Check for updates

OPEN ACCESS

EDITED BY Peter Hart, Roosevelt University College of Pharmacy, United States

REVIEWED BY Lin Wu, Central South University, China Nan Ma, First Affiliated Hospital, Guangxi Medical University, China

*CORRESPONDENCE Caicun Zhou, drcaicunzhou@163.com Chunyu Li, lichunyu@tmu.edu.cn

^tThese authors have contributed equally to this work

SPECIALTY SECTION

This article was submitted to Cancer Genetics and Oncogenomics, a section of the journal Frontiers in Genetics

RECEIVED 25 July 2022 ACCEPTED 18 October 2022 PUBLISHED 28 October 2022

CITATION

Tang Z, Wang Q, Chen P, Guo H, Shi J, Pan Y, Li C and Zhou C (2022), Computational recognition of LncRNA signatures in tumor-associated neutrophils could have implications for immunotherapy and prognostic outcome of non-small cell lung cancer. *Front. Genet.* 13:1002699. doi: 10.3389/fgene.2022.1002699

COPYRIGHT

© 2022 Tang, Wang, Chen, Guo, Shi, Pan, Li and Zhou. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms. Computational recognition of LncRNA signatures in tumor-associated neutrophils could have implications for immunotherapy and prognostic outcome of non-small cell lung cancer

Zhuoran Tang^{1†}, Qi Wang^{2†}, Peixin Chen¹, Haoyue Guo¹, Jinpeng Shi¹, Yingying Pan¹, Chunyu Li³* and Caicun Zhou²*

¹Tongji University Medical School Cancer Institute, Tongji University, Shanghai, China, ²Department of Medical Oncology, Tongji University Affiliated Shanghai Pulmonary Hospital, Tongji University Medical School Cancer Institute, Tongji University, Shanghai, China, ³Department of Integrated Traditional Chinese and Western Medicine, International Medical School, Tianjin Medical University, Tianjin, China

Cancer immune function and tumor microenvironment are governed by long noncoding RNAs (IncRNAs). Nevertheless, it has yet to be established whether IncRNAs play a role in tumor-associated neutrophils (TANs). Here, a computing framework based on machine learning was used to identify neutrophil-specific IncRNA with prognostic significance in squamous cell carcinoma and lung adenocarcinoma using univariate Cox regression to comprehensively analyze immune, IncRNA, and clinical characteristics. The risk score was determined using LASSO Cox regression analysis. Meanwhile, we named this risk score as "TANIncSig." TANIncSig was able to distinguish between better and worse survival outcomes in various patient datasets independently of other clinical variables. Functional assessment of TANIncSig showed it is a marker of myeloid cell infiltration into tumor infiltration and myeloid cells directly or indirectly inhibit the anti-tumor immune response by secreting cytokines, expressing immunosuppressive receptors, and altering metabolic processes. Our findings highlighted the value of TANIncSig in TME as a marker of immune cell infiltration and showed the values of lncRNAs as indicators of immunotherapy.

KEYWORDS

non-small cell lung cancer, tumor-associated neutrophils, long noncoding RNA, immunotherapy, computational recognition

Introduction

Lung cancer is related with high mortality rates in China with non-small cell lung cancer (NSCLC) accounting for >80% of lung cancers (Zhu et al., 2017). The administration of immune checkpoint inhibitors (ICIs) in cancer therapy has had remarkable results (Yue et al., 2018; Dolladille et al., 2020; Galluzzi et al., 2020). For advanced non-small cell lung cancer (NSCLC), several clinical trials have confirmed that as first- or second-line treatment, ICIs are superior to platinum-based chemotherapy (Ko et al., 2018; Vansteenkiste et al., 2019; Chen et al., 2020). However, only 20%-40% of advanced NSCLC patients achieve sustained clinical benefits from PD-(L)1 inhibitor therapy, with most patients having primary or acquired resistance to immunotherapy (Socinski, 2014). Moreover, those who do not respond to immunotherapy may suffer immune-related adverse events (IRAE) and the high costs of anti-PD-(L)1 monoclonal antibody therapy (Khoja et al., 2017; Das and Johnson, 2019; Schoenfeld et al., 2019). Thus, effective biomarkers that distinguish potential responders from non-responders, and indicate patient clinical response in real-time are urgently needed to improve treatment outcomes.

The TME is comprised of a complex cell population that includes tissue-resident lymphocytes, fibroblasts, endothelial cells, and neurons that are present before tumorigenesis, as well as bloodderived cells recruited to tumor sites (Butturini et al., 2019). Immune cells are the main cellular components in tumors. Tumor-infiltrating myeloid cells, including tumor-associated macrophages (TAM), regulatory dendritic cells, tumor-associated neutrophils (TAN), myeloid-derived suppressor cells (MDSC), as well as tolerogenic dendritic cells (TOL-DC), facilitate the formation of immunosuppressive microenvironments (Schupp et al., 2019). These cells directly or indirectly inhibit the antitumor immune response by secreting cytokines, expressing immunosuppressive receptors, and altering metabolic processes, leading to tumor immune escape. Tumor-associated neutrophils (TANs) are a key part of tumor-infiltrating myeloid cells and are regularly detected in the TME. Clinically, TANs can be used to predict treatment outcomes and immunotherapy response (Nielsen et al., 2021). Transcriptomic studies have identified gene expression biomarkers as well as signatures for quantitative assessment of TANs, as well as for stratification based on prognoses and immunotherapeutic response (Lecot et al., 2019; Wu and Zhang, 2020).

Long non-coding RNA (lncRNAs) influence almost all biological processes and pathways, and their dysregulation is associated with various diseases. Additionally, lncRNAs have wide functional diversity due to their influence on gene expression levels at transcriptional, post-transcriptional and epigenetic levels (Rinn and Chang, 2012; Fatica and Bozzoni, 2014; Marchese et al., 2017; Bao et al., 2020). The correlation between lncRNAs and immune function has been reported. Recent studies have shown that lncRNAs are abundant with cell type specificity in various immune cell subsets (Rinn and Chang, 2012; Atianand et al., 2017; Chen et al., 2017; Zhou et al., 2017; Zhou et al., 2017; Zhou et al., 2018). LncRNAs expression pattern has been correlated with infiltrations of immune cells into the TME (Hu et al., 2013; Ranzani et al., 2015; Sage et al., 2018; Wang et al., 2018; Zhao et al., 2021). Nevertheless, neutrophil-specific lncRNAs as well as their significance in assessing TANs and prediction of clinical outcomes and immunotherapeutic responses require further study.

