapses and to excitatory synaptic transmission. In the central nervous system (CNS) nAChRs have mostly modulatory roles, including a role in regulating release of neurotransmitters [5]. Moreover, these receptors are expressed in many non-excitable cells where they have been shown to affect migration, differentiation and proliferation [6].

CNS-expressed nAChRs have been implicated in several neurodegenerative diseases. Evidence supporting involvement of nAChRs in these diseases includes epidemiological



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**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). studies linking some of these diseases to tobacco smoking and gene expression studies. Specifically, smoking reduces the prevalence of Parkinson's disease (PD) [7]; schizophrenia is associated with heavy smoking [8]; and decreased brain expression of nAChRs was shown in Alzheimer's disease (AD), PD, Lewy-body dementia, and schizophrenia [9–12], but the mechanisms underlying involvement of nAChRs in these diseases are, yet, unclear.

This review will focus on the Immunomodulatory roles of nAChRs. Neuronal nAChR subunits (subunits that are expressed in neurons and are not part of the skeletal muscle nAChR) are expressed in immune cells and in glial cells (microglia and astrocytes). Specifically, expression analysis in mice showed that microglia express all nine neuronal nAChR subunits examined while other immune cells express subsets of these subunits. Two of these subunits  $\alpha 9$  and  $\beta 2$  showed high expression in all of the cells examined except regulatory T-cells (Treg).  $\alpha 5$ ,  $\alpha 6$ ,  $\alpha 7$  and  $\beta 4$  subunits showed significant, although weaker, expression in several cell types (CD4<sup>+</sup>, CD8<sup>+</sup>, CD11c<sup>+</sup> and CD11b<sup>+</sup>), while the expression of  $\alpha 3$  and  $\beta 3$  was limited to microglia [13].

In immune cells, nAChRs have immunomodulatory roles and among them the best studied role is of the  $\alpha$ 7 nAChR as part of the cholinergic anti-inflammatory pathway. This pathway was first described in the experimental model of septic shock. In this model, inflammatory activation is induced by lipopolysaccharides (LPS) injection leading to increased production of pro-inflammatory cytokines (such as IL-1 and TNF- $\alpha$ ), micro-circulatory failure and systemic hypotension. Vagal stimulation attenuated the pathology associated with septic shock an effect requiring  $\alpha$ 7 nAChR, whose activation in macrophages inhibited pro-inflammatory cytokine production from these cells. Importantly,  $\alpha$ 7 nAChR was required for the inflammation reducing effects of vagal stimulation (Figure 1A left and [14,15]). More details on the cholinergic anti-inflammatory pathway are provided below.



**Figure 1.** Immunomodulation mediated by nAChRs. (**A**) Left pathway, the splenic vagal nerve activity via the cholinergic anti-inflammatory pathways reduces inflammation within and outside the spleen. Right pathway, the sympathetic splenic nerve via neuroadrenaline (NA) release in the spleen promotes antibody production. (**B**) An anti-inflammatory negative feedback loop involving monocyte expressed nAChRs.

Additional immunomodulatory effects of activating immune cell-expressed nAChRs are emerging. One such pathway depends on  $\alpha$ 9 and  $\alpha$ 10, with  $\alpha$ 7 having a minor role. This pathway inhibits the pro-inflammatory effects of ATP on monocytes thus reducing IL-1 $\beta$  release from these cells [16]. Interestingly, C-reactive protein (CRP)–a blood marker for inflammation that is produced by the liver in response to IL-1 $\beta$ –is an agonist for this nAChR [4]. These findings suggest a nAChR-dependent negative feedback loop reducing IL-1 $\beta$  release and the deleterious effects resulting from its overproduction (Figure 1B).

Recent findings identified a pro-inflammatory pathway involving nAChRs and acetylcholine. Activity of this pathway is initiated by neurons in the central nucleus of the amygdala (CeA) and in the paraventricular nucleus (PVN) of the hypothalamus. Both types of neurons express the stress related corticotropin-releasing hormone (CRH). Activation of these neurons via stimulation of the sympathetic splenic nerve leads to acetylcholine (ACh) release from T-cells which activates B-cell expressed  $\alpha$ 9 nAChR to promote plasma cell formation and IgG antibody production (Figure 1A right and [17]).

## 2. α7. nAChR and the Cholinergic Anti-Inflammatory Pathway

 $\alpha$ 7 nAChR, a key player in the cholinergic anti-inflammatory pathway, was first identified as the  $\alpha$ -bungarotoxin ( $\alpha$ -BTX) binding receptor in the CNS [18]. This receptor has high permeability to calcium, likely to enable some of its effects on cellular functions [19]. In addition,  $\alpha$ 7 receptors are activated by ACh and by choline, a precursor and breakdown product of ACh as well as a breakdown product of membrane phospholipids. This enables  $\alpha$ 7-mediated responses to both targeted transmission and to localized tissue damage.

In immune cells  $\alpha$ 7 nAChR activation leads to phosphorylation and activation of Janus kinase 2 (JAK2) and to phosphorylation and nuclear entry of Signal Transducer and Activator of Transcription 3 (STAT3) which inhibits expression of pro-inflammatory cytokines. In parallel  $\alpha$ 7 nAChR activation reduces degradation of IkB $\alpha$  therefore inhibiting nuclear translocation of NF-k $\beta$  and the expression of pro-inflammatory cytokines. Interestingly, multiple lines of evidence suggest metabotropic activity of  $\alpha$ 7 nAChR in immune cells. Mechanisms enabling the immune modulating effects of  $\alpha$ 7 nAChR are reviewed in [20].

# 3. Cholinergic Signaling via nAChRs in Immune Cells

The cholinergic anti-inflammatory pathway was initially described by its ability to inhibit inflammation through activation of macrophage expressed  $\alpha$ 7 nAChR [14,15,21]. Below we provide detailed description on the immunomodulatory effects of activating nAChRs in different immune cell types, including CNS resident glial cells having immune functions. A summary of this information is provided in Figure 2.



Figure 2. Cholinergic effects of  $\alpha$ 7 nAChR on immune and glial cells.

# 3.1. $\alpha$ 7. *nAChR* in the Periphery

# 3.1.1. Macrophages

The  $\alpha$ 7 nicotinic receptor expressed in macrophages is involved in the regulation of inflammatory processes. Activation of  $\alpha$ 7 nAChR using acetylcholine, nicotine or other agonists decreases pro-inflammatory cytokine production in response to LPS. Unlike production of anti-inflammatory cytokines which is unaffected by these agonists. The mechanism implicated was reduced NF- $\kappa$ B-mediated transcription. [14,22] Involvement of NF- $\kappa$ B-mediated transcription in this pathway is consistent with its anti-inflammatory effects, as many pro-inflammatory mediators are transcribed by this transcription factor [23].

In addition to suppressing the inflammatory response to LPS, activation of  $\alpha$ 7 nAChR suppressed pro-inflammatory cytokine production in response to activation of various toll-like receptors (TLRs) such as TLR2, TLR3, and TLR9 [24].

The anti-inflammatory effects of ACh are mimicked by nicotine, but not by muscarine, an agonist of muscarinic acetylcholine receptors, and these effects depend on  $\alpha$ 7 nAChR. These effects are eliminated when macrophages were exposed to antisense oligonucleotides specific for the  $\alpha$ 7 nAChR or in loss of function mutants for the gene that codes for this receptor (*CHRNA7*) [15,22].

Further evidence for anti-inflammatory effects of activating macrophage expressed  $\alpha$ 7 nAChRs were provided in experiments using a specific activator of this receptor, the allosteric agonist and positive allosteric modulator (ago-PAM) GAT107. Application of this drug following LPS stimulation of a murine macrophages cell line (RAW264.7) significantly reduced IL-6 secretion by these cells. These anti-inflammatory effects were eliminated by methyllycaconitine (MLA), a specific antagonist of  $\alpha$ 7 nAChR [25].

## 3.1.2. Dendritic Cells

Mashimo et al. (2020) [26] established the role of the cholinergic inflammatory system in dendritic cells (DCs), reporting that these cells express nAChR subunits, including  $\alpha$ 7 nAChR. DCs can produce ACh and express the enzymes for ACh synthesis and degradation, choline acetyltransferase (ChAT) and acetylcholine esterase (AChE). In addition, they express Secreted Ly-6/uPAR-related protein 1 (SLURP-1), the endogenous peptide that acts as a positive allosteric ligand for  $\alpha$ 7 nAChR. Overall, DCs express most of the components necessary for a functional cholinergic system [26,27].

We and others have shown decreased antigen presentation following cholinergic activation in vivo [28,29]. Additionally, following nicotine treatment, the expression of co-stimulatory molecules such as CD80, CD86 and MHCII on antigen presenting cells (APCs) was reduced [30]. In vitro exposure of human DCs to nicotine decreased secretion of pro-inflammatory cytokines and reduced DCs phagocytic activity. Moreover, a direct effect of nicotine on DCs was also found when DCs were cultured with nicotine and failed to mount an effective Th<sub>1</sub> response in T-cells [31].

## 3.1.3. T-Cells

Proliferation of specific T-Cells is crucial to mount an effective adaptive immune response. Murine and human T-cells express  $\alpha$ 7 nAChR, and this expression is increased following antigenic or mitogenic stimulation [28,32]. Our recent study on Experimental Autoimmune Encephalomyelitis (EAE) mice treated with the  $\alpha$ 7 nAChR ago-PAM, GAT107 showed reduced ex-vivo antigen specific induced T-cell proliferation and reduced secretion of pro-inflammatory cytokines (IL-17, IL-6 and IFN $\gamma$ ) while secretion of the anti-inflammatory cytokine, IL-10, was increased [25]. A similar result of reduced T-cell activity was shown when cholinergic agents such as Rivastigmine, an acetylcholine esterase inhibitor (ACHEI), or EN101 (an anti-sense oligonucleotide to exon 2 of AChE) were used. Blocking of  $\alpha$ 7 nAChR by using pharmacology and anti-sense oligonucleotides eliminated this effect [28,33]. Exposure to AChEI increases extracellular ACh concentration and induces cholinergic receptor activation. This mechanism is similar to the therapeutic mechanism by which AChEIs function in myasthenia gravis (MG) or Alzheimer's disease (AD).

