

## REVIEW

# An Insight into the Potentially Old-Wonder Molecule—Quercetin: the Perspectives in Foresee

Nidhi Rani<sup>1</sup>, Lakshmi Palanisamy Thanga Velan<sup>1</sup>, Saravanan Vijaykumar<sup>1</sup> and Annamalai Arunachalam<sup>2</sup>

**ABSTRACT** Use of phyto-medicine and digitalization of phyto-compounds has been fallen enthralling field of science in recent years. Quercetin, a flavonoid with brilliant citron yellow pigment, is typically found in fruits and leafy vegetables in reasonable amount. Quercetin's potentials as an antioxidant, immune-modulator, anti-inflammatory, anti-cancer, and others have been the subject of interest in this review. Although, profiling the insights in to the molecular characterization of quercetin with various targets provided the loop-holes in understanding the knowledge for the aforementioned mechanisms, still necessitates research globally to unearth it completely. Thus, the available science on the synthesis and significant role played by the old molecule - quercetin which does wonders even now have been vividly explained in the present review to benefit the scientific community.

**KEYWORDS** quercetin, antioxidant, flavonoids, mechanism of action, molecular characterization

In recent years, the values of synthetic drugs have almost lost due to its significant side effects and most of them have been withdrawn from the market because of their high toxicity. In fact, researchers have realized the potentials of herbs, which have been very well documented even before thousands of years. Generally, synthetic drugs are synthesized by pure ingredients, while herbal medicines are made up of complex ingredients. Those different ingredients in one or many herbs may balance each other, or in other words buffer each other, and act synergistically to make the systemic effect more potent. Moreover, some synthetic drugs may target only one molecule and therefore, its effect on other molecules in the pathways or systems are unknown and this often could lead to severe side effects. However, in contrast to this, herbs act on multiple targets, for example, one formulation may have effects on major diseases such as anti-inflammatory, anti-allergic,<sup>(1)</sup> anti-oxidant, anti-toxic, including improvement of cardiovascular functions, anti-cancerous,<sup>(2)</sup> anti-diabetic,<sup>(3)</sup> and anti-osteoporotic.<sup>(4)</sup>

Generally, bioflavonoids are a large group of non-nitrogenous class of plant secondary metabolites that provide pigmentation to flower and protect from ultraviolet light, microbes, and insects, apart from imparting flavors to the fruits and vegetables of the plants.<sup>(5)</sup> They are identified as a good alternative to the synthetic drugs. One of the most valuable bioflavonoid is quercetin, which needs to be explored to a larger extent as it is reported to be the highly competitive and potential component in drug formulations.<sup>(6)</sup> Therefore, the present

review was written to emphasize the information on quercetin's structure, synthesis, metabolism, medicinal value, proteomic, and genomic information, which in turn may provide evidence about its role as a therapeutic and outlines gap in the available data that need to be filled in order to determine the quercetin's appreciable role in future disease therapy.

### Classification of Bioflavonoids

Bioflavonoid shares a common flavones backbone; three ringed with hydroxyl [OH] groups. According to the international union of pure and applied chemistry (3-hydroxy-2-phenylchromen-4-one: IUPAC) nomenclature, bioflavonoids are classified into flavonoid, isoflavonoid, and neo-flavonoid (Figure 1). The flavonoids are further grouped in to flavanones, flavones, flavonols, flavans, and anthocyanins, based on the position of functional groups. However, among them flavonols that enrolls the 3-hydroxyflavone backbone plays a vital role in imparting taste, flower color and flavor to fruits and green leafy vegetables.<sup>(7)</sup> Many different flavonols such as kaempferol, morin, spirenoside, fisetin, quercetin, and Galanginetc, have been identified in a wide variety of

---

©The Chinese Journal of Integrated Traditional and Western Medicine Press and Springer-Verlag Berlin Heidelberg 2015

1. Centre for Bioinformatics, School of Life science, Pondicherry University, Pondicherry (605014), India; 2. Department of Botany, Sethupathy Government Arts and Science Collage, Alagappa University, Ramanathapuram, Tamil Nadu (632502), India  
Correspondence to: Dr. Lakshmi Palanisamy Thanga Velan, Tel: 91-413-2654947, Fax: 91-413-2655211, E-mail: lakshmiptv@yahoo.co.in; lakanna@bicpu.edu.in  
DOI: 10.1007/s11655-015-2073-x

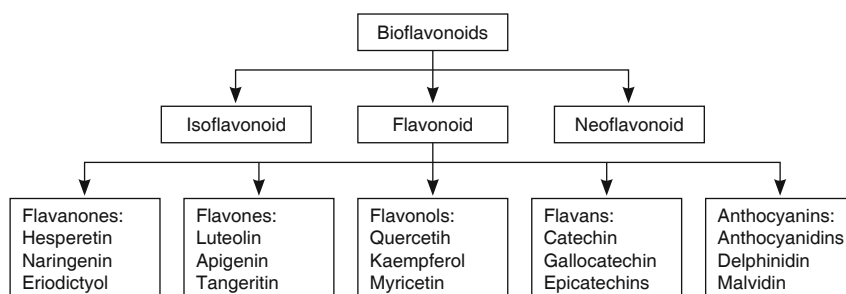


Figure 1. IUPAC Classification of Bioflavonoids

plants and are reported to exhibit the difference by the position of their functional group (Table 1).

Table 1. Position of the Functional Group in Flavonols

Examples	Position of functional group in carbon chain						
	3	5	7	2'	3'	4'	5'
Kaempferol	OH	OH	OH	H	H	OH	H
Morin	OH	OH	OH	OH	H	OH	H
Rutin	O-R	OH	OH	H	OH	OH	H
Myricetin	OH	OH	OH	H	OH	OH	OH
Quercetin	OH	OH	OH	H	OH	OH	H
Quercetrin	O-R <sup>a</sup>	OH	OH	H	OH	OH	H
Myricitrin	O-R'	OH	OH	H	OH	OH	OH
Spirenoside	OH	OH	OH	H	OH	O-Gluc	H
Galangin	OH	OH	OH	H	H	H	H
Robinin	O-R	OH	OH	H	H	OH	H
Kaempferide	OH	OH	OH	H	H	O-Me	H
Fisetin	OH	H	OH	H	OH	OH	H
Rhamnetin	OH	OH	O-Meb	H	OH	OH	H

