

REVIEW

Targeting immune checkpoints: how to use natural killer cells for fighting against solid tumors

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Abstract

Natural killer (NK) cells are unique innate immune cells that mediate anti-viral and anti-tumor responses. Thus, they might hold great potential for cancer immunotherapy. NK cell adoptive immunotherapy in humans has shown

List of abbreviations: mAb, monoclonal antibody; CTLA-4, cytotoxic T-lymphocyte associated protein; PD-1, programmed cell death protein 1; ICIs, immune checkpoint inhibitors; LAK, lymphokine-activated killer; TILs, tumor-infiltrating lymphocytes; TCRs, T cell receptors; CARs, chimeric antigen receptors; FDA, Food and Drug Administration; GVHD, graft-versus-host disease; TME, tumor microenvironment; NK, natural killer; BM, bone marrow; LNs, lymph nodes; NCR, natural cytotoxicity receptor; PB, peripheral blood; IFN- γ , interferon gamma; MHC-I, major histocompatibility complex class I; KIRs, killer cell immunoglobulin-like receptors; NKG2A, natural killer group 2 A; LIRs, leukocyte immunoglobulin-like receptors; Ig, immunoglobulin; HLA, human leukocyte antigen; ILTs, immunoglobulin-like transcripts; TIGIT, T cell immunoreceptor with Ig and ITIM domains; LAG-3, lymphocyte activation gene-3; TIM-3, T cell immunoglobulin- and mucin-domain-containing molecule-3; DNAM-1, DNAX accessory molecule-1; FasL, Fas ligand; TRAIL, tumor necrosis factor-related apoptosis-inducing ligand; ADCC, antigen-dependent cell-mediated cytotoxicity; TNF- α , tumor necrosis factor- α ; GM-CSF, granulocyte macrophage colony-stimulating factor; MIP, macrophage inflammatory protein; PRRs, pattern recognition receptors; aKIRs, activating KIRs; iKIRs, inhibitory KIRs; ITIM, immunoreceptor tyrosine-based inhibitory motif; MM, multiple myeloma; EGFR, epidermal growth factor receptor; HMGB1, high mobility group box 1; Gal-9, galactin-9; CEACAM-1, carcinoembryonic antigen-related cell adhesion molecule 1; ITT, immunoreceptor tyrosine tail; FGL1, fibrinogen-like protein 1; PTLN, post-transplant lymphoproliferative disease; TM, transmembrane; scFvs, single-chain variable fragments; DARPins, designed ankyrin repeat proteins; ITAMs, immunoreceptor tyrosine-based activation motifs; TRUCKS, T cells redirected for universal cytokine-mediated killing; ACT, adoptive cell therapy; GvT, graft-versus-tumor; HSCT, hematopoietic stem cell transplantation; AML, acute myeloid leukemia; HPCs, hematopoietic progenitor cells; iPSCs, induced pluripotent stem cells; hESCs, human embryonic stem cells; UCB, umbilical cord blood; CSCs, cancer stem cells; TGF- β , transforming growth factor- β ; PBMCs, peripheral blood mononuclear cells; EBV, Epstein-Barr Virus; CMV, cytomegalovirus; aAPCs, artificial antigen-presenting cells; Eomes, eomesodermin; iC9, inducible caspase 9; G-CSF, granulocyte colony stimulating factor; FLT-3L, FMS-like tyrosine kinase 3 ligand; EBs, embryonic bodies; GBM, glioblastoma; CRISPR, clustered regularly interspaced short palindromic repeats; BMP4, bone morphogenetic protein 4; VEGF, vascular endothelial growth factor; ZFN, zinc finger nuclease; GPC3, glypican-3; PSCA, prostate stem cell antigen; HCC, hepatocellular carcinoma; CCCR, chimeric costimulatory converting receptor; TNBC, triple negative breast cancer; ADAM17, A Disintegrin And Metalloproteinase-17; rAAV6, recombinant adeno-associated virus serotype 6; ABE, adenine base editor; DTCR, dual targeting chimeric receptor; PDAC, pancreatic ductal adenocarcinoma; CIS, cytokine-inducible SH2-containing protein; DNMT1, DNA methyltransferase 1; TAP-1, transporter associated with antigen processing 1; TALENs, transcriptional activator-like effector nucleases; NPs, nanoparticles; CTC, circulating tumor cell; BiKEs, bi-specific natural killer cell engagers; TriKEs, tri-specific natural killer cell engagers; NSCLC, non-small cell lung cancer; SCLC, small cell lung cancer; IAP, integrin-associated protein; TSP-1, thrombospondin-1; SIRP α , signal regulatory protein alpha; HNSCC, head and neck squamous cell carcinoma; LDH, lactate dehydrogenase; Bi-apt, bi-specific aptamer; KI, knock in; HI, hepatic intrasinusoidal; EMA, European Medicines Agency; MMPs, matrix metalloproteinases; CRS, cytokine release syndrome; SCF, stem cell factor; NB, neuroblastoma; GSDME, gasdermin E; IND, investigational new drug; GMP, Good Manufacturing Practice; PD-L1, programmed death-ligand 1.

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modest efficacy. In particular, it has failed to demonstrate therapeutic efficiency in the treatment of solid tumors, possibly due in part to the immunosuppressive tumor microenvironment (TME), which reduces NK cell immunotherapy's efficiencies. It is known that immune checkpoints play a prominent role in creating an immunosuppressive TME, leading to NK cell exhaustion and tumor immune escape. Therefore, NK cells must be reversed from their dysfunctional status and increased in their effector roles in order to improve the efficiency of cancer immunotherapy. Blockade of immune checkpoints can not only rescue NK cells from exhaustion but also augment their robust anti-tumor activity. In this review, we discussed immune checkpoint blockade strategies with a focus on chimeric antigen receptor (CAR)-NK cells to redirect NK cells to cancer cells in the treatment of solid tumors.

KEYWORDS

natural killer cell, immune checkpoint, chimeric antigen receptor-natural killer cell, immunotherapy, tumor

1 | BACKGROUND

Following cardiovascular disease, cancer is the second cause of mortality worldwide [1]. As a result, several treatment options have been developed to restrict disease development, including surgery, chemotherapy, radiation, targeted therapy, and immunotherapy. The relevance of new cancer treatment approaches, such as immunotherapy, has been recognized due to the negative effects of traditional cancer treatment methods. Although cancers use a variety of tactics to delay, alter or stop immune responses, various strategies have been devised to improve the immune response. Generally, cancer immunotherapy is classified into passive and active approaches. In passive immunotherapy, the host's immune response is not stimulated directly. This approach involves the administration of ex vivo-generated cells or molecules (tumor-specific monoclonal antibodies (mAbs), recombinant cytokines, and adoptive cell transfer) to patients. On the other hand, active immunotherapy promotes the patient's immune response and leads to effector immunological responses [2]. Active cancer immunotherapy includes Abs targeting immune checkpoints, oncolytic viruses, and vaccines (such as peptide vaccines, DC vaccines, and allogeneic whole cell vaccines) [2, 3].

A breakthrough in immunotherapy was achieved when James P. Allison and Tasuku Honjo received the Nobel Prize in Physiology or Medicine for their discovery of new targets for cancer therapy. They discovered two critical immune checkpoints, including the cytotoxic T-lymphocyte-associated protein (CTLA-4) and programmed cell death protein 1 (PD-1) as T cells brake [4]. By

inhibiting or blocking these checkpoints, they hoped that T cells would be able to find cancer cells and kill them.

Natural killer (NK) cells are unique innate immune cells that play crucial roles in anti-viral and anti-tumor responses. This suggests that they might hold great promise for cancer immunotherapy. NK cell adoptive immunotherapy in humans has shown modest efficacy [5]. It has failed to demonstrate therapeutic efficacy in the treatment of solid tumors in particular [6, 7]. This is due in part to the immunosuppressive tumor microenvironment (TME), which reduces NK cell immunotherapy's efficiencies. It is known that immune checkpoints play a prominent role in creating an immunosuppressive TME, leading to NK cell exhaustion and tumor immune escape. Therefore, NK cells must be reversed from their dysfunctional status and increased in their effector roles in order to improve the efficiency of cancer immunotherapy. Blockade of immune checkpoints can not only rescue NK cells from exhaustion but also augment their robust anti-tumor activity [8].

Immune checkpoint inhibitors (ICIs), which are mAbs, have been shown to be effective in clinical trials, defeating tumor immune evasion mechanisms [9]. While some patients significantly respond to ICIs, most cancers are either resistant at first or develop resistance after the initial response. Therefore, treatment strategies must be developed to overcome ICI resistance and other approaches.

Activation of NK cells by Abs has been a hot research topic in recent years. Because cancer is a multifactorial disease, targeting two or more of the involved molecules will aid in effective treatment. Bi/tri-specific Abs, which encompass two or three target-binding units, have

been explored as a therapeutic point of view in cancer immunotherapy [10]. Bi and tri-specific natural killer cell engagers (BiKEs and TriKEs) are bi/tri-specific Abs that specifically target NK cell CD16 and tumor antigens. In this setting, CD16 interaction causes NK cells to become more cytotoxic and improves their anti-cancer actions [11, 12]. Therefore, there is no competition for binding to CD16 with the physiologic serum IgG compared to therapeutic mAbs [13–15]. Due to their smaller size, BiKEs and TriKEs may have better biodistribution than mAbs, particularly in the treatment of solid tumors [16–18]. In addition, they are non-immunogenic, have rapid clearance properties, and can be engineered quickly to target known tumor antigens.

Aptamers, which are short single-strand oligonucleotides (single-stranded DNA or RNA), are emerging as a promising molecular tool in targeted immunotherapy. They encompass Ab-mimicking functions and can attach to specific targets and regulate biological processes. They are more stable than Abs, smaller, less immunogenic, and can be produced without the use of biological systems [19, 20]. Aptamers are currently divided into three groups in cancer immunotherapy: aptamers against immunoregulatory factors, aptamers against cancer-specific proteins, and aptamers used as a drug delivery system for anti-cancer agents [21].

In immune cell therapy, which is an adoptive immunotherapy strategy, large numbers of autologous or allogeneic immune cells are injected into a patient to target cancer cells. In clinical trials, immune cell therapies using non-genetically modified lymphokine-activated killer cells (LAKs) and tumor-infiltrating lymphocytes (TILs) showed relatively minor benefits [22]. Therefore, engineered immune cells have been created to express antigen-specific T cell receptors (TCRs) or chimeric antigen receptors (CARs). Adoptive cell therapy (ACT) with T cells expressing CARs has been widely used to redirect autologous T cells' specificity against lymphoid malignancies. While CAR-T cell treatment has been demonstrated to be highly beneficial in individuals with hematologic malignancies, it has not been proven to be effective in patients with solid tumors. Furthermore, large-scale usage of CAR-T cells confronts various hurdles, including the generation of autologous products from patients, which takes several weeks to manufacture, and the inability to produce clinical doses of CAR-T cells from T cell lymphopenia patients who have been highly pretreated [23]. In addition, graft-versus-host disease (GVHD) can occur even in human leukocyte antigen (HLA)-matched patients when an allogeneic source is used [24]. Antigen escape, trafficking and tumor infiltration, immunosuppressive TME, toxicities [25], and sub-optimal persistence and potency are some of the other constraints [26]. Although

many innovative strategies to improve the function and to overcome the toxicity of CAR-T cells are being explored [27, 28], an alternative approach is to use other immune cell types that are more efficient and safe, such as NK cells. Figure 1 shows blockade of immune checkpoints to improves anti-tumor efficacy of NK cells. In this review, we discuss strategies for immune checkpoint blockade with a focus on CAR-NK cells to redirect NK cells to cancer cells in the treatment of solid tumors.

2 | WHAT ARE NK CELLS?

In 1976, NK cells were defined as a defense against pathogen invasion and malignant transformation [29]. These cells were initially thought to develop in the bone marrow (BM), but recent evidence suggests that they can also develop and mature in secondary lymphoid tissues, including tonsils, lymph nodes (LNs), and spleen [30]. These lymphocytes are located in the blood, BM, LNs, skin, gut, tonsils, liver, uterus (during pregnancy), and lungs [31]. NK cells are licensed and avoid attacking normal self-cells through a process known as education, which helps them acquire functional maturation and self-tolerance during development [32]. NK cells are CD3⁻CD56⁺ lymphocytes in humans, whereas they are recognized in C57BL/6 and SJL mice by the presence of NK1.1 and natural cytotoxicity receptor 1 (NCR1 or NKp46), as well as CD49b but not CD3 [33] and in mice strains without NK1.1 expression (e.g., BALB/c mice), CD49b is used to identify NK cells [33, 34]. Human NK cells are further divided depending on the density of CD56 on the cell surface, which differ in phenotype, tissue placement, and immunological effects: CD56^{bright}CD16^{dim} NK cells (usually known as immature NK cells), constitute about 5%-15% of total peripheral blood (PB) NK cells, preferably reside in secondary lymphoid organs, such as LNs, produce cytokines (especially interferon gamma (IFN- γ)) and play a key role in immunomodulation; CD56^{dim}CD16^{bright} NK cells (also known as mature and cytotoxic NK cells), make up about 90% of NK cells in PB and primarily mediate NK cells immune function [35]. CD56 is not only a marker for NK cells, but it also plays a significant role in their final differentiation because its blockade by mAbs impedes the transition from CD56^{bright} to CD56^{dim}, decreasing their cytotoxic capacity [36]. Furthermore, almost all human NK cells express the activating receptor NKp46 [37, 38].