A recent study using an experimental inflammatory bowel disease model described nicotine-induced suppression of the Th<sub>2</sub> lineage activity and an increase in Th<sub>1</sub> and Th<sub>17</sub> activity [34]. Notably, this effect of nicotine was correlated with the expression level of the  $\alpha$ 7 nAChR: the inhibitory effect of nicotine on Th<sub>2</sub> lineage was correlated with higher expression of  $\alpha$ 7 nAChR on these T-cells, while the failure to suppress the Th<sub>1</sub> lineage correlated with lower expression of  $\alpha$ 7 nAChR on these cells. Thus, it seems that nicotine does not suppress a specific Th lineage, but rather may inhibit any active lineages as long as  $\alpha$ 7 nAChR is expressed on these cells.

Mashimo et al. demonstrated that  $\alpha$ 7 nAChRs expressed on T-cells and APCs have different effects on naïve CD4+ T-cell differentiation. Initially, activation of  $\alpha$ 7 nAChRs on APCs reduced antigen processing and thereby resulted in suppression of T-cell differentiation, but activation of  $\alpha$ 7 nAChRs on T-cells increased T-cell differentiation [35]. Immune activation of both naïve  $\alpha$ 7 nAChR knockout (KO) and of wild-type CD4+ T-cells via TCR/CD3 activation increased their differentiation into T regulatory cells (Tregs) and effector T-cells. Furthermore, the  $\alpha$ 7 nAChR agonist, GTS-21, further enhanced the differentiation of wild-type naïve CD4+ T-cells into Tregs and effector T-cells, but without any effect on cells that do not express the  $\alpha$ 7 nAChR. These results confirm that these effects on T-cell differentiation are mediated by  $\alpha$ 7 nAChR. T-cells and APCs (DCs and macrophages) express ChAT and therefore synthesize ACh, and their immunological activation enhances ACh synthesis [26,36]. ACh released from these activated immune cells acts via  $\alpha$ 7 nAChRs in an autocrine and/or paracrine manner. Thus,  $\alpha$ 7 nAChRs on T-cells and APCs is likely to be part of an auto-inhibitory mechanism [37].

Tregs from naive mice express  $\alpha$ 7 nAChR, and its activation by nicotine enhanced the inflammation suppressing capacity of these cells. Nicotine stimulation up-regulated the expression of cytotoxic T-lymphocyte-associated antigen (CTLA)-4 and forkhead/winged helix transcription factor p3 (Foxp3) on Tregs [38]. Our recent study also shows that activation of  $\alpha$ 7 nAChR up-regulated Foxp3 expression and, therefore, supports the role of  $\alpha$ 7 nAChR in regulating the immunomodulatory effects of Tregs during neuroinflammation [25].

## 3.1.4. B-Cells

Several studies have showed that B-cells express nAChR subunits [39–41]. Mice deficient for both  $\alpha$ 7 or  $\beta$ 2 nAChR subunits had less IgG-producing cells, and less serum IgG. However, cells from deficient mice produce more antibodies in response to activation, caused by increased CD40 expression. Interestingly, the inhibitory effects of nicotine on antibody production could only be demonstrated in the absence of the  $\beta$ 2 subunit [42]. In another study,  $\alpha$ 7 deficient mice had increased serum IgG1 levels, increased specific IgG1 production in response to antigen, and higher production of IL-6, TNF- $\alpha$  and IFN- $\gamma$  by T-cells from the same mice. This increased antibody production can be attributed to the effect of  $\alpha$ 7 on CD4 T-helper cells, which induce B-cell antibody production [43]. Overall, there is evidence that different nAChR subtypes play non-identical roles in B-lymphocytes [44].

Recently, we reported that the  $\alpha$ 7 ago-PAM GAT107 also affects B-cell function [25]. Mice with EAE showed a significant reduction in B-cell markers and lower pathogenic antibody production, following GAT107 treatment. B-cell activating factor (BAFF) and A proliferation-inducing ligand (APRIL) as well as B220, were significantly reduced by the activation of  $\alpha$ 7 nAChR. BAFF is a potent survival factor for B-cells that plays an essential role in the preservation and maturation of peripheral B-cells and APRIL is important for B-cell development. In addition to expressing nAChRs B-cells express ChAT mRNA, AChE mRNA and contain ACh. Also, activation of TLR by LPS in murine B cells, led to activation of cholinergic function by enhancing ACh synthesis [26].

## 3.1.5. Natural Killer Cells

Natural killer (NK) cells play a cytotoxic role against target cells and modulate the innate and the adaptive response via dendritic cells. Zanetti et al. (2016) showed that NK cells express the  $\alpha$ 7 nAChR, and that its expression was increased following activation by cytokines (IL-12, IL-18, and IL-15). It is important to note that the  $\alpha$ 7 nAChR is functional in NK cells, since activation with a  $\alpha$ 7 nAChR agonist increases intracellular calcium concentration, mainly released from intracellular stores. In addition, activation of  $\alpha$ 7 nAChR in Human NK Cells regulates NKG2D expression (a NK cell activating receptor).

NK cells that were exposed to specific  $\alpha$ 7 nAChR agonist-PNU-282987 display significantly reduced cytotoxic activity. Also, while NK cells are stimulated with cytokines (IL-12, IL-18, and IL-15) and exposed to  $\alpha$ 7 nAChR agonist- PNU-282987, the percentage of IFN $\gamma$  producing NK cells was lower. Those effects of PNU-282987 were abolished in the presence of  $\alpha$ -BTX, an antagonist of  $\alpha$ 7 nAChR [45].

## 3.2. a7 nAChR in CNS Glial Cells

#### 3.2.1. Microglia

Similar to the findings in macrophages, the same effects of nicotine can be demonstrated on microglia, the brain-resident macrophages [46]. Microglia express both  $\alpha$ 7 nAChR mRNA and protein. Furthermore, microglia in co-culture with neurons increased pro inflammatory factor secretion following LPS stimulation. And, ACh reduced secretion of pro inflammatory factors and reduced the pro-apoptotic effects of activated microglia on neurons. These effects of ACh were abolished when  $\alpha$ 7nAChR was silenced using  $\alpha$ 7 nAChR-shRNA [47].

LPS-stimulated microglia that were also exposed to nicotine or ACh showed reduced TNF- $\alpha$  release. This reduction was mediated through MAP-kinase inhibition. Of note, reduction in TNF- $\alpha$  release by microglia was demonstrated for  $\alpha$ 7 nAChR selective ligands that are poor channel activators or even antagonists of channel activity; these effects suggest that in microglia  $\alpha$ 7 nAChR does not function as an ion channel [48].

Neuroinflammation induced by LPS in primary fetal sheep microglia cultures resulted in pro-inflammatory microglial phenotype. Exposure to an agonist of  $\alpha$ 7 nAChR reversed this phenotype. Conversely, an antagonist of  $\alpha$ 7 nAChR potentiated the pro-inflammatory microglial phenotype. Hepcidin-ferroprotein signaling and iron homeostasis were implicated in maintaining the microglial inflammatory phenotype and in the effects of  $\alpha$ 7 nAChR signaling on this phenotype [49].

Over activation of microglia is associated with aggravation of EAE in mice [50]. Ke et al., showed that activating  $\alpha$ 7 nAChR can inhibit NLRP3 inflammasome via the regulation of  $\beta$ -arrestin-1 in microglia, contributing to the suppression of neuroinflammation and attenuation of EAE severity [51]. Similarly, we have shown that treatment of EAE mice with the ACEI, rivastigmine, led to clinical improvement that was associated with reduced activation of microglia and preservation of neurofilaments [28].

#### 3.2.2. Astrocytes

The  $\alpha$ 7 nAChRs are also expressed by astrocytes [52] and there is evidence that those receptors can affect synaptic transmission in the CNS [53]. As activation of  $\alpha$ 7 nAChR increased expression of post synaptic  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors in hippocampal GABAergic neurons. This led to increased AMPA receptor mediated synaptic currents which in turn can reinforce the activity of specific GABAergic synapses. In addition to these neuromodulatory effects, astrocyte expressed  $\alpha$ 7 nAChR has immunoregulatory roles; using a  $\alpha$ 7 nAChR agonist, GTS21, in astrocyte cultures stimulated with LPS, attenuated IL-6 and TNF- $\alpha$  secretion, as well as reduced inflammatory astrocyte induced apoptosis of neuronal cells. This effect was reversed by the  $\alpha$ 7 nAChR antagonist MLA, indicating the specificity of this response [52].

Further support for the neuroprotective effect of  $\alpha$ 7 nAChR in astrocytes is provided by the  $\alpha$ 7 nAChR agonist PNU-282987, which reduced apoptosis of neurons in glia-neuron co-culture and reduced glutamate induced toxicity of these neurons. These effects were achieved via inhibition of TGF $\beta$  signaling. In addition, activation of the  $\alpha$ 7 nAChR on astrocytes resulted in decreased neurotoxicity of the chemotherapeutic agent (oxaliplatin) [54].

Cao et al. performed RNA-Seq analysis in primary fetal astrocyte cultures exposed to LPS together with a selective  $\alpha$ 7 nAChR agonist or antagonist. Stimulation by a  $\alpha$ 7 nAChR agonist promoted the neuroprotective profile of astrocytes, and  $\alpha$ 7 nAChR inhibition reduced it; LPS double-hit (first in vivo, then in vitro) on astrocytes exacerbates these effects [55].