Notes: a-O-R' = alkoxy; b-O-Me = methoxy; c-O-Glu = glucosyl

### Structural Organization and Implications of Quercetin

Quercetin (molecular formula  $C_{15}H_{10}O_7$ ; molecular weight of 302.236 g/mol) is a heterocyclic pyrone ring (aromatic trimeric heterocyclic) with two benzene rings. According to IUPAC, quercetin is a 3,3',4',5,7-pentahydroxyflavanone (synonym 3,3',4',5,7-pentahydroxy-2-phenylchromen-4-one), a compound with five hydroxyl groups attached in the ring structure at the position 3, 5, 7, 3', and 4' (Figure 2), giving it a status of amphipathic, i.e., both lipophilic and hydrophilic in nature. Those quercetin derivatives which have O-methyl, C-methyl, and prenyl derivatives are lipophilic and are reported in glands located on the surface of the leaves, flowers, or fruits in members of Labiatae or Compositae families.<sup>(7)</sup> Basically, it is a brilliant citron yellow color compound which is often found in plant as either glycosides [with attached

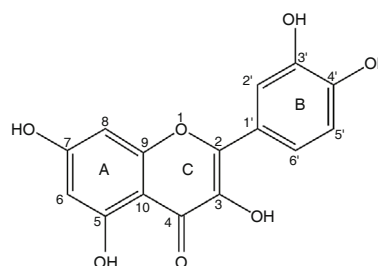
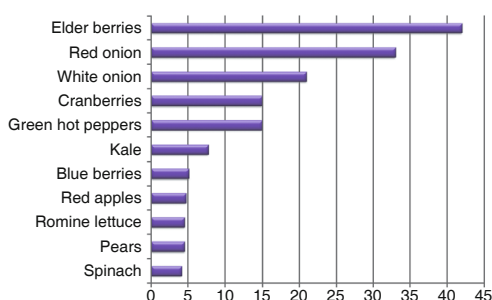


Figure 2. Molecular Structure of Quercetin

sugars (glycosyl groups)] or as aglycones (without attached sugars). The hydroxyl group at the C-3 carbon is easily glycosylated to form quercetin glycosides. The monosaccharides such as glucose, galactose, rhamnose, or xylose are attached with quercetin at C-3 carbon and form quercetin 3-O-glycosides,<sup>(8)</sup> as found in sage (*Salvia officinalis*) and mango fruit (*Mangifera indica*),<sup>(9,10)</sup> while quercetin 3-O-rhamnoside is found in olive (*Olea europaea*) oil,<sup>(11)</sup> peppers (*Piper nigrum*),<sup>(12)</sup> and spinach (*Spinacia oleracea*).<sup>(13)</sup> During glycosylation of the hydroxyl group (commonly at position 3), quercetin derivatives undergo a change from lipophilic to hydrophilic to form glycosylated quercetin, which is cytosol-soluble and could be easily transported to various parts of the plant.<sup>(14,7)</sup> In general the physical and chemical properties (such as absorption, solubility, and *in-vivo* effects) of the glycosylated quercetin differ from aglycosylated quercetin.<sup>(15,16)</sup> Disaccharides are also found to be attached with quercetin molecule such as rutin3-O-rhamnosylglucoside as in tea (*Camellia sinensis*),<sup>(17)</sup> spinach,<sup>(13)</sup> chokeberries (*Aronia arbutifolia*),<sup>(18)</sup> and buckwheat (*Fagopyrum esculentum*).<sup>(19)</sup> Another glycosylation site (hydroxyl group) is observed at C-7 as 3-O-rhamnoside-7-O-glucoside as seen in peppers<sup>(12)</sup> and quercetin 7-O-glucoside in beans<sup>(20)</sup> respectively. Perhaps, C-glycosides and sulphate derivatives of quercetin such as 3, 4, 7, 3', 4'-pentahydroxy-6-glucose flavon and quercetin 3-O-glucoside-3'-sulfate were also found in *Ageratina calophylla*<sup>(21)</sup> and cornflower (*Centaurea cyanus*)<sup>(22)</sup> respectively, but these compounds occur relatively rare in nature.

### Metabolism of Quercetin

Quercetin is consumed daily by millions of people as a dietary source due to its presence in vegetables and fruits such as beans, onions, grapes, apple, green tea, berries, vegetables, nuts either in the fruits, flowers, barks, and leaves (Figure 3). Digestion of quercetin begins from oral cavity by cleavage of glycosides molecule by  $\beta$ -glycosidases, while aglycosylated molecule become more lipophilic and could be absorbed into the epithelial cell of the colon.<sup>(23)</sup> Generally, absorbed quercetins are metabolized in liver and unabsorbed in intestine involving four processes namely, glucuronidation, methylation, hydroxylation, and sulfonation.



**Figure 3. Amount of Quercetin (mg/100 g edible portion) in Food**

Glucuronidation is one of the detoxifying mechanisms in liver which play a major role in metabolism of xenobiotic compound with the help of uridine 5'-diphospho (UDP)-glucuronosyltransferase enzyme.<sup>(24)</sup> Quercetin is reported to be glucuronidized during passage across the epithelium in the liver by UDP-glucuronosyl transferase<sup>(25,26)</sup> and further conjugates with glucose transporter receptor in small intestine.<sup>(27)</sup> For instance, a study on a rat's small intestine revealed the uptake of quercetin conjugate that interacted with intestinal hexose transport pathway through glucose transporter receptor and competitively inhibited the uptake of galactose, due to the presence of quercetin-3-glucoside in the mucosal medium, indicating its interaction with the sodium-dependent glucose transporter pathway (SGLT1).<sup>(28)</sup> However, several observations suggested that quercetin could be absorbed by two ways; as stated above, one in the small intestine by SGLT1 with subsequent deglycosylation within the enterocyte by cytosolic  $\beta$ -glucosidase, or luminal hydrolysis of the glucoside by lactase phlorizin hydrolase (LPH); and the other is absorption by passive diffusion of the released aglycone.<sup>(29)</sup> However, the first hypothesis was evaluated with the use of phlorizin (the inhibitor of SGLT1) and N-(n-butyl)-deoxy

