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Synthesis, Inhibitory Activity, and In Silico Modeling of Selective COX-1 Inhibitors with a Quinazoline Core

Marcela Dvorakova,* Lenka Langhansova, Veronika Temml, Antonio Pavicic, Tomas Vanek, and Premysl Landa



KEYWORDS: Cyclooxygenase, inhibitor, quinazoline, selectivity, docking

selectivity were elucidated using in silico modeling.

quinazoline inhibitor was 64 nM. The structural features ensuring COX-1

yclooxygenase (COX) is a bifunctional, membranebound homodimer with fatty acid oxygenase and peroxidase function. It is a key enzyme of the inflammation process and the main target of nonsteroidal anti-inflammatory drugs (NSAIDs). Two catalytically functional COX isoforms are present in mammals, COX-1 and COX-2. These isoforms share a considerable protein sequence, and their catalytic mechanism is very similar. Both are activated by peroxides; however, COX-1 requires much higher concentrations for activation than COX-2.¹ Both also consist of three domains, one of which is the catalytic domain that contains both the fatty acid oxygenase and peroxidase active sites. The oxygenase site is buried deep in the catalytic domain, and its entrance is governed by Arg120, Tyr355, and Glu524 amino acids. The volume of the COX-1 oxygenase site is about 20% smaller than that of COX-2. In addition, the COX-2 oxygenase site contains an additional hydrophilic side-pocket in the proximity of Phe518. In COX-1, an access to this pocket is hindered by bulky amino acids, Ile523 and Ile434, which in COX-2 are replaced by smaller valines.²

The oxygenase site catalyzes the synthesis of prostaglandins from arachidonic acid (AA). COX-1 is a primary contributor to the synthesis of thromboxane A2 (TXA2), while COX-2 furnishes predominantly prostaglandin E2 (PGE2) and prostacyclin (PGI2).³ Thus, deletion of the COX-1 gene causes predominantly impaired platelet aggregation, whereas COX-2 gene deletion seems to be detrimental to an organism.⁴ Moreover, because COX-2 cannot be usually detected under normal conditions and is rapidly induced during inflammation, it was long believed that COX-2 is the main proinflammatory isoform and COX-1 provides gastrointestinal tract protection

and modulates platelet function.^{4,5} This oversimplified view has been lately challenged by a number of studies.⁶⁻⁹ Today, it is believed that COX-1 is responsible for the primary prostanoid response to inflammatory stimuli, especially in these cells and tissues, where it is constitutively expressed, whereas COX-2 contributes to prostanoid synthesis later in the inflammation process.^{4–6,10–12} Moreover, COX-1 upregulation was found in various cancers, such as skin, breast, colorectal, or epithelial ovarian cancer^{10,11} as well as during atherosclerosis,¹ tocolysis,¹⁴ or neuroinflammation.^{12,15,16} Consequently, selective COX-1 inhibition is a promising target for the treatment of cancers and neurodegenerative disorders. Furthermore, the cardioprotective role of COX-1 in control of thrombocyte aggregation has been reported.^{17,18} On the top of that, COX-1 has been identified as one of two main enzymes involved in the synthesis of chemoprotective factors derived from mesenchymal stem cells, which are able to modulate the response to chemotherapy, suggesting that the inhibition of COX-1 may reverse drug resistance.¹⁹ Therefore, selective COX-1 inhibitors may provide a new direction in the development of antiinflammatory compounds with potential activity toward diseases other than those treated with NSAIDs.

 IC_{50} (COX-1) = 64 nM/ IC_{50} (COX-2) = > 50 μ M

Relatively few selective COX-1 inhibitors have been developed so far, such as SC-560, mofezolac, or FR122047

Received: January 3, 2021 Accepted: March 5, 2021 Published: March 12, 2021



ACS Medicinal Chemistry Letters

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Figure 1. Examples of selective COX-1 inhibitors.



Figure 2. Quinazoline anti-inflammatory drugs on market.

Scheme 1. Synthesis of Quinazolines 2a and 2b^a



^{*a*}Reagents and conditions: (a) acetic anhydride, 120 $^{\circ}$ C, 3 h; (b) aqueous ammonia, reflux, 3 h; (c) benzaldehyde, acetic acid, reflux, 12 h; (d) POCl₃, 4-(dimethylamino)pyridine (DMAP), toluene, reflux, 5 h.

