


Detection of *Ehrlichia* spp. and *Theileria* spp. in *Hyalomma anatolicum* ticks collected in Tajikistan

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Abstract. The objectives of our study were to survey the prevalence of genetic markers for *Rickettsia* spp., *Ehrlichia* spp., *Anaplasma* spp., *Babesia* spp., and *Theileria* spp. in *Hyalomma anatolicum* ticks collected in southwestern Tajikistan and to perform sequencing and phylogenetic analysis of fragments of the 16S rRNA gene and *groESL* operon from *Ehrlichia* spp. and fragments of the 18S rRNA gene of *Theileria* spp. detected in *H. anatolicum* ticks. *Hyalomma anatolicum* ticks collected in the Tursunzade and Rudaki districts of Tajikistan were tested for DNA of *Rickettsia* spp., *Ehrlichia* spp., *Anaplasma* spp., *Babesia* spp., and *Theileria* spp. by PCR with specific primers. The amplified fragments were sequenced and analyzed. DNA of *Ehrlichia* spp. (3.3 %) and *Theileria* spp. (3.3 %) was detected only in *H. anatolicum* ticks collected from the Rudaki district, and DNA of *Ehrlichia* spp. (0.7 %) was found in *H. anatolicum* ticks from the Tursunzade district. Sequence analysis of fragments of the 16S rRNA gene and *groESL* operon from *Ehrlichia* spp. revealed high similarity to *Ehrlichia* spp. The Tajik isolates of *Theileria* spp. were genotyped as *Theileria annulata* based on the analysis of 18S rRNA gene sequences. The phylogenetic analysis demonstrates that *Ehrlichia* spp. isolates are highly similar to *Ehrlichia* spp. circulating in China and Brazil. The isolate Tajikistan-5 is closely related to the putative novel species *Ehrlichia mineirensis*. The Tajik isolates of *Theileria* spp. were clustered with *T. annulata* isolates from Turkey, Iran, Pakistan, and China by phylogenetic analyses.

Key words: *Hyalomma anatolicum*; tick-borne infections; *Ehrlichia* spp.; *Theileria* spp.; Tajikistan.

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Выявление *Ehrlichia* spp. и *Theileria* spp. в клещах *Hyalomma anatolicum*, собранных в Таджикистане

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Аннотация. Исследовано наличие генетического материала *Rickettsia* spp., *Ehrlichia* spp., *Anaplasma* spp., *Babesia* spp. и *Theileria* spp. в клещах *Hyalomma anatolicum*, собранных в юго-западной части Таджикистана, с последующим секвенированием и филогенетическим анализом фрагментов 16S рРНК гена и *groESL* оперона для *Ehrlichia* spp. и фрагмента гена 18S рРНК для *Theileria* spp., обнаруженного в изученных клещах. Клещи *H. anatolicum* были собраны в районах Турсунзаде и Рудаки и исследованы с использованием специфичных праймеров с помощью ПЦР на наличие генетического материала *Rickettsia* spp., *Ehrlichia* spp., *Anaplasma* spp., *Babesia* spp. и *Theileria* spp. Выделенные ПЦР-фрагменты генов 16S рРНК, *groESL* оперона и 18S рРНК были секвенированы и проведен их филогенетический анализ с целью генотипирования обнаруженных изолятов клещевых патогенов. В клещах, собранных в районе Рудаки, обнаружена ДНК *Ehrlichia* spp. (3.3 %) и *Theileria* spp. (3.3 %), а в Турсунзаде – ДНК *Ehrlichia* spp. (0.7 %). Секвенирование фрагментов гена 16S рРНК и *groESL* оперона *Ehrlichia* spp. показало высокий уровень гомологии нуклеотидной последовательности с известными последовательностями *Ehrlichia* spp. Таджикские изоляты *Theileria* spp. были генотипированы как *Theileria annulata* на основе анализа последовательности гена 18S рРНК. Филогенетический анализ показал,

что таджикские изоляты *Ehrlichia* spp. очень близки с изолятами *Ehrlichia* spp., циркулирующими в Китае и Бразилии. Изолят Таджикистан-5 кластеризуется с предполагаемым новым видом *Ehrlichia mineirensis*. Таджикские изоляты *Theileria* spp. были генетически схожи с вариантами *T. annulata*, циркулирующими в Турции, Иране, Пакистане и Китае.

Ключевые слова: *Hyalomma anatolicum*; клещевые инфекции; *Ehrlichia* spp.; *Theileria* spp.; Таджикистан.