galactonojirimycin (as an inhibitor of the lactase domain of LPH) in a rat everted-jejunal sac model.<sup>(30)</sup> Quercetin-3-glucuronides and quercetin-7-glucuronides, the major product of quercetin metabolism in the intestine are further absorbed or excreted by two pathways: (i) methylation of both quercetin-3-glucuronides and quercetin-7-glucuronides by methyltransferases, and (ii) hydroxylation of endogenous  $\beta$ -glucuronidase followed by sulfonation to quercetin-3'-sulfate.<sup>(31,32)</sup> Quercetin aglycones were found in human plasma as main circulating metabolite in an un-conjugated form, which resulted in deconjugation of quercetin glucuronides by the enzyme  $\beta$ -glucuronidase.<sup>(33)</sup> Nevertheless, an alternate mechanism for quercetin absorption was also revealed in small intestinal tract of human, where the flavonoid degrading strict anaerobic microorganism *Eubacterium ramulus* resides and cleaves quercetin ring structure into 3, 4-dihydroxy phenyl acetic acid.<sup>(34)</sup>

### Pharmaceutical Accomplishment of Quercetin in Human Diseases

Mainly, by the fact that the quercetin has all the right structural features for free radical scavenging activity, they exert beneficial health effects such as anti-inflammatory, anti-allergic, anti-oxidant, anti-toxic, and anti-viral against reverse transcriptase of human immuno deficiency virus (HIV) and other retroviruses including *Herpes simplex* virus type 1, polio-virus type 1, parainfluenza virus type 3, respiratory syncytial virus (RSV), and HIV-1 integrase.<sup>(35)</sup> Additionally, it is also reported to interact with cyclin-dependent kinases such as CDK6, CDK5, and CDK1,<sup>(36)</sup> fatty acid synthesizing enzyme (enoyl-acyl carrier protein reductase),<sup>(37)</sup> and G protein-coupled receptor<sup>(38)</sup> to label a few. Moreover, it has remarkably proven to improve the cardiovascular functions, reducing the risk for cancer. Although potential benefits are extraneous, it is not possible to highlight all of them individually and therefore, with much care, effort has been undertaken to cover at most under the four major headings.

#### Effects on Allergy

An immune response produces effector molecules that act to remove antigen by various mechanisms. Generally these effector molecules induce a subclinical and localized inflammatory response that eliminates antigen without extensively damaging the host tissue. An allergy is a hypersensitivity disorder of the immune system. An excessive activation of white blood cells, mast cells and basophils, through the immunoglobulin

E antibody (IgE) is a symptom of allergic reaction. Quercetin exerts many effects on anti-allergic response; to inhibit histamine release in rat connective tissue mast cells, mucosal mast cells,<sup>(39)</sup> human lung, and intestinal mast cells.<sup>(40)</sup> In fact, quercetin isolated from *Gingko biloba* is reported to inhibit the lipopolysaccharide (LPS)-induced tumor necrosis factor (TNF)- $\alpha$ , interleukin (IL)-6, IL-1 $\beta$  transcription by inhibiting the activation of ERK1/2 and p38 MAPK in macrophages.<sup>(41)</sup> The pathologic role of TNF- $\alpha$  and IL-4, which are involved in the onset of various allergic diseases including atopic dermatitis, atopic rhinitis, and asthma, were arrested even at the lower (100  $\mu$ mol/L) concentration of quercetin, when applied on human umbilical cord blood-derived cultured mast cells (hCBMCs).<sup>(42)</sup> In another study, IgE or phorbol-12-myristate 13-acetate and calcium ionophore A23187 (PMACI)-mediated histamine release were blocked by quercetin in RBL-2H3 cells and also it inhibited the elevation of intracellular calcium as well as gene expression and production of all the pro-inflammatory cytokines.<sup>(43)</sup> In an experiment, quercetin also inhibited the expression of CD63 and CD203c and the histamine release by the basophils that were activated with anti-IgE.<sup>(44)</sup> Yet another investigation reported that quercetin inhibited the process of degranulation and suppressed the CD23 mRNA expression in RBL-2H3 cells at 10  $\mu$ mol/L concentration.<sup>(45)</sup>

Likewise, when Fc $\epsilon$ RI- anti-IgE activated model was treated with 1.8–20  $\mu$ mol/L quercetin, it interacted with catalytic pocket of the enzyme and inhibited P13K, consequently leading to the loss of phosphorylation of kinases (such as Bruton's tyrosine kinase)<sup>(46)</sup> which otherwise would phosphorylate phosphoinositide phospholipase C- $\gamma$  (PLC $\gamma$ ) and lead to the production of inositol trisphosphate (IP3) and diacylglycerol (DAG) that may be responsible for the activation of membrane markers up-regulation and histamine production.<sup>(47)</sup> Moreover, quercetin is also reported to be effective on N-formyl-methionine-leucine-phenylalanine (fMLP) triggered basophil function, which activate the P13K $\gamma$  and G-coupled receptor kinase (GRK) that are basically responsible for degranulation event by IP3-calcium signaling or by the activation of diacylglycerol-protein kinase C-PKC pathway. The calcium ionophore A23187 induced the expression of CD63 and CD203c and these markers promote the activation of Ca<sup>2+</sup>/calmodulin pathway, which is inhibited by quercetin.<sup>(48)</sup> Thus collectively, quercetin acts as a strong inhibitor of components those involved

in allergic reaction and found to be functional even at the micromolar concentrations and thereby arising as a novel alternative for allergic treatments.

