

COMMENTARY

Mining new sources of natural history observations for disease interactions

Allyson Kido¹ and Michael E. Hood^{1,2}

Manuscript received 30 September 2019; revision accepted 7 November 2019.

¹ Department of Biology, Amherst College, Amherst, Massachusetts, USA

²Author for correspondence (e-mail: mhood@amherst.edu)

Citation: Kido, A. and M. E. Hood. 2020. Mining new sources of natural history observations for disease interactions. American Journal of Botany 107(1): 3-11.

doi:10.1002/aib2.1409

KEY WORDS anther smut; biogeography; Caryophyllaceae; citizen science; disease ecology; epidemiology; herbarium; Microbotryum; natural history collections; phenology.

Natural history collections provide resources to investigate specimens from diverse and distant places, shedding light on trait variation and distributions of species at scales not possible for individual researchers in the field. Additionally, insights can be gained on the biotic interactions of wild species, among which infectious diseases are known to have a major influence on the ecology and evolution of natural populations (Dobson and Crawley, 1994; Burdon and Thrall, 2008; Ghelardini et al., 2016). However, our knowledge of disease distributions, in terms of spatial, temporal, and host-specific patterns, remains significantly lacking outside of humans and agriculturally important organisms. Wild plant populations often occur across a broader range of densities, environmental gradients, and ecological communities than do crop plants. Understanding disease distributions in these populations can yield critically important information on pathogen transmission modes (Alexander et al., 1996, 2007; Bruns et al., 2018) and factors leading certain host species to be more or less heavily impacted (Burdon and Chilvers, 1982; Eastburn et al., 2011).

Multiple studies have utilized museum collections to investigate wild plant-pathogen systems (Ristaino et al., 2001; Li et al., 2006; Alexander et al., 2007; Saville et al., 2016; Andrew et al., 2018), among other plant ecological interactions (e.g., plant-animal interaction, Beauvais et al., 2017, or phytosociology, Hanan-A et al., 2016). There are two types of natural history collections relevant to such disease-related studies: pathogen-centered collections that directly target and preserve the disease-causing organism and broader

plant-centered collections that nonetheless may retain evidence of the disease occurrence and cause. While diverse phytopathogenic fungi are documented in fungaria (i.e., mycological herbaria) (Brock et al., 2009; Heberling and Burke, 2019), surveys of general plant collections for symptoms and signs of pathogens (Alexander et al., 2007; Yoshida et al., 2014) can additionally reveal the patterns of infection incidence based on the proportions of plants affected (Hood et al., 2010). Such ecological insights beyond the immediate intention of collectors (exaptations, sensu Heberling et al., 2017) have been highlighted as an important yet underutilized benefit of natural history collections (Heberling et al., 2017; Meineke et al., 2018a, b).

Despite the tremendous value of herbarium specimens, depositions of new specimens have substantially declined over the last several decades (Prather et al., 2004; Lavoie and Lachance, 2006; Renner and Rockinger, 2016; Beauvais et al., 2017; Romberg and Rivera, 2017), limiting our ability to assess whether historic distributions are impacted by ongoing environmental changes or to find and study extant populations (Applequist et al., 2007; Meineke et al., 2018a; Lang et al., 2019). The rise in citizen science platforms, such as iNaturalist (www.inaturalist.org), eBird (www. ebird.org), or Zooniverse (www.zooniverse.org), holds new potential for assessing the ecology of wildlife species. iNaturalist alone currently has over 10 million plant observations, primarily from within the past decade; other online platforms (e.g., calflora.org) add to the available data. Several recent studies have emphasized

American Journal of Botany 107(1): 3–11, 2020; http://www.wileyonlinelibrary.com/journal/AJB © 2019 The Authors. American Journal of Botany is published by Wiley Periodicals, Inc. on behalf of the Botanical Society of America. This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original

work is properly cited and is not used for commercial purposes. • 3

the value of such platforms as complementary to museum collections for documenting species distributions (e.g., Heberling and Isaac, 2018). Some citizen science projects have targeted diseases (e.g., Meentemeyer et al., 2015; Brown et al., 2017), which are particularly informative in the case of emerging diseases that threaten domestic or wildlife populations. Such data are similar to the pathogen-centered natural history collections mentioned above in reporting the occurrence of disease. Diseases are less often a desired subject of citizen scientists documenting general biodiversity inventories, but a similar strategy of quantitatively surveying online plant observations for telltale signs of pathogens may be feasible and could contribute valuable and current information on how often plants are affected by of these important ecological interactions. Such insights are especially important in this era of strong environmental change when access to information at broad geographic scales is urgently needed.

Here, we aimed to assess the utility of recent citizen-driven natural history observations for gaining insights into the ecological interactions of wild plant populations. We compared different sources of information on disease occurrence, illustrated by using and advancing knowledge on anther-smut disease affecting a diverse range of plant taxa. We compared data on the distribution of anther smut from fungaria, surveys of general collections in plant herbaria, and surveys of plant observations deposited to the iNaturalist online platform. We show the effectiveness of the new online resources for quantifying variation in disease incidence across members of the Caryophyllaceae host family and demonstrating species-specific results for disease occurrence across temporal (i.e., phenological) and spatial scales (i.e., within native and introduced ranges) that complement prior studies. While pointing to other diseases where similar approaches are reasonable, we show that the tremendous efforts in citizen-driven observations can be extended beyond their original aims regarding plant distributions, enriching our understanding of important ecological interactions and integrating with approaches based on herbarium collections.