Introduction

Ixodid ticks transmit various pathogens to both humans and animals in Asia (Tishkova et al., 2012; Wu et al., 2013). Twenty-three species of ixodid ticks have been described in this region of Central Asia, with the predominant ixodid tick species being *Hyalomma anatolicum* Koch, 1844 (Rasulov, 2007). The Crimean-Congo hemorrhagic fever, Sindbis, and Wad Medani viruses were previously detected in ixodid ticks in Tajikistan and other Asian countries (Begum et al., 1970; Gresikova et al., 1978; Petrova et al., 2013). *Hyalomma anatolicum* ticks are also known to transmit bacterial and parasite infections such as Lyme disease, babesiosis, piroplasmosis, theileriosis, and anaplasmosis (Tishkova et al., 2012; Wu et al., 2013). *Theileria annulata* (Piroplasmida: Family Theileriidae, Genus *Theileria*) is the causative agent of theileriosis in domestic animals, which is transmitted by 15 species of ixodid ticks of the genus *Hyalomma* (Robinson, 1982). *Ehrlichia* spp. (Family Anaplasmataceae, Genus *Ehrlichia*) are intracellular Gram-negative bacteria, ecologically associated with ixodid ticks and their animal hosts (Parola et al., 2001). The pathogenicity to domestic and wild animals, as well as to humans, has been demonstrated in *Ehrlichia canis*, *E. chaffeensis*, *E. ewingii*, *E. muris* and *E. ruminantium* (Aguiar et al., 2014; Cabezas-Cruz et al., 2014). Currently, there are no published studies on genetic markers and genotyping of *Rickettsia* spp., *Ehrlichia* spp., *Anaplasma* spp., *Babesia* spp., and *Theileria* spp. in *H. anatolicum* ticks in Tajikistan.

The objectives of this study were to survey the prevalence of genetic markers for these tick-borne infections in *H. anatolicum* ticks collected in southwestern Tajikistan, and to perform sequence and phylogenetic analysis of *Ehrlichia* spp. and *Theileria* spp. detected in the ticks.

Materials and methods

Tick harvesting. Adult ticks were collected from domestic animals in several villages of the Rudaki district (Somoniyon N 38°26'27", E 68°46'28") and the Tursunzade district (Tursunzade N 38°30'39", E 68°13'49") in southwestern Tajikistan in July 2009 (Fig. 1). The ticks were transported and samples for analysis were prepared as described in (Petrova et al., 2013). Tick species were identified by morphological examination with subsequent confirmation by PCR and sequencing of PCR products of a 16S rRNA fragment of the mitochondrial genome of the ticks.

PCR detection of genetic markers. DNA was isolated from tick homogenates by phenol/chloroform extraction using a commercial kit (Lytech, Moscow, Russia) following manufacturer's instructions. It was kept at -20 °C until use. The genetic markers of *Rickettsia* spp., *Ehrlichia* spp., *Anaplasma* spp., *Babesia* spp. and *Theileria* spp. in ticks were detected by PCR with specific primers (see the Table). The PCR fragments were purified using Wizard SV Gel and a PCR Clean-Up System kit (Promega, USA) according to manu-

facturer's instructions. All PCR fragments were sequenced in a 3130 Genetic Analyzer automated capillary sequencer (Applied Biosystems Inc.). DNA sequencing reactions were performed with BigDyeTerminator v3.1 Cycle Sequencing Kits (Applied BioSystems, USA). Both strands of each gene fragment were directly sequenced; each sample was sequenced twice. Precautions were taken at all steps of analysis to avoid cross-contamination among samples.

Nucleotide sequences and phylogenetic analyses. DNA sequences were compared with sequences available in GenBank using the Basic Local Alignment Search Tool (BLAST) on <http://blast.ncbi.nlm.nih.gov>. Evolutionary analyses were conducted with MEGA5 software (Tamura et al., 2011). Multisequence alignments were performed using ClustalX. For each analyzed gene a phylogram was constructed by the maximum likelihood method. Phylogenetic distances between homologous sequences were calculated using Kimura's two-parameter model. Confidence levels for individual branches of the resulting tree were determined by bootstrap analysis with 1000 replicates.