Here, a computational framework is proposed for determining neutrophil-specific lncRNA expression levels and lncRNA signatures for TANs (TANLncSig) *via* integrative immune, lncRNA, and clinical profiling analyses. The TANLncSig's ability to predict clinical outcome and response to immunotherapy by NSCLC patients was also investigated.

Materials and methods

Neutrophil-specific long noncoding RNAs screening

The data set can be obtained from the GEO database with series accession number GSE28490 (https://www.ncbi.nlm.nih. gov/geo/query/acc.cgi?acc=GSE28490), These included chip data on the expression of nine human immune cells (neutrophils, monocytes, B cells, eosinophils, CD4 T cells, NK cells, mDCs, CD8 T cells, and pDCs). The GEO2R tool from GEO was used for differential expression analysis. Using adjusted p = <0.05 and logFC >1 as cutoff thresholds identified 17 lncRNAs with high neutrophil-specific expression.

Construction of risk scoring model

Clinical data and TCGA RNA-seq datasets for LUSC and LUAD were downloaded by the UCSC Xena browser (https:// xenabrowser.net/). Lusc-LINC01272-neutrophils malignant/ Luad-LINC01272-neutrophils malignant results from single cell sequencing datasets. First, a monovariate Cox regression analysis was used to find neutrophil-specific lncRNAs with prognostic value in LUSC and LUAD, and LASSO Cox regression was used to determine their risk scores. The multivariate Cox regression analysis (age, risk score, tumor stage, gender), Kaplan-Meier (KM) survival analysis and 3, 5, and 10 years survival AUCs were used to evaluate risk score.

Correlation analysis between risk score and tumor clinical phenotype

Multivariate ANOVA was used to analyze differences between neutrophil-specific, highly expressed lncRNA and risk score in LUSC and LUAD samples at various TNM stages.



Analysis of risk score related pathways

In LUSC and LUAD samples, genes with mean expression levels >1 were identified and their correlation with risk score analyzed. 1,000 genes with the highest absolute correlation coefficient value were selected from those with positive correlation coefficients (>0, p = <0.05) and those with negative correlation coefficients (<0, p = <0.05). ClusterProfiler for R was used to analyze GO terms of biological process (BP), Molecular function (MF), cellular component (CC), and KEGG pathway enrichment analyses. After gene enrichment, the adjusted *p*-value < 0.05 and the smallest TOP10 was selected for mapping.

Development of tumor-associated neutrophils-derived long noncoding RNAs signature to judge the prognosis of immunotherapy for non-small cell lung cancer using machine learning

Pearson correlation analysis was used to determine correlations between risk score, neutropen-specific lncRNAs, and the expression of common immune checkpoint inhibitors and correlation heat maps drawn, with * denoting $p \le 0.01$ while + denotes $p \le 0.05$.

Results

Prognostic significance of neutrophilspecific long noncoding RNAs

To recognize neutrophil-specific lncRNAs, dataset GSE28490 was downloaded from GEO (https://www.ncbi.nlm. nih.gov/geo/query/acc.cgi?acc=GSE28490). This dataset includes

chip data on expressions of nine human immune cells (CD4⁺ T cells, neutrophils, monocytes, B cells, eosinophils, CD8⁺ T cells, NK cells, mDCs, and pDCs). Using GEO2R, 17 lncRNA specifically highly-expressed in neutrophils (p = <0.05, log2>1) were identified. These neutrophil-specific lncRNAs are referred to as TAN-associated lncRNAs (TANlncRNA) (Figure 1).

Construction of a risk score based on neutrophil-specific long noncoding RNAs for prognosis prediction

To develop a neutrophil-specific lncRNA risk score for predicting prognosis, the TCGA NA-SEQ dataset, TCGA lung squamous cell carcinoma (LUSC) as well as adenocarcinoma (LUAD) gene expression data, clinical features, and prognosis data were downloaded from UCSC Xena. First, univariate Cox regression analyses were used to establish neutrophil-specific lncRNAs with prognostic value in LUSC and LUAD. The final signature named TANIncSig (Table 1). This analysis identified three lncRNAs with prognostic value in LUSC (LINC01272, LINC00261, LINC00668, $p = \langle 0.05 \rangle$. Using these three lncRNAs, the expression value of lncRNA was weighted using multivariate Cox regression coefficient to obtain risk scores via the formula: risk score = 0.09 * LNC00668 + 0.17 * LNC00261. Then, TANIncSig scores for every patient in the discovery dataset were determined, after which the 542 patients were grouped into the high (n = 271) or low (n = 271) risk groups. Low risk group patients were found to have longer overall survival (OS) relative to the high-risk group patients (p = 0.039, ≤ 0.05 , Figure 2A). Multivariate Cox regression analyses revealed that risk score (p <0.001), stage (*p* < 0.001), age (*p* = 0.037, ≤0.05), and gender (*p* = 0.007, \leq 0.01) significantly affected the prognostic outcomes of LUSC patients. The *p*-value and hazard ratio of TANIncSig were better than those of stage and age (Figure 2B). That said, TANIncSig has the potential to be a good predictor of

	Ensembl ID	Gene symbol	Location (GRCh37/hg19)	HR	Lower 0.95	Upper 0.95	<i>p</i> -value
LUAD	ENSG00000259974	LINC00261	chr20:22,541,191–22,559,280	0.8726407	0.7771	0.98	0.021359
	ENSG00000269220	LINC00528	chr22:18,260,056–18,262,247	0.5049413	0.2662	0.9577	0.036422
	ENSG00000253138	LINC00967	chr8:67,104,349–67,109,554	0.0026941	9.72E-06	0.7471	0.039252
LUSC	ENSG00000259974	LINC00261	chr20:22,541,191–22,559,280	1.2676382	1.1	1.461	0.001087
	ENSG00000265933	LINC00668	chr18:6,925,473–6,929,868	0.8529687	0.7512	0.9685	0.01412
	ENSG00000224397	LINC01272	chr20:48,884,015–48,896,333	1.14005	1.022	1.272	0.018818





squamous cell carcinoma patients. (B) Multivariate Cox regression analyses of patients with LUSC based on TCGA dataset. (C) ROC curve analyses of patients with LUSC using the TCGA dataset.