ANTHER SMUT AS A MODEL FOR BROADLY DISTRIBUTED DISEASES

Anther smut has been a particularly informative model for pathogen surveys in general herbarium collections. This disease is caused by fungi that replace the host's pollen with dark-colored spores (Schäfer et al., 2010). Although anther smut can be clearly ascertained upon examination of the anthers in comparison to healthy flowers (Fig. 1), multiple studies have noted the general lack of recognition of the disease by plant collectors (Antonovics et al., 2003; Hood and Antonovics, 2003; Rabeler and Hartman, 2005; Bueker et al., 2016) or the assumption that appearance is part of the plant species natural phenotypic variation (Hood and Antonovics, 2003; Antonovics and Hood, 2018). This lack of recognition by collectors has allowed for comparison of disease rates based upon the proportions of specimens with anther smut for a species (Hood et al., 2010) or comparisons of disease incidence across hosts' geographic distributions or time periods of collections (Antonovics et al., 2003; Bueker et al., 2016).

Herbarium surveys for this disease have included broad groups of host taxa (Hood et al., 2010) and targeted host species (Antonovics et al., 2003; Rabeler and Hartman, 2005; Bueker et al., 2016). A nearly global disease distribution has been revealed, which



FIGURE 1. *Silene virginica* observations showing signs of anther-smut disease as dark, powdery, spore-filled anthers. (A) Herbarium sample of *S. virginica* (Collector: M. Nee, 1983; image courtesy of the C. V. Starr Virtual Herbarium, http://sweetgum.nybg.org/science/vh/; barcode NY 331579). (B) iNaturalist observation of diseased *S. virginica* (observer: M. Rung, 'wildflowerenthusiast5', 2014; http://www.inaturalist.org/ observations/1084300). (C). iNaturalist observation of healthy *S. virginica* (observer: J. Gallagher, 'judygva', 2016; https://www.inaturalist.org/observations/4283886).

often extends to the limits of individual host species ranges and is associated with particular host traits (e.g., perennial life history; Thrall et al., 1993; Hood et al., 2010). Herbarium surveys for anther smut are facilitated by the main symptom being in the flowers, which are often the target of preservation due to their importance in taxonomic determination. Similarly, photographs of flowers are the focus of natural history observations of plants recently made available through online platforms. The best studied of these anther-smut pathogens are members of the basidiomycete genus *Microbotryum* (subphylum Pucciniomycotina) occurring on plants in the carnation family, the Caryophyllaceae. The fitness consequences of infection are large because the hosts are sterilized and the infection is systemic and persistent (Alexander and Maltby, 1990). Sporulation in the anthers can facilitate pathogen spread via pollinators, and anther smut has been studied as a model of frequency-dependent and vector-borne disease transmission (Alexander and Antonovics, 1988; Bucheli and Shykoff, 1999). The most affected members of the Caryophyllaceae include the tribes Sileneae and Caryophylleae (Thrall et al., 1993), but with wide variation in occurrence among the species even within plant genera (Hood et al., 2010).

Microbotryum species can also occur on several other plant families (Piątek et al., 2005; Kemler et al., 2009) and, in some cases, cause other forms of smut diseases (e.g., ovary smut, whole-flower smut or leaf smut) (Kashefi and Vánky, 2004; Lui et al., 2009). Additionally, there are a few instances of convergence for the anther-smut disease syndrome, caused by distantly related fungi in the subphylum Ustilagomycotina (Bauer et al., 2008; Roets et al., 2008).

SURVEYS OF NATURAL HISTORY OBSERVATIONS OLD AND NEW

The availability of rapidly accumulating online natural history observations presents the opportunity to compare them with independent data sets based upon specimen collections. These include specimens of *Microbotryum* (formerly *Ustilago violacea*) in fungarium collections, prior published herbarium surveys of general plant collection, and a survey of plant observations deposited in the iNaturalist website.

Fungarium data were retrieved from the Mycology Collections data Portal (MyCoPortal; http://mycoportal.org; accessed 18 June 2019), integrating records from ca. 120 institutions, and from examination of collections at the Royal Botanic Gardens, Kew (K). Specimen details (i.e., date, locality, host species) were used to remove duplicate depositions; a few recognized instances of anther smut in agricultural greenhouses (Conners, 1933; Spencer and White, 1951) and experimental inoculation studies (Kniep, 1919) were also removed. A total of 1394 specimens remained, of which 846 contained decipherable and informative location data.

Data from prior herbarium surveys of general plant collections included studies that focused on the tribe Sileneae, representing ca. 39,000 specimens from a dozen major herbaria (Hood et al., 2010) and targeted surveys of the North American native host species *Silene virginica* and *Silene caroliniana* from regional collections (Antonovics et al., 2003).

iNaturalist observations within the Caryophyllaceae (n = 79,801) were examined (by M. E. Hood) for anther smut as the presence of dark-colored fungal spores in place of pollen (Fig 1) (http://inatu ralist.org; accessed 2 July 2017 through 24 June 2019). Plant identifications are often evaluated by the online community, and those ascertained at least to the level of tribe were used for quantifying disease incidence (Table 1). Summary statistics on observation numbers per species, geolocation data, and phenology were collected from iNaturalist main webpages for each species. An iNaturalist project, called Wildflower Health Watch, was created for adding observations that included anther smut; a small number of observations did not permit assembly into projects and were not included in further analyses. iNaturalist entries explicitly for Microbotryum species were few and were not included in this study. Plant species in other families known to harbor anther smut were also examined to assess detectability of the disease among iNaturalist observations.

This iNaturalist survey produced a quantity of anther smut observations comparable to those from the fungaria or the prior survey of herbarium general plant collections. However, this quantity of observations has been achieved in iNaturalist over the much narrower observation timespan of a decade, compared to two centuries for the specimen collections, and in a period when the number of observations in the other databases has markedly declined (Fig. 2). The number of diseased species in the Caryophyllaceae found in the iNaturalist observations (n = 43) is smaller than in the other databases (fungaria, N = 97; herbarium general plant collection, N = 111) (Appendix S1). Still, 13 host-pathogen combinations in the iNaturalist survey had not been recorded in either the fungarium data or the general plant herbarium collection survey (Table 2). Overall, the incidence of disease in iNaturalist is very similar to herbarium survey of the Sileneae specimens (iNaturalist disease incidence = 0.95%; herbarium survey = 0.89%; χ^2 = 0.689, df = 1, *P* = 0.407).