Results and discussion

Tick harvesting

Adult *H. anatolicum* ticks (138 females and 244 males) were collected and grouped in 137 pools. Tick species were identified by sequencing a fragment of 16S rRNA mitochondrial gene for all pools. Two original variants of 16S rRNA mitochondrial gene fragment sequences found in these ticks were submitted to GenBank (accession numbers KP059123 and KP059124). The nucleotide fragments showed 99.9 % similarity to the corresponding *H. anatolicum* sequences from GenBank. These tick pools were tested by PCR for genetic markers of *Rickettsia* spp., *Ehrlichia* spp., *Anaplasma* spp., *Babesia* spp., and *Theileria* spp. and other ticks were used for genotyping. Of those ticks, 290 (179 males and 111 females)



Fig. 1. Tick collection sites in Tajikistan: 1, Tursunzade district; 2, Rudaki district.

Primers used for PCR identification of ticks and tick-borne infections in the present study

Species detected	Gene	Primers	Primer sequence (5'→3')	Amplicon size, bp	Reference
<i>Hyalomma</i> spp. (ticks)	16S rRNA gene (mitochondrial)	H16Mf	GATTCTCATCGGTCTAACTCAG	425	This study
		H16Mr	AGTATTTTGACTATACAAGGTATTG		
<i>Rickettsia</i> spp.	<i>gltA</i>	CS409d	CCTATGGCTATTATGCTTGC	765	Roux et al., 1997
		RP1258n	ATTGCAAAAAGTACAGTGAACA		
<i>Babesia</i> spp./ <i>Theileria</i> spp.	18S rRNA gene	BS1	GACGGTAGGGTATTGGCCT	1120	Rar et al., 2005
		BS2	ATTCACCGGATCACTCGATC		
<i>Ehrlichia</i> spp./ <i>Anaplasma</i> spp.	16S rRNA gene	Erl1	GAACGAACGCTGGCGGCAAGC	1375	Rar et al., 2010
		Erl6	GACCCAACCTTAAATGGCTGC		
		Erl7	TAACACATGCAAGTCGAACG		
		Erl8	CTTCGAGTTAAGCCAATTCC		
	<i>groESL</i> operon	HS1-f	CGYCACTGGGCTGTAATGAA	1340	Sumner et al., 1997
		HS6-r	CCWCCWGGTACWACACCTTC		

were collected in the Tursunzade district and 92 (65 males and 27 females) from the Rudaki district. The PCR tests for *Ehrlichia* spp. and *Theileria* spp. were positive in the range 0.7–3.3%. The PCR tests for *Rickettsia* spp., *Anaplasma* spp., and *Babesia* spp. were negative in all tick samples.

Theileria identification

Theileria spp. was detected in 3.3% ticks from Rudaki but not in ticks from Tursunzade. The amplified PCR fragments of 18S rRNA (1090–1092 bp) were isolated and sequenced (GenBank accessions KM288517–KM288519). The sequences were 100% identical to isolates of *Theileria annulata* circulating in Turkey (AY508463) and Iran (KF429799, HM628581), similar by 99.9% to isolated from Pakistan (JQ743630) and China (EU073963) and by 99.7% to isolates from Spain (DQ287944). Phylogenetic analysis confirmed that *Theileria* spp. isolates from the Rudaki district of Tajikistan belonged to *Th. annulata* (Fig. 2). The analysis of 18S rRNA gene fragment for three isolates of *Th. annulata* from southwestern Tajikistan showed that all isolates were genetically identical (100% similarity).

Ehrlichia identification

The presence of *Ehrlichia* spp. has not been previously documented in ticks and animals in Tajikistan. DNA of *Ehrlichia* spp. was detected in five pools of *H. anatolicum* ticks collected in the Rudaki and Tursunzade districts. The infection rates for *Ehrlichia* spp. were 3.3% in Rudaki and 0.7% in Tursunzade. The fragments of the 16S rRNA gene (1291–1352 bp) and *groESL* operon (1248–1315 bp) were sequenced (KM995818–KM995821, KP059122, KJ930191–KJ930195). The nucleotide sequences of 16S rRNA gene fragments were highly conserved (99.5–100%) among studied isolates. The similarity levels of the studied 16S rRNA fragments to *E. chaffeensis* (CP007478), *E. canis* (KJ513197), and *E. muris* (NR121714) were 99.2, 99.2, and 99.3%, respectively.

