

The changes of T lymphocytes and cytokines in ICR mice fed with Fe₃O₄ magnetic nanoparticles

Jun Wang¹
Baoan Chen¹
Nan Jin¹
Guohua Xia²
Yue Chen¹
Ying Zhou¹
Xiaohui Cai^{1,2}
Jiahua Ding¹
Xiaomao Li³
Xuemei Wang⁴

¹Department of Hematology, Zhongda Hospital, ²Department of Medical Laboratory, Medical School, Southeast University, Nanjing, People's Republic of China; ³Department of Physics, University of Saarland, Saarbruecken, Germany; ⁴National Key Laboratory of Bioelectronics (Chien-Shiung Wu Laboratory), Southeast University, Nanjing, People's Republic of China

Abstract: The aim of this article is to study the changes inhibited T lymphocytes and cytokines related to the cellular immunity in ICR (imprinting control region) mice fed with Fe₃O₄ magnetic nanoparticles (Fe₃O₄-MNPs). The Fe₃O₄-MNPs were synthesized, and their characteristics such as particle size, zeta potential, and X-ray diffraction patterns were measured and determined. All ICR mice were sacrificed after being exposed to 0, 300, 600, and 1200 mg/kg of Fe₃O₄-MNPs by single gastric administration for 14 days. Splenocyte proliferation was indicated with stimulate index by MTT assay; release of cytokines in the serum of ICR mice was detected by enzyme-linked immunosorbent assay, and the phenotypic analyses of T-lymphocyte subsets were performed using flow cytometry. Our results indicated that there were no significant differences in splenocyte proliferation and release of cytokines between exposed and control groups. Furthermore, there was no significant difference in the proportions of T-lymphocyte subsets in the low-dose Fe₃O₄-MNPs group when compared to the control group, but the proportions of CD3⁺CD4⁺ and CD3⁺CD8⁺ T-lymphocyte subsets both in the medium- and high-dose Fe₃O₄-MNPs groups were higher than those in the control group. It is concluded that a high dose of Fe₃O₄-MNPs, to some extent, could influence in vivo immune function of normal ICR mice.

Keywords: Fe₃O₄, magnetic nanoparticles, splenocyte proliferation, release of cytokines, T-lymphocyte subsets, ICR mice

Introduction

Recently, nanometer-sized magnetic particles have been intensively investigated due to their particularly large surface:volume ratio, quantum-size effect, magnetic character as well as their potential application in the areas of bioscience and medicine. Very promising nanoparticles are magnetic iron oxide (Fe₃O₄) nanoparticles with appropriate surface modification, which have been widely used experimentally for numerous in vivo applications, such as magnetic resonance imaging, tissue repair, immunoassay, detoxification of biological fluids, drug delivery, hyperthermia, cell separation, and so on.^{1,2}

In our previous studies,^{3,4} we demonstrated that Fe₃O₄ magnetic nanoparticles (Fe₃O₄-MNPs) combined with chemotherapy drugs could inhibit tumor proliferation and induce apoptosis of tumor cells in a dose- and time-dependent manner, which may be related to the change in drug formulations caused by magnetic nanomaterials. As a targeted therapeutic drug agent for malignant tumor due to its high specificity, Fe₃O₄-MNPs may become the mainstay for tumor-targeted therapy.

So far, clinical application of magnetic nanoparticles has focused mainly on the areas of magnetic ferrite. Compared with other magnetic materials, magnetic iron

Correspondence: Baoan Chen
Department of Hematology, Zhongda Hospital, School of Clinical Medicine, Southeast University, Nanjing 210009, People's Republic of China
Tel +86 25 83272006
Fax +86 25 83272011
Email cba8888@hotmail.com

oxide nanoparticles (mainly Fe_3O_4 and $\gamma\text{-Fe}_2\text{O}_3$), which have good chemical stability, magnetic responsiveness, biocompatibility, easy preparation, and significant features in the diagnosis and treatment of diseases, have attracted the attention of researchers worldwide.