Geographic distributions of anther-smut observations differed somewhat among three sources of data (Fig. 3). The fungarium collections were focused mostly in western Europe, with the largest

TABLE 1.	Observations of	f anther-smut diseas	e in the Caryoph	nyllaceae among i	iNaturalist data.
----------	-----------------	----------------------	------------------	-------------------	-------------------

Family	Subfamily	Tribe	Total count	Diseased count	Proportion diseased
Caryophyllaceae			79,801	380	0.00476
	Caryophylloideae		38,011	357	0.00939
		Sileneae	26,946	257	0.00954
		Caryophylleae	11,065	100	0.00904
	Alsinoideae		35,205	23	0.00065
		Alsineae	27,102	11	0.00041
		Arenarieae	2,472	12	0.00485
		Sagineae	2,884	0	0.00000
		Sclerantheae	2,117	0	0.00000
		Eremogoneae	630	0	0.00000
	Paronychioideae		6,585	0	0.00000
		Sperguleae	2,986	0	0.00000
		Polycarpeae	2,366	0	0.00000
		Paronychieae	1,233	0	0.00000
		Corrigioleae	0	0	0.00000

Note: Tribe-level data are summed to produce subfamily- and family-level statistics.



Decade Intervals

FIGURE 2. Anther-smut disease observations over time and by data collection type. The blue dashed line represents fungaria (i.e., mycological herbaria) for hosts in the Caryophyllaceae (N = 846, number of observations with year data [w/yr] = 599), red dash-dotted line represents the survey of herbarium general collections of hosts primarily in the Sileneae from Hood et al. (2010) (N = 373, w/yr = 357), and green solid line represents the survey of iNaturalist observations of hosts in the Caryophyllaceae (N = 361, w/yr = 361). Herbarium general collection survey includes data obtained before the last interval from 2010 to 2020.

numbers from Germany, Sweden, and Austria (Fig. 3A), likely reflecting the intense activity of fungal biologists in Germany, especially coinciding with the peak in collections during the pre-war period of the 20th century (Ainsworth, 1976). The survey of general plant collections displayed the broadest distribution among continents, and indeed, the underlying aim of that study was to cover herbaria from distant regions (Hood et al., 2010) (Fig. 3B). The iNaturalist distribution was more restricted in geographic coverage, with fewer records from southern or extreme far northern latitudes (Fig. 3C).

The differences seen between the iNaturalist and historic collections may be due to differing goals of the observers. In general, herbaria contain specimens primarily collected to be representative of diversity from a comprehensive range of geographic locations. On the other hand, iNaturalist data are being generated by citizen scientists, probably more locally and without particular research aims in mind. As a result, observed distributions from any particular source of data represent where the observers tend to go, and they do not necessarily reflect unbiased ranges of the organisms at broad geographic scales.

RESOLVING AMONG-SPECIES VARIATION IN DISEASE INCIDENCE

Pathogens are expected to cause disease on a fraction of their host species, and thus surveys of disease distributions are facilitated by large numbers of host observations. Indeed, on the iNaturalist platform, anther smut was infrequent overall, occurring in 0.48% of 79,801 Caryophyllaceae observations (Table 1). However, the disease incidence varied greatly among taxonomic divisions within this plant family. Among subfamilies, incidence of disease ranged from 0.00% to 0.95%. Disease incidence was highest in the Caryophylloideae, low in the Alsinoideae, and no disease was recorded in the Paronchioideae, consistent with a prior review of the literature (Thrall et al., 1993). These differences among subfamilies were highly significant (χ^2 = 329, df = 2, *P* < 0.001). Within the Caryophylloideae subfamily, the Sileneae and Caryophylleae tribes were nearly equal in disease incidence at 0.95% and 0.90% ($\chi^2 = 0.211$, df = 1, P = 0.646), including in these tribes the genera Silene and Dianthus, respectively, that are known to be commonly diseased (Thrall et al., 1993). Results so similar to prior herbarium surveys lend confidence in the ability of citizen observations to be informative of real differences among host groups.

Among the 43 plant species in the Caryophyllaceae where anther smut was found in the iNaturalist survey (Table 2), all have perennial life histories, consistent with conclusions of prior studies (Thrall et al., 1993; Hood et al., 2010). There was large variation among plant species in the proportions of observations with anther smut, and disease-rich species were not restricted to one section of the family (Table 2). Several species were found to have significantly higher disease incidence than the average for their subfamily, most notably including (in descending order of the significance of the binomial probabilities, analyzed like in the herbarium survey of Hood et al., 2010): Silene virginica (Sileneae), Moehringia lateriflora (Arenarieae), and Dianthus carthusianorum (Caryophylleae). Of particular note was S. virginica, with 6.91% of 1723 plants observed being diseased, and these constituted nearly one third of all disease observations within the family. This level of disease was not statistically different from the 8.32% among 1022 herbarium specimens surveyed in a prior study (Antonovics et al., 2003) ($\chi^2 = 1.855$, df = 1, *P* < 0.173). The statistical significance of the absence of disease was not assessed for all perennial species (see Hood et al., 2010), although some species known to be diseased in nature were notable for the significantly low rate of disease among iNaturalist observations, including Silene flos-cuculi, Silene laciniata, and Silene vulgaris as examples (Table 2).