efficacy. The predictive capacity of TANIncSig was authenticated using the TCGA internal testing dataset and revealed the 3-, 5-, and 10-year OS rates for low-risk group patients to be 60.42, 54.47, and 54.23%, respectively (Figure 2C). Indicating that risk score significantly correlates with OS in LUSC. Similar analysis was done for LUAD. First, three lncRNAs with prognostic values (LINC00528, LINC00967, and LINC00261) were identified using univariate Cox analysis. Using the above three lncRNAs, lncRNAs expression value was weighted by multivariate Cox



regression coefficient to determine risk score using the formula: risk score = -5.32 * LINC00967-0.16 * LINC00261-0.74 * LINC00528. Patients with LUAD in the low-risk group had longer OS relative to high-risk group LUAD patients (p = 0.0029, ≤ 0.01 , Figure 3A). Cox multivariate regression analyses revealed that risk score (p < 0.001) and stage (p < 0.001) significantly correlated with LUAD prognosis. In lung adenocarcinoma, the p-value and hazard ratio of TANIncSig were equally better than those of stage and age (Figure 3B). The 3-, 5-, and 10-year OS rates in low-risk group patients were 61.01, 61.20, and 65.30%, respectively (Figure 3C). These results indicate that risk scores in the LUAD dataset significantly correlate with patients' OS.

Correlation analysis between risk score and tumor clinical phenotype

Clinical phenotypic correlation analysis of single prognostic lncRNA and risk score (tumor stage, T, N, and M staging) was performed in lung adenocarcinoma as well as squamous cell carcinoma. According to statistical analysis, the risk score in different tumor stages of lung squamous cell carcinoma showed significant statistical differences, and the statistical results showed that p = 0.0013, <0.01 (Figure 4A). The risk score in different tumor stages of lung adenocarcinoma also showed significant statistical differences (p = 0.0081, <0.01) (Figure 4B).

The TNM staging system is the most widely used tumor staging system, worldwide. T denotes tumor sizes and local invasion range,





N denotes lymph node involvement, and M denotes distant metastasis. TNM staging has great clinical value in prognosis prediction (Ficarra et al., 2007; Moch et al., 2009). The risk score lack of significance in different T stages and N stages of lung squamous cell carcinoma (Figures 5A,B). The risk score has significant statistical difference in different M stages of lung squamous cell carcinoma (p = 0.011, <0.05) (Figure 5C). The

risk score has significant statistical difference in different T stages (T1, T2, T3, and T4 stages) of lung adenocarcinoma (p = 0.0023, <0.01) (Figure 5D). Similarly, the risk score has significant statistical difference in different N stages (N0, N1, N2, and N3 stages) of lung adenocarcinoma (p = 0.013, <0.05) (Figure 5E). The risk score lack of significance in different M stages of lung adenocarcinoma (Figure 5F).



(B) Enrichment of negative correlation gene pathways in lung squamous cell carcinoma. (C) Enrichment of positively correlation gene pathways in lung adenocarcinoma.

Riskscore correlation pathway analysis

In LUSC and LUAD samples, genes with average expression levels >1 were identified and their risk scores analyzed. 1,000 genes with the largest absolute correlation coefficient values were selected from positive (correlation coefficient >0, $p \le 0.05$) and negative (correlation coefficient <0, $p \le 0.05$) and correlation genes and pathway enrichment analysis done using cluster profiler on R. In LUSC, positive correlation genes are mainly associated with biological processes (BP) associated with T-cell activation, leukocyte proliferation, and leukocyte cell-cell adhesion. For cellular component (CC) they were enriched in endocytic vesicle, tertiary granule, and secretory granule membrane. For molecular function (MF), they were enriched in immune receptor activity and cytokine binding. KEGG pathway analysis revealed enrichment mainly for cell adhesion molecules cams (Figure 6A). Negative correlation genes in lung squamous cell carcinoma are mainly enriched for biological processes (BP) associated with skin development, epidermis development, and cornification. For cellular component (CC),

they were enriched for cornified envelope, desmosome, and cellcell junction. For molecular function (MF), they were enriched for microtubule binding and tubulin binding. For KEGG pathways, they were enriched for basal cell carcinoma (Figure 6B). Positive correlation genes in lung adenocarcinoma were mainly enriched in biological processes (BP) associated with translational termination and adenocarcinoma. For cellular component (CC), they were enriched for ribosomal subunits, ribosome and large ribosomal subunit. For molecular function (MF) they were enriched for structural constituent of ribosome and cadherin binding. For KEGG pathways, they were enriched for ribosome and cell cycle (Figure 6C). Genes associated with negative correlations in LUAD are involved in biological processes (BPs) associated with lymphocyte differentiation, leukocyte proliferation, and antigen receptor-mediated signaling. For cellular component (CC), they were enriched for external side of plasma membrane and immunological synapse. For molecular functions (MFs), they were enriched for guanyl-nucleotide exchange factor activity. For KEGG pathways, they were



enriched for primary immunodeficiency and B-cell receptor signaling pathway (Figure 6D).

The TANIncSig associates with tumorassociated neutrophils

In accordance with previously reported expression levels of the immune cell specific marker genes, cibersort (https:// cibersort.stanford.edu/) was further used to evaluate the levels of immune infiltration of 22 immune subpopulations in high-risk and low-risk patient groups. *t*-test was performed to determine the difference in lymphocyte infiltration levels between the two groups. As shown in Figures 7A,B, in both LUSC and LUAD, high-risk patients were significantly enriched in 12 immune subpopulations, while low-risk patients were enriched in 10 immune subpopulations. Additionally, mononuclear immune cells, including neutrophils, were found to infiltrate significantly more in the high-risk patient group than in several other groups. Single-cell sequencing data of LUSC and LUAD downloaded from GSE127465, cell type notes downloaded from TISCH (http://tisch.comp-genomics.org/). The homologous expression levels of LINC01272 of the TANIncSig in neutrophil cell lines differed significantly from those of malignant cell lines (Figures 7C,D). This indicates that these



The TANIncSig was independently validated in the GSE30219 dataset (A). Kaplan–Meier survival curves of OS were plotted between high- and low-risk groups stratified by the TANIncSig. (B) Time ROC curve of luad patients the GSE30219 dataset. (C) Visualization of the HRs from a multivariate Cox analysis of the TANIncSig and clinicopathological factors in GSE30219.