## 4. α7 nAChR and CNS Diseases

In mice loss of function mutations in subunits contributing to major CNS expressed nAChRs ( $\alpha$ 7,  $\alpha$ 4 and  $\beta$ 2) have no easily discernible phenotypes [56]. But, in humans copy number variations of the gene encoding for  $\alpha$ 7 nAChR subunit (*CHRNA7*) are associated with brain diseases such as epilepsy or autism [57]. In addition, a 2bp deletion in of the gene encoding for a human-specific chimeric protein, CHRFAM7A, containing a duplicate of exons 5-10 of *CHRNA7*, shows strong linkage to schizophrenia [58]. Instability of the genomic region harboring the *CHRNA7* complicates its analysis, as disease causing deletions encompassing this gene also cover additional genes. Since, heterozygosity for rare small deletions covering *CHRNA7* alone is associated with similar phenotypes to those associated with larger deletions in this region it is likely that haplo-insufficiency of this gene is a cause for disease in humans [57], but some heterozygous carriers of *CHRNA7* containing deletions are phenotypically normal. This confounding finding may be explained by incomplete penetrance of these deletions due to interactions with the genetic background or with the environment [57].

The CHRFAM7A chimeric protein, which is unique to humans, was shown to interact with and inhibit surface expression of  $\alpha$ 7 nAChRs [58,59]. This chimeric protein expresses in immune cells and its expression in these cells is regulated by LPS, nicotine and IL1 $\beta$ . Thus, this protein is likely to be a human specific regulator of the cholinergic anti-inflammatory pathway [58,59].

Of note, involvement of  $\alpha$ 7 nAChR in neurodegenerative diseases may stem, at least partly, from its immunomodulatory effects, which in the CNS are likely to involve astrocytes and microglia [60]. As described above,  $\alpha$ 7 nAChR is expressed in glial cells and was shown to affect their activity.

#### 5. RIC-3 and $\alpha$ 7 nAChRs in Neurodegenerative Diseases

RIC-3 was first characterized in *C. elegans* as an ER-resident protein that is needed for the maturation of multiple nAChRs but not for the maturation other ligand-gated ion channels [1]. Later analysis showed conservation of RIC-3 sequence and function during evolution [61]. Moreover, RIC-3 is required for heterologous expression of the  $\alpha$ 7 nAChR in non-neuronal mammalian cells [62].

Analysis of *ric-3* loss-of-function mutants in *C. elegans* demonstrated reduced functional expression of multiple nAChRs in these mutants. Therefore, RIC-3 was suggested to promote surface expression of co-expressed nAChRs [1], but later analysis demonstrated both positive and negative effects of RIC-3 on co-expressed receptors depending on identity of the co-expressed receptor and on the expression system [61,63]. One explanation for these different effects is the finding that altering RIC-3-to-nAChR ratio shifts its effects from positive to negative [64,65].

RIC-3 was expected to promote expression of  $\alpha$ 7 nAChR in-vivo, as functional expression of  $\alpha$ 7 nAChR in non-neuronal cells requires co-expression with RIC-3 [62] and expression patterns of *ric3* and the  $\alpha$ 7 nAChR subunit overlap [61]. However, knockout of *ric3* did not significantly affect  $\alpha$ 7 nAChR expression in the brains of mice [66]. Nevertheless, its knockdown using siRNA-mediated silencing eliminated the anti-inflammatory effects of GAT107, an  $\alpha$ 7 nAChR-specific agonist [67]. This suggests cell-specific requirement of RIC-3 for functional expression of  $\alpha$ 7 nAChR.

RIC-3 like  $\alpha$ 7 nAChR has been implicated in diseases of the central nervous system: *ric3* expression was increased in bipolar disease and in Schizophrenia [68]. Genome wide association studies (GWAS) identified polymorphisms in the *ric3* region in association with multiple sclerosis (MS) [69,70]. GWAS also suggested that ric3 variants modify cognitive maintenance in the elderly [71]. Mutations in *ric3* were found in PD patients [72].

# 6. α7 nAChR and RIC-3 in Neurodegenerative Diseases Involving Neuroinflammation

The well-established role of  $\alpha$ 7 nAChR in the cholinergic anti-inflammatory pathway suggests that  $\alpha$ 7 may be involved in neuroinflammatory diseases, as reviewed by [20,60]. RIC-3, like  $\alpha$ 7 nAChR is expressed in immune cells. Expression of *ric3* is regulated by pro-inflammatory stimuli, and its knockdown eliminates the anti-inflammatory effects of a  $\alpha$ 7 nAChR-specific agonist [65,67]. Thus, RIC-3 like  $\alpha$ 7 nAChR is likely to be involved in neuroinflammatory diseases. Below we will provide evidence implicating the cholinergic anti-inflammatory pathway,  $\alpha$ 7 nAChR and RIC-3 in three neurodegenerative diseases: MS, AD and PD. While neuroinflammation is causal for MS its role in AD and PD is less clear. Nevertheless, neuroinflammation is likely to contribute to progression of these diseases [73]. Therefore, we will also describe attempts to harness the cholinergic anti-inflammatory as therapy for these diseases.

# 7. Involvement of α7 nAChR and RIC-3 in EAE

EAE is the animal model used to study MS, and for the study of neuroinflammation. Using the chronic EAE model in C57bl mice we showed that treatment of EAE with EN101, an anti-sense oligodeoxynucleotide, targeted to AChE mRNA, reduced EAE clinical severity and CNS inflammation intensity. This clinical amelioration was accompanied by reduction in lymphocyte proliferation and reduced inflammatory infiltrates to the spinal cord. These results suggest that AChEIs increase the concentration of extracellular ACh, rendering it available for interaction with a nicotinic receptor expressed on lymphocytes [32].

In addition, a marked improvement in EAE severity was found by treatment with rivastigmine. Rivastigmine is a pseudo-irreversible AChEI that was shown to ameliorate cognitive dysfunction in AD [74]. Rivastigmine treatment of EAE mice ameliorated disease severity, reduced demyelination, and showed a multi-level activity on various stages of the immune response in EAE, as well as improving cognitive function [28].

In another set of experiments, we found that treatment with bifunctional compounds comprising both a cholinergic up-regulation moiety (the AChEI pyridostigmine or cytisine) and a nonsteroidal anti-inflammatory moiety (ibuprofen), ameliorated clinical symptoms in the same EAE model [32,75]. In this setting the presence of the cholinergic up-regulation moiety increased the anti-inflammatory effects and reduced the pro-inflammatory effects.

Nicotine administration also ameliorated EAE severity and reduced T-cell proliferation in response to encephalitogenic antigens, as well as the production of cytokines by Th1 cells (TNF $\alpha$  and IFN $\gamma$ ) and Th17 cells (IL-17, IL-17F, IL-21, and IL-22) [29].

Recently, we examined effects of GAT107, a  $\alpha$ 7 nAChR specific ago-PAM, on EAE mice. Administration of GAT107 to EAE mice, resulted in EAE amelioration, accompanied by reduced inflammatory infiltrates to the spinal cord. In addition, the expression of immune cell markers was altered by GAT107 treatment, which induced a significant reduction in macrophages, dendritic cells, and B cells, as well as a reduction in the production of specific antibodies [25].

In addition to our results, others have reported that treatment with nicotine ameliorates EAE severity, reduced CNS infiltrates, and reduced T-cell reactivity in this model [13,76]. Of note, some effects of nicotine on EAE were attributed to  $\alpha$ 9 and  $\beta$ 2 containing nAChRs. These receptors were suggested to play different roles from  $\alpha$ 7 nAChR in endogenous ACh-and in nicotine-dependent modulation of immune functions [77]. Table 1 summarizes information from studies using the EAE model to examine the therapeutic effects of activating the cholinergic anti-inflammatory pathway.

Cholinergic Agent Used for EAE Treatment	CNS Pathology	T-Cell Activity	B Cells	Reference
EN101	Reduced inflammatory Infiltrates in the spinal cord	Reduced proliferation and proinflamma- tory cytokine secretion		Nizri et al., 2005
IBU-PO IBU-PD	Reduced inflammatory activation of astrocytes in vitro	Reduced proliferation		Nizri et al., 2005
IBU-Octyl- Cytisine	Reduced inflammatory Infiltrates in the spinal cord	Reduced proliferation and proinflamma- tory cytokine production		Niziri et al., 2007
Rivastigmine (AChEI)	Reduced demyelination, microglia activation and axonal damage	Reduced proliferation and proinflamma- tory cytokine production		Nizri et al., 2008
Nicotine	Reduced CNS inflammatory infiltrates, decreased demyelination and reduced axonal loss	Reduced proliferation and proinflamma- tory cytokine secretion		Nizri et al., 2009 Hao et al., 2011
GAT107	Reduced inflammatory infiltrates in the spinal cord	Reduced proliferation and proinflamma- tory cytokine production	Reduced B- cell number Reduction of B-cell markers' expression Reduced ab production	Mizrachi et al., 2021

**Table 1.** EAE amelioration via  $\alpha$ 7 nAChR activation. Summary of studies using the EAE model to examine the therapeutic effects of activating the cholinergic anti-inflammatory pathway.

Besides, therapeutic effects of cholinergic agents on EAE, we showed that induction of EAE in mice is associated with dynamic changes in expression of *ric3*, encoding for RIC-3, in the periphery and in the CNS. Furthermore, similarities in the expression dynamics of *ric3* and *CHRNA7*, encoding for the  $\alpha$ 7 nAChR subunit, were found. Moreover, *ric3* was found to be required for the anti-inflammatory effects of cholinergic agonists in RAW264.7 cells. [67].