The prospects of extensive application of magnetic nanoparticles, account for their impact on human health, living environment, and even life in all aspects of the process are the focus of researchers,^{5,6} and there are high expectations for their magnetic characteristics and lower cytotoxicity. However, the risk assessment of the biological safety of magnetic nanoparticles has just started and remains in the initial stage of accumulation of controversial data. As a result of their short period of use in human activities, there are few reports on the toxicity of nanomaterials.⁷⁻⁹ One experiment reported that nanosilver at a high concentration had a significant cytotoxic effect on the healthy human peripheral blood mononuclear cells and significantly inhibited the phytohemagglutinin-induced cytokine production.¹⁰ Therefore, a safety evaluation, especially of the in vitro and in vivo immunological effects of Fe_3O_4 -MNPs is urgently needed.

According to the requirements of the State Drug Administration in 2002 as stated in 'Guiding Principles for the Chemical and Therapeutic Biological Products Study',¹¹ clinical treatment with Fe_3O_4 -MNPs is <5 days, and the experimental period is 14 days. In our study, Fe_3O_4 -MNPs were orally administered once as a single dose imprinting control region (ICR) mice, and in their general performance was observed. Furthermore, all mice were sacrificed separately and used to assess splenocyte proliferation, release of cytokines, and T-lymphocyte subsets and also to understand whether Fe_3O_4 -MNPs are toxic to ICR mice, thus providing a basis for its safe use in clinical settings.

Material and method

Animals and animal care

Female and male ICR mice, which were age-matched (8 weeks of age) and weight-matched (18–22 g), were purchased from Shanghai National Center for Laboratory Animals. The mice were kept in a 12-hour dark/light cycle and received water and food ad libitum in a semibarrier system.

Preparation of Fe_3O_4 -MNPs

As previously described,¹² Fe_3O_4 -MNPs were produced by electrochemical deposition under oxidizing conditions. Briefly, a quantity of ferric chloride hexahydrate (0.02 mol/L) and ferrous sulfate heptahydrate (0.012 mol/L) was dissolved in 200 mL deionized water under nitrogen protection,

and then about 50 mL of 25% ammonium hydroxide was added slowly with violent magnetic force stirring until the pH was 9. The procedure took 30 minutes to generate a dark precipitate. Next, the resultant precipitate was separated using a permanent magnet and then washed repeatedly by deionized water 5–7 times until the pH of the supernatant was 7. Finally, the characteristics of Fe_3O_4 -MNPs were analyzed.

Determination of Fe_3O_4 -MNPs size and zeta potential

To measure the Fe_3O_4 -MNPs particle size, the sample was dispersed in deionized water and then the particle size was measured using a glass cuvette by the ZetaPlus particle seizer (Brookhaven Instruments Corporation, Holtsville, NY). The same suspension was used for measuring the zeta potential of particles. The average zeta potential and the standard deviation of the three analyses were reported.

Analysis of X-ray diffraction patterns

X-ray diffraction (XRD) pattern of lyophilized sample of Fe_3O_4 -MNPs was carried out using an X-ray diffractometer (ARL X'TRA; Thermo Electron Co, Ecublens, Switzerland). The equipment used a copper target X-ray tube with $\text{Cu K}\alpha$ radiation at a scan speed of 5°/minute. The acceleration voltage and the current flux chosen for the measurement were 45 kV and 35 mA, respectively.

Experimental groups

Forty ICR mice, half male and half female, were individually and randomly divided into four groups: control group (0.5 mL of sterile physiologic saline), low-dose group (300 mg/kg Fe_3O_4 -MNPs), medium-dose group (600 mg/kg Fe_3O_4 -MNPs), and high-dose group (1200 mg/kg Fe_3O_4 -MNPs). The mice in the exposed group were given different concentrations of Fe_3O_4 -MNPs, and those in the control group were given RPMI-1640 (Gibco Chemical Co, Carlsbad, CA) medium by single gastric perfusion and observed for the poisoning symptoms after administration of the drug; finally, they were all sacrificed under Avertin® anesthesia by cervical dislocation after 14 days and used to measure various immune indicators.

Preparation of blood samples

Blood samples were obtained from ICR mice by extirpated eyeballs after being exposed for 14 days. Blood was centrifuged (1500 rpm) for 5 minutes at 4°C to separate plasma from blood cells. The blood mononuclear cells were used for analyzing surface markers of T lymphocytes, and the plasma was prepared for quantitative determination of cytokines.