It is the balance of germline-encoded activating and inhibitory receptors (without undergoing somatic recombination) that controls whether NK cells are activated or inhibited. The activation of NK cells is usually initiated in one of two ways: “missing-self” recognition or “induced-self” recognition [39–42]. “Missing-self”

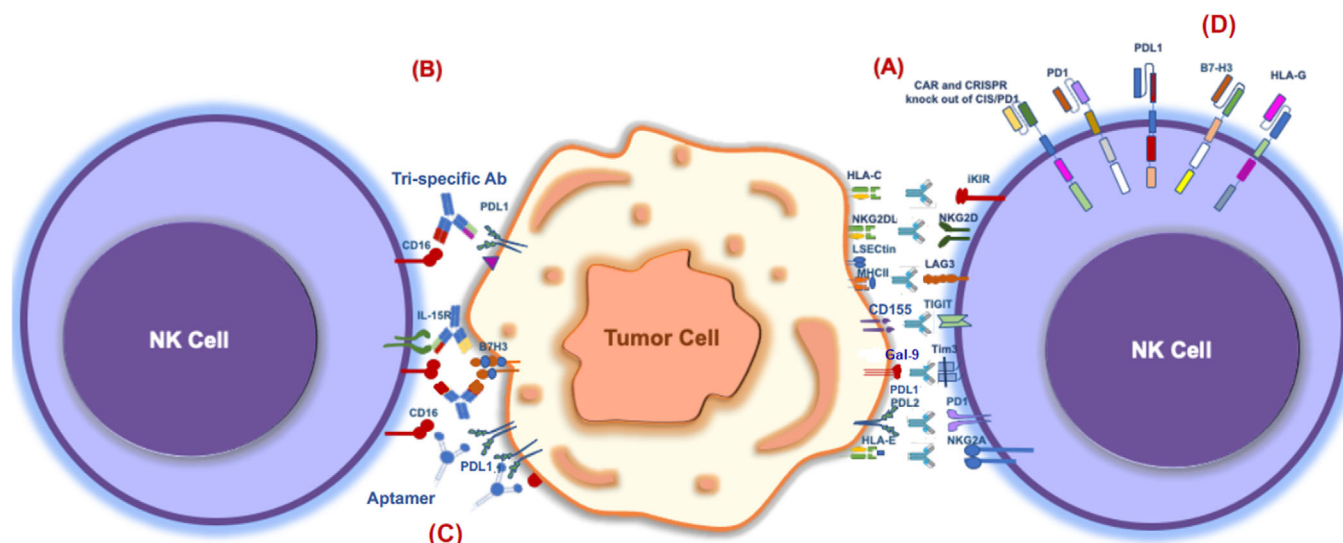


FIGURE 1 Blockade of immune checkpoints improves anti-tumor efficacy of NK cells. NK cell dysfunction is caused by multiple factors in the tumor microenvironment (TME), as demonstrated by the upregulation of inhibitory checkpoint receptors and the downregulation of activating receptors on tumor-infiltrating NK cells, resulting in the exhaustion of these cells. **(A)** Immune checkpoint inhibitors (ICIs), **(B)** Bi and tri-specific natural killer cell engagers (BiKEs and TriKEs), **(C)** Aptamers, and **(D)** Chimeric antigen receptor (CAR)-NK cells targeting immune checkpoints would restore NK cell antitumor activity. (The purple triangle and red circle on the tumor cell indicate tumor antigens [in part B and C]). Abbreviations: Ab, antibody; iKIRs, inhibitory killer cell immunoglobulin-like receptors; NKG2A, natural killer group 2 A; TIM-3, T cell immunoglobulin- and mucin-domain-containing molecule-3; TIGIT, T cell immunoreceptor with Ig and ITIM domains; LAG-3, lymphocyte activation gene-3; PD-1, programmed cell death protein 1; PD-L1, programmed death-ligand 1; HLA, human leukocyte antigen; CAR, chimeric antigen receptor; CRISPR, clustered regularly interspaced short palindromic repeats; CIS, cytokine-inducible SH2-containing protein; MHC-II, major histocompatibility complex class II; IL-15R, IL-15 receptor; NKG2D, natural killer group 2 D.

recognition occurs when the target cells downregulate or lose surface expression of major histocompatibility complex class I (MHC-I) molecules, which is usually associated with virally infected cells or cellular transformation to evade T cell anti-tumor immunity. This induces NK cell activation by reducing inhibitory signals from the MHC-I-binding killer cell immunoglobulin-like receptors (KIRs) or CD94/natural killer group 2 A (NKG2A) in human and Ly49 receptors in mice. Subsequent studies have revealed that the absence of MHC expression is neither sufficient nor necessary for NK activation, and signaling from activating receptors is also required. “Induced-self” recognition occurs when stress or virus-related ligands on target cells engage with germ-line activating receptors, resulting in activating signals.

Leukocyte immunoglobulin-like receptors (LIRs), NKG2A, and some KIRs are inhibitory receptors on the surface of NK cells that detect MHC-I [43–48]. KIRs, which belong to the immunoglobulin (Ig) superfamily, include inhibitory and activating receptors and detect classical HLA-A, -B, and -C (HLA class Ia) in different ways [43, 44, 46]. LIRs, also known as immunoglobulin-like transcripts (ILTs), are Ig superfamily members that include both activating and inhibitory receptors [49–51]. From a total of 11 LIR members, 5 inhibitory receptors,

including LIRB1-5, have been identified [52]. Among them, LIRB1 (ILT2) and LIRB2 (ILT4), in addition to other ligands, recognize HLA-G as their main ligand, resulting in immunological tolerance [50, 53]. NKG2A, another inhibitory receptor, belongs to the C-type lectin family of receptors and is a member of the CD94/NKG2 family of receptors, which includes A, B, C, D, E, F, and H with activating or inhibitory potential. NKG2A pairs with CD94 to form the NKG2A/CD94 receptor and recognizes non-classical MHC-I molecule, HLA-E, as the ligand [54]. In addition, emerging immune checkpoints such as CTLA-4 and PD-1 that are from the B7-CD28 family of receptors, T cell immunoreceptor with Ig and ITIM domains (TIGIT), CD96, lymphocyte activation gene-3 (LAG-3), and T cell immunoglobulin-and mucin-domain-containing molecule-3 (TIM-3) have been identified to mediate NK cell dysfunction in the TME [55]. On the other hand, dominant activating receptors of NK cells, including NCRs (NKp46, NKp30, and NKp44), NKG2D, and DNAX accessory molecule-1 (DNAM-1, also known as CD226), recognize ligands on the surface of virus-infected or malignant cells [56–58]. Coreceptors such as 2B4 (CD244), NKp80, NTB-A, and CD59 are also expressed and begin their function when co-engaged with other activating receptors [57, 59].

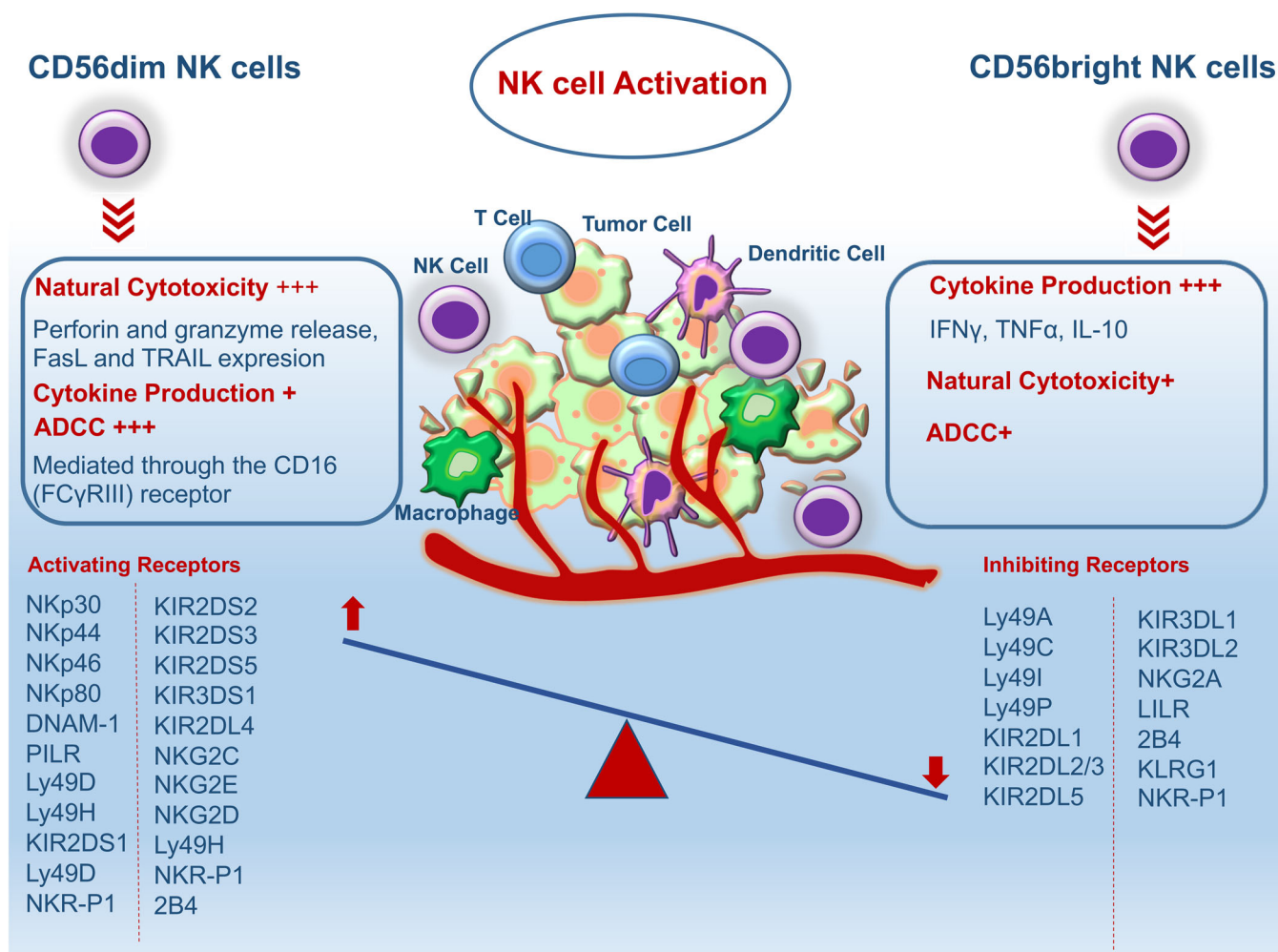


FIGURE 2 Schematic diagram of types and functions of NK cells for killing tumors. It is the balance of germline-encoded activating and inhibitory receptors that control whether NK cells are activated or inhibited. Abbreviations: NK, natural killer; ADCC, antigen-dependent cell-mediated cytotoxicity; FasL, Fas ligand; TRAIL, tumor necrosis factor-related apoptosis-inducing ligand; KIR, killer cell immunoglobulin-like receptor; NKG2, natural killer group 2; DNAM-1, DNAX accessory molecule-1; LILR, leukocyte immunoglobulin-like receptors; KLRG1, killer cell lectin like receptor G1; IFN- γ , interferon gamma; TNF- α , tumor necrosis factor- α .

The effector function of NK cells is similar to that of CD8⁺ cytotoxic T cells, but NK cells do not require prior antigen exposure or MHC class I limitation to carry out their functions [60]. When exposed to their targets, NK cells can cause cytotoxicity through a variety of mechanisms, including expression of Fas ligand (FasL) and tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) [61], the release of preformed cytotoxic granules containing perforin and granzyme B [62–64], secretion of extracellular vesicles, such as exosomes, with cytotoxic potential [65], and participation in antigen-dependent cell-mediated cytotoxicity (ADCC) by detecting IgG Abs attached to the tumor cell surface by low-affinity Fc activating receptor CD16 (or Fc γ RIII) [66]. NK cells are also polyfunctional and can secrete multiple cytokines and chemokines, such as IFN- γ , tumor necrosis factor- α (TNF- α), granulocyte-

macrophage colony-stimulating factor (GM-CSF), IL-5, IL-10, IL-13, macrophage inflammatory protein (MIP)-1 α , MIP-1 β , IL-8, and CCL5 (RANTES) that help modulate other innate and adaptive immune cells function [67–69]. NK cells express cytokine receptors (including IL-2R, IL-12R, IL-15R, IL-18R and IL-21R) that allow them to respond to cytokines secreted by other cells [70]. Also, other receptors, such as chemokine receptors (including CXCR1-4 and CCR5) [71], and a diverse repertoire of pattern recognition receptors (PRRs), such as TLRs, are expressed by NK cells and play an important role in their migration and pathogen recognition, respectively [72]. NK cells have recently been revealed to have adaptive immune cell properties, especially memory-like responses [73–79]. Figure 2 shows a schematic diagram of the types and functions of NK cells for killing tumors.

In naive NK cells, ligation of individual activating receptors (except for CD16) is insufficient to activate them; cytokine exposure is necessary to preactivate them [80]. Cytokines such as IL-2, IL-15, IL-12, IL-18, IL-21, and type I IFNs all play an important role in regulating NK cell activation, maturation, and survival. Improved NK cell performance has been achieved by administering cytokines *in vivo* or pre-treating NK cells before adoptive transfer [81]. In NK cells, IL-12 enhances signaling from activating receptors [82]. IL-2 increases NKG2D expression [83], whereas IL-15 is a powerful driver of NK cell differentiation and proliferation, and is shown to improve NK cell viability *ex vivo* when combined with IL-2 [84]. The production of IFN- γ by NK cells was demonstrated to be stimulated by IL-18, which also provides co-stimulatory activation [85], while the maturation of NK cells without promoting proliferation was enhanced by IL-21 [86]. The proinflammatory cytokine type I IFN potentially leads to the pre-activation of NK cells, readying them for activation by activating receptors [87]. Combination of IL-15 with IL-12 and IL-18 results in significant biological alterations and a status reminiscent of memory NK cells, including improved function upon re-stimulation, hyper-responsiveness to IL-2, increased IFN- γ production and increased cytotoxicity [87–89].

3 | TARGETING OF NK CELL-BASED IMMUNE CHECKPOINTS BY mAbs

3.1 | KIRs

Based on the number of extracellular Ig-like domains and a cytoplasmic tail that dictate the function of the molecule, KIRs are divided into two categories: activating KIRs (aKIRs) and inhibitory KIRs (iKIRs). aKIRs have a shorter cytoplasmic tail than iKIRs and lack an immunoreceptor tyrosine-based inhibitory motif (ITIMs). Binding of a positively charged lysine residue to the KARAP/DAP12 adapter molecule results in activated NK cell-mediated lysis [90, 91]. iKIRs contain two (KIR2DL) or three (KIR3DL) extracellular Ig domains that confer specificity to HLAs. As a result, they can detect changes in HLA molecules caused by a viral infection or cancer [92]. iKIRs are upregulated in tumor malignancy, while aKIRs are downregulated in multiple tumors, such as breast cancer, lymphoma, and non-small cell lung cancer (NSCLC) [93, 94]. These alterations reduce NK cell activation and anti-cancer activity, allowing the tumor to evade immunosurveillance. The anti-tumor activity of NK cells may be induced by mAbs that inhibit the interactions of iKIRs by blocking the signaling pathway. Thus, several anti-KIR mAbs, including those targeting KIR2DL1-3, have been evaluated in several clinical

trials for treating patients with leukemia, lymphoma, multiple myeloma (MM) and some solid tumors [95]. Lirilumab (IPH2101, formerly called 1-7F9) is a humanized IgG4 mAb that blocks KIR2DL1, KIR2DL2, KIR2DL3, KIR2DS1 and KIR2DS2 interacting with HLA-C [96]. However, lirilumab's results in many clinical trials were disappointing. Increasingly, clinical trials of combination blocking strategies are being conducted.

3.2 | CD94/NKG2A

Monalizumab is a humanized IgG4 Ab that blocks NKG2A binding to its HLA-E ligand, which is overexpressed in tumor cells, as well as initiates an NK and CTL-mediated immune response against cancer cells [97]. The findings of published studies from phase I clinical trial NCT02459301 show that monalizumab prescribed in patients with advanced gynecological cancers with minimal therapeutic toxicity is well tolerated [98]. In phase II of the NCT02643550 clinical trial, monalizumab and cetuximab, an epidermal growth factor receptor (EGFR) inhibitor, were tested in patients with squamous cell carcinoma of the head and neck. According to preliminary findings, the most common adverse effects were fatigue, fever, and headache, while the objective response rate was 31% [99]. In clinical trials, NCT05061550, NCT03822351, NCT03794544 and NCT03833440, the efficacy of monalizumab plus anti-PD-L1 blocking mAb (durvalumab) was studied. In several trials, monalizumab and anti-PD-L1 have demonstrated to have a synergistic effect on cancers that express HLA-E and PD-L1 [55].