# 8. α7 nAChR, RIC-3 and Multiple Sclerosis

The beneficial, clinical, immunological and neuropathology reducing effects of activating the cholinergic anti-inflammatory pathway in EAE mice, led to several studies examining this pathway in MS patients. This analysis showed increased responsiveness of peripheral blood mononuclear cells (PBMCs) to the anti-inflammatory effects of nicotine, as shown by its effects on PHA-dependent pro-inflammatory cytokine release. This enhanced responsiveness correlated with increased expression of  $\alpha$ 7 nAChR in PBMCs stimulated with nicotine and PHA in MS patients but not in healthy donors [78] Moreover, treatment of MS patients with IFN- $\beta$  led to reduced serum levels of pro-inflammatory cytokines and increased serum ACh levels; suggesting that circulating cytokines and ACh are co-regulated [79]. In addition, levels of ACh and of the enzymes responsible for its synthesis and degradation were altered in the serum of MS patients [79,80]. Analysis of NK cells from MS patients showed increased intracellular ACh levels. This increase correlated with disease severity as seen by the EDSS score, lesion number and lesion volume [81]. Together, these changes suggest compensatory mechanisms occurring in MS patients aimed at reducing inflammation via enhanced activity of the cholinergic anti-inflammatory pathway. Previous reports showed low levels of ACh in the CSF and in the serum of MS patients when compared to healthy controls; this effect is opposite to the high levels of pro-inflammatory cytokines detected in these patients, supporting a relationship between pro-inflammatory cytokines and ACh levels in MS patients [82].

The beneficial effects of nicotine on disease pathology in the animal model for MS suggested therapeutic potential for nicotine in MS patients. Tobacco smoking, however, is a well-known risk factor for MS [83]. Although, an epidemiological study undertaken in Sweden showed reduced MS risk in users of moist snuff, also a source of nicotine [84]. Immunomodulatory effects of tobacco smoke-derived nicotine are implicated as a risk factor for MS by a study showing that polymorphisms in loci encoding for  $\alpha$ 7 and  $\alpha$ 9 nAChRs modify the association between smoking and MS [85]. Alternatively, it was suggested that while tobacco-derived nicotine ameliorates MS pathology other components of tobacco smoke have detrimental effects. Evidence supporting this suggestion were provided from analysis of the EAE model for MS [86].

RIC-3 the chaperone for α7 nAChR was also implicated in MS. Genome-wide association studies detected an association between single nucleotide polymorphisms in the region encoding for this protein and MS or specific types of MS lesions [69,70]. The known role of the RIC-3 protein in  $\alpha$ 7 nAChR's maturation, combined with the known roles of  $\alpha$ 7 nAChR in inflammation and its likely involvement in MS, suggest that genetic variants in the RIC-3 encoding gene (*ric3*) may affect MS via their effects on  $\alpha$ 7 nAChR's maturation in immune cells. Support for this hypothesis comes from our recent experiments showing that siRNA-mediated knockdown of ric3 in a macrophage cell line eliminated the antiinflammatory effects of GAT107, a  $\alpha$ 7 nAChR specific agonist. In addition, in support of variable expression and function of *ric-3* and *CHRNA7* in some MS patients', it should be noted that ric-3 expression is increased in PBMCs from some MS patients and this increase correlates with increased expression of CHRNA7 (encoding for  $\alpha$ 7 nAChR) [67]. Besides, GAT107 reduced pro-inflammatory cytokine (IL-6 and IL-17) secretion from PBMCs derived from MS and HD, and this cholinergic effect was more variable in the MS patients [25]. Future, work should examine how this increased expression of ric3 and CHRNA7 affects MS progression and the relationship between the *ric3* polymorphisms associated with MS and ric3 expression.

Currently treatment of MS at its neurodegenerative stage needs improvement. Findings showing impairment of the cholinergic anti-inflammatory pathway in MS and studies showing that activation of this pathway ameliorates inflammatory and neurodegenerative components of this disease (Table 1) support the cholinergic anti-inflammatory pathway as target for new MS drugs.

# 9. Parkinson's Disease, Neuroinflammation and the $\alpha$ 7 nAChR

Parkinson's disease is a neurodegenerative disease whose hallmark is degeneration of dopaminergic neurons in the substantia nigra. A1 astrocytes which are a marker for neuroinflammation are detected in the substantia nigra of PD patients [73]. Besides, blocking microglia-dependent astrocyte activation into A1 astrocytes is neuroprotective in an animal model for PD [87]. Therefore, one explanation for the reduced prevalence of PD among tobacco smokers [7] is reduced neuroinflammation due to activation of the cholinergic anti-inflammatory pathway. Specifically, tobacco-derived nicotine was suggested to protect substantia nigra dopaminergic neurons via activation of microglia- or astrocyte-expressed  $\alpha$ 7 nAChRs leading to reduced neuroinflammation [88]. In support for the above explanation, nicotine protects substantia nigra dopaminergic neurons in animal models for PD in which dopaminergic neurons are selectively targeted using the neurotoxins 6-OHDA or MPTP. However, reduction of PD symptoms, can also be explained by effects of nicotine on nigro-striatal dopamine release which is mediated primarily by  $\alpha \beta \beta^2$ -containing receptors expressed in dopaminergic neurons; expression of this receptor is significantly reduced in animal models for PD [89].

 $\alpha$ 7 nAChR activation ameliorates both the degeneration of dopaminergic neurons and the associated neuroinflammation. For example, in mice nicotine reduced the motor deficits induced by MPTP and these effects were associated with reduced dopaminergic neuron degeneration. In addition, nicotine reduced astrocyte and microglial activation in the substantia nigra. Effects of nicotine on dopaminergic neuron degeneration and on inflammatory markers were abolished by an  $\alpha$ 7 nAChR antagonist [90]. Also,  $\alpha$ 7 nAChR specific agonists reduced degeneration of dopaminergic neurons and neuroinflammation in the striatum of rats treated with 6-OHDA [91] and in the subtantia nigra of mice treated with MPTP [92]. In addition, nicotine pretreatment decreased microglial activation and significantly reduced LPS-induced TNF- $\alpha$  release. Also, co-cultures of microglia and mesencephalic neurons, pretreated with nicotine had significantly decreased loss of tyrosine hydroxylase-immunopositive (TH-ip) cells (tyrosine hydroxylase is a marker for dopaminergic neurons). This effect was blocked by  $\alpha$ -bungarotoxin, an  $\alpha$ 7 nAChR selective blocker. Besides, chronic nicotine pretreatment in rats had the same protective effect in the substantia nigra, suggesting that nicotine protects dopaminergic neurons via an anti-inflammatory mechanism [93].

# 10. Alzheimer's Disease Neuroinflammation and the $\alpha$ 7 nAChR

Alzheimer's disease is a neurodegenerative disease and the most common cause of dementia in the elderly. Neurodegeneration in AD patients is associated with neuroinflammation. Specifically, reactive microglia and astrocytes migrate to and surround amyloid beta (A $\beta$ ) deposits, a hallmark of AD pathology. The roles of astrocytes, microglia and of neuroinflammation in AD progression is complex. While, microglia are responsible for clearance of toxic A $\beta$  deposits and thus have neuroprotective roles, reactive microglia and astrocytes are likely to exacerbate pathology. Accordingly, various anti-inflammatory drugs have been tested for their therapeutic effects on AD, as reviewed in [94].

An early pathology associated with AD is reduced cholinergic signaling. This reduction led to the "cholinergic hypothesis of AD" and to the development of AChEI drugs as a therapy for AD. One explanation for the role of cholinergic reduction in AD pathology is reduced activity of the cholinergic anti-inflammatory pathway in these patients. This hypothesis suggests that treatment with AChEIs improves AD symptoms via reduced neuroinflammation. In support, AD patients treated with AChEIs show altered immune profile which is consistent with inflammation reducing effects of these drugs [95–98].

One AChEI used to treat AD, galantamine, is a relatively weak AChEI also having allosteric potentiating activity on nAChRs. This potentiation of nAChR activity may contribute to galantamine's therapeutic effects. Indeed, galantamine increased A $\beta$  phagocytosis by microglia, an effect depending on  $\alpha$ 7 nAChR and on the allosteric binding site for galantamine present in this receptor. Interestingly, in vitro depletion of choline, the degradation product of ACh and an agonist of  $\alpha$ 7 nAChR, abolished the A $\beta$  phagocytosis inducing effects of galantamine. This result suggests that effects of galantamine on AD can be, at least partially, explained by potentiation of choline dependent enhancement of A $\beta$  clearance by microglia [99].

Recently the therapeutic effects of activating the cholinergic anti-inflammatory pathway on AD pathology were examined in-vivo, in an APP/PS1 mouse model for AD [100]. This research showed that the positive allosteric modulator, JWX-A0108, improved spatial memory, reduced neurodegeneration and reduced neuroinflammation in this AD model. These effects were eliminated by a  $\alpha$ 7 nAChR specific antagonists, MLA. Together these results support activation of the cholinergic anti-inflammatory pathway as a therapy for

AD. In support of the involvement of  $\alpha$ 7 nAChR in AD, treatment with an  $\alpha$ 7 nAChR specific antibody that reduces its expression induced inflammation, A $\beta$ 42 accumulation and impaired memory in the brains of mice [101].

A $\beta$  was shown to interact with and activate  $\alpha$ 7 nAChR leading to investigations into the role of this receptor in AD, reviewed in [102]. Studies examining effects of  $\alpha$ 7 nAChR knockout on pathology and behavior of AD models led to conflicting results [103–105]. One recent paper suggested that oligomeric A $\beta$ <sub>42</sub> activated  $\alpha$ 7 $\beta$ 2 nAChRs to enhance excitability of basal forebrain cholinergic neurons, a possible explanation for the network hyperexcitability and cognitive decline seen in AD patients [106]. Another study showed increased age dependent AD-like pathology and increased A $\beta$  levels in  $\alpha$ 7 nAChR knockout mice; this suggests that  $\alpha$ 7 nAChR malfunction precedes A $\beta$  pathology [105]. This study also showed increased astrocyte activation in  $\alpha$ 7 nAChR knockout mice [105]. The role of the cholinergic anti-inflammatory pathway in effects of  $\alpha$ 7 nAChR knockout on AD-like pathology is, yet, unknown.

Contribution of the cholinergic anti-inflammatory pathway to AD pathology in humans is, yet, unclear. Evidence supporting such involvement include: The proportion of  $\alpha$ 7 nAChR-expressing astrocytes, but not of  $\alpha$ 4 nAChR-expressing astrocytes, was increased in the hippocampus and entorhinal cortex of AD patients [107]. Furthermore,  $\alpha$ 7 nAChR expressing microglia were shown to accumulate around A $\beta$  plaques in AD brains [99]. These findings suggest that in AD brains the cholinergic anti-inflammatory pathway functions to reduce A $\beta$  deposits. We note, however, that tobacco smoking is a risk factor for AD [108,109]. But, there is no evidence linking tobacco smoking associated AD risk to alterations in the cholinergic anti-inflammatory pathway.