(Figure 1). From these, only mofezolac is used clinically as an analgesic drug and only in Japan.^{4,5} Other compounds, often abundantly used as reference compounds in *in vitro* and *in vivo* studies, have poor pharmacodynamic or pharmacokinetic profiles to enter clinical trials.^{4,5} Cingolani et al. studied the structural basis for COX-1 selectivity of two inhibitors, P6 and mofezolac (Figure 1).² They concluded that the COX-1

Table 1. First Series of Quinazoline Derivatives^a





^aReagents and conditions: (a) urea, 200 °C, 3 h; (b) POCl₃, N_1 , N_2 dimethylaniline, 120 °C, 4 h; (c) 4-methoxybenzylamine or 3,4-difluorobenzylamine, sodium acetate, THF, 65 °C, 3 h.

selectivity is due to their tighter fit within the COX-1 binding site caused by the nature of substituents on heterocycle core, while carboxylic acid and chlorine in mofezolac and P6, respectively, made inevitable contact with Arg120 and Tyr355 at the entrance to the catalytic domain.

Quinazolines are heterocyclic compounds with numerous therapeutic applications such as anti-inflammatory, analgesic, anticonvulsant, or anticancer applications. There are a few quinazoline compounds on the market used as anti-inflammatory drugs, for example, tryptanthrin, proquazone, or fluproquazone (Figure 2). Proquazone and fluproquazone were developed as third-generation NSAIDs with outstanding safety and efficacy.²⁰ Still, the potential of other quinazolines to inhibit COXs is weakly explored. Only several quinazolinones were identified as selective COX-2 inhibitors.^{21–26}

Therefore, three series of 2,4,7-substituted quinazolines as potential COX-1 inhibitors were designed and synthesized.



^aReagents and conditions: (a) corresponding amine or aniline, DMAP, triethylamine, THF, or toluene, reflux, 20 h.

Table 2. Second Series of Quinazoline Derivatives^a



entry	reactant	R ₃ (intermediate)	R_4	product (yield %)
1	5a	4-methoxyphenyl (6a)	morpholin-1-yl	7a (89.8)
2	5a	4-methoxyphenyl (6a)	pyrrolidin-1-yl	7 b (80.2)
3	5a	4-methoxyphenyl (6a)	piperazin-1-yl	7c (79.6)
4	5a	4-methoxyphenyl (6a)	diethylamino	7d (40.7)
5	5a	4-fluorophenyl (6b)	morpholin-1-yl	7e (94.9)
6	5a	4-fluorophenyl (6b)	pyrrolidin-1-yl	7f (96.6)
7	5a	4-fluorophenyl (6b)	piperazin-1-yl	7g (90.8)
8	5a	4-fluorophenyl (6b)	diethylamino	7h (34.7)
9	5a	thiophen-2-yl (6c)	morpholin-1-yl	7i (82.6)
10	5a	thiophen-2-yl (6c)	pyrrolidin-1-yl	7j (87.6)
11	5b	4-methoxyphenyl (6d)	morpholin-1-yl	7k (90.1)
12	5b	4-methoxyphenyl (6d)	pyrrolidin-1-yl	7 l (91.8)
13	5b	4-methoxyphenyl (6d)	piperazin-1-yl	7m (84.9)
14	5b	4-methoxyphenyl (6d)	diethylamino	7n (54.4)
15	5b	thiophen-2-yl (6e)	morpholin-1-yl	70 (90.5)

^{*a*}Reagents and conditions: (a) corresponding boronic acid or ester, K_2CO_3 , PdCl₂(PPh₃)₂, toluene/dioxane/water (10:5:8, v/v/v), 90 °C, overnight; (b) corresponding secondary amine, KI, K_2CO_3 , *N*,*N*diisopropylethylamine, dimethylformamide, 110 °C, overnight.

The derivatization of the quinazoline core in positions 2, 4, and 7 was chosen in order to resemble the shape of a letter "V", which is typical for numerous diarylheterocyclic COX-1 inhibitors. Although several of the selective COX-2 inhibitors (coxibs) also possess the diarylheterocyclic moiety, they always contain either a sulfonamide or methylsulfamoyl group. Instead, we decided to bet on the substitution with a styryl group to partly resemble the structure of stilbenes, a class of selective COX-1 inhibitors.²⁷ Other substituents were chosen with the aim to offer distinct electronic and steric properties of the final compounds as well as potential variability in H-bonding with the amino acids within the COX-1 binding site. The synthesized compounds were evaluated *in vitro* for their ability to inhibit COX-1 and COX-2 isoenzymes. The results were also supported by *in silico* modeling.