3.3 | TIM-3

TIM-3 is an inhibitory receptor of the TIM protein family that can detect multiple ligands. The extracellular regions of TIMs include a variable IgV domain that binds to the high mobility group box 1 (HMGB1) and galactin-9 (Gal-9). It also detects phosphatidylserine and carcinoembryonic antigen-related cell adhesion molecule 1 (CEACAM-1) [100, 101]. The intracellular domain of TIMs consists of 5 conserved tyrosine residues that interact with several components of the TCR complex [102]. TIM-3 is considered a marker for the maturation or activation of NK cells because it is expressed on almost all mature CD56^{dim}CD16⁺ NK cells. NK cell-mediated cytotoxicity can also be inhibited by TIM-3 cross-linking. Upregulation of circulating and/or tumor-infiltrating TIM-3⁺ NK cells has been elevated in various types of malignant tumors such as gastric cancer, lung cancer, follicular B-cell non-Hodgkin lymphoma, and colorectal cancer [103, 104]. Anti-TIM-3 Abs such

as Sym023, LY3321367, MGB453, TSR-022, and BGB-A425 alone or in combination with anti-PD-1 or anti-LAG-3 Abs in patients with advanced solid tumors (NCT02608268) are being evaluated. Previous studies have shown that increased TIM-3 expression on NK cells improves cytotoxicity and IFN- γ production [105, 106]. Therefore, due to the use of TIM-3 blockade, there is a risk of decreased NK cell-mediated cytotoxicity in pancreatic cancer cell lines, so caution is required [107]. Anti-Gal-9 Abs, on the other hand, reduce IFN- γ production in NK cells in response to acute myeloid leukemia (AML) blast cells, complicating the effects of TIM-3 inhibition [108]. Therefore, more studies need to be done to comprehend the role of TIM-3 and its potential application in the management of NK cell-mediated malignancy.

3.4 | TIGIT and CD96

TIGIT and CD96 are inhibitory receptors that compete for PVR (CD155) and Nectin-2 (CD112) binding with the activating receptor DNAM-1 [100]. TIGIT and CD96 share the same ligand and are composed of an immunoreceptor tyrosine tail (ITT) and an ITIM. ITT-like motifs play an important role in inhibiting signals. TIGIT interaction with CD155 induces phosphorylation via Fyn and Lck, which recruits SHIP1 in T cells and reduces signaling pathways in immune cells. Human NK cytotoxicity and cytokine production are also prevented by this interaction [100]. TIGIT and its ligands are highly expressed in a variety of cancers, and TIGIT expression has been demonstrated to decrease the activity of NK and CD8⁺ T cells in colorectal cancer patients as well as being linked to the depletion of tumor-infiltrated NK cells in a mouse model of colon cancer [109]. CD155 is highly expressed in many types of tumor cells. TIGIT inhibition could help to reverse NK cell exhaustion and restore efficient anti-tumor immunity. Moreover, NK cells in TME express lower levels of PD1 and CTLA-4 in comparison to T cells. Therefore, it has been suggested that PD-1 Ab treatment is less effective on NK cells than anti-TIGIT blockade as a supplemental treatment because it improves tumor clearance and provides substantial immunity in tumor rechallenged models [109]. PVR expression, on the other hand, has been linked to a poor prognosis in many solid tumors, including colon, breast, lung, and pancreatic cancers. Accordingly, the PVR/TIGIT immune checkpoint axis has been proposed as a novel target for cancer immunotherapy, and several TIGIT-targeting clinical trials are currently undergoing regulatory approval [110]. Tiragolumab, an anti-TIGIT Ab, is currently being evaluated in combination with atezolizumab (anti-PD-L1) for use in small cell lung cancer (SCLC) (NCT04256421) and NSCLC (NCT04513925

and NCT03563716). In addition, two other anti-TIGIT Abs, ociperlimab and domvanalimab, are being investigated in combination with tislelizumab (anti-PD-1) in phase III and zimberelimab (anti-PD-1) in phase II in solid malignancies, respectively (NCT04746924 and NCT04791839). The role of CD96 in NK cells is relatively less clear. One study found that using CD96 Ab-mediated blockade increases IFN- γ production by NK cells and controls cancer in a mouse model of metastatic lung tumor [111–113]. However, the efficiency of CD96 Ab-mediated blockade on the performance of NK cells and their effects on human cancer patients remains unknown. Therefore, further studies are required to grasp its potential as a target molecule for immunotherapy.

3.5 | LAG-3

LAG-3 is a member of the Ig family of receptors with inhibitory properties that are structurally similar to CD4; however, it binds to MHC class II molecules with a higher affinity than CD4. Fibrinogen-like protein 1 (FGL1) is a recently identified ligand for LAG-3 [114]. LSECTin, a member of the DC-SIGN family, has also been described as a potential ligand for LAG-3-expressing immune cells [115]. LAG-3 is expressed as an upregulated molecule on T and NK cells but is also expressed on other immune cells, including TILs, Treg cells, iNKT cells, B cells, and DCs [116, 117]. High expression of LAG-3 has been indicated in patients with breast cancer, gastric cancer, B-cell lymphoma, and lung cancer [118]. Because of LAG-3's role in T cell exhaustion and Treg inhibitory activity, targeting it in combination with anti-PD-1 has a synergistic effect on T cell function restoration [119, 120]. However, the precise role of LAG-3 in regulating NK cell function has yet to be determined, demanding further investigation. Notably, Abs that block the LAG-3 pathway are not able to induce human NK cell cytotoxicity, or even soluble LAG-3, able to bind to MHC-II molecules, has no effect on the NK cells' ability to kill cancer cells [121]. However, targeting LAG-3 in immunotherapy may be beneficial because of its effect on the effective function of T and NK cells. Eftilagimod alpha (IMP321), a soluble version of the recombinant LAG-3-Ig fusion protein, activates human NK cells to produce IFN- γ and TNF- α in vitro and has been utilized as a safety aid for immunization against several diseases and malignancies. IMP321 has been investigated as a monotherapy for advanced renal cell carcinoma and in combination with chemotherapy for metastatic breast cancer in clinical trials [122, 123]. Relatlimab (BMS-986016) is a new anti-LAG-3 blocking Ab that is currently being evaluated in phase I clinical trials, such as solid tumors and lymphomas (NCT03489369) or in combination with

other ICIs such as nivolumab and ipilimumab in advanced cancer (NCT02488759) or either alone or combined with PD-1 blockade, in various cancers, such as in patients with advanced solid tumors (NCT05134948 and NCT01968109).

3.6 | PD-1/PD-L1

PD-1 is expressed on various immune cells, including NK cells. PD-1's cognate ligands are PD-L1 (B7-H1) and PD-L2 (B7-DC), which suppress immune cells such as T cells and NK cells, resulting in immunological escape [124]. PD-1 expression in mature NK cells, CD56^{dim} NKG2A⁻KIR⁺CD57⁺, is found in one-fourth of healthy people's PB but not in CD56^{bright} NK cells [125]. PD-1 overexpression on NK cells is related to NK cell dysfunction in cancer patients with ovarian cancer, Kaposi sarcoma and MM, presumably due to the presence of MHC-deficient tumor cells [126–128]. In comparison to PD-1⁻ NK cells, PD-1⁺ NK cells are functionally exhausted [129]. The blockade of PD-L1 on hepatocellular carcinoma (HCC) cells did not significantly increase the cytotoxicity of hepatic intrasinusoidal (HI) NK cells. [130]. Also, in organoid culture, colorectal cancer cells were effectively killed by primary human-activated NK cells, regardless of PD-L1 expression [131]. On the other hand, anti-PD-1 inhibition may induce the production of cytokines like IFN- γ , which may indirectly boost NK cells [59]. NK cell function is restored using anti-PD-1 mAbs. In mice, tumor-infiltrating NK cells express PD-1, and PD-1 blocking in NK cells promoted an anti-tumor immune response [132]. In post-transplant lymphoproliferative disease (PTLD) patients, disruption of the PD-1 pathway enhances IFN- γ release but has no impact on cytotoxicity, demonstrating a relative dependence on the PD-1 pathway and calling for additional research to determine the function of PD-1 inhibition in NK cells [133]. PD-1 blockade, on the other hand, has been demonstrated to improve NK cell-based anti-tumor responses. The safety and efficacy of NK cells in combination with anti-PD-1 or PD-L1 are being evaluated in clinical trials NCT03841110, NCT04847466 and NCT03439501. Anti-PD-L1 Abs such as atezolizumab, avelumab, and durvalumab have been approved for use as monotherapy or in combination with other ICIs in the treatment of certain types of tumors. By overcoming T cell exhaustion, the use of an anti-PD-L1 mAb promises to increase the effectiveness of cancer treatment. However, the function of PD-L1 on NK cells and the anti-PD-L1 mAb effects on PD-L1⁺ NK cells remain unknown. One study found that NK cell function in some tumors was enhanced by induction of PD-L1 by AKT signaling on NK cells and prevented cell exhaustion [134]. Also, another study found that NK cells induce ADCC against PD-L1-positive tumors in the presence of anti-PD-L1 mAbs,

indicating the efficacy of NK cell immunotherapy in targeting cancers with high PD-L1 expression in combination with anti-PD-L1 mAbs [135]. Avelumab is a fully human IgG1 anti-PD-L1 Ab, with the ability to induce ADCC, was evaluated in patients with non-resectable advanced mesothelioma with or without PD-L1 expression in phase I of the NCT01772004 clinical trial. The findings revealed that avelumab was both effective and safe in clinical trials. The results of the NCT01693562 clinical trial show that durvalumab has clinical benefits in enhancing NK cell-specific genes expression, which leads to NK cell priming of the adaptive immunity response.

3.7 | CD73

During hypoxia, apoptosis, and inflammatory response in TME, the levels of ATP and adenosine as a derivative of ATP are increased dramatically [136, 137]. CD39 and 5'-nucleotidase CD73, which are highly expressed on a variety of cells, including T cells, tumor cells, stromal cells, Tregs, and MDSCs, mediate ATP catabolism in the tumor environment [138, 139]. Interestingly, NK cells in the TME achieve the CD73 molecule and contribute to immune suppression through adenosine production. In this state, adenosine accumulation inhibits NK cell activation by binding to A2ARs on the surface of NK cells [140]. Therefore, CD73 inhibition and A2A receptor blocking have been considered effective treatments to decrease tumor metastasis and improve survival. Recently, anti-CD73 mAbs (oleculumab, NZV930) alone or in combination with other immunosuppressive drugs like anti-PD-1 and A2AR antagonists have been assessed in the treatment of different solid tumors in several phase I/II studies (NCT03381274, NCT03454451 and NCT03549000). However, the impact of anti-CD73 treatment on NK cell function requires more detailed studies.

3.8 | CD47

CD47 was first identified on the leukocytes' surface as a part of the $\beta 3$ integrin pathway. This molecule, which is also known as integrin-associated protein (IAP), is a glycoprotein that belongs to the Ig superfamily [141]. In addition to integrins, CD47 interaction with thrombospondin-1 (TSP-1) and signal regulatory protein alpha (SIRP α) has been indicated in previous studies [142, 143]. The inhibitory function of CD47 is performed through engagement with SIRP α and TSP-1 molecules, which is accompanied with suppression of phagocytic cells, T cells, and NK cells [144, 145]. Kim *et al.* [144] have shown that high CD47-expressing head and neck squamous cell carcinoma

(HNSCC) cell lines induce lower NK cell cytotoxicity compared to low-expressing cells. Findings of another study have indicated the inhibitory effects of CD47 in NK cells are independent of SIRP α [146]. In several phase I and II clinical trials magrolimab (Hu5F9-G4), an anti-CD47 Ab, is under investigation in various combinations with other therapeutic agents such as cetuximab and atezolizumab [55]. Nevertheless, there are still unknown aspects worth investigating to deepen our understanding of the CD47 targeting effects on NK cells.

3.9 | B7-H3 (CD276)

The B7 homolog 3 protein (B7-H3) is an immune checkpoint that regulates the function of T and NK cells [147]. Overexpression of B7-H3 in several cancers has been indicated and is related to tumor metastasis and evasion [148]. Upregulation of B7-H3 suppresses NK mediated cell lysis [149]. The expression of this molecule by the tumor vasculature has also been indicated in previous studies [150]. B7-H3 has been proposed as a suitable target for cancer immunotherapy [148]. Several phase I/II clinical trials have been conducted to assess the impact of anti-B7-H3 in combination with other mAbs in different cancers. To improve the ADCC function of NK cells, B7-H3 binding Fc-optimized humanized IgG1 mAb (enoblituzumab) has currently been designed. It has been shown to suppress tumor development in B7-H3 positive renal and bladder carcinoma [151]. In an ongoing dose-escalating phase I study, the safety and efficacy of a Fc-optimized humanized anti-B7-H3 mAb (MGA271) have been indicated [152]. The pharmacological characteristics of enoblituzumab and also its safety, efficacy, and anti-tumor capacity are being assessed in a phase I study (NCT02982941). The safety, anti-tumor effect, and immunogenicity of enoblituzumab given before radical prostatectomy are being evaluated in a phase II study (NCT02923180). Table 1 shows clinical studies of ICIs for the treatment of solid tumors.

4 | BiKEs AND TriKEs

The GTB-4550 TriKE molecule (CD16a/IL-15/PD-L1) is being investigated in preclinical studies for application in the treatment of solid cancers. This TriKE specifically targets PD-L1⁺ tumors, and the IL-15 part of it enhances NK cell activation and proliferation [153]. The preclinical studies of CD16/B7-H3 BiKE [154] and CD16a/IL-15/B7-H3 TriKE [153] are also ongoing. Additionally, a humanized TriKE (EGFR, CD16a, and PD-L1) with a shared light chain has recently been developed [155]. Therefore, in addition to activating NK cells and inhibiting inhibitory molecules,

this TriKE will likely impede the growth and survival of tumor cells.