IncRNAs are expressed differently in neutrophils compared with malignant cells. In the above study, we found that the TANIncSig was not only associated with patient prognosis but also as a TAN indicator.

TANIncSig was validated over several independent datasets using a microarray platform for prognostic value

TANIncSig was further validated in independent datasets by the microarray platform in order to verify versatility and robustness of TANIncSig. The Affymetrix HG-U133 Plus 2.0 platform was used to

analyze 83 LUAD patients from the GSE30219 dataset. As demonstrated again, TANIncSig can distinguish between patients who have high and low survival risk. A total of 83 patients were stratified into 41 high-risk patients and 42 low-risk patients in the GSE30219 dataset. Furthermore, patients in the high-risk group had a marginally poorer outcome than those in the low-risk group (p = 0.0024, ≤ 0.01 ; log-rank test) (Figure 8A). The AUC of ROC curve at 3, 5, and 10 years were 64.13, 66.87, and 60.58% respectively (Figure 8B). The results show that TANIncsig can accurately predict the 5-year overall survival of patients, indicating that TANIncsig has good efficacy and certain stability. In order to investigate whether TANIncSig is an independent prognostic factor, a multivariate Cox regression analysis was conducted in



patient cohorts. In the independent GSE30219 dataset, the TANIncSig still maintained a significant correlation with OS in the multivariate analysis (HR = 6.74, 95% CI 1.283-35.5, *p* = 0.024, \leq 0.01). Thus, these results demonstrate that the TANIncSig helps predict OS independently of other conventional clinical factors (Figure 8C).

Significance of TANIncSig as a marker of immunotherapy

Next, prognostic lncRNAs and risk score were correlated with immune checkpoint molecules expression in LUSC and LUAD patients. In LUSC, risk score, LINC01272, and LINC00261 positively correlated with the expression of most ICBs, while LINC00668 had negative correlations with the expression of most ICBs (Figure 9A). In LUAD, risk score had negative correlations with expression levels of most ICBs, while LINC00528 positively correlated with expression levels of most ICBs (Figure 9B). The expressions of risk score were divided into high and low groups and combined according to the median. The combination was used to analyze the prognosis of immunotherapy for non-small cell lung cancer. In lung squamous cell carcinoma, the combination of CEACAM1, TNFSF4, gem, CD47, vtcn1 and risk score can well stratify the prognosis of patients. In lung adenocarcinoma, all ICB molecules combined with risk score can well predict the prognosis of patients. These results suggest that risk score can be used as an index to predict the response of patients to immunotherapy.

Discussion

In the peripheral blood, neutrophils are the most abundant white blood cells (Dinh et al., 2020). They have a central role in human non-specific immunity. Previous studies suggest that neutrophils inhibit tumors by secreting cytokines and producing reactive oxygen species (Vaughan and Walsh, 2005; Mishalian et al., 2013; Coffelt et al., 2015; Ponzetta et al., 2019). However, other studies indicate that neutrophils in the tumor microenvironment (TME) promote tumorigenesis. Cytokines and chemokines production by invasive neutrophils might affect the recruitment and activation of inflammatory cells in the TME, create an immunosuppressive microenvironment that is conducive for tumorigenesis, regulate tumor growth, metastasis and angiogenesis, and influence patient prognosis. Traditional methods for quantifying tumor immune cells infiltration based on histology or immunohistochemistry may have bias and variabilities (Yoshihara et al., 2013; Gibney et al., 2016; Spranger and Gajewski, 2018; Zhang et al., 2020; Sanchez-Pino et al., 2021). More recently, RNA-seq analyses have shown that lncRNAs exhibit a better degree of cell type specificity, relative to protein-coding genes in immune cells, highlighting their potential as subpopulation-specific immune cells molecular markers (Huang et al., 2018; Chen et al., 2019; Zhou et al., 2021).

Here, we used a machine learning-based computational framework to identify lncRNA features for evaluating TANs and explored their clinical significance using a combination of lncRNA, immune, and clinical spectrum analyses. The computational framework was used on the TCGA discovery dataset of NSCLC to identify a lncRNA signature (TANIncSig) comprised of 17 lncRNAs obtained from a list of neutrophilspecific lncRNAs using machine learning. Functional enrichment analysis of TANIncSig-related mRNAs showed that TANIncSig is highly correlated with cancer markers of immune response and sustained proliferative signals. Recent experimental evidence on some TANIncSig components is consistent with functional annotations using bioinformatics. It appears that Mir-1303, which is upregulated in tumor tissues, acts as a sponge for LINC01272 and negatively correlates with its expression. A reduction in LINC01272 expression in tissues and cells of NSCLC patients may serve as an independent prognostic marker. LINC01272 overexpression may inhibit NSCLC cells proliferation, migration, and invasion by inhibiting MI-1303 (Zhang and Zhou, 2021). LINC00261 downregulation in gastric cancer is associated with poor prognosis. Ectopic LINC00261 expression disrupts cell migration and invasion, inhibiting metastasis in vitro as well as in vivo. LINC00261 downregulation promotes cell migration and invasion in vitro. LINC00261 overexpression influences epithelial-mesenchymal transition (EMT) through the regulation of E-cadherin, Vimentin and N-cadherin (Liu et al., 2020; Zhai et al., 2021). LINC00668 expression is significantly upregulated via STAT3 signaling in NSCLC tissues as well as cell lines. Clinical studies show that upregulated LINC00668 correlates with histological grade, advanced TNM stage, and lymph node metastasis. Additionally, multivariate analyses established that LINC00668 as an independent marker of overall survival (OS) in patients with NSCLC. LINC00668 downregulation inhibits proliferation, migration, and invasion of NSCLC cells and promotes apoptosis. Mechanistically, LINC00668 is a direct target of miR-193a, leading to down-regulation in the expression of its target gene KLF7. STAT3-initiated LINC00668 promotes NSCLC progression by upregulating KLF7 via sponging Mir-193a. Therefore, it may serve as a prognostic marker and therapeutic target for NSCLC (An et al., 2019). From the perspective of lncRNA, TANIncSig seems to be a transcriptional marker as a potentially measurable indicator of neutrophil activity and prognosis.