The synthesis of the first series started with the preparation of (E)-4-chloro-2-styrylquinazoline (2a) from the commercially available anthranilic acid (1a), following a literature procedure.²⁸ Shortly, the treatment of 1a with acetic anhydride, followed by the reaction with benzaldehyde in acetic acid gave an intermediate, which upon exposure to

Table 4. IC₅₀ Values for COX-1 and COX-2 Enzymes

compound	IC ₅₀ COX-1 (μM)	IC ₅₀ COX-2 (μM)	selectivity index (COX-2/ COX-1)
3b	1.57 ± 0.54	43.0 ± 10.3	27.4
3c	1.89 ± 0.63	37.3 ± 9.36	19.7
3k	1.90 ± 0.69	10.1 ± 1.38	5.32
3v	3.14 ± 0.99	>50	COX-1-selective
6c	0.376 ± 0.189	>50	COX-1-selective
6e	0.142 ± 0.014	>50	COX-1-selective
7 o	1.39 ± 0.72	>50	COX-1-selective
8a	0.78 ± 0.64	>50	COX-1-selective
8b	1.58 ± 0.89	>50	COX-1-selective
9a	0.141 ± 0.045	>50	COX-1-selective
9b	0.064 ± 0.044	>50	COX-1-selective
ibuprofen	2.19 ± 0.78	3.30 ± 0.96	1.51
SC-560	0.006 ± 0.003	1.03 ± 0.40	179.5

phosphoryl chloride (POCl₃) afforded (*E*)-4-chloro-2-styrylquinazoline (2a) (Scheme 1). Final compounds (3a-v) were prepared *via* amination of quinazoline 2a with variously substituted anilines or amines (Table 1).

The synthesis of the second series began with the preparation of 7-bromo-2,4-dichloroquinazoline (4) from the commercially available 2-amino-4-bromobenzoic acid (1b), by the reaction with urea followed by a treatment with POCl₃, according to published procedure (Scheme 2).²⁹ The step-by-step modification of three functional groups in quinazoline 4 afforded the desired final quinazolines. Shortly, the 4-chloro position was aminated using two different benzylamines to give the intermediates 5a,b (Scheme 2), which underwent a Suzuki cross-coupling reaction³⁰ of the 7-bromo position with three aromatic boronic acids to afford intermediates 6a–e. Finally, the 2-chloro position was aminated using four different secondary amines to give the final quinazoline derivatives 7a-o (Table 2).

Out of the synthesized compounds, seven exerted potent activity toward COX-1. For these, IC_{50} values were determined on both COX isoforms, while compounds exhibiting less than 50% inhibition on both COX isoforms were considered inactive (for percent inhibition, see Table 1S in the Supporting Information B). From the first series, three compounds (3b, 3c, 3k) exerted single-digit micromolar inhibitory activity toward COX-1, which was comparable with the inhibitory activity of the reference inhibitor ibuprofen, while their inhibitory activity toward COX-2 was rather moderate (Table 4). Their selectivity index was between 5 and 27. On the other hand, one compound from the first series (3v) and three compounds from the second series (6c, 6e, and 7o) were totally COX-1-selective, as they did not inhibit COX-2 even at



^{*a*}Reagents and conditions: (a) Corresponding aniline or benzylamine, DMAP, Et₃N, toluene, reflux, 3 h; (b) thiophene-2-boronic acid pinacol ester, K_2CO_3 , PdCl₂(PPh₃)₂, toluene/dioxane/water (10:5:8, v/v/v), 90 °C, overnight.



Figure 3. (A,C) **9b** in the binding site of COX-1. The amine functionality acts as a hydrogen bond donor (green arrow) to Tyr355. The yellow spheres represent hydrophobic contacts within the binding pocket. Within COX-1, **9b** assumes an orientation, where the fluorinated benzyl fills a pocket leading up to Met113 and the styryl moiety rests in a pocket leading up to Tyr385. The thiophene moiety adds additional stability by filling a channel leading to Phe518. (B,D) The key hydrogen bond interaction is lost in COX-2, where **9b** assumes a flipped orientation.