## 11. Conclusions

This review highlights the importance of  $\alpha$ 7 nAChR and its chaperon RIC-3 in the regulation of immune responses as part of the cholinergic anti-inflammatory pathway.

We describe the immune functions of this pathway on multiple types of immune cells and on glial cells. We also describe evidence for involvement of the non-neuronal cholinergic receptors in regulation of inflammatory processes in the experimental model of MS in which  $\alpha$ 7 nAChR agonists and antagonists have been shown to modify disease progression. Thus,  $\alpha$ 7 nAChR is a promising target for drugs aimed at downregulating inflammatory processes leading to or exacerbating various diseases including neurodegenerative diseases. Since AChEIs, (which induce cholinergic upregulation and activation of  $\alpha$ 7 nAChRs), are already in clinical use for several neurological indications, such as myasthenia gravis and AD, these agents can also be used for immonomodulation of other neurodegenerative diseases involving neuroinflammation. Furthermore, the ago-PAM GAT107 is promising candidates for treatment in these conditions, as we describe in this review. The widespread effects of the cholinergic anti-inflammatory pathway on inflammation in the periphery and in the CNS suggest the therapeutic potential of harnessing this pathway in diseases involving neuroinflammation. Better understanding of the mechanism of action of  $\alpha 7$ nAChR in immune cells and of proteins, such as RIC-3, affecting  $\alpha$ 7 nAChR's activity may facilitate development of treatments for neurodegenerative and other diseases involving inflammatory processes.

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## References

- 1. Halevi, S.; McKay, J.; Palfreyman, M.; Yassin, L.; Eshel, M.; Jorgensen, E.; Treinin, M. The C. elegans ric-3 gene is required for maturation of nicotinic acetylcholine receptors. *EMBO J.* **2002**, *21*, 1012–1020. [CrossRef] [PubMed]
- Gu, S.; Matta, J.A.; Lord, B.; Harrington, A.W.; Sutton, S.W.; Davini, W.B.; Bredt, D.S. Brain α7 Nicotinic Acetylcholine Receptor Assembly Requires NACHO. *Neuron* 2016, 89, 948–955. [CrossRef] [PubMed]
- Dani, J.A. Neuronal Nicotinic Acetylcholine Receptor Structure and Function and Response to Nicotine. *Int. Rev. Neurobiol.* 2015, 124, 3–19. [CrossRef] [PubMed]
- Richter, K.; Sagawe, S.; Hecker, A.; Küllmar, M.; Askevold, I.; Damm, J.; Heldmann, S.; Pöhlmann, M.; Ruhrmann, S.; Sander, M.; et al. C-Reactive Protein Stimulates Nicotinic Acetylcholine Receptors to Control ATP-Mediated Monocytic Inflammasome Activation. *Front. Immunol.* 2018, *9*, 1604. [CrossRef] [PubMed]
- 5. Dani, J.A.; Bertrand, D. Nicotinic acetylcholine receptors and nicotinic cholinergic mechanisms of the central nervous system. *Annu. Rev. Pharmacol. Toxicol.* 2007, 47, 699–729. [CrossRef]
- 6. Kurzen, H.; Wessler, I.; Kirkpatrick, C.J.; Kawashima, K.; Grando, S.A. The Non-neuronal Cholinergic System of Human Skin. *Horm. Metab. Res.* 2007, *39*, 2191–2194. [CrossRef]
- 7. Li, X.; Li, W.; Liu, G.; Shen, X.; Tang, Y. Association between cigarette smoking and Parkinson's disease: A meta-analysis. *Arch. Gerontol. Geriatr.* **2015**, *61*, 510–516. [CrossRef]
- Šagud, M.; Vuksan-Ćusa, B.; Jakšić, N.; Mihaljević-Peleš, A.; Rojnić Kuzman, M.; Pivac, N. Smoking in Schizophrenia: An Updated Review. *Psychiatr. Danub.* 2018, 30, 216–223.
- 9. Freedman, R.; Hall, M.; Adler, L.E.; Leonard, S. Evidence in postmortem brain tissue for decreased numbers of hippocampal nicotinic receptors in schizophrenia. *Biol. Psychiatry.* **1995**, *38*, 22–33. [CrossRef]
- 10. Lange, K.W.; Wells, F.R.; Jenner, P.; Marsden, C.D. Altered muscarinic and nicotinic receptor densities in cortical and subcortical brain regions in Parkinson's disease. *J. Neurochem.* **1993**, *60*, 197–203. [CrossRef]
- Gotti, C.; Moretti, M.; Bohr, I.; Ziabreva, I.; Vailati, S.; Longhi, R.; Riganti, L.; Gaimarri, A.; McKeith, I.G.; Perry, R.H.; et al. Selective nicotinic acetylcholine receptor subunit deficits identified in Alzheimer's disease, Parkinson's disease and dementia with Lewy bodies by immunoprecipitation. *Neurobiol. Dis.* 2006, 23, 481–489. [CrossRef] [PubMed]
- 12. Perry, E.K.; Morris, C.M.; Court, J.A.; Cheng, A.; Fairbairn, A.F.; McKeith, I.G.; Irving, D.; Brown, A.; Perry, R.H. Alteration in nicotine binding sites in Parkinson's disease, Lewy body dementia and Alzheimer's disease: Possible index of early neuropathology. *Neuroscience* **1995**, *64*, 385–395. [CrossRef]
- 13. Hao, J.; Simard, A.R.; Turner, G.H.; Wu, J.; Whiteaker, P.; Lukas, R.J.; Shi, F.D. Attenuation of CNS inflammatory responses by nicotine involves a7 and non-a7 nicotinic receptors. *Exp. Neurol.* **2011**, 227, 110–119. [CrossRef] [PubMed]
- Borovikova, L.V.; Ivanova, S.; Zhang, M.; Yang, H.; Botchkina, G.I.; Watkins, L.R.; Wang, H.; Abumrad, N.; Eaton, J.W.; Tracey, K.J. Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature* 2000, 405, 458–462. [CrossRef] [PubMed]
- 15. Wang, H.; Yu, M.; Ochani, M.; Amella, C.A.; Tanovic, M.; Susarla, S.; Li, J.H.; Yang, H.; Ulloa, L.; Al-Abe, Y.; et al. Nicotinic acetylcholine receptor alpha7 subunit is an essential regulator of inflammation. *Nature* **2003**, *421*, 384–388. [CrossRef]
- Zakrzewicz, A.; Richter, K.; Agné, A.; Wilker, S.; Siebers, K.; Fink, B.; Krasteva-Christ, G.; Althaus, M.; Padberg, W.; Hone, A.J.; et al. Canonical and Novel Non-Canonical Cholinergic Agonists Inhibit ATP-Induced Release of Monocytic Interleukin-1β via Different Combinations of Nicotinic Acetylcholine Receptor Subunits α7, α9 and α10. *Front Cell Neurosci.* 2017, 11, 189. [CrossRef]
- 17. Zhang, X.; Lei, B.; Yuan, Y.; Zhang, L.; Hu, L.; Jin, S.; Kang, B.; Liao, X.; Sun, W.; Xu, F.; et al. Brain control of humoral immune responses amenable to behavioural modulation. *Nature* **2020**, *581*, 204–208. [CrossRef]
- 18. Alkondon, M.; Albuquerque, E.X. Diversity of nicotinic acetylcholine receptors in rat hippocampal neurons. I. Pharmacological and functional evidence for distinct structural subtypes. *J. Pharmacol. Exp. Ther.* **1993**, *265*, 1455–1472.
- 19. Sequela, P.; Wadiche, J.; Dineley-Miller, K.; Dani, J.A.; Patrick, J.W. Molecular cloning, functional properties, and distribution of rat brain a7: A nicotinic cation channel highly permeable to calcium. *J. Neurosci.* **1993**, *13*, 596–604. [CrossRef]
- 20. Treinin, M.; Papke, R.L.; Nizri, E.; Ben-David, Y.; Mizrachi, T.; Brenner, T. Role of the α7 Nicotinic Acetylcholine Receptor and RIC-3 in the Cholinergic Anti-inflammatory Pathway. *Cent Nerv. Syst. Agents. Med. Chem.* **2017**, *17*, 90–99. [CrossRef]
- Nizri, E.; Brenner, T. Modulation of inflammatory pathways by the immune cholinergic system. *Amino Acids* 2013, 45, 73–85. [CrossRef]
- 22. Báez-Pagán, C.A.; Delgado-Vélez, M.; Lasalde-Dominicci, J.A. Activation of the Macrophage α7 Nicotinic Acetylcholine Receptor and Control of Inflammation. *J. Neuroimmune Pharm.* **2015**, *10*, 468–476. [CrossRef] [PubMed]
- 23. Barnes, P.J. Nuclear factor-kappa B. Int. J. Biochem. Cell Biol. 1997, 29, 867-870. [CrossRef]
- Rosas-Ballina, M.; Goldstein, R.S.; Gallowitsch-Puerta, M.; Yang, L.; Valdés-Ferrer, S.I.; Patel, N.B.; Chavan, S.; Al-Abed, Y.; Yang, H.; Tracey, K.J. The selective alpha7 agonist GTS-21 attenuates cytokine production in human whole blood and human monocytes activated by ligands for TLR2, TLR3, TLR4, TLR9, and RAGE. *Mol. Med.* 2009, *15*, 195–202. [CrossRef]