MTT assays for splenocyte proliferation

Single-cell suspensions were prepared aseptically from ICR mice spleens in RPMI-1640 medium. Briefly, a single-cell suspension was prepared by puncturing the spleen with a 20-gauge needle and gently flushing the organ with ice-cooled (4°C) RPMI-1640. The remaining splenocyte suspension, which was freed from debris by centrifugation at 1000 rpm for 20 minutes at 4°C, was washed twice and adjusted to 2×10^6 cells/mL with RPMI-1640. The adjusted cells were placed in a 96-well microtiter plate in 200 μ L aliquots and cultured in the presence of T-lymphocytes mitogen (5 μ g/mL Concanavalin A [ConA]) (Sigma Chemical Co, St. Louis, MO) for 68 hours at 37°C in a 5% humidified CO₂ atmosphere. The cells that received complete RPMI-1640 medium in the place of mitogen were regarded as the control group. Following this, 10 μ L 3-2 (4, 5-dimethylthiazol-2-yl) 2, 5-diphenyltetrazolium bromide (MTT) (0.5 mg/mL) (Pharmingen, San Diego, CA) was added to each well and cultured for another 4 hours. The formazan was dissolved with 150 μ L dimethyl sulfoxide after blotting the culture medium. The plates were shaken lightly for 10 minutes, and then the reduction of methyl thiazol tetrazolium (MTT) was quantified by absorbance at a wavelength of 570 nm using a microplate reader (Model-550; Bio-Rad Laboratories, Hercules, CA). The results were expressed as a mean differential stimulate index (SI) (SI = optical density with ConA/optical density without ConA) of each animal.

ELISA assays for quantitative determination of cytokines

The release of interleukin 2 (IL-2), interleukin 4 (IL-4), interleukin 10 (IL-10), and interferon- γ (INF- γ) in serum were measured in duplicate using enzyme-linked immunosorbent assay (ELISA) kit (Gibco, Carlsbad, CA) according to the manufacturer's instructions. Briefly, 50 μ L samples or standard control was added to 50 μ L assay diluents in each well, incubated at room temperature for 2 hours, and after thorough washing, 100 μ L conjugate was added to each well for incubation for 2 hours. A 100 μ L substrate solution was then added to each well and incubated for a further 30 minutes. Finally, a 100 μ L stop solution was added to each well, and the optical density was measured using an ELISA reader (Bio-Rad Laboratories) with dual wavelength of 450 nm.

FCM assays for T-lymphocyte subsets

Phenotypic analyses of blood lymphocytes were performed using flow cytometry. Briefly, blood mononuclear

cells were separated on Ficoll-Hypaque, washed twice in phosphate-buffered saline (PBS), and adjusted to a density of 1×10^7 cells/mL. The cells were incubated with PE or FITC-conjugated monoclonal antibodies (antimouse CD3 [PE-Cy5], antimouse CD4 [FITC], or antimouse CD8 [PE]) (all sourced from Pharmingen) for 10 minutes, washed three times, and resuspended in PBS immediately before determination. At least 10,000 cells in fluorescent channel 3 for CD3, fluorescent channel 1 for CD4, and fluorescent channel 2 for CD8 were detected using FACScan flow cytometer (Becton Dickinson, Franklin Lakes, NJ) and analyzed with CELLQuest®. Results were expressed as mean fluorescence intensity for a given molecule per cell.

Statistical analysis

Data were expressed as means \pm standard deviation ($\bar{x} \pm S$). Statistical analyses were done using SAS software (version 9.1; SAS Institute, Cary, NC). One-way analysis of variance (ANOVA) and rank sum test were carried out to compare the normal data distribution between the exposed and control groups. Wilcoxon rank test was carried out to compare the nonnormal data distribution between the exposed and control groups. Statistical significance for all tests was $P = 0.05$.

Results

Physical characteristics of magnetic nanoparticles

The particle size of the analyzer showed that the size of Fe₃O₄-MNPs ranged from 20.3 to 95.5 nm and the median particle size was 44.0 nm (Figure 1). The zeta potential of Fe₃O₄-MNPs in water was -24.71 mV.