50 μ M. It should be noted that two of these COX-1-selective compounds, **6c** and **6e**, were just intermediates, and only one compound, **7o**, belonged to the final derivatives. In addition, these two intermediates exhibited IC₅₀ values in the nanomolar range (1 order of magnitude better than ibuprofen), while the IC₅₀ value of the final compound **7o** was in micromolar range, and two other corresponding final derivatives, **7i** and **7j**, were found inactive. From these results, it is obvious that the presence of an amine moiety in position 2 decreases substantially the inhibitory activity toward COX-1.

Structurally, all active compounds possessed either a chlorine or styryl group in position 2 and in position 4 was a phenyl or benzyl ring substituted in the *para*-position with an electron-donating group (F, CH₃, NH₂, OCH₃). Except for fluorine, which was tolerated also in the *meta*-position, the presence of other substituents, either an electron-donating (EDG) or electron-withdrawing (EWG), in the *meta*- or *ortho*-position caused inactivity in COX assays. Similarly, with aliphatic or heterocyclic substitution in position 4, the inhibitory activity vanished. Thiophene ring in position 7 significantly enhanced COX-1-selective inhibitory activity and was even indispensable for this activity when the styryl group in position 2 was missing.

Based on these results, two other quinazoline derivatives were prepared as examples of a third series. A combination of the substituents from the active compounds of both previous series was employed. Thus, the styryl group was placed in position 2, a thiophene ring was placed in position 7, and position 4 was substituted with 3,4-difluorobenzylamine or 4-methylaniline (Table 3). The synthesis followed the procedure for the first series,²⁸ while the starting material from the second series, 2-amino-4-bromobenzoic acid (1b), was used. Thus, (*E*)-7-bromo-4-chloro-2-styrylquinazoline 2b was formed (Scheme 1). The reaction of 2b with 3,4-difluorobenzylamine or 4-methylaniline, respectively, afforded intermediates 8a,b, which upon Suzuki cross-coupling³⁰ with thiophene-2-boronic acid pinacol ester gave the final quinazolines 9a,b (Table 3).

Both intermediates 8a,b as well as both final compounds 9a,b exerted selective COX-1 inhibitory activity. As expected, the IC₅₀ values of intermediates 8a,b were comparable with ibuprofen, as their structure resembled that of the final compounds of the first series, 3b and 3k (Table 4). Moreover, the comparison of the IC₅₀ values of the final compounds 9a,b (141/64 nM, respectively) with 6c and 6e (376 and 142 nM, respectively) demonstrated the importance of the 2-styryl group for the inhibitory activity toward COX-1. Furthermore, similarly to compounds of the second series, the presence of the thiophene ring in compounds 9a,b significantly augmented the COX-1 inhibitory activity, causing a decrease in IC₅₀ values by 1 to 2 orders of magnitude, with the best value observed for compound **9b** (IC₅₀ = 64 nM). Even though this value was 1 order of magnitude higher than the IC₅₀ value determined for the reference COX-1-selective inhibitor, SC-560, such activity together with total COX-1 selectivity renders quinazoline 9b as



Figure 4. (A,C) **3k** also forms the key hydrogen bond with Tyr355 in COX-1. The fluorinated benzyl group is oriented toward Met113, while the styryl moiety fills the channel leading up to Tyr385. (B,D) In COX-2, the molecule is flipped, but the pyrimidine ring can act as a hydrogen bond acceptor to Tyr355, explaining the residual COX-2 activity in the smaller molecules of the first series. Here, the fluorinated benzyl group fills the channel leading up to Tyr385, while the styryl is oriented toward Phe518. The yellow spheres represent hydrophobic contacts with the binding pocket.

a promising compound for further evaluation of its biological activity and pharmacological profile.

To determine whether the synthesized compounds compete with the substrate (AA) in binding to the binding site of the COX-1 enzyme, **9b** was selected as a representative compound and tested in COX-1 assays with different concentrations of AA. The ability of compound **9b** to inhibit COX-1 was reduced when the concentration of AA increased in the reaction. IC₅₀ values increased from 21.1 nM in the presence of 250 nM AA, through 72.8 nM in the presence of 1250 nM AA, to 254 nM in the presence of 6250 nM AA. It suggests that compound **9b** competed with AA in binding to COX-1. Ibuprofen, which is a known competitive COX-1 inhibitor,³¹ also exhibited such a tendency, as its IC₅₀ values increased from 4.00 μ M with 250 nM AA to 70.1 μ M with 6250 nM AA (Table 2S in the Supporting Information B).