The phylogenetic tree generated using *Ehrlichia* spp. *groESL* operon fragment sequences was markedly different

from the tree based on 16S rRNA sequences (Fig. 2, b, c). The 16S rRNA gene fragment analysis (1140 nucleotides) showed that all isolated *Ehrlichia* spp. were genetically close (see Fig. 2, b). The studied isolates grouped in the same branch of the phylogenetic tree as isolates from the Fujian province in Southeastern China (DQ324547) and the Tibet Autonomous Region of China (AF414399). The Tibetan isolate was grouped with the *E. canis* branch, which is genetically close to the species *E. chaffeensis*, pathogenic for humans (Wen et al., 2002). We note that the Tajik isolates were most similar to Chinese isolates from regions of China that do not border Tajikistan. The phylogenetic tree generated using *Ehrlichia* spp. *groESL* operon fragment sequences was markedly different from the tree based on 16S rRNA sequences (see Fig. 2, c). The nucleotide sequences of *groESL* operon *Ehrlichia* spp. found in Tajikistan are separated into three groups. Tajikistan 1 and 2 isolates were closest to two isolates *Ehrlichia* spp. from different regions of China (Xinjiang, *Hyalomma asiaticum*; Yunnan, *Rhipicephalus microplus*), Tajikistan 3 and 4 cluster with a different Chinese isolate (Xinjiang, *Hyalomma asiaticum*). Tajikistan 5 showed high similarity to *Ehrlichia* spp. (JX629806) isolated in Brazil from a *Rhipicephalus microplus* tick (Cruz et al., 2012). Tajikistan 5 has 13 nucleotide and 2 amino acid substitutions in comparison to the Brazilian isolate. The American isolate was previously identified as a new species of *Ehrlichia* spp. named *E. mineirensis*. It causes clinical manifestations associated with ehrlichiosis in experimentally infected calf (Aguiar et al., 2014).

Tajikistan 1–4 isolates clustered with Chinese isolates from Xinjiang and Yunnan Provinces. Xinjiang Province shares borders with Tajikistan in southwestern China, unlike Yunnan. Tajikistan 5 isolate was the most genetically distinct from other *Ehrlichia* spp. grouping with UFMG-EV and UFMT-BV isolates from Brazil and BOV2010 isolate from Canada (Gajadhar et al., 2010; Aguilar et al., 2014; Cabezas-Cruz et al., 2014). We infer that Tajikistan 5 isolate belongs to the putative novel species of *Ehrlichia* spp. previously named *E. mineirensis*.