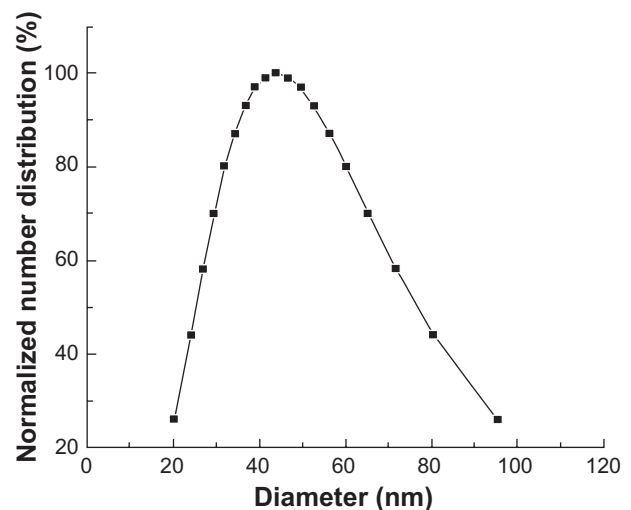


Figure 1 Particle size of Fe₃O₄-MNPs.

Abbreviation: Fe₃O₄-MNPs, Fe₃O₄ magnetic nanoparticles.

XRD pattern of prepared Fe_3O_4 -MNPs reveal diffraction peaks at (111), (220), (311), (400), (422), (511), (440), and (533), which are the characteristic peaks of the Fe_3O_4 crystal with a cubic spinel structure (Figure 2).

Splenocytes proliferation

Wilcoxon rank test was carried out to compare the data of splenocyte proliferation from the ICR mice in the groups fed with a different dose of Fe_3O_4 -MNPs, which were not subject to the normal distribution. Our results showed that there was no significant difference in SI between the exposed and control groups ($P > 0.05$) (Figure 3).

The level of serum cytokine in ICR mice

With the exception of $\text{INF-}\gamma$, the data on cytokine production from the ICR mice in these four groups fed with a different doses of Fe_3O_4 -MNPs showed a normal distribution. One-way ANOVA and rank sum test were carried out to compare the data on IL-2, IL-4, and IL-10 production between the exposed and control groups. Wilcoxon rank test was carried out to compare the data on $\text{INF-}\gamma$ production between the exposed and control groups. We found that the various differences of cytokine production from the ICR mice were not statistically significant in the four groups fed with a different dose of Fe_3O_4 -MNPs (Table 1).

T-lymphocyte subsets

A Wilcoxon rank test was carried out to compare the data of T-lymphocyte subsets from the ICR mice in the four groups fed with a different dose of Fe_3O_4 -MNPs, which were not subject to a normal distribution. We found that there was no difference between the low-dose group and the control group in the proportion of $\text{CD3}^+\text{CD4}^+$ and $\text{CD3}^+\text{CD8}^+$

T lymphocytes in peripheral blood of ICR mice ($P > 0.05$), but a greater proportion of $\text{CD3}^+\text{CD4}^+$ and $\text{CD3}^+\text{CD8}^+$ T lymphocytes in peripheral blood of ICR mice was detected in both the medium- and high-dose groups than the control group ($P < 0.05$) (Table 2).

Discussion

Fe_3O_4 -MNPs with excellent properties have been successfully prepared by electrochemical deposition under oxidizing conditions and confirmed by XRD. The positions of the characteristic peaks agreed well with Joint Committee on Powder Diffraction Standards (JCPDS) card (No. 19-0629), which showed that the product is in the pure Fe_3O_4 phase and that there are no other detectable phases. Based on the characteristics of prepared Fe_3O_4 -MNPs, they can be used to study its effect on the immune function in vivo.

Splenic lymphocytes contain T lymphocytes and B lymphocytes, and both contents are approximate. ConA is a mitogen of T lymphocytes, which promotes the proliferation of only T lymphocytes, and lipopolysaccharides is a mitogen of B lymphocytes, which promotes the proliferation of only B lymphocytes. Lymphocytes proliferation is the most direct reflection of the cellular immunity. T lymphocytes and B lymphocytes stimulated by antigens can split and proliferate, resulting in a specific immune response and producing antibodies or cytokines. Therefore, we have investigated splenocyte proliferation of ICR mice fed with a different dose of Fe_3O_4 -MNPs, following exposure to ConA.