In order to elucidate the structural causes of the selective binding mechanism, all compounds (3-9) were docked into the binding sites of COX-1 using the pdb entry 3n8w, cocrystallized with flurbiprofen,³² and COX-2 using the pdb entry 6COX, cocrystallized with SC-558.³³ An analysis of the resulting docking poses revealed a hydrogen bridge between the secondary amine functionality and Tyr355, a key residue for electron transfer in the COX mechanism of action, as the primary indicator of activity within the compound class. The

most active quinazoline derivative **9b** is placed in the COX-1 binding site so that the amine functionality is located within binding vicinity of Tyr355. The three ring substituents fill different hydrophobic parts of the pocket. The fluorinated benzyl fills a pocket leading up to Met113, and the styryl moiety rests in a pocket leading up to Tyr385. The thiophene moiety is oriented toward Phe518 (Figure 3). In the binding site of COX-2, compound **9b** is flipped and can no longer interact with Tyr355 (Figure 3). The loss of this key interaction could explain the loss of *in vitro* activity.

A similar effect, but less pronounced, occurs for the smaller molecules from the first series, which still display dual activity. Compound **3k** binds to COX-1 in the same orientation as **9b**, also forming the hydrogen bond with Tyr355, but as it lacks the thiophene ring, the channel leading to Phe518 is empty causing a reduced activity. In COX-2, the molecule is again flipped, but due to the smaller size, the pyrimidine ring can act as an interaction partner for Tyr355 (Figure 4).

In summary, three series of quinazoline derivatives were synthesized, and their inhibitory activity toward COX-1 and COX-2 isoenzymes was evaluated *in vitro*. From the synthesized compounds, 11 quinazoline derivatives exhibited good to excellent inhibitory activity toward COX-1 (IC₅₀ = 0.064–3.14 μ M). Out of these, seven compounds did not inhibit a COX-2 isoform even at 50 μ M, making these

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compounds totally COX-1-selective. The IC₅₀ values of the most active compounds were 1 to 2 orders of magnitude lower than the inhibitory activity of the reference compound, ibuprofen (IC₅₀ = 2.19 μ M). According to our results, the compounds compete with the substrate in the binding to COX-1 binding site. All the active compounds possessed in the 4-position of the quinazoline ring either para-EDG-substituted aniline or benzylamine and in the 2-position a chlorine or styryl group. The presence of a thiophene ring in the 7-position markedly enhanced the inhibitory activity as well as COX-1 selectivity. In the docking study, it was shown that the activity hinges on the formation of a hydrogen bond between the secondary amine and key enzyme residue Tyr355. This interaction is only formed in the binding site of COX-1 and thus may be the cause of the selectivity.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsmedchemlett.1c00004.

Supporting Information A: Detailed synthetic procedures, complete spectroscopic data of synthesized compounds, experimental procedure of docking experiments and presentation of docking scores, and experimental procedure of the determination of COX inhibition (PDF)

Supporting Information B: ¹H and ¹³C NMR spectra of all newly synthesized compounds, percentual inhibition of COX-1 and COX-2 isoenzymes by tested compounds at 20 μ M, COX-1 inhibition by compound **9b** and ibuprofen at different substrate concentrations, percentual inhibition of COX-1 by active compounds at different concentrations graphically depicted as % inhibition vs log(concentration), UPLC/UV–vis chromatograms of the active compounds showing their purity (PDF)

AUTHOR INFORMATION

Corresponding Author

Marcela Dvorakova – Laboratory of Plant Biotechnologies, Czech Academy of Sciences, Institute of Experimental Botany, 165 02 Prague 6 - Lysolaje, Czech Republic; o orcid.org/ 0000-0002-6664-6870; Email: dvorakova@ueb.cas.cz

Authors

- Lenka Langhansova Laboratory of Plant Biotechnologies, Czech Academy of Sciences, Institute of Experimental Botany, 165 02 Prague 6 - Lysolaje, Czech Republic; orcid.org/ 0000-0002-3689-8786
- Veronika Temml Department of Pharmaceutical and Medicinal Chemistry, Paracelsus Medical University of Salzburg, 5020 Salzburg, Austria; orcid.org/0000-0001-7662-9882
- Antonio Pavicic Laboratory of Plant Biotechnologies, Czech Academy of Sciences, Institute of Experimental Botany, 165 02 Prague 6 - Lysolaje, Czech Republic
- **Tomas Vanek** Laboratory of Plant Biotechnologies, Czech Academy of Sciences, Institute of Experimental Botany, 165 02 Prague 6 - Lysolaje, Czech Republic
- **Premysl Landa** Laboratory of Plant Biotechnologies, Czech Academy of Sciences, Institute of Experimental Botany, 165