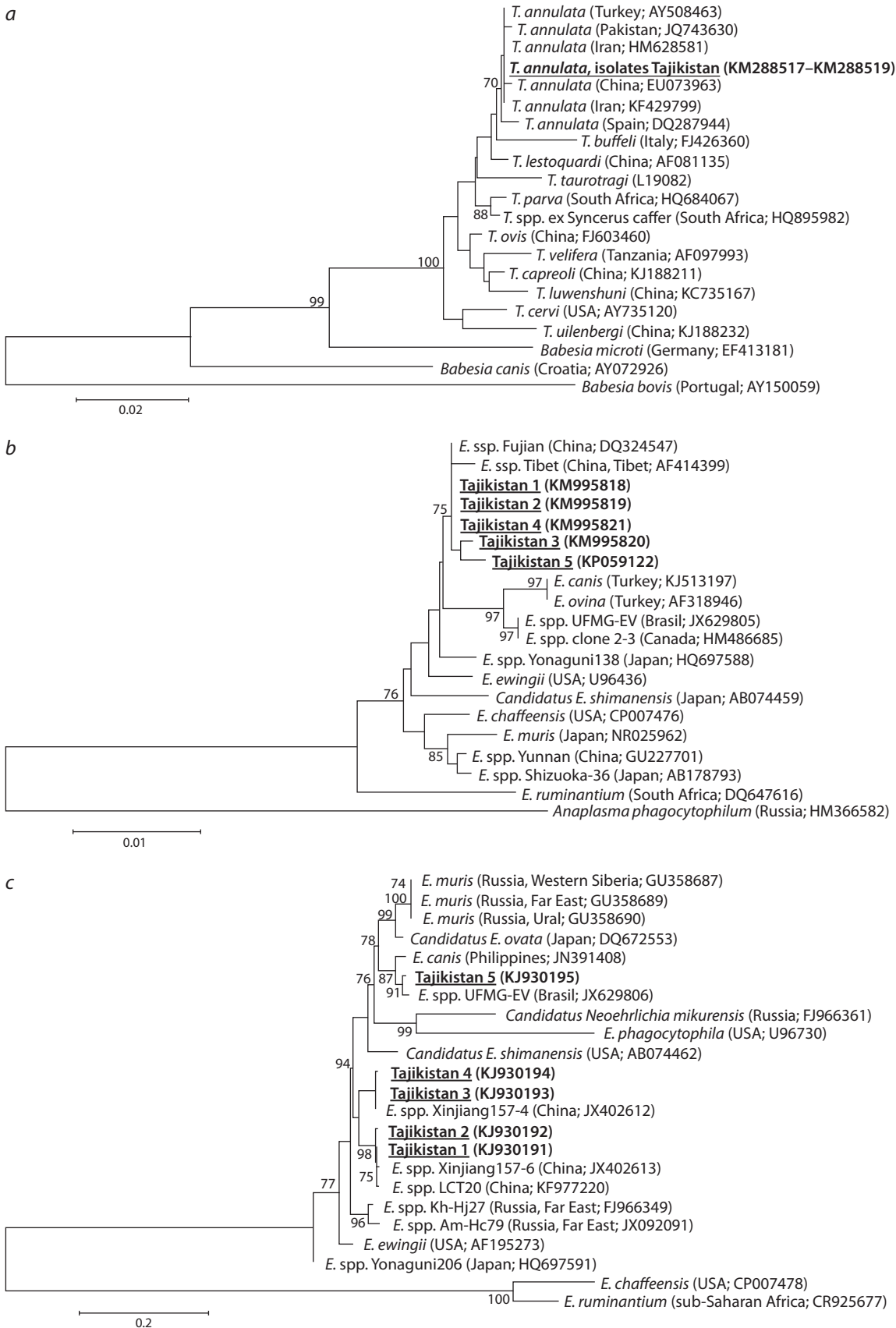


Fig. 2. Phylogenetic tree of *Theileria* spp. and *Ehrlichia* spp. isolates: *a*, based on 18S rRNA of *Theileria* spp.; *b*, based on the 16S rRNA gene of *Ehrlichia* spp.; *c*, based on the *groESL* operon of *Ehrlichia* spp.

For each gene analyzed, a phylogram was constructed by the maximum likelihood method. Phylogenetic distances between homologous sequences were calculated using Kimura's two-parameter model. Confidence values for individual branches of the resulting tree were determined by bootstrap analysis with 1000 replicates.

Conclusions

Hyalomma anatolicum ticks collected in Tajikistan were tested by PCR for markers of tick-borne bacterial and protozoan infections. DNA of *Ehrlichia* spp. and *Theileria* spp. was detected in ticks collected from the Rudaki and Tursunzade districts. The infection rates for *Ehrlichia* spp. and *Theileria* spp. DNA markers ranged within 0.7–3.3 % according to PCR. Fragments of the 16S rRNA gene and *groESL* operon from *Ehrlichia* spp. and of the 18S rRNA gene from *Theileria* spp. were isolated and sequenced from *H. anatolicum* ticks. Phylogenetic analysis demonstrated that *Ehrlichia* spp. isolates were highly similar to *Ehrlichia* spp. circulating in China and Brazil. Isolate Tajikistan 5 was closely related to the putative novel species *E. mineirensis*. The Tajik isolates of *Theileria* spp. were genotyped as *Theileria annulate*, and fragments of the 18S rRNA gene from these isolates were highly similar to the 18S rRNA gene of *T. annulata* isolates from Turkey, Iran, Pakistan and China.