5 | APTAMER

Passariello *et al.* [156] have investigated the impact of anti-CTLA4 Ab (ipilimumab) and anti-EGFR CL4 nuclease-resistant RNA-aptamer combinatorial treatment on the anti-cancer function of immune cells, including NK cells. They constructed the Fc region of ipilimumab mAb with the amino-terminated CL4 aptamer and produced an immunoconjugate (CL4-ipilimumab). CL4-ipilimumab activated NK cells and induced IL-2 and IFN- γ production by these cells more efficiently than the parental mAb used alone or in combination with the CL4 aptamer, according to their findings. Also, NK cells' cytotoxic function against SK-BR-3 tumor cells increased in the presence of CL4-ipilimumab with a higher lactate dehydrogenase (LDH) release by tumor cells. They have suggested that expression of CTLA-4 affects the anti-tumor activity of NK cells and CL4-ipilimumab restores it. In another study, Zhang *et al.* [157] produced dual aptamer-equipped NK cells (T-P NK cells) by using TLS11a aptamer targeting HepG2 cells (hepatocellular carcinoma cell line) and PD-L1-specific aptamer without genetic modification. They observed significant cytotoxicity against HepG2 cells following treatment with T-P NK cells and hypothesized that the ease of NK cell-target interaction likely boosts the NK cells' capacity to identify the target cells. Simultaneously, PD1-PD-L1 interaction blockade by PD-L1 aptamer improves NK cell cytotoxicity. They tracked T-P NK cells in tumor-bearing mice and found significant tumor penetration and accumulation of T-P NK cells. Moreover, increased IFN- γ production by T-P NK cells enhanced the expression of PD-L1 on cancer cells. To improve NK cell migration into the tumor environment, CP-bi-apt, a stable CD16/PD-L1 bi-specific aptamer, has been designed. CP-bi-apt has increased NK cell cytotoxicity and promoted NK cell accumulation in the tumor site [158]. Bi-apt seem to be effective in cancer immunotherapy for activating NK cells, but more preclinical and trial research is needed before aptamers can be used as immunotherapy drugs.

6 | CAR-NK CELL THERAPY

While NK cell therapy has a lot of benefits, it also has a number of drawbacks that restrict its effectiveness. NK cells have a short lifespan without cytokine assistance, lasting only one to two weeks. Ex vivo expansion and activation of a limited number of cells is also required, and NK cells may be vulnerable to the immunosuppressive

TABLE 1 Clinical studies of ICIs for the treatment of solid tumors

Immune checkpoint	Clinical trials identifier	Phase	Status	Malignancy	Combinations	Ab/drug	Number enrolled
KIRs	NCT03203876	I	Completed	Advanced and/or metastatic solid tumors	Nivolumab Ipilimumab	Lirilumab (BMS-986015)	10
	NCT01750580	I	Completed	Advanced tumor	Ipilimumab	Lirilumab (BMS-986015)	22
	NCT01714739	I/II	Completed	Advanced solid tumors	Nivolumab Ipilimumab	Lirilumab (BMS-986015)	337
	NCT03532451	I	Active, not recruiting	Bladder cancer	Nivolumab	Lirilumab	43
	NCT03347123	I/II	Terminated	Advanced or metastatic malignancies	Epacadostat Nivolumab Ipilimumab	Lirilumab	11
	NCT03341936	II	Active, not recruiting	Squamous cell carcinoma of the head and neck	Nivolumab	Lirilumab	29
CD94/NKG2A	NCT02459301	I	Completed	Gynecologic cancer	Single	Monalizumab (IPH2201)	59
	NCT02671435	I/II	Completed	Advanced solid tumors	Durvalumab	Monalizumab (IPH2201)	383
	NCT02643550	I/II	Active, not recruiting	Head and neck neoplasms	Cetuximab Anti-PD(L)1	Monalizumab (IPH2201)	143
	NCT05061550	II	Recruiting	Non-small cell lung cancer	Durvalumab Oleclumab Chemotherapy	Monalizumab (IPH2201)	140
	NCT05221840	III	Recruiting	Non-small cell lung cancer	Durvalumab Oleclumab Placebo	Monalizumab (IPH2201)	999
	NCT02331875	I/II	Terminated	Squamous cell carcinoma of the oral cavity	Standard Surgery Postsurgical Adjuvant Therapy	Monalizumab (IPH2201)	3
	NCT03822351	II	Active, not recruiting	Non-small cell lung cancer	Durvalumab Oleclumab	Monalizumab (IPH2201)	189
	NCT03833440	II	Recruiting	Non-small cell lung cancer	Durvalumab Oleclumab Ceralasertib Docetaxel	Monalizumab (IPH2201)	120
	NCT03088059	II	Recruiting	Squamous cell carcinoma of the head and neck	Afatinib Palbociclib Durvalumab Niraparib	Monalizumab (IPH2201)	340
TIM-3	NCT03744468	I/II	Recruiting	Advanced or metastatic solid tumors	Tislelizumab	BGB-A425	162
	NCT03680508	II	Recruiting	Liver cancer	Dostarlimab (TSR-042)	Cobolimab (TSR-022)	42
	NCT02608268	I/II	Active, not recruiting	Advanced malignancies	Decitabine Spartalizumab (PDR001)	Sabatolimab (MBG453)	252
	NCT03961971	I	Recruiting	Glioblastoma multiforme	Anti-PD-1 Stereotactic radiosurgery	Sabatolimab (MBG453)	15

(Continues)

TABLE 1 (Continued)

Immune checkpoint	Clinical trials identifier	Phase	Status	Malignancy	Combinations	Ab/drug	Number enrolled
	NCT03099109	I	Active, not recruiting	Solid tumor	Anti-PD-L1 (LY3300054)	LY3321367	275
	NCT03489343	I	Completed	Solid tumor malignancies or lymphomas	Single	Sym023	24
	NCT02817633	I	Recruiting	Neoplasms	Nivolumab Docetaxel Cisplatin Carboplatin Pemetrexed TSR-033 TSR-042	TSR-022	369
TIGIT	NCT02794571	I	Recruiting	Advanced/metastatic tumors	Atezolizumab Carboplatin Cisplatin Pemetrexed Paclitaxel Etoposide Capecitabine Bevacizumab Pembrolizumab	Tiragolumab	660
	NCT04256421	III	Active, not recruiting	Small cell lung cancer	Atezolizumab Carboplatin Etoposide Placebo	Tiragolumab	490
	NCT03119428	I	Terminated	Locally advanced cancer metastatic cancer	Nivolumab	OMP-313M32	33
	NCT03563716	II	Active, not recruiting	Non-small cell lung cancer	Atezolizumab Placebo	Tiragolumab	135
	NCT03628677	I	Active, not recruiting	Solid Tumor	Zimberelimab (AB122)	Domvanalimab (AB154)	74
	NCT05211895	III	Recruiting	Non-small cell lung cancer	Durvalumab Placebo	Domvanalimab (AB154)	860
LAG-3	NCT00349934	I	Completed	Metastatic breast cancer	Paclitaxel	Eftilagimod alpha (IMP321)	33
	NCT03252938	I	Recruiting	Solid Tumors	Avelumab	Eftilagimod alpha (IMP321)	45
	NCT04811027	II	Recruiting	Squamous cell carcinoma of the head and neck	Pembrolizumab	Eftilagimod alpha (IMP321)	154
	NCT03625323	II	Active, not recruiting	Non-small cell lung cancer Squamous head and neck cancer	Pembrolizumab	Eftilagimod alpha	189
	NCT02614833	II	Completed	Metastatic breast carcinoma	Paclitaxel Placebo	Eftilagimod alpha	242
	NCT02996110	II	Active, not recruiting	Advanced renal cell carcinoma	Nivolumab Ipilimumab BMS-986205 BMS-813160	Relatlimab	155

(Continues)

TABLE 1 (Continued)

Immune checkpoint	Clinical trials identifier	Phase	Status	Malignancy	Combinations	Ab/drug	Number enrolled
	NCT02750514	II	Terminated	Advanced non-small cell lung cancer	Nivolumab Ipilimumab Dasatinib BMS-986205	Relatlimab	295
	NCT02935634	II	Active, not recruiting	Advanced gastric cancer	Nivolumab Ipilimumab BMS-986205 Rucaparib	Relatlimab	190
	NCT04095208	II	Recruiting	Soft tissue sarcoma adult advanced cancer	Nivolumab	Relatlimab	67
	NCT03489369	I	Completed	Solid tumor Lymphoma	Single	Sym022	15
	NCT03044613	I	Active, not recruiting	Gastro/esophageal cancer	Nivolumab Carboplatin Paclitaxel Radiation	Relatlimab	32
	NCT03623854	II	Recruiting	Advanced chordoma	Nivolumab	Relatlimab	20
	NCT03459222	I/II	Recruiting	Solid tumor	Nivolumab Ipilimumab BMS-986205	Relatlimab	255
	NCT03493932	I	Active, not recruiting	Glioblastoma	Nivolumab	Relatlimab	20
	NCT02658981	I	Active, not recruiting	Glioblastoma, recurrent brain neoplasm	Nivolumab or urelumab	Relatlimab	63
	NCT02966548	I	Active, not recruiting	Solid tumor	Nivolumab	Relatlimab	35
PD-1	NCT03590054	I	Active, not recruiting	Advanced solid tumor	Abexinostat	Pembrolizumab	42
	NCT03815084	I	Unknown	Solid tumor	DC-NK Cells	Pembrolizumab	100
	NCT03841110	I	Recruiting	Advanced solid tumor	iPSC-derived NK Cells Nivolumab Atezolizumab Cyclophosphamide Fludarabine IL-2	Pembrolizumab Nivolumab	37
	NCT02660034	I	Completed	Solid tumor	BGB-290 (PARP inhibitor)	Tislelizumab	229
	NCT03707808	I	Completed	Solid tumor	CD1c (BDCA-1)+myeloid DC Ipilimumab Avelumab	Nivolumab	9
	NCT03958097	II	Unknown	Non-small cell lung cancer	NK Cells	PD-1 Ab	20
	NCT04080804	II	Recruiting	Head and neck squamous cell carcinoma	Relatlimab (anti-LAG-3 Ab) Ipilimumab (anti-CTLA-4 Ab)	Nivolumab	60

(Continues)

TABLE 1 (Continued)

Immune checkpoint	Clinical trials identifier	Phase	Status	Malignancy	Combinations	Ab/drug	Number enrolled
	NCT02913313	I/II	Recruiting	Advanced solid tumor	Ipilimumab (anti-CTLA-4 Ab) BMS-986207 (anti-TIGIT Ab)	Nivolumab	241
	NCT04047862	I	Recruiting	Advanced solid tumor	BGB-A1217 (anti-TIGIT Ab) Pemetrexed Paclitaxel Nab paclitaxel Carboplatin Cisplatin Etoposide 5fluorouracil Oxaliplatin Capecitabine	Tislelizumab	444
	NCT02857166	I	Completed	Advanced solid tumors	Single	Toripalimab	24
	NCT03374007	I	Recruiting	Solid tumor lymphoma	Single	Geptanolimab(G.	72
	NCT03468751	I	Unknown	Solid tumors	Single	HLX10	30
	NCT03474640	I	Active, not recruiting	Advanced malignancies	Single	Toripalimab	198
	NCT02715284	I	Recruiting	Advanced solid tumors	Single	Dostarlimab (TSR-042)	740
PD-L1	NCT03518606	I/II	Active, not recruiting	Advanced solid tumors	Tremelimumab Metronomic vinorelbine	Durvalumab	150
	NCT03430466	II	Terminated	Breast cancer	Tremelimumab Fulvestrant	Durvalumab	1
	NCT03084471	III	Active, not recruiting	Advanced solid tumors	Tremelimumab	Durvalumab	867
	NCT03217747	I/II	Active, not recruiting	Advanced malignancies	Utomilumab PF-04518600 (Ivuxolimab) Radiation Therapy	Avelumab	173
	NCT02862275	I	Recruiting	Active hematologic or solid tumor malignancies	Single	Atezolizumab	40
	NCT03212404	I	Recruiting	Advanced cancers	Single	CK-301	500
	NCT03588650	I	Completed	Advanced solid tumors	Single	HLX20	30
	NCT03275597	I	Active, not recruiting	Non-small cell lung cancer	Tremelimumab Stereotactic body radiotherapy	Durvalumab	17
CD73	NCT03381274	Ib/II	Active, not recruiting	Non-small cell lung cancer	Osimertinib AZD4635	Oleclumab	43
	NCT03454451	I/Ib	Active, not recruiting	Advanced cancers	Ciforadenant Pembrolizumab	CPI-006	378
	NCT03549000	I/Ib	Active, not recruiting	Advanced malignancies	PDR001 NIR178	NZV930	127

(Continues)

TABLE 1 (Continued)

Immune checkpoint	Clinical trials identifier	Phase	Status	Malignancy	Combinations	Ab/drug	Number enrolled
CD47	NCT02953782	Ib/II	Completed	Solid tumors and advanced colorectal cancer	Cetuximab	Magrolimab (Hu5F9-G4)	78
	NCT03869190	Ib/II	Recruiting	Urothelial carcinoma	Atezolizumab Enfortumab Vedotin Niraparib Hu5F9-G4 Tiragolumab Sacituzumab Govitecan Tocilizumab Cisplatin Gemcitabine	Magrolimab (Hu5F9-G4)	645
B7-H3	NCT02923180	II	Active, not recruiting	Localized intermediate and high-risk prostate cancer	Single	Enoblituzumab (MGA271)	33
	NCT02982941	I	Completed	Malignant solid tumors	Single	Enoblituzumab	25

Abbreviations: KIRs, killer cell immunoglobulin-like receptors; NKG2A, natural killer group 2 A; TIM-3, T cell immunoglobulin- and mucin-domain-containing molecule-3; TIGIT, T cell immunoreceptor with Ig and ITIM domains; LAG-3, lymphocyte activation gene-3; PD-1, programmed cell death protein 1; PD-L1, programmed death-ligand 1; NK, natural killer; iPSCs, induced pluripotent stem cells; PARP, poly (ADP-ribose) polymerase; Ab, antibody.

TME, which could limit their effector function and trafficking. Exosome shedding, proteolytic cleavage, internalization and degradation, decreased transcript stability, decreased translation, differential glycosylation and lipidation, increased intracellular retention, impaired protein maturation and refolding, and affecting splicing and alternative adenylation are all pathways by which tumors reduce activating receptor ligand expression [159]. Tumors can potentially evade the function of NK cells by upregulating MHC, which engages their inhibitory receptors [160]. Engineering advancements have helped to overcome some of these constraints. One of these, genetic alterations of NK cells with CAR constructs, has received a lot of attention recently.

CAR is an artificially modified fusion protein. It can recognize antigens in the absence of MHC presentation, unlike innate TCRs [161]. CAR-NK cells have extracellular antigen recognition domain, hinge, transmembrane (TM) and intracellular domains just like CAR-T cells. T cells that have been genetically engineered to produce CAR can directly detect the CAR-targeted antigen, enabling them to activate, proliferate, release cytokines, and exhibit cytotoxicity against tumor cells that express the CAR-specific antigen.