02 Prague 6 - Lysolaje, Czech Republic; ^(a) orcid.org/0000-0003-0364-2247

Complete contact information is available at: https://pubs.acs.org/10.1021/acsmedchemlett.1c00004

Author Contributions

M.D. synthesized the compounds and wrote the paper, V.T. did the molecular docking studies, L.L., A.P., and P.L. carried out the COX inhibition experiments, P.L. did the substrate competition experiment, and T.V. supervised the work.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

The authors gratefully acknowledge the financial support by the grant LTC17048 from the Ministry of Education, Youth and Sports of the Czech Republic. V.T. is funded by a FWF Hertha Firnberg fellowship (T-942).

ABBREVIATIONS

AA, arachidonic acid; COX, cyclooxygenase; DMAP, 4-(dimethylamino)pyridine; EDG, electron-donating group; EWG, electron-withdrawing group; NSAIDs, nonsteroidal anti-inflammatory drugs; PDB, protein data bank; PdCl₂(PPh₃)₂, bis(triphenylphosphine)palladium(II) dichloride; PGE2, prostaglandin E2; PGI2, prostacyclin; POCl₃, phosphoryl chloride; THF, tetrahydrofuran; TXA2, thromboxane A2

REFERENCES

(1) Lu, G.; Tsai, A. L.; Van Wart, H. E.; Kulmacz, R. J. Comparison of the Peroxidase Reaction Kinetics of Prostaglandin H Synthase-1 and -2. *J. Biol. Chem.* **1999**, *274*, 16162–16167.

(2) Cingolani, G.; Panella, A.; Perrone, M. G.; Vitale, P.; Di Mauro, G.; Fortuna, C. G.; Armen, R. S.; Ferorelli, S.; Smith, W. L.; Scilimati, A. Structural Basis for Selective Inhibition of Cyclooxygenase-1 (COX-1) by Diarylisoxazoles Mofezolac and 3-(5-Chlorofuran-2-yl)-5-methyl-4-phenylisoxazole (P6). *Eur. J. Med. Chem.* **2017**, *138*, 661–668.

(3) Desai, S. J.; Prickril, B.; Rasooly, A. Mechanisms of Phytonutrient Modulation of Cyclooxygenase-2 (COX-2) and Inflammation Related to Cancer. *Nutr. Cancer* **2018**, *70* (3), 350–375.

(4) Perrone, M. G.; Scilimati, A.; Simone, L.; Vitale, P. Selective COX-1 inhibition: A Therapeutic Target to Be Reconsidered. *Curr. Med. Chem.* **2010**, *17*, 3769–3805.

(5) Perrone, M. G.; Lofrumento, D. D.; Vitale, P.; De Nuccio, F.; La Pesa, V.; Panella, A.; Calvello, R.; Cianciulli, A.; Panaro, M. A.; Scilimati, A. Selective Cyclooxygenase-1 Inhibition by P6 and Gastrotoxicity: Preliminary Investigation. *Pharmacology* **2015**, *95*, 22–28.

(6) Ornelas, A.; Zacharias-Millward, N.; Menter, D. G.; Davis, J. S.; Lichtenberger, L.; Hawke, D.; Hawk, E.; Vilar, E.; Bhattacharya, P.; Millward, S. Beyond COX-1: The Effects of Aspirin on Platelet Biology and Potential Mechanisms of Chemoprevention. *Cancer Metastasis Rev.* **2017**, *36*, 289–303.

(7) Choi, S. H.; Aid, S.; Choi, U.; Bosetti, F. Cyclooxygenases-1 and -2 Differentially Modulate Leukocyte Recruitment into the Inflamed Brain. *Pharmacogenomics J.* **2010**, *10*, 448–457.

(8) Khan, A. A.; Iadarola, M.; Yang, H. Y.; Dionne, R. A. Expression of COX-1 and COX-2 in a Clinical Model of Acute Inflammation. *J. Pain* **2007**, *8*, 349–354.

(9) Li, X.; Mazaleuskaya, L. L.; Ballantyne, L. L.; Meng, H.; FitzGerald, G. A.; Funk, C. D. Genomic and Lipidomic Analyses

ACS Medicinal Chemistry Letters

Differentiate the Compensatory Roles of Two COX Isoforms During Systemic Inflammation in Mice. J. Lipid Res. 2018, 59, 102–112.