#### Effect on Inflammation

Inflammation is a mechanism of innate immunity which act as a first response from immune system against harmful stimuli, such as injury caused by pathogens, damaged cells, and irritation.<sup>(49)</sup> It is characterized by increased blood flow to the tissue, raise in temperature, redness, swelling, and pain. It may involve in developing various diseases such as allergy, asthma, arthritis, atherosclerosis, cancer, aging, etc.<sup>(50)</sup> Inflammation is a complex response which is caused by numerous biological factors such as LPS (major component of the Gram-negative bacteria cell wall),<sup>(51)</sup> enzymes [cyclooxygenase (COX) and lipoxygenase (LOX)],<sup>(52)</sup> nitric oxide production, and nitric oxide synthase (NOS) expression.<sup>(53)</sup>

LPS is one of the major factor for inflammation which is recognized by Toll-like receptor (TLR4) receptors that is found on the immune cells, including macrophages.<sup>(54)</sup> When LPS bind with specific TLR4 receptor, it can trigger signaling pathways and activate nuclear factor (NF)- $\kappa$ B.<sup>(55)</sup> Under normal conditions, NF- $\kappa$ B occurs in cytoplasm in an inactive state, bound to the inhibitory  $\kappa$ B (I $\kappa$ B) proteins. The NF- $\kappa$ B is activated by I $\kappa$ B kinase (IKK) complex, that are composed of Ser/Thr kinases IKK $\alpha$  and IKK $\beta$  associated with other signal transducers IKK $\gamma$  and IKAP. Signal components activate Ser/Thr kinases in IKK complex and activated IKK complex phosphorylates I $\kappa$ B and then followed by proteasome-mediated degradation of I $\kappa$ B.<sup>(56,57)</sup> After I $\kappa$ B degradation NF- $\kappa$ B enters into the nucleus and bind to the promoter regions of immune genes including IL-6 for transcriptional activation.<sup>(58)</sup> IL-6 is a pleiotropic interleukin that acts as both pro-inflammatory and anti-inflammatory cytokine. It is produced by T cells and macrophages as well as varieties of other cell types including adipocytes and microglial cell.<sup>(59)</sup> However, the effect of quercetin 3-O- $\beta$ -(2"-galloyl)-glucopyranoside (QG-32) from *Persicaria lapathifolia* (polygonaceae) was realized when it inhibited reactive superoxide (ROS) production in human monocytes.<sup>(60)</sup> Perhaps, studies showed that ROS could increases the LPS-induced IL-6 expression at the transcription level. In spite of ROS's independent production, it could still amplify TLR4-mediated

inflammatory responsiveness.<sup>(61)</sup> When endotoxin LPS-activated macrophages RAW 264.7 were treated with various concentrations (10–100  $\mu$  mol/L) of QG-32 or pyrrolidine dithiocarbamate (PDTC) within 24 h, it inhibited the production of IL-6 as well as down-regulated the LPS-induced IL-6 expression at the transcription level.<sup>(62)</sup> Similarly, NF- $\kappa$ B, a transcription factor that is involved in proteolytic degradation of I $\kappa$ B was also inhibited by the administration of quercetin.<sup>(62)</sup> In fact, quercetin inhibits cyclooxygenase and lipoxygenase at concentration of 10–20  $\mu$  mol/L, which is an important mediator in inflammation and tumor promotion.<sup>(52)</sup> The NOS expression is also found to be suppressed by administration of 100  $\mu$  mol/L quercetin, resulting in inhibition of nitric oxide (a pro-inflammatory mediator) production.<sup>(53)</sup> Hence these factors may attribute a major role in numerous chronic diseases such as allergy,<sup>(50)</sup> diabetes,<sup>(63)</sup> atherosclerosis,<sup>(64)</sup> depression,<sup>(65)</sup> Alzheimer's disease,<sup>(66)</sup> systemic lupus erythematosus,<sup>(67)</sup> prostate cancer,<sup>(68)</sup> and rheumatoid arthritis.<sup>(69)</sup> Since these studies have given lots of pharmacological potential of quercetin in the inflammatory disorders, the action of quercetin against numerous inflammatory factors may provide a better option to cure above mentioned diseases in the future.

#### Quercetin as an Antioxidant

The antioxidant activity of a compound is determined by the presence of free hydroxyl groups as well as position of double bond<sup>(14)</sup> that can donate electron through resonance to stabilize the free radicals.<sup>(70)</sup> The radical scavenging property defends the body against oxidative stress, reduces heart disease, prevents cancer, and slows the aging process in cells.<sup>(71)</sup> Lipid peroxidation is an oxidative degradation of lipid in which unsaturated fatty acids are converted to free radicals via the abstraction of hydrogen and further these free radicals are oxidized by molecular oxygen to create lipid peroxy radicals.

Quercetin shows inhibitory effect against human lipoxygenase (hLO) isozymes<sup>(71)</sup> that catalyzes the dioxygenation of polyunsaturated fatty acids to their hydroperoxy acids that have been implicated in several diseases including inflammation, immune disorders, and various types of cancers.<sup>(72)</sup> The low-density lipoprotein (LDL) is another reason for cardiovascular disease. While, studies have shown the ability of quercetin to inhibit LDL oxidation,<sup>(73)</sup> it not only stops the lipid peroxidation but also increases the glutathione

(GSH) level,<sup>(74)</sup> which is a tripeptide that acts as an antioxidant in our body and neutralizes the free radicals by regulating the nitric oxide cycle<sup>(75)</sup> and other biochemical reactions involved in DNA synthesis and repair, protein synthesis, prostaglandin synthesis, amino acid transport, and enzyme activation.

Administration of quercetin-3'-glucuronide quercetin is also reported to inhibit xanthine oxidase,<sup>(76)</sup> which catalyzes the oxidation of hypoxanthine and xanthine to uric acid and superoxide radicals. The former plays a crucial role in gout, while the latter is involved in oxidative stress including inflammation, atherosclerosis, cancer, and aging. So, an increase in xanthine oxidase influences the rate of hepatitis and the degree of brain edema and its control by derivative of quercetin could accomplish a good measure for treating hepatitis, brain edema and also reduce the oxidative stress.<sup>(77,78)</sup>

In view of the number of studies, it is clear that quercetin possesses the structure that act as an effective and powerful antioxidants and since it is playing a major role in preventing the above mentioned diseases, and hence quercetin could be a subject of interest to control them naturally.

#### Quercetin as an Anti-cancer Agent

Oxidative DNA damage by oxygen species superoxide, hydroxyl, peroxy, and alkoxy, and reactive nitrogen species play a key role in human cancer development. The hydroxyl groups of quercetin have electron accepting capacity, while the catechol group chelate with metal ions.<sup>(23)</sup> *In-vitro* studies indicate that quercetin plays an important role in cancer treatment with the ability to act as potential antioxidants and there by inducing numerous molecular pathways such as apoptotic pathway, down-regulation of mutant P53 protein, G<sub>1</sub>-phase arrest, inhibition of tyrosine kinase, inhibition of heat shock proteins, inhibition of ras protein expression, and estrogen receptor binding capacity.