Single-chain variable fragments (scFvs), which include variable sections of both the light and heavy chains of a target-reactive Ig, are separated by a flexible linker in the

extracellular binding domain of a CAR [162]. Alternative binding domains, such as ligands (ICAM-1-specific CAR-T cells built using the I domain derived from LFA-1) [163], physiological receptors (chimeric PD-1 receptor) [164], peptides (using NK1 as a targeting moiety for generating MET-specific CAR) [165], nanobodies (single domain antibodies (VHH)), [166] and DARPins (designed ankyrin repeat proteins) are still being investigated in preclinical studies [167]. These recognition sequences establish CAR specificity and affinity for target antigens. Evidence suggests that the antigen binding affinity and stability of the construct are affected by the order of the variable fragments and the length of the linker. Both epitope location and abundance should be considered while constructing scFv [168]. The CAR's hinge domain connects the extracellular domain to the TM domain, providing it enough orientation and flexibility to bind to tumor antigens and influence CAR-NK cell activity. The size of the hinge area has been shown to affect the CAR-T cell function. Longer hinges allow for more flexibility in membrane-proximal antigens, whereas shorter hinges suffice for membrane-distal antigens [169, 170]. According to several studies, CAR-T cells with a shorter hinge region had more anti-tumor efficacy [171, 172]. A TM domain is located between the hinge and the intracellular signaling domain, which is required for surface expression and stability of the receptor. The structure of the intracellular

signaling domain, which consists of immunoreceptor tyrosine-based activation motifs (ITAMs), determines the intensity of the CAR-NK activation signal. The backbone of all CAR generations is the same.

Based on the structure and composition of the endodomain, the evolution of CARs during the last three decades can be approximately divided into five generations [173]. In the first generation of CARs, a single CD3 ζ intracellular domain was presented. Early experiments of first-generation CAR-T cells demonstrated low cytotoxicity and proliferation due to the lack of co-stimulatory (e.g., CD27, CD28, OX40 [CD134], 4-1BB [CD137]) and cytokine (e.g. IL-2) signaling [173, 174]. The second generation of CARs was generated by adding a costimulatory domain to the intracellular signaling domain, such as parts of CD28 or 4-1BB, to increase T cell proliferation and cytotoxicity [175–177]. A third intracellular signaling sequence with an additional co-stimulatory domain, such as OX40 or 4-1BB, was added to the third generation of CARs to improve on the second generation [173]. In contrast to the second-generation CAR, there is no consistent evidence that performance has increased in the third-generation CAR. The CAR-mediated effector functions can be dramatically influenced by the type of co-stimulatory domain used. Co-activation of 4-1BB leads to the formation of T memory cells, whereas co-activation of CD28 leads to enhanced T cell activation and expansion [178]. T cells redirected for universal cytokine-mediated killing (TRUCKs), the fourth generation of CARs, are based on the second generation, but they release transgenic cell products that are expressed either constitutively or inducible when the CAR is activated to induce tumor death [179, 180]. A fifth generation of CARs is currently being investigated; these are based on the second generation of CARs but have a truncated cytoplasmic IL-2 receptor β (IL-2R β)-chain domain with a STAT3 binding site, which effectively provides all three synergistic signals needed for complete T cell activation and proliferation [181]. SUPRA CAR, Tandem CAR, Dual CAR, split CAR, and SynNotch CAR are among the newer generations of CARs [182, 183].

6.1 | CAR-NK cell advantages in tumor immunotherapy

The use of allogeneic haploidentical NK cells in ACT has been clinically proven to be safe, without the risk of causing GVHD. Using NK cells, an increased graft-versus-tumor (GvT) response has been seen after hematopoietic stem cell transplantation (HSCT) for AML [184, 185]. T cells have been proven to be less effective at driving CARs than NK cells. CAR-expressing NK cells are deemed safer in

clinical application than CAR-T cells, as evidenced by the results of many clinical trials, and NK cell immunotherapy is a safe and practical therapeutic method [186]. In some phase I/II trials, allogeneic NK cell administration has been well tolerated and has not caused GVHD or other severe unwanted events [187–189] if the infused product is sufficiently depleted of T cells, indicating that NK cells are general CAR drivers that are not restricted to autologous cells. Unlike CAR-T cells, CAR-NK cells have a short lifespan in circulation; therefore, there is a low chance of on-target/off-tumor damage to normal organs [190]. NK cells also generate different cytokines from T cells. When stimulated, they produce IFN- γ and GM-CSF, whereas CAR-T cells produce proinflammatory cytokines such as TNF- α , IL-1, and IL-6 [191, 192]. While their anti-cancer activity is based on the CAR-specific mechanism, which involves the identification of tumor-related antigens via scFv, NK cells also eliminate malignant cells by detecting a variety of ligands through their receptors like NCRs (NKp46, NKp44, and NKp30), NKG2D, and DNAM-1 [193]. In addition, CD16 facilitates ADCC by NK cells [194]. CAR-modified NK cells are therefore capable of effectively eliminating heterogeneous tumors in which some tumor cells do not express CAR-targeted antigens through both CAR-dependent and NK cell receptor-dependent mechanisms. The CAR-NK cells do not need to be matched to the patient and can be derived from autologous or allogeneic sources, including PB [195], BM-derived hematopoietic progenitor cells (HPCs) [196], induced pluripotent stem cells (iPSCs) [197], human embryonic stem cells (hESCs) [198], umbilical cord blood (UCB) [199], post-partum placenta [200], NK cell lines (such as NK-92) [201] or memory-like NK cells [87], each with its own advantages and disadvantages (Figure 3). Compared to the manufacturing of autologous CAR-T cells, the development of “off-the-shelf” allogeneic NK cells takes less time. Additionally, CAR-NK therapy can be used to treat solid tumors more effectively because of its lower expression of PD-1 [202]. The resistance of cancer stem cells (CSCs) to chemotherapy, radiotherapy, and immunotherapy can lead to relapses and metastasis. So, since CSCs are quiescent and express low levels of MHC-I, NK cells can target and kill them by default [203, 204].

6.2 | NK cell sources of CAR generation

Autologous NK cells activated by IL-2 were employed in early trials of NK cell immunotherapy. Autologous NK cell infusions in combination with IL-2 were examined in other trials after high-dose chemotherapy conditioning [205, 206]. However, this approach led to poor clinical outcomes due to the inhibition of autologous NK cells because ligands presented by self HLA molecules on tumor cells

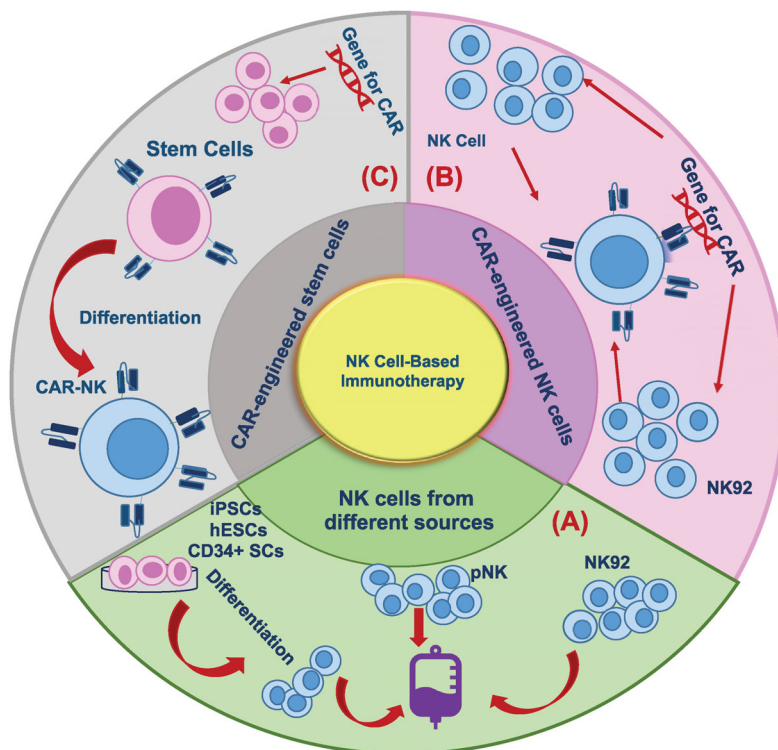


FIGURE 3 The schematic diagram of the various sources for CAR-NK cell production. (A) NK cells from peripheral blood mononuclear cells (PBMCs), umbilical cord blood (UCB), post-partum placenta, and memory-like NK cells are used in NK therapy. It is also possible to differentiate NK cells from CD34⁺ hematopoietic progenitor cells (HPCs). It has recently become more attractive to use induced pluripotent stem cells (iPSCs) for NK cell products. (B) Production of CAR-engineered NK cells by delivery of CAR construct gene into NK cells and NK-92 cell lines. (C) A variety of CAR-expressing vectors can be used to genetically engineer stem cells, and these cells can be differentiated into CAR-NK cells. Abbreviations: NK, natural killer; CAR, chimeric antigen receptor; iPSCs, induced pluripotent stem cells; hESCs, human embryonic stem cells; SCs, stem cells; pNK, peripheral blood NK.

bound to KIR on NK cells [205, 206]. Additionally, IL-2 induces the expansion of Tregs, which can inhibit NK cell expansion indirectly by depriving them of IL-2 or directly by inhibiting NK cell function in a transforming growth factor (TGF)- β -dependent manner [207, 208]. Endogenous NK cells may not retain sufficient cytotoxicity against cancer cells because of the overall immunosuppressive tumor milieu with impaired functions [209]. Another aspect contributing to this limitation could be the difficulty of obtaining pure and enhanced expansion of autologous NK cells from cancer patients. Furthermore, autologous NK cells are vulnerable to the presence of resistant tumor cells, allowing tumor cells that are resistant to autologous NK cells to survive and develop [210]. Patients who received NK cell infusions were heavily pretreated before their cells were collected and used, which might have adversely affected their function and expansion [211]. While it is also possible that anti-KIR Abs may block KIR-HLA matching in order to increase NK function [96], the transfer of allogeneic or haploidentical NK cells with KIR ligand-mismatch prevents this suppression due to the “missing self” recognition of tumor cells [184]. Allogeneic NK cells may also be less likely to be rejected by the recipient’s alloreactive T cells.

6.2.1 | PB-NK cells

The PB-NK cells are phenotypically mature and highly functional. However, they need blood or leukapheresis

from a healthy person who is willing to donate. Even though PB-NK cells are readily available, their low transduction efficiency, coupled with poor expansion, limits their application [212]. However, this source has certain limitations. The process of selecting an appropriate allogeneic NK cell donor is time-consuming, which may cause the patient’s treatment to be delayed. The number of peripheral blood mononuclear cells (PBMCs) that can be extracted and the proportion of NK cells contained in the PBMCs vary considerably among donors, all of which can have an impact on the amount of purified NK cells available for transplant. Furthermore, NK cell phenotypes and functions differ between donors (depending on age, sex, weight, and other factors), reducing consistency in the expansion rate and final product of NK cells.

6.2.2 | UCB-NK cells

Another interesting source of NK cells, UCB, has a number of advantages, such as its ready availability “off-the-shelf”. UCB collection is easy and harmless to the mother or baby, and they are almost never infected with Epstein-Barr Virus (EBV) or cytomegalovirus (CMV). UCB is easily frozen [213]. The benefit of UCB banks is the ability to select donors with certain HLA types and specific NK receptor profiles. UCB also has fewer T cells than PB, and these cells have a naive phenotype, lowering the risk of GVHD from contaminated T cells infused into patients via an NK cell product infusion [214]. The NK

cells derived from the UCB are younger and have a greater potential for proliferating than those derived from the PB [215]. By using feeder cells and cytokines, UCB-NK cells can be expanded and genetically modified to express CARs. Because of their higher proliferative capacity, NK cells from this source are easier to engineer, as demonstrated in the first reported clinical trial of CAR-NK cells [216]. In UCB, NK cells comprise up to 30% of lymphocytes [217], with a higher proportion of CD56^{bright} cells and hence inferior cytotoxic capabilities [215, 217, 218]. Unexpanded UCB-derived NK cells have some limitations, including availability in low numbers due to the UCB unit's small volume and immature function [217]. UCB-NK cells express lower levels of certain adhesion molecules, along with KIRs, CD16, perforin, and granzyme B, compared to PB-NK cells, as well as higher levels of inhibitory molecules, including NKG2A [219]. One study used Good Manufacturing Practice (GMP)-grade K562-based artificial antigen-presenting cells (aAPCs) that express membrane-bound IL-21 and 4-1BB ligand to develop a GMP-compliant procedure that reliably produces therapeutically relevant amounts of GMP-grade NK cells from a CB unit for the purposes of adoptive immunotherapy [199]. When activated and expanded *ex vivo*, CB-derived NK cells express the full array of activating and inhibitory receptors, strongly express eomesodermin (Eomes) and T-bet, two factors needed to mature NK cells, and have cytotoxic properties similar to PB-NK cells [220, 221]. However, unlike PBMCs, CAR-NK cells produced from UCBs are not homogeneous, making standardisation challenging [222].

6.2.3 | NK cell lines

In most clinical trials with CAR-NK cells, the NK-92 cell line is used because it shows clinical benefits and has minimal side effects, and it is the only one that has been approved for clinical application [223, 224]. It demonstrates unlimited proliferation *in vitro* and is less sensitive to repeated freeze/thaw cycles [225]. These properties will make it easier and cheaper to manufacture “off-the-shelf” CAR-NK products for clinical use. Furthermore, they are a genetically modifiable NK cell population. In addition, NK-92 cells have higher transduction efficiency than primary NK cells [226, 227]. However, this cell line has some limitations: neither CD16 (are hence unable to trigger ADCC) nor NKp44 are expressed due to their tumor cell line origin, they must be irradiated before infusion and they can't be expanded *in vivo*, owing to the lethal irradiation required before their infusion [228], rendering them unsuitable for CAR-NK cell therapy. Genetically modifying NK-92 cells to express the high-affinity V158 variant of the FcγRIIIa/CD16a (termed haNKTM) and endogenous,

intracellularly retained IL-2 helped overcome these cytotoxic limitations [224, 229]. NK-92 also lacks some KIRs, with the exception of KIR2DL4 (CD158d), which may promote GVHD [230–233]. A recent study has suggested that the host PBMCs might show cytotoxicity against CAR-transduced NK-92 cells [234]. Moreover, they are EBV-positive and exhibit multiple cytogenetic abnormalities similar to those of NK lymphoma [235]. While repeated infusions of irradiated NK-92 cells are possible and may be utilized to overcome their short-term persistence, such an approach is likely to result in the production of Abs and cellular immunity against the allogeneic cell line, as well as faster rejection with each infusion. Since IL-2 is essential to the growth and survival of these cells in culture, it must be accompanied by any additional stimulation factors [225, 236]. NK-92MI cells were created by genetically engineering NK-92 cells to produce soluble IL-2, thus eliminating the need for exogenous IL-2. In addition to NK-92 cells, other human cell lines have also been evaluated as alternatives, such as NKL, KHYG-1, YTS or NKG [237]. Irradiation reduces the anti-tumor potency of this product when compared to NK cells from other sources, and it may not be clinically relevant.