The disease incidence among herbarium general collections and among iNaturalist data was remarkably similar despite being recorded by different sets of observers and at different times. This similarity in disease incidence is consistent with prior indications that anther smut generally goes unrecognized by a botanist when making field collections for herbarium use (Antonovics et al., 2003), and thus disease proportion among total observations can be a relatively objective measure for comparisons of disease incidence among host taxa. Flower size and color have been evaluated for potentially influencing disease recognition by collectors, but no significant relationship was found between these two traits and disease incidence among host species (Hood et al., 2010). Differing incidence of disease between very similar-appearing species support the neutrality of general collections with regard to anther smut. A striking example is Silene uniflora and S. vulgaris that are so similar as to previously be considered conspecific forms (Marsden-Jones and Turrill, 1957), but the iNaturalist survey showed statistically significant higher and lower disease incidence, respectively, for these two hosts compared to the other Sileneae taxa. Finally, multiple aspects of anther smut were confirmed by the iNaturalist data, supporting this application to understanding the ecology of disease interactions. For instance, the subfamily differences in incidence

TABLE 2. Caryophyllaceae species with anther-smut disease among iNaturalist observations.

Host species	Tribe	Total count	Diseased count	Proportion diseased	Deviation	Binomial distribution probability
Moehringia lateriflora	Ar	463	12	0.02592	high	0.0000
Myosoton aquaticum	Al	452	7	0.01549	high	0.0000
stellaria borealis	Al	34	1	0.02941	high	0.0219
Mononeuria groenlandica	Al	195	1	0.00513	low	0.9926
Dianthus carthusianorum	С	485	24	0.04948	high	0.0000
Saponaria pumila	С	10	3	0.30000	high	0.0001
Dianthus arenarius ⁺	С	39	4	0.10256	high	0.0005
Dianthus superbus	С	175	7	0.04000	high	0.0014
Dianthus pavonius	С	29	3	0.10345	high	0.0025
Dianthus pygmaeus ⁺	С	31	3	0.09677	high	0.0031
Dianthus graniticus ⁺	С	1	1	1.00000	high	0.0094
Dianthus saxicola [†]	С	3	1	0.33333	high	0.0279
Dianthus hyssopifolius ⁺	С	28	2	0.07143	high	0.0284
Saponaria ocymoides	С	253	6	0.02372	high	0.0335
Dianthus acicularis ⁺	С	4	1	0.25000	high	0.0370
Dianthus capitatus†	С	9	1	0.11111	high	0.0814
Dianthus sylvestris	С	113	3	0.02655	high	0.0910
Dianthus repens ⁺	С	11	1	0.09091	high	0.0986
aponaria officinalis	С	2492	17	0.00682	low	0.1062
, Dianthus gallicus [†]	С	13	1	0.07692	high	0.1154
Dianthus monspessulanus	С	63	2	0.03175	high	0.1185
Dianthus sequierit	С	16	1	0.06250	high	0.1401
Dianthus deltoids	С	632	7	0.01108	high	0.3831
Gypsophila repens	С	74	1	0.01351	high	0.5025
Dianthus chinensis [†]	С	201	2	0.00995	low	0.7073
ilene virginica	S	1723	119	0.06907	high	0.0000
/iscaria vulgaris	S	345	14	0.04058	high	0.0000
ilene flos-cuculi	S	1577	2	0.00127	low	0.0000
ilene laciniata	S	1916	4	0.00209	low	0.0001
ilene uniflora	S	462	13	0.02814	high	0.0005
ilene dioica	S	3240	47	0.01451	high	0.0030
ilene latifolia	S	4741	28	0.00591	low	0.0053
ilene acaulis	S	927	17	0.01834	high	0.0079
ilene atropurpurea [†]	S	3	1	0.33333	high	0.0279
Silene italica	S	32	2	0.06250	high	0.0363
ilene vallesia	S	4	1	0.25000	high	0.0370
ilene grayi	S	5	1	0.20000	high	0.0461
ilene saxifraga	S	12	1	0.08333	high	0.1070
ilene parryi	S	41	1	0.02439	high	0.3208
ilene fulgens [†]	S	50	1	0.02000	high	0.3761
ilene caroliniana	S	195	1	0.00513	low	0.4525
ilene baccifera	S	74	1	0.01351	high	0.5025
ilene lemmonii	S	106	1	0.00943	low	0.7375
Silene vulgaris*	S	2727	0	0.00000	low	0.0000

Note: Species are ranked within tribes according to the significance of the deviation of observed diseased observations from expectations based upon subfamily diseased proportions (Table 1). Deviation indicates high or low rates of disease relative to tribe-level proportions. **Silene vulgaris* is also included as an example of statistically assessing disease absence. [†]Indicates host–pathogen combinations not found in prior surveys of herbarium general collections or fungaria data. Tribes are abbreviated as Ar = Arenarieae, AI = Alsinoideae, C = Caryophylloideae, S = Sileneae. Bold binomial distribution probabilities indicate statistical significance at the level of alpha = 0.05 following the Bonferroni corrected threshold of 0.0014.

were consistent with disease incidence based on a review of previous publications (Thrall et al., 1993). There was also a consistent association of the disease with perennial host life histories across data sets as previously reported (Hood et al., 2010).

WITHIN-SPECIES VARIATION IN DISEASE INCIDENCE

Spatial and temporal patterns provide further information about the dynamics of disease as threats to natural populations and could be detected within some species having a larger number of iNaturalist

observations. Within-species patterns included assessment of disease incidence caused by *Microbotryum lychnidis-dioicae* in the native (Europe) and introduced (North America) ranges of *Silene latifolia*. In North America, only one specimen was diseased among 2727 observations, significantly fewer than the 27 of 1923 observations in the native range ($\chi^2 = 35.6$, df = 1, P < 0.001). This difference is consistent with the escape from enemies hypothesis, which says there should be lower disease incidence in the introduced range (Wolfe, 2002).