Quercetin is reported to induce cell death by apoptosis in leukemia, lung, hepatoma, oral, and colon cancer cell lines.<sup>(79)</sup> For instance, administration of 40–50  $\mu$  mol/L of quercetin induced the mitochondrial apoptotic pathway through initiating Bcl-2-associated X protein (Bax) and/or Bcl-2 homologous antagonist/killer (Bak) proteins. These proteins are involved in increasing the size of outer mitochondrial membrane pore and cytochrome C leakage into

the cytoplasm which further activates the apoptotic protease activating-factor 1 (APAF-1) and produces apoptosome.<sup>(80)</sup> The P53 is another important tumor suppression protein which activates the Bax and initiate cell death. When human hepatocellular carcinoma cell was treated with 40–120  $\mu$  mol/L of quercetin, p53 expression was increased, while down-regulating the anti-apoptotic protein survivin that regulates the caspase activation and Bcl-2 that prevents mitochondrial mediated apoptosis.<sup>(81)</sup>

TNF- $\alpha$ -related apoptosis-inducing ligand (TRAIL or Apo2L) that belongs to the TNF cytokine family is produced by the activated macrophages and it is responsible for inducing inflammation, apoptotic cell death, and inhibiting tumorigenesis through enhancing the transcription of Bcl-2.<sup>(82)</sup> TRAIL, however, binds with the death receptors DR4/DR5, which further interact with the adaptor protein Fas-associated death domain (FADD) and procaspase-8 and form the death-inducing signaling complex (DISC). Procaspase-8 activation and DISC lead to cleavage of procaspase-3 and engagement of the cellular machinery associated with the type I extrinsic apoptotic pathway. However, evidence claim that these attributes by the TRAIL proves futile under glioma cells and many cancer cell lines became more or less resistant to the apoptotic effect.<sup>(83)</sup> Thus the administration of quercetin especially 250  $\mu$  mol/L was revealed to reducing the viability of U251, LN229, U87-MG, MDA-MD-231 and A172 glioma cells and also affecting the estrogen receptor  $\alpha$  (ER- $\alpha$ ) by inducing cytotoxicity in some cancer cell lines.<sup>(84)</sup> Moreover, quercetin is also reported to prevent the ROS production in the human cervix epithelial carcinoma cell line (HeLa) and stimulate the activation of p38/MAPK,<sup>(85)</sup> which are responsible for proapoptotic caspase-3 activation and mediate poly (adenosine diphosphate-ribose) polymerase (PARP) cleavage. In another study quercetin (50  $\mu$  mol/L) in combination with ascorbate bound to the estrogen receptor (ER- $\beta$ ) and induced apoptosis of breast cancer (T47D-ER- $\alpha$ ) and osteosarcoma (U2OS-ER- $\alpha$  and -ER- $\beta$ ) by increasing the intracellular pH through the modulation of the cells Na<sup>+</sup>/H<sup>+</sup> exchanger.<sup>(86)</sup>

Quercetin reportedly evidenced the inhibition of malfunction of protein chaperons that are basically responsible for protein folding and maintenance of protein structure in our body. The disturbed chaperons are unable to perform their function and eventually result is death. Moreover, the heat shock proteins

(HSP) such as HSP90, HER2, and IGF2BP-2 allowed tumor cells to bypass normal mechanisms of cell cycle and allowed survival of cancer cell in unfavorable condition viz., hypoxic condition, low circulation, high temperature etc. However, these conditions were reported to be suppressed by quercetin (1–100  $\mu$  mol/L) in several malignant cell lines namely colon cancer,<sup>(87,88)</sup> breast cancer, and prostate cancer.<sup>(89)</sup> Thus, the ability of quercetin to interact with electrons even at lesser concentrations plays a central role in its mechanism of action, mainly by the activation of proteins and DNA damage, leading to the induction of many downstream pathways of the cancer.

Thus quercetin is the subject of intense research on the basis of its anti-inflammatory, anti-allergic, antioxidant, and anti-cancer activities, as well as many therapeutic targets to cure different kinds of diseases such as Alzheimer's disease, diabetes, malaria, Chagas' disease, Schizophrenia etc. Apart from this, studies also suggested that quercetin is effective against antibiotic resistance bacteria. For instance, the antibacterial activities of quercetin has been tested on anti-methicillin resistant *Staphylococcus aureus* (MRSA), which uncovered the unique antibacterial properties of quercetin against *Staphylococcus aureus* (*S. aureus*).<sup>(90)</sup> The study was further substantiated by in-silico approach, which showed a strong interaction of quercetin and kaempferol with multidrug resistant  $\beta$  lactamase of *S. aureus*.<sup>(91)</sup> Thus, collectively, one could say that the quercetin has risen as a novel alternate to the synthetic molecules as evidenced by the enlisted literature presented in Table 2.

### Biosynthesis

Realizing the potentials, it has become essential to know about the synthesis of quercetin, which involves in multiple enzymatic processes in the cytoplasm and associated with endoplasmic reticulum<sup>(137)</sup> via phenyl propanoid pathway (Figure 4). The first step of this pathway is deamination of phenylalanine by the enzyme L-phenylalanine ammonia-lyase (PAL),<sup>(138)</sup> which acts as a precursor molecule to synthesize 4-coumarate that further acts as a substrate for formation of 4-coumaroyl coenzyme (CoA) with the help of 4-coumarate-CoA ligase using 1 ATP molecule. The 4-coumaroyl CoA also participates in 6'-deoxychalcone metabolism and in isoflavonoid biosynthesis I, in addition to the synthesis of naringenin chalcone, which involves in the precipitation of bioflavonoid or may form many