6.2.4 | NK cells derived from stem cells

NK cell products derived from UCB stem cells, hESCs and iPSCs are currently being tested in clinical trials. Because ESCs are more difficult to obtain and their applications raise ethical concerns, they are used less frequently than CB stem cells and iPSCs. It is possible to obtain large numbers of NK cells by differentiating them from CD34⁺ HPCs. This approach provides virtually unlimited numbers of homogeneous NK cells, as well as the advantage of allowing their genetic manipulation over primary NK cells. CD34⁺ HPCs can be isolated from BM, mobilized PB, or UCB, expanded and then differentiated into mature NK cells in a culture system using a cocktail of cytokines [238]. As a result, a more homogeneous and well-defined NK cell product can be produced. Although PB apheresis after granulocyte colony-stimulating factor (G-CSF) stimulation is a well-established alternative to extracting CD34⁺ progenitors from BM, it may influence the NK cell phenotype [239]. It was previously common to use BM CD34⁺ cells for the generation of NK cells, but UCB CD34⁺ cells have become more frequently used as UCB is easier to obtain and contains higher levels of HSCs [240–245]. In this way, various combinations of growth factor and cytokine mixtures, BM stroma cells, and culture media with or without animal or human serum were used in the different studies. oNKord, an allogeneic partial HLA-matched NK cell product derived from UCB CD34⁺

progenitors, has been designated as an orphan medication by the European Medicines Agency (EMA) and the Food and Drug Administration (FDA) for treating AML patients who are not candidates for allogeneic stem cell transplantation. CD56⁺CD3⁻ NK cells produced from this process are largely similar to PB-NK cells, expressing activating receptors, and exhibiting potent cytotoxicity against leukemia cells *in vitro* and *in vivo* [246]. On the other hand, it has been shown that while CD34⁺ cells from CB can be employed to produce a homogeneous population of CD56⁺ NK cells on a large scale, they do not appear to be as mature as PB-NK cells [238] and have little ADCC activity [247] even after expansion, necessitating additional measures to improve potency for clinical application. In addition, the differentiation process is labor-intensive, requiring frequently changing media. In order to develop clinically applicable NK cell expansion protocols, shorter expansion times and less operator involvement are likely to be preferred.

PSCs, such as ESCs and iPSCs, have the ability to self-renew while maintaining pluripotency and could provide limitless supplies for target cells [248]. It has been established that clinical-scale NK cell generation can be achieved using hESC or iPSC [198, 249, 250]. The use of commonly available hESC/iPSC cell lines is often preferred to the use of stem cells from BM biopsy, G-CSF mobilization, or human embryos [251, 252]. To differentiate these cell sources into NK cells, xenogeneic stromal feeder cell lines [253] or human spin embryoid bodies (EBs) [197, 254, 255] are required, which eliminate xenogeneic cells for more defined NK cell development conditions and can therefore be scaled up clinically. hESC/iPSC were centrifuged to form spin EBs [256], which gave rise to HPCs expressing CD34 and CD45, which were then differentiated into mature NK cells by using a specific cytokine cocktail. The choice of target donor somatic cell type and the reprogramming protocol, which includes the nature and combination of genes as well as the method used to deliver transcription factors into somatic cells, have an impact on the pluripotency and differentiation abilities of reprogrammed cells in iPSC cells [257]. NK cells derived from iPSC/hESC expressed common NK cell markers, such as KIRs, CD16, NKp44, NKp46, NKG2D, and TRAIL, and were cytotoxic against several hematological and solid tumor cells *in vitro* [198, 249]. NK cells produced from iPSCs or hESCs combine the advantages of PB-NK and NK-92 cells, as they have a similar phenotype to PB-NK cells and are a homogeneous population. NK cells derived from the H9 hESC cell line had a lower allogeneic immunological response, but they had a more mature cytotoxic profile than CB-NK cells [249]. In contrast to PB-NK cells, iPSC-derived NK cells, similar to UCB-NK cells, have an immature phenotype, with lower KIR and CD16 expression

and higher NKG2A expression [197, 254, 258]. Because of their unlimited proliferation capacity, iPSCs have emerged as an attractive source of CAR-NK cells [259]. Using iPSCs to produce NK cells allows for more gene modification, repeat dosing, and the generation of standardized products, allowing for more successful treatment of refractory solid tumors [260]. Another advantage of using iPSC-NK products over iPSC-T cell therapies is that iPSC-NK products can be truly “off-the-shelf” because they don’t require HLA matching between donors and patients.

There are still obstacles to overcome before iPSC-NK cells can be safely employed to generate CAR-NK for clinical usage. iPSC-derived cells are always capable of malignant transformation and have the potential to be immunogenic, causing effector cell death or even adverse immunological reactions like cytokine release storms. The 5-week biomanufacturing time for iPSC-NK cells, on the other hand, could be a barrier to establishing and maintaining iPSC-NK cell banks, especially if genetic modifications such as CARs are required. It would be useful to shorten and streamline the production process.

6.2.5 | Memory-like NK cells

Memory-like NK cells were first discovered as a result of CMV infection. CMV was observed to cause the expansion of an NK cell subpopulation with CD94/NKG2C overexpression as well as decreased expression of PD-1, TIGIT, and NKG2A [261, 262]. When activated, CD56^{dim}/CD16^{bright}/CD94/NKG2C cells showed enhanced proliferation, degranulation, and elevated IFN- γ and TNF- α production [263]. In addition to living longer *in vivo*, CMV-induced adaptive memory NK cells can withstand the suppressive effects of Tregs and MDSCs [264, 265]. Incubation of NK cells with IL-12, IL-15, and IL-18 for 16 hours stimulates the generation of memory-like NK cells with abilities and features comparable to CMV-induced cells [87]. On the other hand, IL-12 has been shown to promote NKG2A expression and inhibit NK cell activation [266]. Cytokine-induced memory-like NK cells demonstrated an excellent safety profile, expanded *in vivo*, and resulted in remission in 44 percent of evaluable patients with AML in a phase I trial [89]. According to preclinical research, inserting CAR into a memory-like NK cell improves its survival and cytotoxicity [267].

6.2.6 | Post-partum placenta

Placenta-derived cells contain a proportion of NK cell progenitors with the ability to develop into NK cells during the maturation and expansion stages. After a 21-day

NK culture of placenta-isolated NKs, an average of 1.2×10^9 NK cells with an 80% viability rate was obtained [200]. Derived placental-NK cells are largely similar to UCB-NK cells phenotypically and functionally. The FDA has approved CYNK-001, an allogeneic “off-the-shelf” cell therapy enriched for $CD56^+/CD3^-$ NK cells generated from human placental $CD34^+$ cells, as an investigational new drug (IND) for the treatment of glioblastoma (GBM).

6.2.7 | CAR-engineered stem cells

A CAR can be expressed by transfecting stem cells with the genetic sequence. It is a newly developed method that allows stem cells to differentiate into CAR-NK cells after being engineered. Stem and progenitor cells can be cryopreserved and have better transduction effectiveness than mature NK cells. Engineered stem cells could potentially produce a cellular product that is “off-the-shelf,” eliminating the necessity for a personalized, patient-specific product. Performing gene engineering or gene editing on a small number of stem cells could reduce the amount of gene-engineering/editing materials, which could be cost-limiting, and allow for the most efficient gene engineering/editing, which could be passed down to subsequent divisions and final cell products. Therefore, there is no need to use viral vectors to transfect NK cells, avoiding high cell mortality from viral infection and uncontrollable variation of gene insertion and expression. Furthermore, the advancement of stem cell-engineered cellular therapy will create new opportunities for product upscaling and banking. In addition, gene-engineering technologies like clustered regularly interspaced short palindromic repeats (CRISPR)/Cas9 can be employed to boost the anti-cancer properties of CAR-engineered stem cells. The genetic changes will be evident in differentiated immune cells since these cells are non-differentiated [258, 268]. One study successfully introduced CD19-targeted CARs into $CD34^+$ progenitors followed by NK differentiation. Gene transfer had no effect on hematopoietic differentiation and cell proliferation when transduced at 1-2 copies/cell [269]. In media supplemented with stem cell factor (SCF), bone morphogenetic protein 4 (BMP4), and vascular endothelial growth factor (VEGF), genetically engineered iPSCs can stably differentiate into HPCs. A substantial number of highly homogeneous CAR-NK cells may be produced for therapeutic usage after activation using aAPCs and stimulation with IL-15, IL-7, SCF, and FMS-like tyrosine kinase 3 ligand (FLT-3L) [197, 254, 258]. Only one CAR-engineered iPSC cell is required for this approach to produce a large number of highly homogeneous CAR-NK cell products for therapeutic usage. Li *et al.* [255] used a CAR genetic sequence to transduce iPSCs.

In the xenograft model of ovarian cancer, iPSC-derived CAR-NK cells showed prolonged survival and better anti-tumor efficacy than iPSC-derived CAR-modified T cells and non-CAR-bearing immune cells. In another study, researchers used zinc finger nuclease (ZFN) technology to genetically modify human iPSCs to insert a cDNA encoding an anti-EpCAM-CAR into the adeno-associated virus integration site 1, a “safe harbour” for transgene insertion into the human genome, and then differentiated the modified iPSCs into CAR-expressing iPSC-derived NK (iNK) cells. The CAR-iNK cells’ lytic activity against NK cell-resistant, EpCAM-positive cancer cells was validated in an in vitro cytotoxicity experiment but not against EpCAM-positive normal cells, confirming the CAR-iNK cells’ retained tolerability towards normal cells [270]. A recent study found that anti-glypican 3 (GPC3) CAR-modified NK/ILC cells made from iPSC had a strong effect against tumors and helped people with ovarian cancer live longer without causing acute systemic toxicity [271]. In a recent phase I trial, iPSC was employed for an “off-the-shelf” CAR-NK product (called FT596) to treat CD19-expressing B cell malignancies (NCT04245722). iPSC cell-derived NK cell products that have been genetically modified are being tested in people with different types of cancers (NCT03841110 and NCT04023071). In conclusion, it is not clear currently, which source is optimal for creating CAR-NK cells with the most potent activity.

6.3 | CAR constructs in NK cells

Most CAR-NK cell research has so far relied on CAR constructs developed for CAR-T cells. Novel CAR constructs for NK cells specifically have been developed. However, diverse CAR constructs have various effects on cytotoxicity and cytokine production in NK cells [272, 273]. T and NK cells share signal domains such as $CD3\zeta$ and 4-1BB, but NK cells lack some costimulatory domains (e.g., CD28) and the $CD8\alpha$ TM domain. Both T and NK cells rely on CD3 for signaling and activation [274]. Despite the fact that CD28 is one of the most commonly used costimulatory domains in CAR-T cells [275], its role in NK cell function is less clear [276]. Despite this, when it was combined with $CD3\zeta$ in a second generation ErbB2-specific CAR-NK-92, it performed better than a $CD3\zeta$ construct alone and was similar to the 4-1BB- $CD3\zeta$ CAR against ErbB2-expressing tumor cells [228]. Another study found that NK-92 cells transduced with a CD19-CAR that expressed CD28- $CD3\zeta$ had better cytotoxicity against CD19-positive targets than cells that expressed a 4-1BB- $CD3\zeta$ -containing CAR [277]. Both primary NK cells and CAR-NK cell lines use a $CD8\alpha$ hinge region. CAR-NK cells also used other spacers, such as CD28, IgG Fc domains, and DAP12 [278]. Some of the most

commonly used CAR-NK TM parts have been adapted from CD3 ζ , CD8 and CD28, but others have been explored as well (e.g., NKG2D, 2B4, DNAM-1). Currently, CD8 α and CD28-derived TM are prevalent in primary CAR-NK cells, while CD28 is the preferred TM region in CAR-NK cell lines [278].

It is critical to build optimized CAR constructs based on NK cell-specific signal activation properties in order to improve the activation and cytolytic capacities of NK cells. Typically, CAR-NK cell studies use first-generation CARs, which have a single signaling domain comprising CD3 ζ , Fc ϵ R1 γ , or DAP-12 [279–281]. A second- and third-generation CAR uses one or two costimulatory domains in conjunction with CD3 ζ to improve cytotoxicity. NK cell-specific adaptor molecules such as DAP12 have ITAMs. DAP10, as the NKG2D receptor's adaptor protein, has not ITAM and can enhance and stabilize NKG2D membrane expression. According to one study, anti-CD19-CAR construct incorporating the intracellular CD3 ζ component or DAP10 showed that the anti-CD19- ζ -CAR caused more NK cell-mediated cytotoxicity than the anti-CD19-DAP10-CAR [282]. DAP12 was found to be preferable to CD3 as an intracellular domain for NKG2D-engineered primary NK cells. In this study, anti-prostate stem cell antigen (PSCA)-CAR-NK cells containing DAP12 showed improved cytotoxicity, cytokine production, and tumor eradication in a mouse model [280, 283]. Furthermore, as compared to anti-CD5-CAR-NK cells created with a 4-1BB-containing construct, anti-CD5-CAR-NK cells engineered with a construct including the 2B4 intracellular domain were observed to have greater anti-tumor activity [284]. In comparison to NK-92 cells transduced with the CD28-containing CAR construct, anti-GPC3-CAR-NK-92 cells engineered with a construct containing NK cell-associated DNAM-1 and/or 2B4 costimulatory domains displayed enhanced proliferative capacity, persistence, and cytotoxicity against HCC cells. The maximum cytolytic activity was seen in CAR-NK-92 cells that contained both the DNAM-1 and 2B4 costimulatory domains [285]. To find the best CAR design for NK cells, Li and colleagues used NK-92 cells to produce ten distinct anti-mesothelin CARs and tested their impact on NK cell-mediated killing. According to the reports, only three CARs (CAR4, CAR7, and CAR9) with an NKG2D TM domain and a 2B4 co-stimulatory domain showed the greatest improvement in anti-tumor activity. The capacity of these NKG2D TM and 2B4 carrying CARs to activate signaling by recruiting endogenous DAP10 under the circumstances of mesothelin^{high} A1847 activation could be one reason for their better performance [255]. In an osteosarcoma xenograft mouse model, a receptor called NKG2D-DAP10-CD3 ζ , containing the NKG2D ectodomain and intracellular DAP10 and CD3 ζ costimulatory domains, has been shown to greatly augment the

activation and anti-tumor capacity of primary NK cells. CAR-NK cells expressing the activating receptor NKG2D were designed by Parihar and colleagues to target human MDSCs. They demonstrated that NKG2D-CAR-NK cells could efficiently reduce human MDSCs in vitro and in a mouse xenograft model grafted with human neuroblastoma and MDSCs. Additionally, CAR-NK cells release proinflammatory cytokines and chemokines that might enhance the infiltration and function of subsequently infused CAR-T cells in the mouse model. For solid tumors, CAR-NK cells have been suggested to be combined with T cell-based therapies [286]. A CAR construct against PD-1 using the NKG2D TM domain fused to 4-1BB and DAP10 found DAP10 to be superfluous, and the same construct lacking DAP10 showed improved NK cell function [287]. Based on these results, it is evident that more studies are needed to optimize NK cell-specific CAR constructs. In addition, similar strategies can be used to design other NK activating receptor-based CAR, with the advantage that a single CAR could be used to attack several different types of cancer.