Two comparisons were performed for the heavily diseased species, *S. virginica*, to assess distributions obtained from the

NEWS & VIEWS Commentary



FIGURE 3. Geographic distribution of anther-smut disease observations. (A) Fungaria (i.e., mycological herbaria), (B) survey of herbarium general collections, and (C) survey of iNaturalist observations.

iNaturalist survey relative to distributions reported in prior studies. The first comparison included geographic distributions of diseased hosts among all host observations of this species (Fig. 4). The current (iNaturalist) distribution is remarkably similar to that obtained from historic collections (Antonovics et al., 2003) (the overall disease incidence between the two data sets was also similar as noted above). The common occurrence of anther smut near the margins of the host distribution was also consistent with the observations of Antonovics et al. (2003). Using the same spatial statistical approach as in that prior study, the iNaturalist data similarly showed a lack of a statistically significant difference in local host densities between observations of diseased hosts versus healthy hosts (mean total observation numbers in 50-km radius from diseased hosts = 40.6 and healthy hosts = 40.2, t-test $F_{_{234}}$ = 1.284, P = 0.258; mean total observation numbers in 100 km radius from diseased hosts = 94.6 and healthy hosts = 89.4; ttest $F_{234} = 0.114$, P = 0.736). Because anther smut has a pollinator-mediated (i.e., frequency-dependent) component to disease transmission, it has been suggested that the disease may persist even at the margins of host ranges where density can be too low

to maintain purely density-dependent transmission (Bruns et al., 2018). The lack of positive association of anther smut in *S. virginica* with regional measures of host density is consistent with findings of prior studies on this and other species (Antonovics et al., 2003; Bueker et al., 2016).

The second comparison was of phenological patterns of diseased versus healthy hosts of S. virginica. The temporal distribution of flowering observations of diseased S. virginica significantly shifted toward the earlier period of the season relative to the distribution of healthy hosts (Fig. 5) (two-sample K-S test, $D_{\text{max}} = 0.328$, P < 0.001). This result is consistent with the earlier flowering of diseased plants documented in field-based studies of other species (Alexander and Antonovics, 1988; Shykoff and Kaltz, 1998; Carlsson and Elmqvist, 1992; Tang et al., 2019) and likely indicates an important role of the flowering timing for pathogen dispersal and epidemiology.

FURTHER UTILITY OF ONLINE NATURAL HISTORY PLATFORMS

Investigation with the anther-smut disease has demonstrated informative and novel ways that online platforms for citizen observations like iNaturalist can add to the resources available in natural history collections, including insights on disease distributions and details of the host-pathogen interactions. Noting that herbaria have faced recent declines

in submissions (Prather et al., 2004; Lavoie and Lachance, 2006; Renner and Rockinger, 2016), here we can see this impact as the general absence of anther-smut observations in the last few decades in data we summarize from both mycological collections and surveys of general plant collections. We show that citizen observations can be valuable for filling a widening gap in our knowledge of recent species distributions and that such online platforms present opportunities for deeper research that quantifies contemporary disease incidence over spatial, temporal, and host-specificity scales.

While the occurrence of anther smut on the Caryophyllaceae provided quantitative insights to several aspects of disease ecology, the approach is not limited to this host–pathogen system (Appendix S2). Anther smut was also detected in the iNaturalist platform for targeted searches of species in other plant families previously known to harbor the fungus. Thus, *Microbotryum* was found infecting *Pinguicula alpina* (Lentibulariaceae), *Calandrinia acaulis*, and *Calandrinia affinis* (Montiaceae) (Kemler et al., 2009; Elvebakk et al., 2015; Ziegler et al., 2018). Based upon similar floral morphology symptoms, anther smut caused by Ustilagomycotina fungi was also detected in several *Scilla* species (fungus *Antherospora*) (Bauer



FIGURE 4. Distribution of anther-smut disease within the geographic range of *Silene virginica*. (A) Survey of general plant collections; image reproduced from Antonovics et al. (2003) (total herbarium sheets = 1022, diseased = 85). Mapped positions for the general plant collection survey are randomly generated locations within the county of the collection's origin. (B) Survey of iNaturalist observations (total observations = 1723, diseased = 119). Black dots indicate observations of anther-smut disease, and gray dots indicate observations of healthy hosts.

et al., 2008) and *Oxalis* species (fungus *Thecaphora*) (Roets et al., 2008).

Extending to other types of plant disease, phyllody symptoms in goldenrod (*Solidago canadensis*) characteristic of phytoplasma infection, fungal anthracnose symptoms on the petals of flowering dogwood (*Cornus florida*), and ergot sclerotia (*Claviceps* fungi) on inflorescences of wild quack grass (*Elymus repens*) can each be found readily in quick surveys of iNaturalist observations of the host plants (Appendix S2). The rates and facility of finding diseased observations will depend upon the particulars of the host–pathogen system, including how conspicuous and unambiguous the symptoms might be. Still, there is strong potential for identifying individual occurrences for a broad range of interactions, especially those that produce observable phenotypes in floral structures. Diseases of leaves or particularly Commentary



FIGURE 5. Phenology of *Silene virginica* from iNaturalist data showing observations with anther-smut disease and healthy observations. The red dash-dotted line represents diseased observations (N = 119), and green solid line represents healthy observations (N = 1659).

fruits also present likely possibilities, such as *Gymnosporangium* rust disease of hawthorns (*Crataegus* sp.) that is easily found among the online postings. In a broader context, observation may be available in sufficient numbers to provide meaningful insights for rare variants in other, non-disease traits (e.g., polymorphism of floral color, or sex ratios in gynodioecious species).