References

- Aguiar D.M., Ziliani T.F., Zhang X., Melo A.L., Braga I.A., Witter R. A novel *Ehrlichia* genotype strain distinguished by the TRP36 gene naturally infects cattle in Brazil and causes clinical manifestations associated with ehrlichiosis. *Ticks Tick Borne Dis.* 2014;5(5):537-544. DOI 10.1016/j.ttbdis.2014.03.010.
- Begum F., Wissemann C.J., Casals J. Tick-borne viruses of West Pakistan: IV. Viruses similar to or identical with, Crimean hemorrhagic fever (Congo-Semunya), Wad Medani and Pak Argas 461 isolated from ticks of the Changa Manga Forest, Lahore District, and of Hunza, Gilgit Agency, W. Pakistan. *Am. J. Epidemiol.* 1970;92(3):197-202.
- Cabezas-Cruz A., Valdes J.J., de la Fuente J. The glycoprotein TRP36 of *Ehrlichia* sp. UFMG-EV and related cattle pathogen *Ehrlichia* sp. UFMT-BV evolved from a highly variable clade of *E. canis* under adaptive diversifying selection. *Parasit. Vectors.* 2014;7:584. DOI 10.1186/s13071-014-0584-5.
- Cruz A.C., Zweygarth E., Ribeiro M.F., da Silveira J.A., de la Fuente J., Grubhoffer L., Valdes J.J., Passos L.M. New species of *Ehrlichia* isolated from *Rhipicephalus* (*Boophilus*) microplus shows an ortholog of the *E. canis* major immunogenic glycoprotein gp36 with a new sequence of tandem repeats. *Parasit. Vectors.* 2012;11(5):291. DOI 10.1186/1756-3305-5-291.
- Gajadhar A.A., Lobanov V., Scandrett W.B., Campbell J., Al-Adhami B. A novel *Ehrlichia* genotype detected in naturally infected cattle in North America. *Vet. Parasitol.* 2010;173(3-4):324-329. DOI 10.1016/j.vetpar.2010.06.034.
- Gresikova M., Sekeyova M., Tempera G., Guglielmino S., Castro A. Identification of a Sindbis virus strain isolated from *Hyalomma marginatum* ticks in Sicily. *Acta Virol.* 1978;22(3):231-232.
- Parola P., Inokuma H., Camicas J.L., Brouqui P., Raoult D. Detection and identification of spotted fever group Rickettsiae and *Ehrlichiae* in African ticks. *Emerg. Infect. Dis.* 2001;7(6):1014-1017. DOI 10.3201/eid0706.010616.
- Petrova I.D., Kononova Iu.V., Chausov E.V., Shestopalov A.M., Tishkova F.Kh. Genetic variants of the Crimean-Congo hemorrhagic fever virus circulating in endemic areas of Southern Tajikistan in 2009. *Molecular Genetics, Microbiology, and Virology.* 2013;(3):29-36. DOI 10.3103/S0891416813030063.
- Rar V.A., Livanova N.N., Panov V.V., Doroschenko E.K., Pukhovskaya N.M., Vysochina N.P., Ivanov L.I. Genetic diversity of *Anaplasma* and *Ehrlichia* in the Asian part of Russia. *Ticks Tick Borne Dis.* 2010;1(1):57-65. DOI 10.1016/j.ttbdis.2010.01.002.
- Rar V.A., Maksimova T.G., Zakharenko L.P., Bolykhina S.A., Dobrotvorskyy A.K., Morozova O.V. *Babesia* DNA detection in canine blood and *Dermacentor reticulatus* ticks in southwestern Siberia, Russia. *Vector-Borne Zoonotic Dis.* 2005;5(3):285-287. DOI 10.1089/vbz.2005.5.285.
- Rasulov I. Ticks status in Central Asia with a special emphasis on Uzbekistan. *Parasitol. Res.* 2007;101(Suppl.2):183-186.
- Robinson P.M. Theileriosis annulata and its transmission – a review. *Trop. Anim. Health Prod.* 1982;14(1):3-12.
- Roux V., Rydkina E., Eremeeva M., Raoult D. Citrate synthase gene comparison, a new tool for phylogenetic analysis, and its application for the Rickettsiae. *Int. J. Syst. Bacteriol.* 1997;47(2):252-261. DOI 10.1099/00207713-47-2-252.
- Sumner J.W., Nicholson W.L., Massung R.F. PCR amplification and comparison of nucleotide sequences from the *groESL* heat shock operon of *Ehrlichia* species. *J. Clin. Microbiol.* 1997;35(8):2087-2092.
- Tamura K., Peterson D., Peterson N., Stecher G., Nei M., Kumar S. MEGA5: Molecular Evolutionary Genetics Analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol. Biol. Evol.* 2011;28(10):2731-2739. DOI 10.1093/molbev/msr121.
- Tishkova F.H., Belobrova E.A., Valikhodzhaeva M., Atkinson B., Hewson R., Mullojonova M. Crimean-Congo hemorrhagic fever in Tajikistan. *Vector-Borne Zoonotic Dis.* 2012;12(9):722-726. DOI 10.1089/vbz.2011.0769.
- Wen B., Jian R., Zhang Y., Chen R. Simultaneous detection of *Anaplasma marginale* and a new *Ehrlichia* species closely related to *Ehrlichia chaffeensis* by sequence analyses of 16S ribosomal DNA in *Boophilus microplus* ticks from Tibet. *J. Clin. Microbiol.* 2002;40(9):3286-3290. DOI 10.1128/jcm.40.9.3286-3290.2002.
- Wu X.B., Na R.H., Wei S.S., Zhu J.S., Peng H.J. Distribution of tick-borne diseases in China. *Parasit. Vectors.* 2013;6:119. DOI 10.1186/1756-3305-6-119.

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