- Mizrachi, T.; Marsha, O.; Brusin, K.; Ben-David, Y.; Thakur, G.A.; Vaknin-Dembinsky, A.; Treinin, M.; Brenner, T. Suppression of neuroinflammation by an allosteric agonist and positive allosteric modulator of the α7 nicotinic acetylcholine receptor GAT107. *J. Neuroinflamm.* 2021, *18*, 99. [CrossRef]
- Mashimo, M.; Takeshima, S.; Okuyama, H.; Matsurida, A.; Murase, M.; Ono, S.; Kawashima, K.; Fujii, T. α7 nAChRs expressed on antigen presenting cells are insensitive to the conventional antagonists α-bungarotoxin and methyllycaconitine. *Int. Immunopharm.* 2020, *81*, 106276. [CrossRef]
- 27. Fujii, T.; Mashimo, M.; Moriwaki, Y.; Misawa, H.; Ono, S.; Horiguchi, K.; Kawashima, K. Physiological functions of the cholinergic system in immune cells. *J. Pharm. Sci* 2017, 134, 1–21. [CrossRef]
- 28. Nizri, E.; Irony-Tur-Sinai, M.; Faranesh, N.; Lavon, I.; Lavi, E.; Weinstock, M.; Brenner, T. Suppression of neuroinflammation and immunomodulation by the acetylcholinesterase inhibitor rivastigmine. *J. Neuroimmunol.* **2008**, 203, 12–22. [CrossRef]
- Nizri, E.; Irony-Tur-Sinai, M.; Lory, O.; Orr-Urtreger, A.; Lavi, E.; Brenner, T. Activation of the cholinergic anti-inflammatory system by nicotine attenuates neuroinflammation via suppression of Th1 and Th17 responses. *J. Immunol.* 2009, 183, 6681–6688. [CrossRef] [PubMed]
- 30. Seyler, L.E.; Fertig, J.; Pomerleau, O.; Hunt, D.; Parker, K. The effects of smoking on ACTH and cortisol secretion. *Life Sci.* **1984**, 34, 57–65. [CrossRef]
- Nouri-Shirazi, M.; Guinet, E. Evidence for the immunosuppressive role of nicotine on human dendritic cell functions. *Immunology* 2003, 109, 365–373. [CrossRef]
- 32. Nizri, E.; Adani, R.; Meshulam, H.; Amitai, G.; Brenner, T. Bifunctional compounds eliciting both anti-inflammatory and cholinergic activity as potential drugs for neuroinflammatory impairments. *Neurosci. Lett.* **2005**, *376*, 46–50. [CrossRef]
- Nizri, E.; Hamra-Amitay, Y.; Sicsic, C.; Lavon, I.; Brenner, T. Anti-inflammatory properties of cholinergic up-regulation: A new role for acetylcholinesterase inhibitors. *Neuropharmacology* 2006, 50, 540–547. [CrossRef]
- Galitovskiy, V.; Qian, J.; Chernyavsky, A.I.; Marchenko, S.; Gindi, V.; Edwards, R.A.; Grando, S.A. Cytokine-induced alterations of α7 nicotinic receptor in colonic CD4 T cells mediate dichotomous response to nicotine in murine models of Th1/Th17- versus Th2-mediated colitis. *J. Immunol.* 2011, *187*, 2677–2687. [CrossRef] [PubMed]
- 35. Mashimo, M.; Komori, M.; Matsui, Y.Y.; Murase, M.X.; Fujii, T.; Takeshima, S.; Okuyama, H.; Ono, S.; Moriwaki, Y.; Misawa, H.; et al. Distinct Roles of α7 nAChRs in Antigen-Presenting Cells and CD4. *Front. Immunol.* 2019, 10, 1102. [CrossRef] [PubMed]
- Mashimo, M.; Iwasaki, Y.; Inoue, S.; Saito, S.; Kawashima, K.; Fujii, T. Acetylcholine released from T cells regulates intracellular Ca. Life Sci. 2017, 172, 13–18. [CrossRef] [PubMed]
- 37. Mashimo, M.; Fujii, T.; Ono, S.; Moriwaki, Y.; Misawa, H.; Kawashima, K. Minireview: Divergent roles of α7 nicotinic acetylcholine receptors expressed on antigen-presenting cells and CD4. *Int. Immunopharmacol.* **2020**, *82*, 106306. [CrossRef]
- Wang, D.W.; Zhou, R.B.; Yao, Y.M.; Zhu, X.M.; Yin, Y.M.; Zhao, G.J.; Dong, N.; Sheng, Z.Y. Stimulation of α7 nicotinic acetylcholine receptor by nicotine increases suppressive capacity of naturally occurring CD4+CD25+ regulatory T cells in mice in vitro. *J. Pharm. Exp.* 2010, 335, 553–561. [CrossRef] [PubMed]
- Sato, K.Z.; Fujii, T.; Watanabe, Y.; Yamada, S.; Ando, T.; Kazuko, F.; Kawashima, K. Diversity of mRNA expression for muscarinic acetylcholine receptor subtypes and neuronal nicotinic acetylcholine receptor subunits in human mononuclear leukocytes and leukemic cell lines. *Neurosci. Lett.* 1999, 266, 17–20. [CrossRef]
- 40. Tarasenko, O.; Voytenko, S.; Koval, L.; Lykhmus, O.; Kalashnyk, O.; Skok, M. Unusual properties of α7 nicotinic acetylcholine receptor ion channels in B lymphocyte-derived SP-2/0 cells. *Int. Immunopharmacol.* **2020**, *82*, 106373. [CrossRef] [PubMed]
- Wu, J.; Liu, Q.; Tang, P.; Mikkelsen, J.D.; Shen, J.; Whiteaker, P.; Yakel, J.L. Heteromeric α7β2 Nicotinic Acetylcholine Receptors in the Brain. *Trends Pharm. Sci.* 2016, 37, 562–574. [CrossRef] [PubMed]
- 42. Skok, M.V.; Grailhe, R.; Agenes, F.; Changeux, J.P. The role of nicotinic receptors in B-lymphocyte development and activation. *Life Sci.* 2007, *80*, 2334–2336. [CrossRef]
- Fujii, Y.X.; Fujigaya, H.; Moriwaki, Y.; Misawa, H.; Kasahara, T.; Grando, S.A.; Kawashima, K. Enhanced serum antigen-specific IgG1 and proinflammatory cytokine production in nicotinic acetylcholine receptor alpha7 subunit gene knockout mice. *J. Neuroimmunol.* 2007, 189, 69–74. [CrossRef] [PubMed]
- Koval, L.; Lykhmus, O.; Zhmak, M.; Khruschov, A.; Tsetlin, V.; Magrini, E.; Viola, A.; Chernyavsky, A.; Qian, J.; Grando, S.; et al. Differential involvement of α4β2, α7 and α9α10 nicotinic acetylcholine receptors in B lymphocyte activation in vitro. *Int. J. Biochem. Cell Biol.* 2011, 43, 516–524. [CrossRef] [PubMed]
- 45. Zanetti, S.R.; Ziblat, A.; Torres, N.I.; Zwirner, N.W.; Bouzat, C. Expression and Functional Role of α7 Nicotinic Receptor in Human Cytokine-stimulated Natural Killer (NK) Cells. *J. Biol. Chem.* **2016**, *291*, 16541–16552. [CrossRef]
- Shytle, R.D.; Mori, T.; Townsend, K.; Vendrame, M.; Sun, N.; Zeng, J.; Ehrhart, J.; Silver, A.A.; Sanberg, P.R.; Tan, J. Cholinergic modulation of microglial activation by alpha 7 nicotinic receptors. *J. Neurochem.* 2004, 89, 337–343. [CrossRef]
- Li, L.; Liu, Z.; Jiang, Y.Y.; Shen, W.X.; Peng, Y.P.; Qiu, Y.H. Acetylcholine suppresses microglial inflammatory response via α7nAChR to protect hippocampal neurons. *J. Integr. Neurosci.* 2019, *18*, 51–56. [CrossRef]
- 48. Thomsen, M.S.; Mikkelsen, J.D. The a7 nicotinic acetylcholine receptor ligands methyllycaconitine, NS6740 and GTS-21 reduce lipopolysaccharide-induced TNF-a release from microglia. *J. Neuroimunol.* **2012**, *251*, 65–72. [CrossRef]