An important benefit of the rise in citizen-driven observations is information on disease in extant populations, whether to be used for comparison with historic data or for directing future field research. While herbarium records have indicated changes in plant distributions within the two centuries of accumulated observations (Antonovics et al., 2003; Meineke et al., 2018a), such cumulative distributions, even partitioned over short time intervals, can sometime not accurately reflect field observations of extant populations (Antonovics et al., 2001; Applequist et al., 2007). Moreover, a rising threat from disease emergence has resulted from increasing rates of climate change and the redistribution of species, by natural and anthropogenic means (Smith et al., 2014; see also Lafferty and Mordecai, 2016). Monitoring the tempo and spatial patterns of disease spread during this period of changes in host geographic ranges would provide critical information to inform conservation and intervention strategies.

We have also found that the ability to quickly contact an observer online is especially helpful in obtaining additional details about disease occurrence or even biological specimens. Particularly for rare or obscure organisms, guidance to extant populations can fundamentally alter the prospect for field studies. Equally as important, this information can be used to target the acquisition of new material for preservation in herbarium collections. The usefulness of herbarium specimens from many years ago for DNA isolation and analysis (e.g., Malmstrom et al., 2007; Andrew et al., 2018; Heberling and Burke, 2019) means that locating extant populations also makes it possible to compare past and current genetic structure. Heberling and Isaac (2018) suggested using iNaturalist as a direct supplement to new herbarium specimens by linking the online observations to the herbarium sample. The reverse also holds potential, connecting new observations to bioinformatics resources on preserved specimens. Readily accessible means for annotating specimens and accessing an assembly of observations as online resources for particular research projects hold great potential to advance studies in natural plant populations.

Finally, the disease surveys from general collections can identify host species deserving of further targeted study, even where rates of infection may not have risen to statistical significance here due to small sample sizes. For example, our field research in the maritime Alps (e.g., Bruns et al., 2017; Petit et al., 2017) was prompted by the observation of several diseased Silene campanula specimens among relatively few herbarium sheets at the Muséum National d'Histoire Naturelle (P). Similarly, Dianthus pavonius had 3 among 29 total iNaturalist observations showing anther smut symptoms, which is suggestive of a heavy disease impact. Confirmation of this suggestion was provided by a recent study by Bruns et al. (2018), which included numerous transects in natural populations of D. pavonius where the product of disease incidence among populations (0.46) and average infection prevalence within diseased populations (0.18) are entirely consistent with the high disease rate among iNaturalist observations.

This study shows that natural history observations currently being generated in high volumes for online platforms can yield valuable insights into the ecological interactions of wild plant species. Where observers are unaware that symptoms are also being recorded, quantitative assessments of disease incidence can also be achieved. This approach is similar to surveys of museum collections. Yet, while herbaria provide essential information on long-term distributions and direct access to specimens for trait measurements, the iNaturalist observations help fill a widening gap in very recent distributions at a time when deposition of new herbarium materials has declined. As citizen-driven plant observations accumulate, there is great potential to discover new and diverse host-pathogen associations and to direct field studies to extant populations. Therefore, the impact of online observations intended to record plant occurrence can be enhanced by mining for disease interactions that are important drivers of plant ecology and evolution.

ACKNOWLEDGMENTS

We thank the U.S. National Institutes of Health for support (award NIGMS-R01GM122061). Thanks is also given to Dr. Michael Nee, Matt Rung, and Judy Gallagher for the use of herbarium and diseased and healthy iNaturalist images, respectively, and to Dr. Janis Antonovics for helpful comments on an earlier version of the manuscript. We also thank the editors and anonymous reviewers, whose comments greatly helped to improve this work.

AUTHOR CONTRIBUTIONS

M.H. originated and designed the project. A.K. and M.H. contributed to data collections and analysis, and both contributed to drafting and revising the manuscript.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

APPENDIX S1. Data on hosts observed with anther smut (*Microbotryum* fungi) on plants in the Caryophyllaceae.

APPENDIX S2. Example iNaturalist observations showing symptoms for a range of disease and host types.