To further assess TANIncSig's role in clinical risk stratification, we evaluated its relationship with survival in patients with NSCLC. When applied to the TCGA RNAseq patient dataset, TANIncSig significantly correlated with patient survival. In TANIncSig, three lung squamous cell carcinoma, neutrophil-specific lncRNAs (LINC01272, LINC00261, and LINC00668) were markedly associated with prognostic outcomes. In lung adenocarcinoma, three neutrophil-specific lncRNAs (LINC00528, LINC00967, and LINC00261) significantly correlated with prognosis. In squamous cell carcinoma and lung adenocarcinoma, correlation analysis of individual lncRNAs and risk score with clinical features (TNM staging) revealed that risk score varied significantly with tumor stage. After adjusting for traditional clinical factors, TANIncSig was verified to be an independent prognostic marker for differentiating between poor and good survival outcomes across patient datasets.

Immune checkpoint inhibitors (ICIs) have emerged as effective lung cancer immunotherapies (Suresh et al., 2018; Iams et al., 2020). Some of the drugs acting on the immune checkpoints, CTLA4 and PD-1/PD-L1, have excellent performance against various tumors. Although significant breakthroughs have been made on CTLA4 and PD-1/PD-L1 antibodies, single-drug effective rates are only about 20%, and they benefit a limited proportion of patients (Magiera-Mularz et al., 2017; Lingel and Brunner-Weinzierl, 2019; Rotte, 2019; Yang and Hu, 2019; Liu and Zheng, 2020). The limited efficacy is attributable to the immune system's complexity. Indeed, immune cells, cytokines, and immune adjuvants in the TME interact with each other, limiting the effects of drugs on individual targets. Thus, drugs that target different links and aspects of tumor immunity are needed to enhance immunotherapy outcomes. Up to 29 immunoglobulin superfamily members and 26 members of the tumor necrosis factor receptor superfamily are expressed on T-cell surfaces alone, and there have been preclinical or clinical studies on related immune targets and drugs. Specific immune checkpoints include lymphocyte activating gene 3 (LAG-3), T-cell immunoglobulin mucin 3 (TIM-3), and V region Ig inhibitor (VISTA). Non-specific immune checkpoints include human killer cell immunoglobulin like receptor (KIR), indoleamine 2, 3-dioxidase (IDO), and CD47, these novel immune checkpoint molecules are expected to provide hints for clinical and basic research (Manser et al., 2015; Munn and Mellor, 2016; Burugu et al., 2018; Huang et al., 2020; Logtenberg et al., 2020). VISTA, (B7-H5, PD-1H) is an immunomodulatory receptor that inhibits T-cell response. VISTA is overexpressed on CD11b myeloid cells (e.g., macrophages, monocytes, neutrophils, and dendritic cells)

and it is found that in humans and mice at a lower level in primitive CD4⁺ and CD8⁺ T-cells as well as Tregs. With two potential protein kinase C binding sites and proline residues acting as docking sites in its cytoplasmic tail domain, VISTA can serve as both a receptor and a ligand (Huang et al., 2020; Mutsaers et al., 2021). OX40 (TNFRSF4) has been found to be expressed in activated NK cells, T-cells, NKT cells, as well as neutrophils, and acts as an auxiliary costimulatory immune checkpoint (Curti et al., 2013; Aspeslagh et al., 2016; Buchan et al., 2018). Combining immune checkpoint genes and TANIncSig showed combined prognostic effects on patient survival, in line with previous findings that immunomotor interactions between neutrophilic infiltration and expression levels of checkpoint genes affect the outcome of cancer patients and immunotherapy may also be associated with this condition. In combination with earlier findings, it TANIncSig is correlated appears that with immunosuppressive phenotypes and could predict ICI response. Together, these results indicate that TANIncSig can complement and/or add information to existing immune checkpoint genetic markers.

Due to few gene mutations, lung squamous cell carcinoma is less selective than adenocarcinoma with regards to treatment options, and its survival time (about 1 year) is shorter than that of adenocarcinoma (Travis et al., 2021). Thus, novel, effective advanced lung squamous cell carcinoma treatments are needed to improve patient outcomes. The emergence of immune checkpoint inhibitors in recent years has markedly improved treatment options for advanced lung squamous cell carcinoma patients. Immune checkpoint inhibitors have substantially changed advanced lung squamous cell carcinoma treatment, leading to a shift from retro line immunotherapy to front-line treatment options. Originally approved as second-line treatment after platinum-based dual therapy, palivizumab is now recommended as a single-agent first-line treatment or in combination with chemotherapy. Although treatments targeting the immune checkpoints PD-1 and CTLA4 are successful in many cancers, not all patients benefit from them. Our findings indicate that the combination of CEACAM1, TNFSF4, GEM, CD47, VTCN1, and TANIncSig in squamous cell carcinoma can effectively stratify patients by prognosis, highlighting these immune checkpoint receptors as potential therapeutic targets against advanced lung cancer.

Conclusion

In conclusion, we used a machine learning-based computational framework to identify lncRNA features of TANs (TANIncSig) *via* comprehensive analyses of lncRNA, immune, as well as clinical features. TANIncSig revealed a substantial and repeatable correlation with outcomes, even after adjustments of clinical covariates. Analysis of correlation between prognostic lncRNAs

and risk score with the expression of immune checkpoint molecules demonstrated that TANIncSig can predict immunotherapy. The study is the first to define lncRNA characteristics of tumor-associated neutrophils, highlighting the importance of lncRNAs in immune responses and the potential for more precise and personalized treatment cancer immunotherapy.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

Author contributions

ZT, QW, PC, and HG designed the experiments; ZT, QW, and JS performed the experiments; PC, HG, and YP prepared figures; ZT, QW, CL, and CZ was responsible for statistical analysis and provided helpful suggestions; ZT and QW wrote the manuscript. All authors read and approved the final version of the article.