Table 2. Responses of Quercetin against Identified Vulnerable Diseases Targets

Target	K <sub>ia</sub> (nmol/L) value	IC <sub>50</sub> (nmol/L) value	Tested on <sup>c</sup>	Effect of the target	References
Dopamine D4 receptor	7.8	-	Homo sapiens	Schizophrenia and antipsychotic action	(38)
Enoyl-acyl-carrier protein reductase, 3-oxoacyl-acyl-carrier protein reductase	22.0	-	<i>Plasmodium falciparum</i>	Fatty acid synthesis	(92,37)
Cytochrome P450 1A1, Cytochrome P450 1B1 (CYP1B1), Cytochrome 1A2, Cytochrome P450 2C9	23.0	-	Homo sapiens	Metabolism of xenobiotics	(93,94,95)
Casein kinase II	1180.0	-	Homo sapiens	Wnt signaling pathway (96)	(97)
Xanthine dehydrogenase	1200.0	-	<i>Bos taurus</i>	Purine catabolism	(77,98)
Multidrug resistance-associated protein 1	2400.0	-	Homo sapiens	Multidrug resistance in tumor cells	(99)
Adenosine A1, A2A, A3 receptor	2470.0	-	<i>Rattus norvegicus</i>	Maintain acid-base balance in blood and other tissues	(100)
Carbonic anhydrase I, II, III, IV, VI, VII, VIII, IX, XII, XIV, VA, VB, 1 (CA I), 2 (CA II)	2540.0	-	Homo sapiens		(101,102)
beta-Secretase (BACE-1)	-	10820.0	Homo sapiens	Alzheimer's disease	(103)
Cyclin-Dependent Kinase1 (CDK1), 5 (CDK5), 6 (CDK6), CDK4/Cyclin D1	-	>20000.0	Homo sapiens	Cell-cycle progression & cellular proliferation	(104,36,105)
Glycogen synthase kinase-3, beta	-	2100.0	<i>Rattus norvegicus</i>	It is implicated in Type 2 diabetes, Alzheimer's disease, inflammation, cancer, and bipolar disorder	(36)
Phosphoinositide 3-kinase (PI3K), alpha Chain A/beta Chain A/beta chain B/delta Chain A/gamma Chain A,	-	3800.0	Homo sapiens	Involved in cell growth, proliferation, differentiation, motility, survival and intracellular trafficking	(106,107)
Alpha-Amylase	-	21400.0	Homo sapiens	Playing role in carbohydrate digestion	(108)
PIM-1 kinase	-	1100.0	Homo sapiens	Involved in cell cycle progression, apoptosis and transcriptional activation	(109)
Aldose reductase	-	50100.0	<i>Sus scrofa</i>	Polyol pathway	(110,111)
Arachidonate 5-lipoxygenase, arachidonate 12-lipoxygenase, arachidonate 15-lipoxygenase, arachidonate 15-lipoxygenase, type II	-	37000.0	<i>Oryctolagus cuniculus</i>	Participates in arachidonic acid metabolism	(112,113,71)
Cyclooxygenase-1/Cyclooxygenase-2	-	50000.0	<i>Rattus norvegicus</i>	Formation of prostaglandins, prostacyclin and thromboxane	(112,114,113)
Tyrosine-protein kinase LCK, Tyrosine-protein kinase SRC, EGF-R Tyrosine Kinase	-	15000.0	Homo sapiens	phosphorylation of tyrosine residues in proteins	(115,116)
HIV-1 integrase	-	13600.0	Human immunodeficiency virus 1	Key component in the retroviral pre-integration complex	(35)
Trypsin	-	7100.0	Homo sapiens	Cleaves peptide chains	(117)
Chymotrypsin, Beta-chymotrypsin	-	100000.0	Homo sapiens	Digestive enzyme component of pancreatic juice	(118)
Protein-tyrosine phosphatase 1B (PTP1B)	-	23300.0	Homo sapiens	Therapeutic target in treating type 2 diabetes	(119)

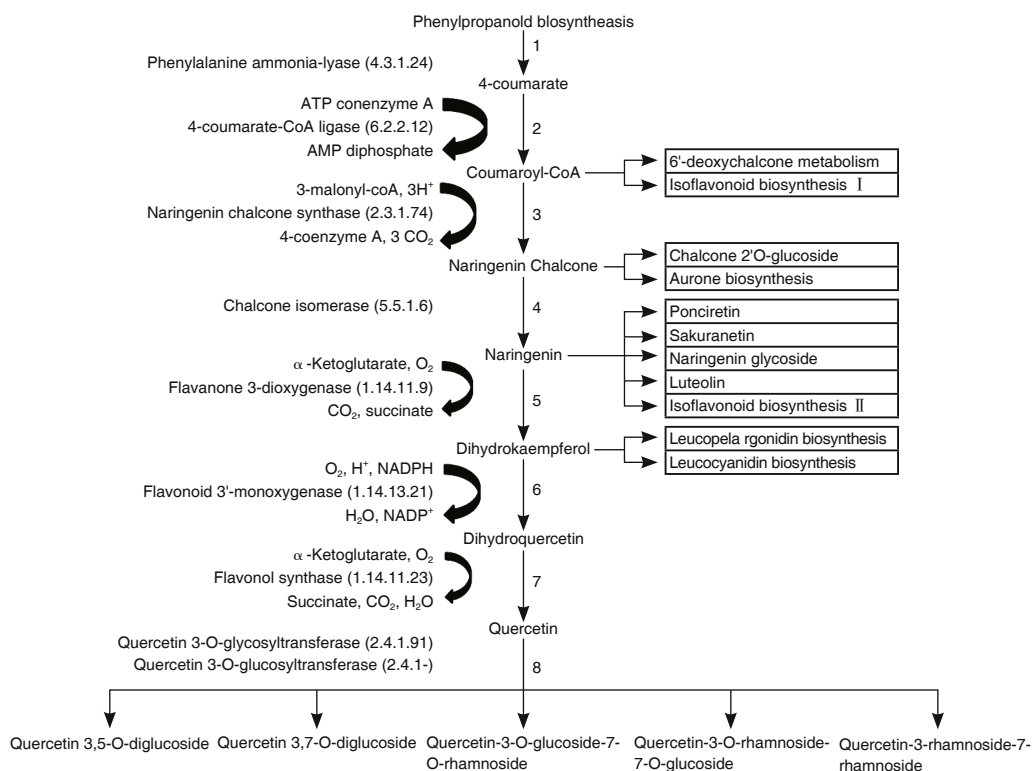
(To Be Continued)