LITERATURE CITED

- Ainsworth, G. C. 1976. Introduction to the history of mycology. Cambridge University Press, Cambridge, UK.
- Alexander, H. M., and J. Antonovics. 1988. Disease spread and population dynamics of anther-smut infection of *Silene alba* caused by the fungus *Ustilago violacea. Journal of Ecology* 76: 91–104.
- Alexander, H. M., and A. Maltby. 1990. Anther-smut infection of Silene alba caused by Ustilago violacea: factors determining fungal reproduction. Oecologia 84: 249–253.
- Alexander, H. M., P. H. Thrall, J. Antonovics, A. M. Jarosz, and P. V. Oudemans. 1996. Population dynamics and genetics of plant disease: a case study of anther-smut disease. *Ecology* 77: 990–996.
- Alexander, H. M., S. Price, R. Houser, D. Finch, and M. Tourtellot. 2007. Is there reduction in disease and pre-dispersal seed predation at the border of a host plant's range? Field and herbarium studies of *Carex blanda*. *Journal of Ecology* 95: 446–457.
- Andrew, C., J. Diez, T. Y. James, and H. Kauserud. 2018. Fungarium specimens: a largely untapped source in global change biology and beyond. *Philosophical Transactions of the Royal Society, B, Biological Sciences* 374: 20170392.
- Antonovics, J., and M. E. Hood. 2018. Linnaeus, smut disease, and living contagion. Archives of Natural History 45: 213–232.
- Antonovics, J., T. J. Newman, and B. J. Best. 2001. Spatially explicit studies on the ecology and genetics of population margins. *In J. Silvertown and J.* Antonovics [eds.], Integrating ecology and evolution in a spatial context, 97–116. Blackwell Science, Oxford, UK.
- Antonovics, J., M. E. Hood, P. H. Thrall, J. Y. Abrams, and G. M. Duthie. 2003. Herbarium studies on the distribution of anther-smut fungus (*Microbotryum violaceum*) and *Silene* species (Caryophyllaceae) in the eastern United States. *American Journal of Botany* 90: 1522–1531.
- Applequist, W. L., D. J. McGlinn, M. Miller, Q. G. Long, and J. S. Miller. 2007. How well do herbarium data predict the location of present populations? A test using *Echinacea* species in Missouri. *Biodiversity and Conservation* 16: 1397–1407.
- Bauer, R., M. Lutz, D. Begerow, M. Piatek, K. Vánky, K. Bacigálová, and F. Oberwinkler. 2008. Anther smut fungi on monocots. *Micological Research* 112: 1297–1306.
- Beauvais, M. P., S. Pellerin, J. Dubé, and C. Lavoie. 2017. Herbarium specimens as tools to assess the impact of large herbivores on plant species. *Botany* 95: 153–162.
- Brock, P. M., H. Döring, and M. I. Bidartondo. 2009. How to know unknown fungi: the role of a herbarium. *New Phytologist* 181: 719–724.
- Brown, N., F. van den Bosch, S. Parnell, and S. Denman. 2017. Integrating regulatory surveys and citizen science to map outbreaks of forest diseases: acute oak decline in England and Wales. *Proceedings of the Royal Society, B, Biological Sciences* 284: 20170547.
- Bruns, E., J. Antonovics, V. Carasso, and M. E. Hood. 2017. Transmission and temporal dynamics of anther-smut disease (*Microbotryum*) on alpine carnation (*Dianthus pavonius*). *Journal of Ecology* 105: 1413–1424.
- Bruns, E., J. Antonovics, and M. E. Hood. 2018. Is there a disease-free halo at species range limits? The co-distribution of anther-smut disease and its host species. *Journal of Ecology* 107: 1–11.
- Bucheli, E., and J. A. Shykoff. 1999. The influence of plant spacing on densitydependent versus frequency-dependent spore transmission of the anther smut *Microbotryum violaceum*. *Oecologia* 119: 55–62.

- Bueker, B., C. Eberlein, P. Gladieuz, A. Schaefer, A. Snirc, D. J. Bennett, D. Begerow, et al. 2016. Distribution and population structure of the anther smut *Microbotryum silenes-acaulis* parasitizing an arctic-alpine plant. *Molecular Ecology* 25: 811–824.
- Burdon, J. J., and G. A. Chilvers. 1982. Host density as a factor in plant disease ecology. Annual Review of Phytopathology 10: 143–166.
- Burdon, J. J., and P. H. Thrall. 2008. Pathogen evolution across agro-ecological interface: implications for disease management. *Evolutionary Applications* 1: 57–65.
- Carlsson, U., and T. Elmqvist.1992. Epidemiology of anther-smut disease (*microbotryum violaceum*) and numeric regulation of populations of *silene dioica*. Oecologia 90: 509–517.
- Conners, I. L. 1933. Carnation (*Dianthus*). Canadian Plant Disease Survey 13: 65.
- Dobson, A., and M. Crawley. 1994. Pathogens and the structure of plant communities. Trends in Ecology & Evolution 9: 393–398.
- Eastburn, D. M., A. J. McElrone, and D. D. Bilgin. 2011. Influence of atmospheric and climatic change on plant–pathogen interactions. *Plant Pathology* 60: 54–69.
- Elvebakk, A., A. R. Flores, and J. M. Watson. 2015. Revisions in the South American Calandrinia caespitosa complex (Montiaceae). Phytotaxa 203: 1–23.
- Ghelardini, L., A. L. Pepori, N. Luchi, P. Capretti, and A. Santini. 2016. Drivers of emerging fungal disease of forest trees. *Forest Ecology and Management* 281: 235–246.
- Hanan-A, A. M., H. Vibrans, N. I. Cacho, J. L. Villaseñor, E. Ortiz, and V. A. Gómez-G. 2016. Use of herbarium data to evaluate weediness in five congeners. AOB Plants 8: plv144.
- Heberling, J. M., and D. J. Burke. 2019. Utilizing herbarium specimens to quantify historical mycorrhizal communities. *Plant Journal* 7: e01223.
- Heberling, J. M., and B. L. Isaac. 2017. Herbarium specimens as exaptations: new uses for old collections. *American Journal of Botany* 104: 963–965.
- Heberling, J. M., and B. L. Isaac. 2018. iNaturalist as a tool to expand research value of museum specimens. *Applications in Plant Sciences* 6: e1193.
- Hood, M. E., and J. Antonovics. 2003. Plant species descriptions show signs of disease. *Proceedings of the Royal Society, B, Biological Sciences* 270: S156–S158.
- Hood, M. E., J. I. Mena-Alí, A. K. Gibson, B. Oixelman, T. Giraud, R. Yockteng, M. T. K. Arroyo, et al. 2010. Distribution of the anther-smut pathogen *Microbotryum* on species of the Caryophyllaceae. *New Phytologist* 187: 217–229.
- Kashefi, J., and K. Vánky. 2004. Microbotryum scolymi, a rare smut fungus new for Greece. Journal of Plant Pathology 86: 157–159.
- Kemler, M., M. Lutz, M. Göker, F. Oberwinkler, and D. Begerow. 2009. Hidden diversity in the non-caryophyllaceous plant-parasite members of *Microbotryum* (Picciniomycotina: Microbotryales). *Systematics and Biodiversity* 7: 297–306.
- Kniep, H. 1919. Untersuchungen uber den Antherenbrand (Ustilago violacea Pers.), Ein Beitrag zum Sexualitatsproblem. Zeitschrift f
 ür Botanik 11: 257–284.
- Lafferty, K. D., and E. A. Mordecai. 2016. The rise and fall of infectious disease in a warmer world. *F1000Research* 5: 2040.
- Lang, P. L., F. M. Willems, J. F. Scheepens, H. A. Burbano, and O. Bossdorf. 2019. Using herbaria to study global environmental change. *New Phytologist* 221: 110–122.
- Lavoie, C., and D. Lachance. 2006. A new herbarium-based method for reconstructing the phenology of plant species across large areas. *American Journal of Botany* 93: 512–516.
- Li, W., R. H. Brlansky, and J. S. Hartung. 2006. Amplification of DNA of Xanthomonas axonopodis pv. citri from historic citrus canker herbarium specimens. Journal of Microbiological Methods 65: 237–246.
- Lui, T., H. Tian, S. He, and L. Guo. 2009. Microbotryum scorzonerae (Microbotryaceae), new to China, on a new host plant. Mycotaxon 108: 245–247.