- Cortes, M.; Cao, M.; Liu, H.L.; Moore, C.S.; Durosier, L.D.; Burns, P.; Fecteau, G.; Desrochers, A.; Barreiro, L.B.; Antel, J.P.; et al. α7 nicotinic acetylcholine receptor signaling modulates the inflammatory phenotype of fetal brain microglia: First evidence of interference by iron homeostasis. *Sci. Rep.* 2017, 7, 10645. [CrossRef]
- 50. Ta, H.M.; Le, T.M.; Ishii, H.; Takarada-Iemata, M.; Hattori, T.; Hashida, K.; Yamamoto, Y.; Mori, K.; Takahashi, R.; Kitao, Y.; et al. Atf6α deficiency suppresses microglial activation and ameliorates pathology of experimental autoimmune encephalomyelitis. *J. Neurochem.* 2016, 139, 1124–1137. [CrossRef]
- 51. Ke, P.; Shao, B.Z.; Xu, Z.Q.; Chen, X.W.; Wei, W.; Liu, C. Activating α7 nicotinic acetylcholine receptor inhibits NLRP3 inflammasome through regulation of β-arrestin-1. *CNS Neurosci.* **2017**, *23*, 875–884. [CrossRef] [PubMed]
- 52. Patel, H.; McIntire, J.; Ryan, S.; Dunah, A.; Loring, R. Anti-inflammatory effects of astroglial α7 nicotinic acetylcholine receptors are mediated by inhibition of the NF-κB pathway and activation of the Nrf2 pathway. *J. Neuroinflamm.* **2017**, *14*, 192. [CrossRef]
- 53. Wang, X.; Lippi, G.; Carlson, D.M.; Berg, D.K. Activation of α7-containing nicotinic receptors on astrocytes triggers AMPA receptor recruitment to glutamatergic synapses. *J. Neurochem.* **2013**, *127*, 632–643. [CrossRef] [PubMed]
- Di Cesare Mannelli, L.; Tenci, B.; Zanardelli, M.; Failli, P.; Ghelardini, C. α7 Nicotinic Receptor Promotes the Neuroprotective Functions of Astrocytes against Oxaliplatin Neurotoxicity. *Neural. Plast* 2015, 2015, 396908. [CrossRef]
- 55. Cao, M.; MacDonald, J.W.; Liu, H.L.; Weaver, M.; Cortes, M.; Durosier, L.D.; Burns, P.; Fecteau, G.; Desrochers, A.; Schulkin, J.; et al. α7 Nicotinic Acetylcholine Receptor Signaling Modulates Ovine Fetal Brain Astrocytes Transcriptome in Response to Endotoxin. *Front. Immunol.* 2019, 10, 1063. [CrossRef] [PubMed]
- Cordero-Erausquin, M.; Marubio, L.M.; Klink, R.; Changeux, J.P. Nicotinic receptor function: New perspectives from knockout mice. *Trends Pharmacol. Sci.* 2000, 21, 211–217. [CrossRef]
- 57. Deutsch, S.I.; Burket, J.A.; Benson, A.D.; Urbano, M.R. The 15q13.3 deletion syndrome: Deficient α(7)-containing nicotinic acetylcholine receptor-mediated neurotransmission in the pathogenesis of neurodevelopmental disorders. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2016**, *64*, 109–117. [CrossRef] [PubMed]
- Sinkus, M.L.; Graw, S.; Freedman, R.; Ross, R.G.; Lester, H.A.; Leonard, S. The human CHRNA7 and CHRFAM7A genes: A review of the genetics, regulation, and function. *Neuropharmacology* 2015, *96*, 274–288. [CrossRef]
- 59. de Lucas-Cerrillo, A.M.; Maldifassi, M.C.; Arnalich, F.; Renart, J.; Atienza, G.; Serantes, R.; Cruces, J.; Sánchez-Pacheco, A.; Andrés-Mateos, E.; Montiel, C. Function of partially duplicated human α77 nicotinic receptor subunit CHRFAM7A gene: Potential implications for the cholinergic anti-inflammatory response. *J. Biol. Chem.* 2011, 286, 594–606. [CrossRef]
- Tata, A.M.; Velluto, L.; D'Angelo, C.; Reale, M. Cholinergic system dysfunction and neurodegenerative diseases: Cause or effect? CNS Neurol. Disord. Drug Targets 2014, 13, 1294–1303. [CrossRef]
- 61. Halevi, S.; Yassin, L.; Eshel, M.; Sala, F.; Sala, S.; Criado, M.; Treinin, M. Conservation within the RIC-3 gene family: Effectors of nAChR expression. J. Biol. Chem. 2003, 278, 34411–34417. [CrossRef]
- Williams, M.E.; Burton, B.; Urrutia, A.; Shcherbatko, A.; Chavez-Noriega, L.E.; Cohen, C.J.; Aiyar, J. Ric-3 promotes functional expression of the nicotinic acetylcholine receptor alpha7 subunit in mammalian cells. *J. Biol. Chem.* 2005, 280, 1257–1263. [CrossRef] [PubMed]
- 63. Lansdell, S.J.; Gee, V.J.; Harkness, P.C.; Doward, A.I.; Baker, E.R.; Gibb, A.J.; Millar, N.S. RIC-3 enhances functional expression of multiple nicotinic acetylcholine receptor subtypes in mammalian cells. *Mol. Pharmacol.* **2005**, *68*, 1431–1438. [CrossRef]
- 64. Alexander, J.K.; Sagher, D.; Krivoshein, A.V.; Criado, M.; Jefford, G.; Green, W.N. Ric-3 promotes alpha7 nicotinic receptor assembly and trafficking through the ER subcompartment of dendrites. *J. Neurosci.* 2010, *30*, 10112–10126. [CrossRef] [PubMed]
- 65. Ben-David, Y.; Mizrachi, T.; Kagan, S.; Krisher, T.; Cohen, E.; Brenner, T.; Treinin, M. RIC-3 expression and splicing regulate nAChR functional expression. *Mol. Brain* **2016**, *9*, 47. [CrossRef]
- 66. Deshpande, A.; Vinayakamoorthy, R.M.; Garg, B.K.; Thummapudi, J.P.; Oza, G.; Adhikari, K.; Agarwal, A.; Dalvi, P.; Iyer, S.; Thulasi Raman, S.; et al. Why Does Knocking Out NACHO, But Not RIC3, Completely Block Expression of α7 Nicotinic Receptors in Mouse Brain? *Biomolecules* 2020, 10, 470. [CrossRef] [PubMed]
- Ben-David, Y.; Kagan, S.; Cohen Ben-Ami, H.; Rostami, J.; Mizrahi, T.; Kulkarni, A.R.; Thakur, G.A.; Vaknin-Dembinsky, A.; Healy, L.M.; Brenner, T.; et al. RIC3, the cholinergic anti-inflammatory pathway, and neuroinflammation. *Int. Immunopharmacol.* 2020, *83*, 106381. [CrossRef]
- Severance, E.G.; Yolken, R.H. Lack of RIC-3 congruence with beta2 subunit-containing nicotinic acetylcholine receptors in bipolar disorder. *Neuroscience* 2007, 148, 454–460. [CrossRef]
- Gourraud, P.A.; Sdika, M.; Khankhanian, P.; Henry, R.G.; Beheshtian, A.; Matthews, P.M.; Hauser, S.L.; Oksenberg, J.R.; Pelletier, D.; Baranzini, S.E. A genome-wide association study of brain lesion distribution in multiple sclerosis. *Brain* 2013, 136, 1012–1024. [CrossRef]
- Wang, J.H.; Pappas, D.; De Jager, P.L.; Pelletier, D.; de Bakker, P.I.; Kappos, L.; Polman, C.H.; Chibnik, L.B.; Hafler, D.A.; Matthews, P.M.; et al. Modeling the cumulative genetic risk for multiple sclerosis from genome-wide association data. *Genome Med.* 2011, 3, 3. [CrossRef]
- Yokoyama, J.S.; Evans, D.S.; Coppola, G.; Kramer, J.H.; Tranah, G.J.; Yaffe, K. Genetic modifiers of cognitive maintenance among older adults. *Hum. Brain Mapp.* 2014, 35, 4556–4565. [CrossRef] [PubMed]
- Sudhaman, S.; Muthane, U.B.; Behari, M.; Govindappa, S.T.; Juyal, R.C.; Thelma, B.K. Evidence of mutations in RIC3 acetylcholine receptor chaperone as a novel cause of autosomal-dominant Parkinson's disease with non-motor phenotypes. *J. Med. Genet.* 2016, 53, 559–566. [CrossRef]