## (Continued)

Target	K <sub>i</sub> a (nmol/L) value	IC <sub>50</sub> (nmol/L) value	Tested on <sup>c</sup>	Effect of the target	References
Sorbitol dehydrogenase	-	177000.0	Homo sapiens	Carbohydrate metabolism	(120)
Aldehyde reductase	-	38400.0	Sus scrofa	Catalyzing the reduction of glucose to sorbitol	(120)
Malate dehydrogenase	-	6000.0	Thermus thermophilus	Reversibly catalyzes the oxidation of malate to oxaloacetate	(107)
Beta-lactamase/Penicillin-binding protein ampH	-	4000.0	Escherichia coli	Responsible for resistance to beta-lactam antibiotics	(107)
Beta-lactamase	-	-	Staphylococcus aureus	Responsible for resistance to beta-lactam antibiotics	(107)
Serine beta-lactamase-like protein	-	4000.0	Homo sapiens	Responsible for the β-lactamase activity	(107)
Glutathione reductase	-	218000.0	Homo sapiens	Targets for aldose reductase inhibitor action	(121,120)
Cell division protein kinase 5	-	-	Rattus norvegicus	Inhibit cell cycle progression	(122)
Fatty acid synthase	-	1500.0	Plasmodium falciparum	Fatty acid biosynthesis	(92)
Serotonin receptor 1A	-	-	Homo sapiens	Target of antidepressants, antipsychotics, anorectics, anti-emetics, gastrokinetic agents, anti-migraine agents, hallucinogens, and entactogens (123)	(124)
Glyoxalase I	-	3200.0	Homo sapiens	Target for anticancer drug	(125)
Dipeptidyl peptidase IV (DPP-IV)	-	130000.0	Homo sapiens	-	(125)
17-beta-hydroxysteroid dehydrogenase 2 (17-beta-HSD2)	-	1540.0	Homo sapiens	Target for anti-breast cancer therapy	(126)
Sialidase (Neuraminidase)	-	1700.0	Clostridium perfringens	Involved in the release of Progeny influenza virus from infected cells	(78)
Glyceraldehyde-3-phosphate dehydrogenase, glycosomal	-	142000.0	Trypanosomacruzi	Target for Chagas' disease	(127)
Replicasepolyprotein 1ab	-	>50000.0	Human SARS coronavirus	Target for human SARS coronavirus	(128)
Monoamine oxidase type A (MAO-A), monoamine oxidase B, amine oxidase, monoamine oxidase A	-	2800.0	Homo sapiens	Central role in the metabolism of monoamine neurotransmitters	(129,130)
Hypoxia-inducible factor 1-alpha inhibitor	-	10200.0	Homo sapiens	Target for treating anemia	(131)
Aromatase (CYP19)	-	12.0	Homo sapiens	Target for anti-breast cancer therapy	(132)
NADPH oxidase 4	-	680.0	Homo sapiens	Contributes in oxidative damage related diseases	(133)
Calmodulin	-	12970.0	Homo sapiens	Target for antihypertensive agents	(134)
ATP-Binding cassette transporter ABCG2	-	6900.0	Homo sapiens	Target for anti-breast cancer therapy	(135)
Mitogen-activated protein kinase p38 alpha, c-Jun N-terminal kinase 3 (JNK3)	-	3450.0	Homo sapiens	Involved in cellular proliferation, differentiation, transcription	(136)

Notes: <sup>a</sup>K<sub>i</sub>- inhibitor constants; <sup>b</sup>IC<sub>50</sub>- half maximal inhibitory concentration; <sup>c</sup>Toxicity tested in the mentioned organism cell line or culture; SARS: severe acute respiratory syndrome; NADPH: nicotinamide adenine dinucleotide phosphate





**Figure 4. Quercetin Glucoside Biosynthesis Pathway in *Arabidopsis thaliana***

Notes: Compiled from Plant Metabolic Pathway Databases; <http://www.plantcyc.org/>

derivatives of quercetin through a series of reactions. Meanwhile, naringenin chalcone may take part in aurone biosynthesis and chalcone 2'-O-glucoside biosynthesis, where the former is important for synthesizing derivatives of auronem that imparts yellow color to flowers,<sup>(139)</sup> while the later is involved in synthesizing chalcone 2'-O-glucoside. The most stereochemically important reaction of flavonoid biosynthesis is conversion of naringenin chalcone to naringenin (2S-flavanones) using chalcone isomerase (CHI, 4th step) or chalcone-flavanone isomerase.<sup>(140)</sup> Precisely, the synthesized naringenin acts as an intermediate for formation of flavones, flavonols, flavan-4-ols, anthocyanins, and isoflavonoids. Thus, naringenin is involved in five different pathways that include ponciretin biosynthesis, sakuranetin biosynthesis, naringenin glycoside biosynthesis, luteolin biosynthesis, and isoflavonoid biosynthesis II, which synthesizes ponciretin, 2S-sakuranetin, naringenin, luteolin, and pratensein respectively. In the meanwhile as a 5th step dihydrokaempferol is synthesized using naringenin 3-dioxygenase, which leads to the leucopelargonidin, leucocyanidin and kaempferolglucoside biosynthesis apart from quercetin/flavonol biosynthesis. Further, dihydrokaempferol in the presence of flavonoid 3'-monooxygenase is converted to dihydroquercetin.

Thus it is evident from the step 7 that the synthesis of quercetin is dependent on 2-oxoglutarate-dependent dioxygenases flavonol synthase (FLS), which is an enzyme that belongs to oxidoreductases family and shows a broad substrate and product selectivity.<sup>(141,142)</sup> Since FLS is an enzyme that catalyzes the formation of quercetin, it is considered to be vital in the quercetin biosynthetic pathway. Eventually the synthesis of glycosylated quercetin from aglycosylated quercetin is catalyzed by the quercetin 3-O-glucosyltransferase and quercetin 3-O-rhamnosyltransferase enzyme (8th and 9th step)<sup>(143)</sup> by the transfer of a glucosyl group from UDP-glucose to the 3'-hydroxy group (of a quercetin molecule). Thus, the glycosylation of aglycosylated quercetin is responsible for the modification of stability, solubility, or localization, and the biological properties of the quercetin glycosides.<sup>(143)</sup>

### Molecular Investigations and Digitalization

Taking into account the importance of each enzyme or the intermediary products formed during the process of secondary metabolite (quercetin) synthesis, it has become necessary to know about the genes involved in the system. With this perception, the literary surfing earmarked the remarkable quantity of work done on this segment. In fact, the genes

responsible for almost all the enzymes involved in the pathway has been studied and very well documented (Table 3). However, when observed keenly, one could realize that the plant systems, that has been explored is very limited (<http://medicinalplantgenomics.msu.edu/>; <http://www.plantcyc.org/>). When the medicinal plants find application in pharmaceutical, cosmetic, agricultural, and food industry right from the pre-historic era and that when the plant diversity is rich in the world,<sup>(144)</sup> why there is scarce research on the molecular investigation of this vital compound in the herbal systems? Digital inventory has a significant role in the pharmaceutical market as many drug interaction studies use these databases for either virtual screening of ligand based on plant origin or proteins, which are of clinical importance (Table 4).<sup>(91)</sup> Although many plant databases are available (many are licensed/commercialized), most of them reveals less information on the important secondary metabolites, which once again give us a scope in the

future for extending research in this frontier area.