Funding

This work was Funded by the National Natural Science Foundation of China (No. 81871865); Science and Technology Commission of Shanghai Municipality (No. 19411950300); Shanghai Innovative Collaboration Project (No. 2020CXJQ02); Natural Science Foundation of Tianjin (No. 19JCQNJC12300); "Dream Tutor" New Person Cultivation Program of Shanghai Pulmonary Hospital (No. fkxr 1907).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

An, Y. X., Shang, Y. J., Xu, Z. W., Zhang, Q. C., Wang, Z., Xuan, W. X., et al. (2019). STAT3-induced long noncoding RNA LINC00668 promotes migration and invasion of non-small cell lung cancer via the miR-193a/KLF7 axis. *Biomed. Pharmacother.* 116, 109023. doi:10.1016/j.biopha.2019.109023

Aspeslagh, S., Postel-Vinay, S., Rusakiewicz, S., Soria, J. C., Zitvogel, L., and Marabelle, A. (2016). Rationale for anti-OX40 cancer immunotherapy. *Eur. J. Cancer* 52, 50–66. doi:10.1016/j.ejca.2015.08.021

Atianand, M. K., Caffrey, D. R., and Fitzgerald, K. A. (2017). Immunobiology of long noncoding RNAs. *Annu. Rev. Immunol.* 35, 177–198. doi:10.1146/annurevimmunol-041015-055459

Bao, S., Zhao, H., Yuan, J., Fan, D., Zhang, Z., Su, J., et al. (2020). Computational identification of mutator-derived lncRNA signatures of genome instability for improving the clinical outcome of cancers: A case study in breast cancer. *Brief. Bioinform.* 21 (5), 1742–1755. doi:10.1093/bib/bbz118

Buchan, S. L., Rogel, A., and Al-Shamkhani, A. (2018). The immunobiology of CD27 and OX40 and their potential as targets for cancer immunotherapy. *Blood* 131 (1), 39–48. doi:10.1182/blood-2017-07-741025

Burugu, S., Dancsok, A. R., and Nielsen, T. O. (2018). Emerging targets in cancer immunotherapy. *Semin. Cancer Biol.* 52 (2), 39–52. doi:10.1016/j.semcancer.2017. 10.001

Butturini, E., Carcereri de Prati, A., Boriero, D., and Mariotto, S. (2019). Tumor dormancy and interplay with hypoxic tumor microenvironment. *Int. J. Mol. Sci.* 20, 4305. doi:10.3390/ijms20174305

Chen, F., Chen, J., Yang, L., Liu, J., Zhang, X., Zhang, Y., et al. (2019). Extracellular vesicle-packaged HIF-1a-stabilizing lncRNA from tumour-associated macrophages regulates aerobic glycolysis of breast cancer cells. *Nat. Cell Biol.* 21 (4), 498–510. doi:10.1038/s41556-019-0299-0

Chen, R., Manochakian, R., James, L., Azzouqa, A. G., Shi, H., Zhang, Y., et al. (2020). Emerging therapeutic agents for advanced non-small cell lung cancer. *J. Hematol. Oncol.* 13 (1), 58. doi:10.1186/s13045-020-00881-7

Chen, Y. G., Satpathy, A. T., and Chang, H. Y. (2017). Gene regulation in the immune system by long noncoding RNAs. *Nat. Immunol.* 18 (9), 962–972. doi:10. 1038/ni.3771

Coffelt, S. B., Kersten, K., Doornebal, C. W., Weiden, J., Vrijland, K., Hau, C. S., et al. (2015). IL-17-producing $\gamma\delta$ T cells and neutrophils conspire to promote breast cancer metastasis. *Nature* 522 (7556), 345–348. doi:10.1038/nature14282

Curti, B. D., Kovacsovics-Bankowski, M., Morris, N., Walker, E., Chisholm, L., Floyd, K., et al. (2013). OX40 is a potent immune-stimulating target in late-stage cancer patients. *Cancer Res.* 73 (24), 7189–7198. doi:10.1158/0008-5472.CAN-12-4174

Das, S., and Johnson, D. B. (2019). Immune-related adverse events and antitumor efficacy of immune checkpoint inhibitors. *J. Immunother. Cancer* 7 (1), 306. doi:10.1186/s40425-019-0805-8

Dinh, H. Q., Eggert, T., Meyer, M. A., Zhu, Y. P., Olingy, C. E., Llewellyn, R., et al. (2020). Coexpression of CD71 and CD117 identifies an early unipotent neutrophil progenitor population in human bone marrow. *Immunity* 53 (2), 319–334. doi:10. 1016/j.immuni.2020.07.017

Dolladille, C., Ederhy, S., Sassier, M., Cautela, J., Thuny, F., Cohen, A. A., et al. (2020). Immune checkpoint inhibitor rechallenge after immune-related adverse events in patients with cancer. *JAMA Oncol.* 6 (6), 865–871. doi:10.1001/jamaoncol. 2020.0726

Fatica, A., and Bozzoni, I. (2014). Long non-coding RNAs: New players in cell differentiation and development. *Nat. Rev. Genet.* 15 (1), 7–21. doi:10.1038/nrg3606

Ficarra, V., Galfano, A., Mancini, M., Martignoni, G., and Artibani, W. (2007). TNM staging system for renal-cell carcinoma: Current status and future perspectives. *Lancet. Oncol.* 8 (6), 554–558. doi:10.1016/S1470-2045(07)70173-0

Galluzzi, L., Humeau, J., Buqué, A., Zitvogel, L., and Kroemer, G. (2020). Immunostimulation with chemotherapy in the era of immune checkpoint inhibitors. *Nat. Rev. Clin. Oncol.* 17 (12), 725–741. doi:10.1038/s41571-020-0413-z

Gibney, G. T., Weiner, L. M., and Atkins, M. B. (2016). Predictive biomarkers for checkpoint inhibitor-based immunotherapy. *Lancet. Oncol.* 17 (12), e542–e551. doi:10.1016/S1470-2045(16)30406-5

Hu, G., Tang, Q., Sharma, S., Yu, F., Escobar, T. M., Muljo, S. A., et al. (2013). Expression and regulation of intergenic long noncoding RNAs during T cell development and differentiation. *Nat. Immunol.* 14, 1190–1198. doi:10.1038/ni. 2712

Huang, D., Chen, J., Yang, L., Ouyang, Q., Li, J., Lao, L., et al. (2018). NKILA lncRNA promotes tumor immune evasion by sensitizing T cells to activation-

induced cell death. Nat. Immunol. 19 (10), 1112–1125. doi:10.1038/s41590-018-0207-y

Huang, X., Zhang, X., Li, E., Zhang, G., Wang, X., Tang, T., et al. (2020). Vista: An immune regulatory protein checking tumor and immune cells in cancer immunotherapy. *J. Hematol. Oncol.* 13 (1), 83. doi:10.1186/s13045-020-00917-y

Iams, W. T., Porter, J., and Horn, L. (2020). Immunotherapeutic approaches for small-cell lung cancer. *Nat. Rev. Clin. Oncol.* 17 (5), 300–312. doi:10.1038/s41571-019-0316-z