6.4 | Preclinical studies of CAR-NK cells targeting immune checkpoints for the treatment of solid tumors

6.4.1 | PD-1

In the study of Guo *et al.* [287], different chimeric receptors that mediate robust NK cell signaling have been designed using NKG2D. The extracellular domain of the inhibitory receptor PD-1 was used in these NKG2D signaling-based chimeric receptors to counteract the immunological escape mediated by PD-1 ligands in solid tumors. They used the PD-1 ectodomain and NKG2D TM domain as a backbone to rationally design four chimeric receptors by TM protein structure modeling. PNBB, a chimeric receptor that was constructed by fusing the human PD-1 extracellular domain to the hinge region, TM and cytoplasmic domains of human NKG2D and human 4-1BB cytoplasmic domain, showed stable surface expression on NK-92 cells and induced significant in vitro cytotoxicity against various tumor cells. As a result, this strategy now offers a promising method for the rational design of NK cell-tailored chimeric receptors with NKG2D signaling. One study created a novel chimeric costimulatory converting receptor (CCCR) by rationally combining the extracellular domain of PD-1, the TM and cytoplasmic domains of NKG2D, and the cytoplasmic domain of 4-1BB created based on the study of Guo *et al.* [287]. This NK-tailored CCCR was able to switch the negative PD-1 signal into an activating signal, reversing PD-1's immune suppressive effects.

When compared to untransduced NK-92 cells, the CCCR-modified NK-92 (CCCR-NK-92) cells retained their typical NK cell features and displayed enhanced anti-tumor activity against human lung cancer H1299 cells *in vitro*. The rapid clearance of H1299 cells was due to the significant gasdermin E (GSDME)-dependent pyroptosis caused by CCCR-NK-92 cells. In a lung cancer xenograft model, CCCR-NK-92 cells dramatically inhibited tumor growth. Their results highlighted the promising immunotherapeutic potential of using NK-tailored CCCR-engineered NK-92 cells for the treatment of human lung cancer [288]. Li *et al.* [289] developed a structure-based rational design approach and created a novel Dual Targeting Chimeric Receptor (DTCR) PD1-DAP10/NKG2D, which consists of the ectodomain of PD-1 fused to a key co-stimulatory receptor DAP10, and subsequently harnessed the activating receptor NKG2D as the second recognition receptor, to assess solid tumor cell killing capacity. Increased PD-1 and NKG2D cell surface expression in NK-92 cells after retroviral DTCR transduction promoted strong cytotoxicity against human gastric cell SGC-7901. TNF- α and TRAIL levels increased significantly after stimulation of the chimeric receptor DTCR, which can cause apoptosis in SGC-7901 cells. DTCR-NK-92 cells had significant anti-tumor activity in the solid tumor cell SGC-7901-bearing mouse model. In another study, a CAR-NK-92 cell line, with an IL-15R α -sushi/IL-15 complex and a PD-1 signal inverter, was created and named SP (Sushi-IL15-PD1). cDNA fragments of a CAR encoded IL-15R α -sushi, TM domain of CD8, signal transduction zone of 4-1BB, and CD3 ζ , 2A signal peptide, PD1 extracellular region, and IL-15. They showed that CAR expression allowed SP cells to proliferate independently of IL-2 and become more resistant to nutrition-starvation-induced apoptosis. Meanwhile, SP cells were more effective in pancreatic ductal adenocarcinoma (PDAC) cell killing assays than NK-92 *in vitro* and *in vivo*, and there was a positive correlation between the killing capability of SP cells and PD-L1 expression in pancreatic cancer cells. The adhesion, responsiveness, degranulation efficiency, targeted delivery of cytotoxic granule content, and cytotoxicity of SP cells were considerably stronger compared to NK-92. Finally, the SP cell line is a promising adoptive immunotherapy cell line that could be useful as an adjuvant treatment for pancreatic cancer, particularly in patients with high PD-L1 expression [290]. In another study, CRISPR/Cas9 was used to develop a highly efficient method for editing the genome of human NK cells to knock out inhibitory signaling molecules. Up to 90% of primary PB-NK cells were efficiently edited by their method. They showed highly efficient knockout of A disintegrin and metalloproteinase-17 (*ADAM17*) and PD1 (*PDCDI*) genes that have a functional impact on NK cells and demonstrated that these

gene-edited NK cells have considerably improved activity, cytokine production, and cancer cell cytotoxicity. In addition, they were able to expand cells to clinically relevant numbers without losing their activity. They also demonstrated that their CRISPR/Cas9 method can be used to efficiently knock in (KI) genes by delivering homologous recombination template DNA using recombinant adeno-associated virus serotype 6 (rAAV6). Their platform represents a feasible method for producing engineered primary NK cells as a universal cancer immunotherapy [291]. They suggested that this simple method for stable gene KI provides a platform for the delivery of other relevant cargo, including CARs and self-stimulating cytokine receptors.

6.4.2 | PD-L1

Other researchers discovered that PD-L1 t-haNK cells expressed PD-L1-targeting CAR (composed of an extracellular scFv that binds to PD-L1, a TM domain, and a Fc ϵ R1 γ intracellular domain for downstream signaling) and CD16, retained the expression of native NK receptors, and had a high content of granzyme and perforin granules. They showed that irradiated PD-L1 t-haNK cells could lyse 20 of the 20 human cancer cell lines studied *in vitro*, including triple-negative breast cancer (TNBC), lung, urogenital, and stomach cancer cells. The cytotoxicity of PD-L1 t-haNK cells was correlated to the PD-L1 expression of the tumor targets and could be improved by pretreating the targets with IFN- γ . It was found that irradiated PD-L1 t-haNK cells inhibited the growth of engrafted TNBC, lung, and bladder tumors in NSG mice. The combination of PD-L1 t-haNK cells with N-803 (formerly ALT-803), an IL-15 super agonist complexed with IL-15R α -Sushi-Fc fusion protein and anti-PD-1 Ab resulted in superior tumor growth control of engrafted oral cavity squamous carcinoma tumors in C57BL/6 mice. Also, when cocultured with human PBMCs, PD-L1 t-haNK cells lysed MDSCs preferentially but not other immune cell types. This study provides a rationale for using these cells in clinical trials [292]. haNKs were engineered by Robbins *et al.* [293] to express a CAR targeting PD-L1. This second-generation CAR included a scFv derived from NANT-601, and an IgG1 mAb targeting PD-L1, along with a CD8 hinge, a CD28 TM domain, and an intracellular Fc ϵ R1 γ signaling domain. They demonstrated that haNKs engineered to express a PD-L1-CAR killed a panel of human and murine head and neck cancer cells at low effector-to-target ratios in a PD-L1-dependent fashion. Treatment of syngeneic tumors led to CD8 and PD-L1-dependent tumor rejection or growth inhibition and a reduction in myeloid cells endogenously expressing high levels of PD-L1. Also, treatment of xenograft tumors resulted in PD-L1-dependent inhibition of tumor

growth. In the PB of patients with head and neck cancer, PD-L1-CAR-haNKs reduced levels of macrophages and other myeloid cells endogenously expressing high PD-L1. Therefore, clinical study of PD-L1-CAR-haNKs is recommended. Another study developed experimental models to investigate mechanisms of T cell escape and demonstrated that resistance to T cell killing can be overcome by adding NK cells engineered to express a CAR targeting PD-L1 developed in two previous studies [292, 293]. In engineered models of tumor heterogeneity, PD-L1-CAR-engineered NK cells (PD-L1 t-haNKs) inhibited the clonal selection of T cell-resistant tumor cells seen with T cell treatment alone in multiple models. T cell treatment of heterogeneous cancer cell populations in vitro and in vivo resulted in IFN- γ release and subsequent PD-L1 upregulation on tumor cells that escaped T cell killing due to defects in antigen processing and presentation, priming escape cell populations for PD-L1 dependent killing by PD-L1 t-haNKs. These findings shed light on the processes underpinning the synergistic anti-cancer effectiveness of T cell-based immunotherapy that result in IFN- γ production, PD-L1 overexpression on T cell escape cells, and the utilization of PD-L1-CAR-engineered NK cells to target and eradicate resistant tumor cell populations [294]. In another study, PD-L1-targeted CAR was generated and introduced into T, NK, or NK-92 cells. They generated a new atezolizumab-based scFv and combined it with a standard second-generation CAR backbone comprised of the IgG4 hinge region, CD28 TM and signaling portions, and the CD3 ζ signaling domain. PD-L1-CAR effector cells degranulated and produced cytokines in response to PD-L1^{high} MDA-MB-231 cells but not to PD-L1^{low} MCF-7 cells. However, in long-term killing assays, both MDA-MB-231 and MCF-7 cells have been killed by the PD-L1-CAR cells, but with a delay in the case of PD-L1^{low} MCF-7 cells. Notably, the coculture of MCF-7 cells with activated PD-L1-CAR cells led to bystander induction of PD-L1 expression on MCF-7 cells and to the unique self-amplifying effect of the PD-L1-CAR cells. In tumor xenograft models, PD-L1-CAR-T cells were active not only against MDA-MB-231 and MCF-7-PD-L1, but also against MCF-7-pLVX (PD-L1^{low/-}) cells. Significantly, PD-L1-CAR cells exhibited potent cytotoxic effects against non-malignant MCF-10A, HMEC, and BM-MSC cells, but not against HEK293T cells that initially did not express PD-L1 and did not respond to the stimulation. Finally, they discovered that HER-2-CAR-T cells stimulate expression of PD-L1 on MCF-7 cells and hence accelerate the functionality of PD-L1-CAR-T cells when used in combination [295].

Combining genetically engineered immune cells with ICIs is an intriguing strategy. A number of studies have shown that co-administration of PD-1/PD-L1 blockers with CAR-T cell therapy enhances anti-tumor activity and pre-

vents T cell exhaustion in preclinical models [296] and patients receiving mesothelin-based CAR-T cell therapy [297]. CAR-T cells were further engineered to produce and secrete an anti-PD-1 scFv for targeting and boosting CAR-T cell activity in order to reduce the systemic effects of ICIs therapy [298]. Cherkassky *et al.* [299] successfully co-transduced anti-MSLN-CAR-T cells with vectors expressing shRNAs targeting *PD-1* genes. As a result of cotransduction with PD-1 shRNAs and subsequent downregulation of PD-1 intrinsic levels, CAR-T cells were able to further expand and kill MSLN/PD-L1-positive cancer cells. Reduced N-linked glycosylation of PD-1 may reduce PD-1 expression and relieve its inhibitory effects on CAR-T cells. Therefore, a preclinical investigation using an adenine base editor (ABE) to decrease PD-1 suppression by changing the glycosylated residue in CAR-T cells downregulated the expression of PD-1 in CAR-T cells and improved cytotoxic functions in vitro and in vivo [300]. These strategies can also be used for CAR-NK cells.

6.4.3 | B7-H3 (CD276)

The B7-H3-CAR was generated by Grote *et al.* [301] by conjugating a B7-H3 clone m851-derived scFv on a second-generation CAR backbone incorporating a CD8 hinge domain, a CD28 TM domain and the cytoplasmic CD28 co-stimulatory as well as CD3 ζ signaling domains. NK-92 cells engineered with B7-H3-CAR could eliminate neuroblastoma (NB) cells in vitro specifically and long-term while sparing B7-H3-negative cancer cells. In addition, B7-H3-CAR-NK-92 cells demonstrated enhanced cytotoxicity in a 3D NB spheroid model recapitulating in vivo morphology as well as cell connectivity, polarity, gene expression, and tissue architecture, thus bridging the gap between in vitro and in vivo studies. A multitude of NK effector molecules were produced by B7-H3-CAR-NK-92 cells as well as pro-inflammatory cytokines that stimulated the immune system. A stable surface expression of B7-H3-CAR and cytotoxic effector function was observed for more than six months. Grote *et al.* [302] also used these CAR-NK-92 cells that target B7-H3, which is abundantly expressed in melanoma, and tested their effectivity in vitro in the presence of low pH, hypoxia, and other features of the TME that affect anti-tumor responses. Additionally, CRISPR/Cas9-induced disruption of NKG2A was tested for its potential to enhance the anti-tumor effect of NK-92. These B7-H3-CAR-NK-92 cells had the ability to cytolyze melanoma cell lines while being able to overcome the immunosuppressive effects typically exerted by TME. Cytotoxicity of CAR-NK-92 cells was not further improved by knocking out NKG2A. This study concluded that B7-H3-CAR-NK-92 cells are a promising cellular product for

treating melanoma and beyond due to the cytotoxic properties of CARs targeting B7-H3 and the capabilities of an “off-the-shelf” NK-92 cell line unaffected by many negative factors associated with the TME. To improve NK cell functions, one study generated NK-92MI cells carrying anti-B7-H3-CAR by lentiviral transduction. The anti-B7-H3-CAR was comprised of an anti-B7-H3 scFv derived from 8H9 Ab, followed by the CD8 TM region, the intracellular domains of 4-1BB and CD3 ζ . The expression of anti-B7-H3-CAR drastically enhanced the cytotoxicity of NK-92MI cells against B7-H3-positive tumor cells. Perforin and granzyme B secretions, as well as CD107a expression, were significantly increased in anti-B7-H3-CAR-NK-92MI cells. Furthermore, anti-B7-H3-CAR-NK-92MI cells efficiently inhibited tumor growth in mouse xenografts of NSCLC and significantly increased mouse survival days when compared to unmodified NK-92MI cells [303].

6.4.4 | HLA-G

Another study established a novel CAR strategy comprised of the HLA-G scFv, TM and cytosolic domains of KIR2DS4, the P2A (porcine teschovirus-1 2A) sequence, the full-length DAP12 sequence, and inducible caspase 9 (iC9) (FK506-binding protein 12 fused to caspase-9). HLA-G-CAR-transduced NK cells showed effective cytolysis of breast, brain, pancreatic, and ovarian cancer cells in vitro, as well as decreased xenograft tumor growth with extended median survival in orthotopic mouse models. In tumor coculture assays, the anti-HLA-G scFv moiety increases Syk/Zap70 activation of NK cells, implying a reversal of the HLA-G-mediated immunosuppression and therefore, restoration of native NK cytolytic functions. Tumor expression of HLA-G can be further promoted using low-dose chemotherapy, which when combined with anti-HLA-G-CAR-NK, leads to extensive tumor ablation both in vitro and in vivo. This upregulation of tumor HLA-G involves the inhibition of DNA methyltransferase 1 (DNMT1) and demethylation of transporters associated with antigen processing 1 promoter (TAP-1) to trigger the translocation of HLA-G [304].