- 73. Liddelow, S.A.; Guttenplan, K.A.; Clarke, L.E.; Bennett, F.C.; Bohlen, C.J.; Schirmer, L.; Bennett, M.L.; Münch, A.E.; Chung, W.S.; Peterson, T.C.; et al. Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 2017, 541, 481–487. [CrossRef] [PubMed]
- 74. Cummings, J.L.; Koumaras, B.; Chen, M.; Mirski, D.; Team, R.N.H.S. Effects of rivastigmine treatment on the neuropsychiatric and behavioral disturbances of nursing home residents with moderate to severe probable Alzheimer's disease: A 26-week, multicenter, open-label study. *Am. J. Geriatr. Pharm.* 2005, *3*, 137–148. [CrossRef]
- 75. Nizri, E.; Irony-Tur-Sinai, M.; Grigoriadis, N.; Abramsky, O.; Amitai, G.; Brenner, T. Novel approaches to treatment of autoimmune neuroinflammation and lessons for drug development. *Pharmacology* **2007**, *79*, 42–49. [CrossRef]
- 76. Shi, F.D.; Piao, W.H.; Kuo, Y.P.; Campagnolo, D.I.; Vollmer, T.L.; Lukas, R.J. Nicotinic attenuation of central nervous system inflammation and autoimmunity. *J. Immunol.* **2009**, *182*, 1730–1739. [CrossRef] [PubMed]
- 77. Simard, A.R.; Gan, Y.; St-Pierre, S.; Kousari, A.; Patel, V.; Whiteaker, P.; Morley, B.J.; Lukas, R.J.; Shi, F.D. Differential modulation of EAE by a9\*- and a2\*-nicotinic acetylcholine receptors. *Immunol. Cell Biol.* **2013**, *91*, 68–74. [CrossRef]
- Reale, M.; Di Bari, M.; Di Nicola, M.; D'Angelo, C.; De Angelis, F.; Velluto, L.; Tata, A.M. Nicotinic receptor activation negatively modulates pro-inflammatory cytokine production in multiple sclerosis patients. *Int. Immunopharmacol.* 2015, 29, 152–157. [CrossRef] [PubMed]
- 79. Gatta, V.; Mengod, G.; Reale, M.; Tata, A.M. Possible Correlation between Cholinergic System Alterations and Neuro/Inflammation in Multiple Sclerosis. *Biomedicines* 2020, *8*, 153. [CrossRef]
- Di Bari, M.; Reale, M.; Di Nicola, M.; Orlando, V.; Galizia, S.; Porfilio, I.; Costantini, E.; D'Angelo, C.; Ruggieri, S.; Biagioni, S.; et al. Dysregulated Homeostasis of Acetylcholine Levels in Immune Cells of RR-Multiple Sclerosis Patients. *Int. J. Mol. Sci.* 2016, 17, 2009. [CrossRef]
- Jiang, W.; Li, D.; Han, R.; Zhang, C.; Jin, W.N.; Wood, K.; Liu, Q.; Shi, F.D.; Hao, J. Acetylcholine-producing NK cells attenuate CNS inflammation via modulation of infiltrating monocytes/macrophages. *Proc. Natl. Acad. Sci. USA* 2017, 114, E6202–E6211. [CrossRef] [PubMed]
- 82. Polachini, C.R.N.; Spanevello, R.M.; Schetinger, M.R.C.; Morsch, V.M. Cholinergic and purinergic systems: A key to multiple sclerosis? *J. Neurol. Sci.* 2018, 392, 8–21. [CrossRef] [PubMed]
- 83. Rosso, M.; Chitnis, T. Association between Cigarette Smoking and Multiple Sclerosis: A Review. JAMA Neurol. 2019. [CrossRef]
- 84. Hedström, A.K.; Hillert, J.; Olsson, T.; Alfredsson, L. Nicotine might have a protective effect in the etiology of multiple sclerosis. *Mult. Scler.* **2013**, *19*, 1009–1013. [CrossRef]
- 85. Briggs, F.B. Nicotinic acetylcholine receptors α7 and α9 modifies tobacco smoke risk for multiple sclerosis. *Mult. Scler.* **2021**, 27, 1166–1174. [CrossRef] [PubMed]
- 86. Gao, Z.; Nissen, J.C.; Ji, K.; Tsirka, S.E. The experimental autoimmune encephalomyelitis disease course is modulated by nicotine and other cigarette smoke components. *PLoS ONE* **2014**, *9*, e107979. [CrossRef] [PubMed]
- Yun, S.P.; Kam, T.I.; Panicker, N.; Kim, S.; Oh, Y.; Park, J.S.; Kwon, S.H.; Park, Y.J.; Karuppagounder, S.S.; Park, H.; et al. Block of A1 astrocyte conversion by microglia is neuroprotective in models of Parkinson's disease. *Nat. Med.* 2018, 24, 931–938. [CrossRef]
- Jurado-Coronel, J.C.; Avila-Rodriguez, M.; Capani, F.; Gonzalez, J.; Moran, V.E.; Barreto, G.E. Targeting the Nicotinic Acetylcholine Receptors (nAChRs) in Astrocytes as a Potential Therapeutic Target in Parkinson's Disease. *Curr. Pharm. Des.* 2016, 22, 1305–1311. [CrossRef]
- 89. Bordia, T.; Grady, S.R.; McIntosh, J.M.; Quik, M. Nigrostriatal damage preferentially decreases a subpopulation of alpha6beta2\* nAChRs in mouse, monkey, and Parkinson's disease striatum. *Mol. Pharm.* **2007**, *72*, *52–61*. [CrossRef]
- Liu, Y.; Hu, J.; Wu, J.; Zhu, C.; Hui, Y.; Han, Y.; Huang, Z.; Ellsworth, K.; Fan, W. α7 nicotinic acetylcholine receptor-mediated neuroprotection against dopaminergic neuron loss in an MPTP mouse model via inhibition of astrocyte activation. *J. Neuroinflamm.* 2012, *9*, 98. [CrossRef]
- 91. Sérrière, S.; Doméné, A.; Vercouillie, J.; Mothes, C.; Bodard, S.; Rodrigues, N.; Guilloteau, D.; Routier, S.; Page, G.; Chalon, S. Assessment of the Protection of Dopaminergic Neurons by an α7 Nicotinic Receptor Agonist, PHA 543613 Using [(18)F]LBT-999 in a Parkinson's Disease Rat Model. *Front. Med.* 2015, 2, 61. [CrossRef]
- 92. Stuckenholz, V.; Bacher, M.; Balzer-Geldsetzer, M.; Alvarez-Fischer, D.; Oertel, W.H.; Dodel, R.C.; Noelker, C. The α7 nAChR agonist PNU-282987 reduces inflammation and MPTP-induced nigral dopaminergic cell loss in mice. *J. Parkinsons Dis.* 2013, *3*, 161–172. [CrossRef] [PubMed]
- 93. Park, H.J.; Lee, P.H.; Ahn, Y.W.; Choi, Y.J.; Lee, G.; Lee, D.Y.; Chung, E.S.; Jin, B.K. Neuroprotective effect of nicotine on dopaminergic neurons by anti-inflammatory action. *Eur. J. Neurosci.* 2007, *26*, 79–89. [CrossRef]
- 94. Minter, M.R.; Taylor, J.M.; Crack, P.J. The contribution of neuroinflammation to amyloid toxicity in Alzheimer's disease. *J. Neurochem.* **2016**, 136, 457–474. [CrossRef] [PubMed]
- 95. Gambi, F.; Reale, M.; Iarlori, C.; Salone, A.; Toma, L.; Paladini, C.; De Luca, G.; Feliciani, C.; Salvatore, M.; Salerno, R.M.; et al. Alzheimer patients treated with an AchE inhibitor show higher IL-4 and lower IL-1 beta levels and expression in peripheral blood mononuclear cells. *J. Clin. Psychopharmacol.* **2004**, *24*, 314–321. [CrossRef]
- Lugaresi, A.; Di Iorio, A.; Iarlori, C.; Reale, M.; De Luca, G.; Sparvieri, E.; Michetti, A.; Conti, P.; Gambi, D.; Abate, G.; et al. IL-4 in vitro production is upregulated in Alzheimer's disease patients treated with acetylcholinesterase inhibitors. *Exp. Gerontol.* 2004, *39*, 653–657. [CrossRef] [PubMed]

- 97. Reale, M.; Iarlori, C.; Gambi, F.; Feliciani, C.; Salone, A.; Toma, L.; DeLuca, G.; Salvatore, M.; Conti, P.; Gambi, D. Treatment with an acetylcholinesterase inhibitor in Alzheimer patients modulates the expression and production of the pro-inflammatory and anti-inflammatory cytokines. *J. Neuroimmunol.* **2004**, *148*, 162–171. [CrossRef]
- 98. Reale, M.; Iarlori, C.; Gambi, F.; Feliciani, C.; Isabella, L.; Gambi, D. The acetylcholinesterase inhibitor, Donepezil, regulates a Th2 bias in Alzheimer's disease patients. *Neuropharmacology* **2006**, *50*, 606–613. [CrossRef]
- 99. Takata, K.; Kitamura, Y.; Saeki, M.; Terada, M.; Kagitani, S.; Kitamura, R.; Fujikawa, Y.; Maelicke, A.; Tomimoto, H.; Taniguchi, T.; et al. Galantamine-induced amyloid-{beta} clearance mediated via stimulation of microglial nicotinic acetylcholine receptors. *J. Biol. Chem.* **2010**, *285*, 40180–40191. [CrossRef]
- 100. Li, H.; Gao, J.; Chang, Y.; Li, K.; Wang, L.; Ju, C.; Zhang, F. JWX-A0108, a positive allosteric modulator of α7 nAChR, attenuates cognitive deficits in APP/PS1 mice by suppressing NF-κB-mediated inflammation. *Int. Immunopharmacol.* 2021, 96, 107726. [CrossRef]
- 101. Lykhmus, O.; Voytenko, L.; Koval, L.; Mykhalskiy, S.; Kholin, V.; Peschana, K.; Zouridakis, M.; Tzartos, S.; Komisarenko, S.; Skok, M. α7 Nicotinic acetylcholine receptor-specific antibody induces inflammation and amyloid β42 accumulation in the mouse brain to impair memory. *PLoS ONE* 2015, *10*, e0122706. [CrossRef] [PubMed]
- Buckingham, S.D.; Jones, A.K.; Brown, L.A.; Sattelle, D.B. Nicotinic acetylcholine receptor signalling: Roles in Alzheimer's disease and amyloid neuroprotection. *Pharmacol. Rev.* 2009, 61, 39–61. [CrossRef] [PubMed]
- 103. Hernandez, C.M.; Kayed, R.; Zheng, H.; Sweatt, J.D.; Dineley, K.T. Loss of alpha7 nicotinic receptors enhances beta-amyloid oligomer accumulation, exacerbating early-stage cognitive decline and septohippocampal pathology in a mouse model of Alzheimer's disease. *J. Neurosci.* **2010**, *30*, 2442–2453. [CrossRef]
- 104. Dziewczapolski, G.; Glogowski, C.M.; Masliah, E.; Heinemann, S.F. Deletion of the 7 Nicotinic Acetylcholine Receptor Gene Improves Cognitive Deficits and Synaptic Pathology in a Mouse Model of AlzheimerÕs Disease. J. Neurosci. 2009, 29, 8805–8815. [CrossRef] [PubMed]
- 105. Tropea, M.R.; Li Puma, D.D.; Melone, M.; Gulisano, W.; Arancio, O.; Grassi, C.; Conti, F.; Puzzo, D. Genetic deletion of α7 nicotinic acetylcholine receptors induces an age-dependent Alzheimer's disease-like pathology. *Prog. Neurobiol.* 2021, 206, 102154. [CrossRef]
- 106. George, A.A.; Vieira, J.M.; Xavier-Jackson, C.; Gee, M.T.; Cirrito, J.R.; Bimonte-Nelson, H.A.; Picciotto, M.R.; Lukas, R.J.; Whiteaker, P. Implications of Oligomeric Amyloid-Beta (oAβ42) signaling through a 7 b 2-Nicotinic Acetylcholine Receptors (nAChRs) on Basal Forebrain Cholinergic Neuronal Intrinsic Excitability and Cognitive Decline. *J. Neurosci.* 2021, 41, 555–575. [CrossRef]
- 107. Teaktong, T.; Graham, A.; Court, J.; Perry, R.; Jaros, E.; Johnson, M.; Hall, R.; Perry, E. Alzheimer's disease is associated with a selective increase in alpha7 nicotinic acetylcholine receptor immunoreactivity in astrocytes. *Glia* 2003, *41*, 207–211. [CrossRef]
- 108. Jiang, Y.; Man, Q.; Liu, Z.; Wang, Y.; Suo, C.; Jin, L.; Dong, Q.; Cui, M.; Chen, X. Temporal trends in the mortality rate of Alzheimer's disease and other dementias attributable to smoking, 1990–2017. *Environ. Res.* **2020**, *184*, 109183. [CrossRef]
- Otuyama, L.J.; Oliveira, D.; Locatelli, D.; Machado, D.A.; Noto, A.R.; Galduróz, J.C.F.; Prince, M.J.; Ferri, C.P. Tobacco smoking and risk for dementia: Evidence from the 10/66 population-based longitudinal study. *Aging Ment. Health* 2019, 1–11. [CrossRef]