Khoja, L., Day, D., Wei-Wu Chen, T., Siu, L. L., and Hansen, A. R. (2017). Tumour- and class-specific patterns of immune-related adverse events of immune checkpoint inhibitors: A systematic review. *Ann. Oncol.* 28 (10), 2377–2385. doi:10. 1093/annonc/mdx286

Ko, E. C., Raben, D., and Formenti, S. C. (2018). The integration of radiotherapy with immunotherapy for the treatment of non-small cell lung cancer. *Clin. Cancer Res.* 24 (23), 5792–5806. doi:10.1158/1078-0432.CCR-17-3620

Lecot, P., Sarabi, M., Pereira Abrantes, M., Mussard, J., Koenderman, L., Caux, C., et al. (2019). Neutrophil heterogeneity in cancer: From biology to therapies. *Front. Immunol.* 10, 2155. doi:10.3389/fimmu.2019.02155

Lingel, H., and Brunner-Weinzierl, M. C. (2019). CTLA-4 (CD152): A versatile receptor for immune-based therapy. *Semin. Immunol.* 42, 101298. doi:10.1016/j. smim.2019.101298

Liu, S., Zheng, Y., Zhang, Y., Zhang, J., Xie, F., Guo, S., et al. (2020). Methylationmediated LINC00261 suppresses pancreatic cancer progression by epigenetically inhibiting c-Myc transcription. *Theranostics* 10 (23), 10634–10651. doi:10.7150/thno.44278

Liu, Y., and Zheng, P. (2020). Preserving the CTLA-4 checkpoint for safer and more effective cancer immunotherapy. *Trends Pharmacol. Sci.* 41 (1), 4–12. doi:10. 1016/j.tips.2019.11.003

Logtenberg, M. E. W., Scheeren, F. A., and Schumacher, T. N. (2020). The CD47sirpa immune checkpoint. *Immunity* 52 (5), 742–752. doi:10.1016/j.immuni.2020. 04.011

Magiera-Mularz, K., Skalniak, L., Zak, K. M., Musielak, B., Rudzinska-Szostak, E., Berlicki, L., et al. (2017). Bioactive macrocyclic inhibitors of the PD-1/PD-L1 immune checkpoint. *Angew. Chem. Int. Ed. Engl.* 56 (44), 13732–13735. doi:10.1002/anie.201707707

Manser, A. R., Weinhold, S., and Uhrberg, M. (2015). Human KIR repertoires: Shaped by genetic diversity and evolution. *Immunol. Rev.* 267 (1), 178–196. doi:10. 1111/imr.12316

Marchese, F. P., Raimondi, I., and Huarte, M. (2017). The multidimensional mechanisms of long noncoding RNA function. *Genome Biol.* 18 (1), 206. doi:10. 1186/s13059-017-1348-2

Mishalian, I., Bayuh, R., Levy, L., Zolotarov, L., Michaeli, J., and Fridlender, Z. G. (2013). Tumor-associated neutrophils (TAN) develop pro-tumorigenic properties during tumor progression. *Cancer Immunol. Immunother.* 62 (11), 1745–1756. doi:10.1007/s00262-013-1476-9

Moch, H., Artibani, W., Delahunt, B., Ficarra, V., Knuechel, R., Montorsi, F., et al. (2009). Reassessing the current UICC/AJCC TNM staging for renal cell carcinoma. *Eur. Urol.* 56 (4), 636–643. doi:10.1016/j.eururo.2009.06.036

Munn, D. H., and Mellor, A. L. (2016). Ido in the tumor microenvironment: Inflammation, counter-regulation, and tolerance. *Trends Immunol.* 37 (3), 193–207. doi:10.1016/j.it.2016.01.002

Mutsaers, P., Balcioglu, H. E., Kuiper, R., Hammerl, D., Wijers, R., van Duin, M., et al. (2021). V-domain Ig suppressor of T cell activation (VISTA) expression is an independent prognostic factor in multiple myeloma. *Cancers (Basel)* 13 (9), 2219. doi:10.3390/cancers13092219

Nielsen, S. R., Strøbech, J. E., Horton, E. R., Jackstadt, R., Laitala, A., Bravo, M. C., et al. (2021). Suppression of tumor-associated neutrophils by lorlatinib attenuates pancreatic cancer growth and improves treatment with immune checkpoint blockade. *Nat. Commun.* 12 (1), 3414. doi:10.1038/s41467-021-23731-7

Ponzetta, A., Carriero, R., Carnevale, S., Barbagallo, M., Molgora, M., Perucchini, C., et al. (2019). Neutrophils driving unconventional T cells mediate resistance against murine sarcomas and selected human tumors. *Cell* 178 (2), 346–360. doi:10. 1016/j.cell.2019.05.047

Ranzani, V., Rossetti, G., Panzeri, I., Arrigoni, A., Bonnal, R. J., Curti, S., et al. (2015). The long intergenic noncoding RNA landscape of human lymphocytes highlights the regulation of T cell differentiation by linc-MAF-4. *Nat. Immunol.* 16, 318–325. doi:10.1038/ni.3093

Rinn, J. L., and Chang, H. Y. (2012). Genome regulation by long noncoding RNAs. *Annu. Rev. Biochem.* 81, 145–166. doi:10.1146/annurev-biochem-051410-092902

Rotte, A. (2019). Combination of CTLA-4 and PD-1 blockers for treatment of cancer. J. Exp. Clin. Cancer Res. 38 (1), 255. doi:10.1186/s13046-019-1259-z

Sage, A., Ng, K., Marshall, E., Enfield, K., Stewart, G., Martin, S., et al. (2018). MA24.06 long non-coding rna expression patterns delineate infiltrating immune cells in the lung tumour microenvironment. *J. Thorac. Oncol.* 13, S443–S444. doi:10. 1016/j.jtho.2018.08.524

Sanchez-Pino, M. D., Gilmore, L. A., Ochoa, A. C., and Brown, J. C. (2021). Obesity-associated myeloid immunosuppressive cells, key players in cancer risk and response to immunotherapy. *Obes. (Silver Spring)* 29 (6), 944–953. doi:10.1002/oby. 23108