T lymphocytes include three major functional subsets: T-helper lymphocytes (Th cells), T-suppressor lymphocytes

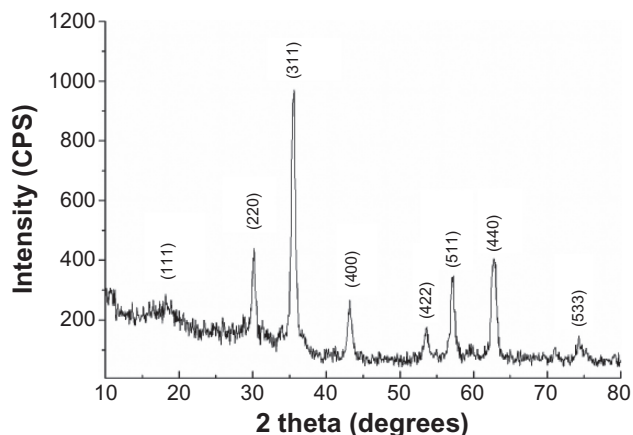


Figure 2 The crystallinity of Fe_3O_4 -MNPs by XRD.

Abbreviations: Fe_3O_4 -MNPs, Fe_3O_4 magnetic nanoparticles; XRD, X-ray diffraction.

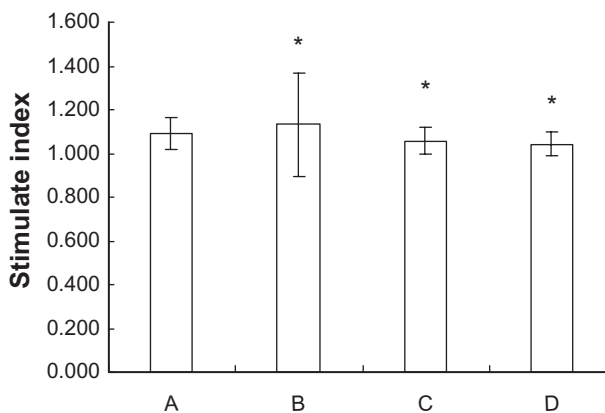


Figure 3 Splenocytes proliferation of ICR mice fed with different doses of Fe_3O_4 -MNPs.

Notes: **A** Control group, **B** low-dose group (300 mg/kg Fe_3O_4 -MNPs), **C** medium-dose group (600 mg/kg Fe_3O_4 -MNPs), **D** high-dose group (1200 mg/kg Fe_3O_4 -MNPs), * $P > 0.05$, there were no difference between exposed groups and control group.

Abbreviations: Fe_3O_4 -MNPs, magnetic Fe_3O_4 nanoparticles; ICR, imprinting control region.

Table 1 The level of serum cytokine in ICR mice fed with Fe₃O₄-MNPs

| Groups | The level of cytokines (pg/mL) (n = 10, $\bar{x} \pm S$) | | | |
|---|---|--------------------|---------------------|----------------------|
| | IL-2 | IL-4 | IL-10 | INF- γ |
| Control | 29.97 \pm 1.93 | 28.90 \pm 1.92 | 28.31 \pm 1.51 | 11.55 \pm 3.55 |
| Low dose of Fe ₃ O ₄ -MNPs | 30.23 \pm 3.02* | 29.18 \pm 2.45** | 29.70 \pm 1.56*** | 13.15 \pm 1.29**** |
| Medium dose of Fe ₃ O ₄ -MNPs | 29.28 \pm 1.14* | 28.14 \pm 0.67** | 29.03 \pm 2.66*** | 12.79 \pm 1.87**** |
| High dose of Fe ₃ O ₄ -MNPs | 29.00 \pm 1.21* | 27.78 \pm 1.55** | 29.27 \pm 3.34*** | 12.57 \pm 2.71**** |

Notes: * $P > 0.05$, ** $P > 0.05$, *** $P > 0.05$ versus control group by one-way analysis of variance (ANOVA) and rank sum test; **** $P > 0.05$ versus control group by Wilcoxon rank test.

Abbreviations: Fe₃O₄-MNPs, Fe₃O₄ magnetic nanoparticles; ICR, imprinting control region.

(T_s cells), and T-cytotoxic lymphocytes (T_c cells). Th cells express CD3⁺CD4⁺, but T_s cells and T_c cells express CD3⁺CD8⁺. CD3⁺CD4⁺ cells can secrete various cytokines and play an important role in regulating the body's immune response. Th1 cells secrete interleukins such as IL-2 and INF- γ , whose main function is to promote cell-mediated immune response; they are involved in proliferation, differentiation, and maturity in late hypersensitivity reacting T cells and cytotoxic T lymphocytes. They can also activate macrophages and natural killer cells. Th2 cells secrete interleukins such as IL-4 and IL-10, which can induce not only the humoral immune mechanism but also the specific cellular immune responses through other ways. Under normal circumstances, Th1/Th2 maintain a balance in the human body, and their imbalance will lead to many diseases.¹³

We chose IL-2 and IL-4, which reflect Th1 cells' function, and IL-10 and INF- γ , which reflect Th2 cells' functions, as our study subjects. With the exception of INF- γ , the quantitative determination of serum cytokines from the ICR mice fed with a different dose of Fe₃O₄-MNPs showed a normal distribution.