- Malmstrom, C. M., R. Shu, E. W. Linton, L. A. Newton, and M. A. Cook. 2007. Barley yellow dwarf viruses (BYDVs) preserved in herbarium specimens illuminate historical disease ecology of invasive and native grasses. *Journal of Ecology* 95: 1153–1166.
- Marsden-Jones, E. M., and W. B. Turrill. 1957. The bladder campions (Silene maritima and S. vulgaris). Ray Society, London, UK.
- Meentemeyer, R. K., M. A. Dorning, J. B. Vogler, D. Schmidt, and M. Garbelotto. 2015. Citizen science helps predict risk of emerging infectious disease. *Frontiers in Ecology and the Environment* 13: 189–194.
- Meineke, E. K., C. C. Davis, and T. J. Davies. 2018a. The unrealized potential of herbaria for global change biology. *Ecological Monographs* 88: 505–525.
- Meineke, E. K., T. J. Davies, B. H. Daru, and C.C. Davis. 2018b. Biological collections for understanding biodiversity in the Anthropocene. *Philosophical Transactions of the Royal Society, B, Biological Sciences* 374: 20170386.
- Petit, E., C. Silver, A. Cornille, P. Gladieux, L. Rosenthal, E. Bruns, S. Yee, et al. 2017. Co-occurrence and hybridization of anther-smut pathogens specialized on *Dianthus* hosts. *Molecular Ecology* 26: 1877–1890.
- Piątek, M., W. Mułenko, J. Piątek, and K. Bacigálová. 2005. Taxonomy and distribution of *Microbotryum pinguiculae*, a species of smut fungi new for the Carpathians. *Polish Botanical Journal* 50: 153–158.
- Prather, L. A., O. Alvarez-Fuentes, M. H. Mayfield, and C. J. Ferguson. 2004. Implications of the decline in plant collecting for systematic and floristic research. *Systematic Botany* 29: 216–220.
- Rabeler, R. K., and R. L. Hartman. 2005. Caryophyllaceae. *In* Flora of North America Editorial Committee [ed.],Flora of North America north of Mexico, 3–215. Oxford University Press, NY, NY, USA.
- Renner, S. S., and A. Rockinger. 2016. Is plant collecting in Germany coming to an end? *Willdenowia* 46: 93–97.
- Ristaino, J. B., C. T. Groves, and G. R. Parra. 2001. PCR amplification of the Irish potato famine pathogen from historic specimens. *Nature* 411: 695–697.
- Roets, F., L. L. Dreyer, M. J. Wingfield, and D. Begerow. 2008. *Thecaphora capensis* sp. nov., an unusual new anther smut on *Oxalis* in South Africa. *Persoonia* 21: 147–152.
- Romberg, M., and Y. Rivera. 2017. Voucher specimens? Yes please! NPDN News USDA APHIS 12: 1–2.
- Saville, A. C., M. D. Martin, and J. B. Ristaino. 2016. Historic late blight outbreaks caused by a widespread dominant lineage of *Phytophthora infestans* (Mont.) de Bary. *PLoS ONE* 11: e0168381.
- Schäfer, A. M., M. Kemler, R. Bauer, and D. Begerow. 2010. The illustrated life cycle of *Microbotryum* on the host plant *Silene latifolia*. *Botany* 88: 875–885.
- Shykoff, J. A., and O. Kaltz. 1998. Phenotypic changes in host plants diseased by Microbotryum violaceum: parasite manipulation, side effects, and trade-offs. International Journal of Plant Sciences 159: 236–243.
- Smith, K. F., M. Goldberg, S. Rosenthal, L. Carlson, J. Chen, C. Chen, and S. Ramachandran. 2014. Global rise in human infectious disease outbreaks. *Journal of The Royal Society Interface* 11: 20140950.
- Spencer, J. L., and H. E. White. 1951. Anther smut of carnation. *Phytopathology* 41: 291–299.
- Tang, H., M. E. Hood, Z.-X. Ren, D.-Z. Li, L. M. Wolfe, and H. Wang. 2019. Specificity and seasonal prevalence of anther-smut disease *Microbotryum* on sympatric Himalayan *Silene* species. *Journal of Evolutionary Biology* 32: 451–462.
- Thrall, P. H., A. Biere, and J. Antonovics. 1993. Plant life-history and disease susceptibility—the occurrence of *Ustilago violacea* on different species within the Caryophyllaceae. *Journal of Ecology* 81: 489–498.
- Wolfe, L. M. 2002. Why alien invaders succeed: support for the escape-fromenemy hypothesis. *American Naturalist* 160: 705–711.
- Yoshida, K., H. A. Burbano, J. Krause, M. Thines, D. Weigel, and S. Kamoun. 2014. Mining herbaria for plant pathogen genomes: back to the future. *PLOS Pathogens* 10: e1004028.
- Ziegler, R., M. Lutz, J. Piątek, and M. Piątek. 2018. Dismantling a complex of anther smuts (*Microbotryum*) on carniverous plants in the genus *Pinguicula*. *Mycologia* 110: 361–374.