(10) Pannunzio, A.; Coluccia, M. Cyclooxygenase-1 (COX-1) and COX-1 Inhibitors in Cancer: A Review of Oncology and Medicinal Chemistry Literature. *Pharmaceuticals* **2018**, *11*, 101.

(11) Perrone, M. G.; Luisi, O.; De Grassi, A.; Ferorelli, S.; Cormio, G.; Scilimati, A. Translational Theragnosis of Ovarian Cancer: Where Do We Stand? *Curr. Med. Chem.* **2020**, *27*, 5675–5715.

(12) Vitale, P.; Panella, A.; Scilimati, A.; Perrone, M. G. COX-1 Inhibitors: Beyond Structure Toward Therapy. *Med. Res. Rev.* 2016, 36, 641–671.

(13) Belton, O.; Fitzgerald, D. J. Cyclooxygenase Isoforms and Atherosclerosis. *Expert. Rev. Mol. Med.* **2003**, *5*, 1–18.

(14) Loftin, C. D.; Trivedi, D. B.; Langenbach, R. Cyclooxygenase-1selective Inhibition Prolongs Gestation in Mice without Adverse Effects on the *Ductus Arteriosus*. J. Clin. Invest. **2002**, 110, 549–557.

(15) Calvello, R.; Lofrumento, D. D.; Perrone, M. G.; Cianciulli, A.; Salvatore, R.; Vitale, P.; De Nuccio, F.; Giannotti, L.; Nicolardi, G.; Panaro, M. A.; Scilimati, A. Highly Selective Cyclooxygenase-1 Inhibitors P6 and Mofezolac Counteract Inflammatory State both *in vitro* and *in vivo* Models of Neuroinflammation. *Front. Neurol.* 2017, 8, 00251.

(16) Calvello, R.; Panaro, M. A.; Carbone, M. L.; Cianciulli, A.; Perrone, M. G.; Vitale, P.; Malerba, P.; Scilimati, A. Novel Selective COX-1 Inhibitors Suppress Neuroinflammatory Mediators in LPSstimulated N13 Microglial Cells. *Pharmacol. Res.* **2012**, *65*, 137–148.

(17) Meek, I. L.; Vonkeman, H. E.; Kasemier, J.; Movig, K. L.; van de Laar, M. A. Interference of NSAIDs with the Thrombocyte Inhibitory Effect of Aspirin: A Placebo-Controlled, ex vivo, Serial Placebo-Controlled Serial Crossover Study. *Eur. J. Clin. Pharmacol.* **2013**, *69*, 365–371.

(18) Mitchell, J. A.; Shala, F.; Elghazouli, Y.; Warner, T. D.; Gaston-Massuet, C.; Crescente, M.; Armstrong, P. C.; Herschman, H. R.; Kirkby, N. S. Cell-Specific Gene Deletion Reveals the Antithrombotic Function of COX1 and Explains the Vascular COX1/Prostacyclin Paradox. *Circ. Res.* **2019**, *125*, 847–854.

(19) Roodhart, J. M.; Daenen, L. G.; Stigter, E. C.; Prins, H. J.; Gerrits, J.; Houthuijzen, J. M.; Gerritsen, M. G.; Schipper, H. S.; Backer, M. J.; van Amersfoeort, M.; Vermaat, J. S. P.; Moerer, P.; Ishihara, K.; Kalkhoven, E.; Beijnen, J. H.; Derksen, P. W. B.; Medema, R. H.; Martens, A. C.; Brenkman, A. B.; Voest, E. E. Mesenchymal Stem Cells Induce Resistance to Chemotherapy through the Release of Platinum-Induced Fatty Acids. *Cancer Cell* **2011**, *20*, 370–383.

(20) Chandrika, P. M.; Yakaiah, T.; Rao, A. R.; Narsaiah, B.; Reddy, N. C.; Sridhar, V.; Rao, J. V. Synthesis of Novel 4,6-Disubstituted Quinazoline Derivatives, Their Anti-Inflammatory and Anti-Cancer Activity (Cytotoxic) Against U937 Leukemia Cell Lines. *Eur. J. Med. Chem.* **2008**, 43, 846–852.