Our results showed that there were no significant changes in the level of IL-2, IL-4, IL-10, and INF- γ in ICR mice fed with Fe₃O₄-MNPs when compared with the control group (Table 1). However, a recent study of Liu et al¹⁴ reported that 100 μ L Fe₃O₄ nanoparticles absorbed autovaccine suspension

(2 mg/mL) were subcutaneously injected into H22 live cancer mice, the mass IL-4 inhibitory rate of mice H22 live cancer was raised, the activity of cytotoxicity was boosted, the level of INF- γ cytokine was enhanced, and the amount of IL-4 was downregulated, which indicates that Fe₃O₄ nanoparticles as adjuvants had great potential to enhance antitumor immune response. Our different results may be related to the different manner and dosage of drug administration.

Previous studies on the toxicity of iron nanoparticles reported that high doses of iron nanoparticles not only cause tissue damage, but also elevate the levels of white blood cell count.¹⁵ A possible explanation for the mechanism of tissue damage could be that large doses of iron nanoparticles cause tissue damage, which then induces cytotoxicity, leading to an inflammatory reaction, eventually causing the change of T-lymphocyte subsets.

Our recent article indicates that the proportion of CD4⁺ and CD8⁺ T-lymphocytes subset in the peripheral blood of the low-dose group was more than that of the control group in ICR mice, though there was no difference in the number of CD4⁺ T cells between the medium-dose group and the low-dose group. Interestingly, the Fe₃O₄-MNPs altered the production of IL-2, INF- γ , and IL-10, but no significant changes of IL-4 were observed in peripheral blood. Therefore, it is concluded that Fe₃O₄-MNPs could influence immune functions of normal ICR mice in a dose-dependent manner.¹⁶

Table 2 The proportion of T-lymphocyte subsets of ICR mice fed with different doses of Fe₃O₄-MNPs (n = 10, $\bar{x} \pm S$)

| T lymphocytes | Groups (%) | | | |
|-----------------------------------|------------------|--|---|---|
| | Control group | Low dose of Fe ₃ O ₄ -MNPs | Medium dose of Fe ₃ O ₄ -MNPs | High dose of Fe ₃ O ₄ -MNPs |
| CD3 ⁺ CD4 ⁺ | 46.01 \pm 4.02 | 40.61 \pm 4.04 | 59.15 \pm 1.59* | 59.66 \pm 1.84* |
| CD3 ⁺ CD8 ⁺ | 20.42 \pm 0.28 | 22.14 \pm 0.60 | 23.24 \pm 0.54** | 23.40 \pm 1.77** |

Notes: * $P < 0.05$ versus control group by Wilcoxon rank test; ** $P < 0.05$ versus control group by Wilcoxon rank test.

Abbreviations: Fe₃O₄-MNPs, Fe₃O₄ magnetic nanoparticles; ICR, imprinting control region.

The results presented in this article are different from these of other recent reports, because we have employed the method of offering oral Fe_3O_4 -MNPs instead of intravenous Fe_3O_4 -MNPs. Oral Fe_3O_4 -MNPs must be affected by the acidic environment in the mice's stomach and absorbed in gastrointestinal organs. How to affect the metabolism of Fe_3O_4 -MNPs at the molecular level by the actions of stomach acidity and gastrointestinal absorption will be investigated in our future research.

Conclusions

Based on the characteristics and the effects of Fe_3O_4 -MNPs on splenocyte proliferation, cytokine production, and T-lymphocyte subsets in ICR mice, our studies conclude that a high dose of Fe_3O_4 -MNPs could influence in vivo immune function of normal ICR mice. However, the reasons why there are changes in splenocyte proliferation, cytokine production, and T-lymphocyte subsets in ICR mice fed with high dose of Fe_3O_4 -MNPs need to be further explored. Moreover, subacute and chronic toxicity of Fe_3O_4 -MNPs in vivo still need to be further explored in the future.