6.4.5 | Cytokine-induced SH2-containing protein (CIS)

Another study developed a strategy that couples targeting of the CIS protein, a major negative regulator of interleukin IL-15 signaling, by CRISPR/Cas9 gene editing with fourth generation ‘armored’ CAR encoded iC9.CAR19.CD28-zeta-2A-IL-15 in CB-derived NK cells. This combined strategy boosted NK cell effector function

via enhancing the Akt/mTORC1 axis and c-MYC signaling and led to increased aerobic glycolysis. When tested in a lymphoma mouse model, this combined approach enhanced NK cell anti-tumor activity more than either alteration alone, eliminating lymphoma xenografts with no signs of any measurable toxicity. They concluded that targeting a cytokine checkpoint further improves the anti-tumor activity of IL-15 secreting armored CAR-NK cells by enhancing their metabolic fitness and anti-tumor activity. This combined approach could pave the way for the next generation of cancer immunotherapy using NK cells [305]. This strategy could also be used against solid tumors.

6.5 | Clinical studies of CAR-NK cells targeting immune checkpoints for the treatment of solid tumors

Because pre-clinical data support the potential therapeutic benefits of CAR-NK-92 and PD-L1 t-haNK cell therapy, these novel cell therapies have a strong rationale for clinical application. A phase I clinical trial (NCT03656705) is being conducted to evaluate the safety and effects of CCCR-modified NK-92 (CCCR-NK-92) infusion with a starting dose of 1×10^7 - 1×10^8 , twice a week, that will be administered intravenously over 1 h in previously treated advanced NSCLC. NCT04050709 is a phase I study to assess the safety and preliminary efficacy of PD-L1 t-haNK and to determine the maximal tolerated dose and designate the recommended phase II dose in subjects with locally advanced or metastatic solid cancers. In a phase II, three-cohort (2 randomized and 1 single-arm), open-label study (NCT04390399) the comparative efficacy and overall safety of standard-of-care chemotherapy versus standard-of-care chemotherapy in combination with aldoxorubicin HCl, N-803, and PD-L1 t-haNK in subjects with locally advanced or metastatic pancreatic cancer are evaluated. Another phase II study (NCT04847466) is testing the effectiveness of irradiated PD-L1 t-haNK cells combined with anti-PD-1 (pembrolizumab) and N-803 in people with advanced forms of gastric or head and neck cancer. The efficacy of PD-L1 t-haNK with a PD-1/PD-L1 checkpoint inhibitor and N-803 in patients with solid tumors that have progressed and/or relapsed after PD-1/PD-L1 ICIs is also being determined in a phase IIb, multicohort, open-label multicenter study (NCT03228667). In another phase Ib/II open-label study (NCT04927884), the safety and efficacy of sacituzumab govitecan-hziy (an Ab-drug conjugate) in combination with chemoimmunotherapy (cyclophosphamide, N-803, and PD-L1 t-haNK) in subjects with TNBC after at least 2 prior treatments for metastatic disease is being evaluated. Table 2 shows clinical studies

TABLE 2 Clinical studies of CAR-NK cells targeting immune checkpoints for the treatment of solid tumors

No. NCT	Status	Phase	Target	Cancer	NK source	Combination agent
NCT03656705	Enrolling by invitation	I	PD-L1	Non-small cell lung cancer	CCCR-NK-92 cells	Not applicable
NCT04050709	Active, not recruiting	I	PD-L1	Locally advanced or metastatic solid cancers	haNK	Not applicable
NCT04390399	Recruiting	II	PD-L1	Pancreatic cancer	haNK	N-803 Aldoxorubicin HCl Nab-paclitaxel Gemcitabine Cyclophosphamide 5-fluorouracil Leucovorin Stereotactic Body Radiation Therapy Irinotecan liposome
NCT04847466	Recruiting	II	PD-L1	Recurrent/metastatic gastric or head and neck cancer	haNK	N-803 Pembrolizumab
NCT03228667	Active, not recruiting	II	PD-L1	Non-small cell lung cancer, small cell lung cancer, urothelial carcinoma, head and neck squamous cell carcinoma, merkel cell carcinoma, melanoma, renal cell carcinoma, gastric cancer, cervical cancer, hepatocellular carcinoma, microsatellite instability, mismatch repair deficiency, and colorectal cancer	haNK	N-803 Pembrolizumab Nivolumab Atezolizumab Avelumab Durvalumab
NCT04927884	Recruiting	I/II	PD-L1	Advanced triple-negative breast cancer	haNK	N-803 Sacituzumab govitecan-hziy Cyclophosphamide

Abbreviations: PD-L1, programmed death-ligand 1; CCCR, chimeric costimulatory converting receptor; hank, high-affinity natural killer.

of CAR-NK cells targeting immune checkpoints for the treatment of solid tumors.

7 | Conclusion and future perspectives

Although checkpoint inhibitor therapy has been demonstrated to restore anti-tumor immune responses, a single checkpoint disruption does not necessarily result in NK or T cell function, and many cancers develop initial resistance or adapt to become resistant to subsequent treatments [306]. To combat this resistance, researchers are currently investigating combinations of ICIs. A growing body of evidence suggests that this approach has the potential to improve anti-tumor activity and response rates dramatically [307]. In clinical trials and/or animal models, checkpoint inhibitors may also be used with

other therapies such as radiation therapy, targeted therapy, chemotherapy, epigenetic modulators, TLR agonists, or other immunotherapies. It's also worth noting that systemic administration of ICIs medications can exacerbate immune-related side effects [308]. As a result, alternate treatments with fewer side effects must be developed.

Currently, the BiKE and TriKE constructs exhibit great translational potential, but efforts are being made to further improve their efficacy. The efficacy of BiKEs and TriKEs may be limited by CD16 expression because by activating NK cells through CD16, matrix metalloproteinases (MMPs), such as ADAM-17, rapidly clip CD16 from their surface [309–312]. Therefore, BiKE/TriKE function may be enhanced by co-treatment with the ADAM-17 inhibitor [313]. Another approach is targeting other receptors on NK cells to circumvent the CD16 problem [314]. BiKE-mediated NK cell function may also be enhanced by the

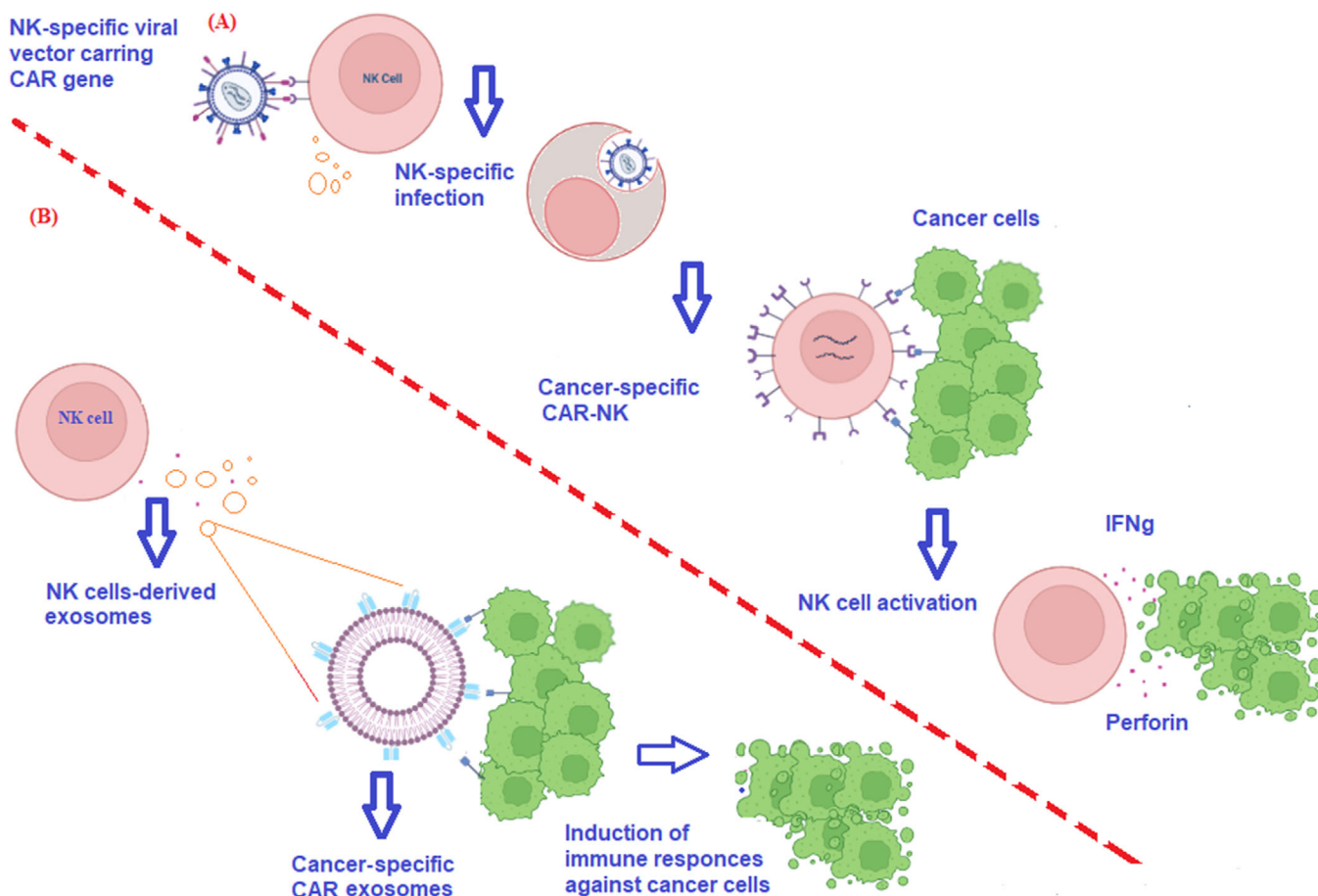


FIGURE 4 Targeting lentiviral for identifying NK cell surface receptor and CAR-NK cell-derived exosome as future directions in CAR-NK cell therapy. **(A)** Targeting lentiviral to identify distinct surface markers on NK cells in vivo as entrance receptors would have significant implications for adoptive immunotherapy development in the future. **(B)** Exosomes expressing CAR derived from CAR-NK cells may offer superior benefits to CAR-NK cells, opening a novel cell-free immunotherapy pathway and suggesting promising treatment for solid tumors. Abbreviations: NK, natural killer; CAR, chimeric antigen receptor; IFN- γ , interferon gamma.

co-administration of cytokines such as IL-15, IL-2, IL-21, and IL-12 [314].

Compared to Abs, aptamers are still in a nascent stage, meaning further development is required to ensure widespread adoption. Clinical trials testing aptamers' therapeutic effects are needed to accurately determine their clinical potential for cancer treatment. Although aptamer-based cancer therapies have huge potential, their disadvantages, such as serum instability and rapid renal clearance, may limit their application. Therefore, modifications are needed to enhance their in vivo half-life and their superior therapeutic benefits [21].

Based on a growing amount of evidence, CAR-NK cell-based cancer therapy has emerged as a promising and advanced research area. However, the field of CAR-NK therapy is still in its infancy, and many questions remain unresolved. To mention a few, these include determining the best NK cell source for immunotherapy, enhancing persistence capacity, determining the best protocol for

introducing foreign genetic material into NK cells due to NK cells' resistance to genetic engineering and optimizing them using viral vectors, nonviral alternatives such as electroporation, nucleofection, lipofection, transposon systems, trogocytosis or nanoparticle treatment; and genome editing using CRISPR/Cas9, ZFNs or transcriptional activator-like effector nucleases (TALENs). Also, determining CARs with the optimum structure for improving NK cell proliferation, activation, cytokine production, and cytolytic activity, the best ex vivo expansion method for acquiring a homogeneous suspension of memory-like, unexhausted NK cells from harvested NK cells to obtain clinically relevant numbers, incorporating a suicide gene into CAR-NK cells as an important point for safety concerns and best freezing media to improve the preservation of NK cells are other challenges in the development of NK cell-based immunotherapy.

Targeting lentiviral to identify distinct surface markers as entrance receptors have enabled precise in vivo

gene delivery into certain cell types [315, 316]. Engineering human NK-targeted vectors capable of efficient and highly selective gene delivery into human NK cells in vivo would have significant implications for adoptive immunotherapy development in the future. The targeted vectors will be employed as a therapeutic tool in this situation, and the cells will not need to be prepared or expanded ex vivo (Figure 4A). Exosomes, which are released by immune cells, can affect tumors or the anti-tumor immune response. Exosomes from NK cells contain FasL, granzysin, perforin, and granzyme A and B, which have cytotoxic properties against tumor cells [317–319]. Researchers have previously demonstrated the safety and efficacy of CAR exosomes in treating tumors. A recent study showed that purified CAR-T cell exosomes expressed CARs, and these CAR⁺ exosomes expressed a high level of cytotoxic molecules and were cytotoxic to tumor cells. As opposed to CAR-T cells, CAR exosomes did not express PD-1, and their anti-tumor effect could not be weakened by recombinant PD-L1 treatment. CAR exosomes were relatively safe compared to CAR-T therapy in a preclinical in vivo model of cytokine release syndrome (CRS) [320]. Exosomes have been shown in multiple studies to have several other benefits, including accessibility and storage, the ability to pass through the blood–brain and blood–tumor barriers, the ability to load various effector molecules, and the ability to combine other therapeutic modalities [321, 322] making them potential alternatives to cell therapies. As a result, exosomes expressing CAR derived from CAR-NK cells may offer superior benefits to CAR-NK cells, opening a novel cell-free immunotherapy pathway that has yet to be explored (Figure 4B). The nanometer size of NK extracellular vesicles as well as their ability to survive in acidic TME makes CAR-NK cell-derived extracellular vesicles a promising treatment for solid tumors. In addition, a novel biomimetic platform, nanoparticles (NPs) camouflaged in the cell membranes, mimics some membrane functions of the cells from which these membranes are derived in biological systems. It is also suggested to design biomimetic nanostructures by coating NPs with NK-secreted exosomes containing identifying markers (e.g., CD226) and cell-death markers (e.g., TRAIL, FAS, and perforin/granzyme B) for targeting and killing circulating tumor cells (CTCs) [323]. It is desired to develop exosome-biomimetic nanoparticles with good biocompatibility and high drug loading for the treatment of cancer.

It is expected that a variety of combinatorial approaches will be developed in the near future to improve NK cell active tumor targeting, persistence and trafficking, higher cytotoxicity while maintaining safety, and improved resistance to immunosuppressive TME. Also, more research needs to be conducted on the safety, tolerability, and effec-

tiveness of checkpoint inhibitors and their combinations. To identify appropriate patient populations for specific therapies, it is important to understand the immune phenotypes of tumors, lymphocyte infiltration, and TME composition. This could assist with deciding which therapies or combinations will be the most effective. Furthermore, identification of biomarkers for the prediction and monitoring of the response to specific therapies or the appropriate combination of therapies is essential.

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AUTHOR CONTRIBUTIONS

All authors contributed to manuscript conception and drafting of the manuscript. All authors provided critical review and revisions, and approved the final version of the manuscript.

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The authors declare that they have no conflicts of interest.

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