(21) Abdel-Aziz, A. A.; Abou-Zeid, L. A.; ElTahir, K. E. H.; Ayyad, R. R.; El-Sayed, M. A.; El-Azab, A. S. Synthesis, Anti-Inflammatory, Analgesic, COX-1/2 Inhibitory Activities and Molecular Docking Studies of Substituted 2-Mercapto-4(3H)-Quinazolinones. *Eur. J. Med. Chem.* **2016**, *121*, 410–421.

(22) El-Feky, S. A.; Imran, M.; Nayeem, N. Design, Synthesis, and Anti-Inflammatory Activity of Novel Quinazolines. *Orient. J. Chem.* **2017**, 33, 707–716.

(23) Aboraia, A. S.; Hara, M. A.; Abdelrahman, M. H.; Amin, M. M.; El-Sabbaghab, O. I. Design, Synthesis and COX1/2 Inhibitory Activity of New Quinazoline-5-one Derivatives. *J. Adv. Chem.* **2017**, *13*, 5923–5931.

(24) Kavitha, K.; Srinivasan, N.; Hari Babu, Y.; Suresh, R. Synthesis and Molecular Docking Study of Novel 2-Phenyl Quinazoline-4-(3*H*)-one Derivative as COX-2 Inhibitor. *Indo Am. J. Pharm. Sci.* **2019**, *06*, 4032–4037.

(25) Kumar, A.; Sharma, S.; Archana; Bajaj, K.; Sharma, S.; Panwar, H.; Singh, T.; Srivastava, V. K. Some New 2,3,6-Trisubstituted Quinazolinones as Potent Anti-Inflammatory, Analgesic and COX-II Inhibitors. *Bioorg. Med. Chem.* **2003**, *11*, 5293–5299.

(26) Sakr, A.; Kothayer, H.; Ibrahim, S. M.; Baraka, M. M.; Rezq, S. 1,4-Dihydroquinazolin-3(2*H*)-yl Benzamide Derivatives as Anti-Inflammatory and Analgesic Agents with an Improved Gastric Profile: Design, Synthesis, COX-1/2 Inhibitory Activity and Molecular Docking Study. *Bioorg. Chem.* **2019**, *84*, 76–86.

(27) Dvorakova, M.; Landa, P. Anti-Inflammatory Activity of Natural Stilbenoids: A Review. *Pharmacol. Res.* **2017**, *124*, 126–145. (28) Okano, M.; Mito, J.; Maruyama, Y.; Masuda, H.; Niwa, T.; Nakagawa, S.; Nakamura, Y.; Matsuura, A. Discovery and Structure-Activity Relationships of 4-Aminoquinazoline Derivatives, A Novel Class of Opioid Receptor Like-1 (ORL1) Antagonists. *Bioorg. Med. Chem.* **2009**, *17*, 119–132.

(29) Jiang, Y.; Chen, A. C.; Kuang, G. T.; Wang, S. K.; Ou, T. M.; Tan, J. H.; Li, D.; Huang, Z. S. Design, Synthesis and Biological Evaluation of 4-Anilinoquinazoline Derivatives as New c-Myc G-Quadruplex Ligands. *Eur. J. Med. Chem.* **2016**, *122*, 264–279.

(30) Miyaura, N.; Suzuki, A. Palladium-Catalyzed Cross-Coupling Reactions of Organoboron Compounds. *Chem. Rev.* **1995**, *95*, 2457–2483.

(31) Knights, K. M.; Mangoni, A. A.; Miners, J. O. Defining the COX Inhibitor Selectivity of NSAIDs: Implications for Understanding Toxicity. *Expert Rev. Clin. Pharmacol.* **2010**, *3*, 769–776.

(32) Sidhu, R. S.; Lee, J. Y.; Yuan, C.; Smith, W. L. Comparison of Cyclooxygenase-1 Crystal Structures: Cross-Talk between Monomers Comprising Cyclooxygenase-1 Homodimers. *Biochemistry* **2010**, *49* (33), 7069–7079.

(33) Kurumbail, R. G.; Stevens, A. M.; Gierse, J. K.; McDonald, J. J.; Stegeman, R. A.; Pak, J. Y.; Gildehaus, D.; iyashiro, J. M.; Penning, T. D.; Seibert, K.; Isakson, P. C.; Stallings, W. C. Structural Basis for Selective Inhibition of Cyclooxygenase-2 by Anti-Inflammatory Agents. *Nature* **1996**, *384*, 644–648.