Acknowledgments

This work was supported by 973 National Key Fundamental Research Project of China (No. 2006CB933205), 863 Project of People's Republic of China (No. 2007AA022007), National Nature Science Foundation of People's Republic of China (No. 30740062, 30872970), and Special-Purpose Science Research Foundation for High School (No. 20070286042).

Disclosure

The authors report no conflicts of interest in this work.

References

1. Singh R, Lillard JW Jr. Nanoparticle-based targeted drug delivery. *Exp Mol Pathol*. 2009;86:215–223.
2. Hong RY, Feng B, Chen LL, et al. Synthesis, characterization and MRI application of dextran-coated Fe_3O_4 magnetic nanoparticles. *Biochem Eng J*. 2008;42:290–300.
3. Chen BA, Cheng J, Wu Y, et al. Reversal of multidrug resistance by magnetic Fe_3O_4 nanoparticle copolymerizing daunorubicin and 5-bromotetrandrine in xenograft nude-mice. *Int J Nanomedicine*. 2009;4:73–78.
4. Chen BA, Cheng J, Shen MF, et al. Magnetic nanoparticle of Fe_3O_4 and 5-bromotetrandrin interact synergistically to induce apoptosis by daunorubicin in leukemia cells. *Int J Nanomedicine*. 2009;4:65–71.
5. Sanhai WR, Sakamoto JH, Canady R, Ferrari M. Seven challenges for nanomedicine. *Nat Nanotechnol*. 2008;3:242–244.
6. Linkov I, Satterstrom FK, Corey LM. Nanotoxicology and nanomedicine: making hard decisions. *Nanomedicine*. 2008;4:167–171.
7. Han Y, Yin LP, Long L, et al. Distribution of nano- Fe_3O_4 and nano- TiO_2 in tissues of mice. *Chin J Public Health*. 2009;25:835–836.
8. Liu L, Tang M, Liu L, et al. The pharmacokinetics study of nanoparticles of Fe_2O_3 coated with glutamic acid. *J Environ Occup Med*. 2006;23:1–3.
9. Wen M, Song L, Bo W, Li SL, Li BB. Preparation of superparamagnetic iron oxide nanoparticles and its acute toxicity to mice. *Acad J Sec Mil Med Univ*. 2007;28:1104–1108.
10. Shin SH, Ye MK, Kim HS, Kang HS. The effects of nano-silver on the proliferation and cytokine expression by peripheral blood mononuclear cells. *Int Immunopharmacol*. 2007;7:1813–1818.
11. Zheng S. *Guiding Principles for the Chemical and Therapeutic Biological Products Study (Trial)*. Beijing, China: China Medical Science and Technology Press; 2002:1–97.
12. Wang J, Chen Y, Chen BA, et al. Pharmacokinetic parameters and tissue distribution of magnetic Fe_3O_4 nanoparticles in mice. *Int J Nanomedicine*. 2010;5:861–866.
13. Gao YJ, Li HW, Dong N. CD4^+ T helper lymphocytes and cytokines participates in the immune tolerance following liver transplantation. *J Clin Rehabilitative Tissue Eng Res*. 2009;13:10521–10524.
14. Liu H, Zhang DS, Du Y. The immunotherapeutic effect of Fe_3O_4 nanoparticles as adjuvants on mice H22 live cancer. *J Nanosci Nanotechnol*. 2010;10:514–519.
15. Liu L, Tang M, Gu N, et al. Study on acute and chronic toxicity to rats of nanopartkks of Fe_2O_3 coated with glutamk add. *J Environ Occup Med*. 2004;21:430–433.
16. Chen BA, Jin N, Wang J. The effect of magnetic nanoparticles of Fe_3O_4 on immune function in normal ICR mice. *Int J Nanomedicine*. 2010;5:593–599.

International Journal of Nanomedicine

Publish your work in this journal

The International Journal of Nanomedicine is an international, peer-reviewed journal focusing on the application of nanotechnology in diagnostics, therapeutics, and drug delivery systems throughout the biomedical field. This journal is indexed on PubMed Central, MedLine, CAS, SciSearch®, Current Contents®/Clinical Medicine,

Submit your manuscript here: <http://www.dovepress.com/international-journal-of-nanomedicine-journal>

Dovepress

Journal Citation Reports/Science Edition, EMBASE, Scopus and the Elsevier Bibliographic databases. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.