Schoenfeld, A. J., Arbour, K. C., Rizvi, H., Iqbal, A. N., Gadgeel, S. M., Girshman, J., et al. (2019). Severe immune-related adverse events are common with sequential PD-(L)1 blockade and osimertinib. *Ann. Oncol.* 30 (5), 839–844. doi:10.1093/annonc/mdz077

Schupp, J., Krebs, F. K., Zimmer, N., Trzeciak, E., Schuppan, D., and Tuettenberg, A. (2019). Targeting myeloid cells in the tumor sustaining microenvironment. *Cell. Immunol.* 343, 103713. doi:10.1016/j.cellimm.2017.10.013

Socinski, M. A. (2014). Update on taxanes in the first-line treatment of advanced non-small-cell lung cancer. *Curr. Oncol.* 21 (5), e691–e703. doi:10.3747/co.21.1997

Spranger, S., and Gajewski, T. F. (2018). Impact of oncogenic pathways on evasion of antitumour immune responses. *Nat. Rev. Cancer* 18 (3), 139–147. doi:10. 1038/nrc.2017.117

Suresh, K., Naidoo, J., Lin, C. T., and Danoff, S. (2018). Immune checkpoint immunotherapy for non-small cell lung cancer: Benefits and pulmonary toxicities. *Chest* 154 (6), 1416–1423. doi:10.1016/j.chest.2018.08.1048

Travis, W. D., Dacic, S., Sholl, L. M., WistubaII, Adusumilli, P., Bubendorf, L., et al. (2021). IASLC multidisciplinary recommendations for pathologic assessment of lung cancer resection specimens after neoadjuvant therapy. *J. Thorac. Oncol.* 16 (1), 709–740. doi:10.1016/j.jtho.2020.01.005

Vansteenkiste, J., Wauters, E., Reymen, B., Ackermann, C. J., Peters, S., and De Ruysscher, D. (2019). Current status of immune checkpoint inhibition in earlystage NSCLC. *Ann. Oncol.* 30 (8), 1244–1253. doi:10.1093/annonc/mdz175

Vaughan, J. E., and Walsh, S. W. (2005). Neutrophils from pregnant women produce thromboxane and tumor necrosis factor-alpha in response to linoleic acid and oxidative stress. *Am. J. Obstet. Gynecol.* 193 (3), 830–835. doi:10.1016/j.ajog.2005.01.057

Wang, L., Felts, S. J., Van Keulen, V. P., Scheid, A. D., Block, M. S., Markovic, S. N., et al. (2018). Integrative genomewide analysis of long noncoding RNAs in diverse immune cell types of melanoma patients. *Cancer Res.* 78, 4411–4423. doi:10. 1158/0008-5472.CAN-18-0529

Wu, L., and Zhang, X. H. (2020). Tumor-associated neutrophils and macrophages-heterogenous but not chaotic. *Front. Immunol.* 11, 553967. doi:10. 3389/fimmu.2020.553967

Yang, J., and Hu, L. (2019). Immunomodulators targeting the PD-1/PD-L1 protein-protein interaction: From antibodies to small molecules. *Med. Res. Rev.* 39 (1), 265–301. doi:10.1002/med.21530

Yoshihara, K., Shahmoradgoli, M., Martínez, E., Vegesna, R., Kim, H., Torres-Garcia, W., et al. (2013). Inferring tumour purity and stromal and immune cell admixture from expression data. *Nat. Commun.* 4, 2612. doi:10.1038/ncomms3612

Yue, P., Harper, T., Bacot, S. M., Chowdhury, M., Lee, S., Akue, A., et al. (2018). BRAF and MEK inhibitors differentially affect nivolumab-induced T cell activation by modulating the TCR and AKT signaling pathways. *Oncoimmunology* 8 (1), e1512456. doi:10.1080/2162402X.2018.1512456

Zhai, S., Xu, Z., Xie, J., Zhang, J., Wang, X., Peng, C., et al. (2021). Epigenetic silencing of LncRNA LINC00261 promotes c-myc-mediated aerobic glycolysis by regulating miR-222-3p/HIPK2/ERK axis and sequestering IGF2BP1. *Oncogene* 40 (2), 277–291. doi:10.1038/s41388-020-01525-3

Zhang, B., Wu, Q., Li, B., Wang, D., Wang, L., and Zhou, Y. L. (2020). m6A regulator-mediated methylation modification patterns and tumor microenvironment infiltration characterization in gastric cancer. *Mol. Cancer* 19 (1), 53. doi:10.1186/s12943-020-01170-0

Zhang, S., and Zhou, J. (2021). Low LINC01272 predicts poor prognosis of nonsmall cell lung cancer and its biological function in tumor cells by inhibiting miR-1303. Oncol. Lett. 22 (3), 652. doi:10.3892/ol.2021.12913

Zhao, R., Li, B., Zhang, S., He, Z., Pan, Z., Guo, Q., et al. (2021). The N6methyladenosine-modified pseudogene HSPA7 correlates with the tumor microenvironment and predicts the response to immune checkpoint therapy in glioblastoma. *Front. Immunol.* 12, 653711. doi:10.3389/fimmu. 2021.653711

Zhou, M., Zhang, Z., Bao, S., Hou, P., Yan, C., Su, J., et al. (2021). Computational recognition of lncRNA signature of tumor-infiltrating B lymphocytes with potential implications in prognosis and immunotherapy of bladder cancer. *Brief. Bioinform.* 22 (3), bbaa047. doi:10.1093/bib/bbaa047

Zhou, M., Zhang, Z., Zhao, H., Bao, S., Cheng, L., and Sun, J. (2018). An immunerelated six-lncRNA signature to improve prognosis prediction of glioblastoma multiforme. *Mol. Neurobiol.* 55, 3684–3697. doi:10.1007/s12035-017-0572-9

Zhou, M., Zhao, H., Xu, W., Bao, S., Cheng, L., and Sun, J. (2017). Discovery and validation of immune-associated long non-coding RNA biomarkers associated with clinically molecular subtype and prognosis in diffuse large B cell lymphoma. *Mol. Cancer* 16, 16. doi:10.1186/s12943-017-0580-4

Zhu, J., Li, R., Tiselius, E., Roudi, R., Teghararian, O., Suo, C., et al. (2017). Immunotherapy (excluding checkpoint inhibitors) for stage I to III non-small cell lung cancer treated with surgery or radiotherapy with curative intent. *Cochrane Database Syst. Rev.* 12 (12), CD011300. doi:10.1002/14651858. CD011300.pub2