### Conclusion

Quercetin derivatives are available not only in dietary vegetables; also it is present in plants that are non-dietary such as *Ginkgo biloba* and *Hypericum perforatum*. Quercetin derivatives are generally nontoxic and manifest a diverse range of beneficial biological activities which are abundantly present in the human diet, as evidenced through the ongoing epidemiological studies, promotion as an effective anti-oxidative agent with scavenging (chelating) capacities and interaction with diverse range of therapeutic target. Therefore, this compound is being intensively investigated, which indicated its role as anti-inflammatory, anti-allergic, antioxidant, anti-cancerous, etc. This has proportionally increased the demand for quercetin from the pharmaceutical industry as an alternate for the synthetic molecules and has given scope for two important concepts to be concentrated in the future. One, in spite of the diverse

**Table 3. Details of the Genes Involved for the Biosynthesis of Quercetin**

Serial No.	Enzyme involved in the biosynthesis of quercetin	Source organism	Gene (mRNA) sequence length (bp)	Gene bank ID
1	phenylalanine ammonia-lyase	<i>Arabidopsis thaliana</i>	2530	30687012
		<i>Arabidopsis lyrata</i>	2436	297827210
		<i>Isatis tinctoria</i>	2490	95020528
		<i>Parrya nudicaulis</i>	2113	323709173
		<i>Brassica rapa</i>	2476	282182892
		<i>Brassica oleracea</i>	2145	269313497
		<i>Thellungiella halophila</i>	2477	312281768
2	4-coumarate-CoA ligase	<i>Arabidopsis thaliana</i>	1648	145336963
		<i>Arabidopsis lyrata</i>	1631	9322511
3	Naringenin chalcone synthase	<i>Arabidopsis thaliana</i>	1491	145357993
		<i>Arabidopsis lyrata</i>	1490	297807414
		<i>Arabidopsis arenosa</i>	1422	333448902
		<i>Arabidopsis halleri</i>	2427	56797558
4	Chalcone isomerase	<i>Arabidopsis thaliana</i>	930	42565949
5	Flavonone/Naringenin 3-dioxygenase	<i>Arabidopsis thaliana</i>	1508	334185877
		<i>Arabidopsis lyrata</i>	1494	297816383
6	Flavonoid 3'-monooxygenase	<i>Arabidopsis thaliana</i>	1835	30682179
		<i>Arabidopsis lyrata</i>	1796	297806828
		<i>Matthiola incana</i>	1748	12231885
		<i>Brassica napus</i>	3038	84380740
7	Flavonol synthase	<i>Arabidopsis thaliana</i>	1323	334187529
		<i>Arabidopsis lyrata</i>	1241	297806938
8	Quercetin 3-O-glucosyltransferase	<i>Arabidopsis thaliana</i>	1672	42569054
		<i>Arabidopsis lyrata</i>	1446	297832031
9	flavonol-3-O-rhamnosyltransferase	<i>Arabidopsis thaliana</i>	1829	42562413

**Table 4. Role of Secondary Metabolites as A Therapeutic Agent for Deadly Diseases**

Name of the herb	Major metabolites
<i>Atropa belladonna</i>	Atropine, scopolamine, calystegine A3, calystegine B2
<i>Camptotheca acuminata</i>	Camptothecin, 10-hydroxycamptothecin, 9-methoxycamptothecin, strictosidine, secologanin, strictosamide
<i>Cannabis sativa</i>	D9-tetrahydrocannabinol, cannabidiol, cannabichromene, cannabigerol, cannabinol
<i>Catharanthus roseus</i>	Vincristine, vinblastine, ajmalicine, serpentine, yohimbine, catharanthine, tabersonine, vindoline, strictosidine, secologanin
<i>Digitalis purpurea</i>	Digitoxin, digoxin, gitoxigenin, gitoxin
<i>Dioscorea villosa</i>	Diosgenin, dioscin, prosapogenin $\alpha$
<i>Echinacea purpurea</i>	Cichoric acid, echinacoside undeca-2E/Z-en 8, 10-diyonic acid, dodeca-2E,4E, 8Z, 10E/Z-tetraenoic acid isobutylamide, dodeca-2E, 4E, 8E, 10E-tetraenoic acid isobutylamide, trideca-2E, 7Z-diene-10,12-diyonic acid isobutylamide, tetradeca-8Z-ene-11,13-diyon-2-one, pentadeca-8Z,11E,13Z-trien-2-one
<i>Ginkgo biloba</i>	Ginkgolide A, B, C, J, M, bilobalide, quercetin 3-methylquercetin, kaempferol, narcissin flavonol
<i>Hoodia gordonii</i>	P57AS3, hoodigoside A-Z, hodigogenin A, hoodistanalosite A, hoodistanalosite B, gordonoside D, F, G, J
<i>Hypericum perforatum</i>	Hypericin, hyperforin, pseudohypericin, amentoflavone, euxanthone
<i>Panax quinquefolius</i>	Ginsenosides Rb1, Rb2, Rb3, Rc, Rd, Re, Rf, Rg1, Rg2, Rh1, Rh2, 20(S)-ginsenoside Rg3 pseudo-ginsenoside F11
<i>Rauvolfia serpentina</i>	Betulinic acid, camosic acid camosol, oleanolic acid rosmarinic acid, ursolic acid
<i>Rosmarinus officinalis</i>	Betulinic acid, camosic acid camosol, oleanolic acid rosmarinic acid, ursolic acid
<i>Valeriana officinalis</i>	Acevaltrate, actinidine hesperidin, isovaleramide linarin, pinosresinol-4,4'-di-O-beta-D-glucoside, valeric acid

medicinal properties upheld by quercetin, the lack of experiments in testing quercetin's efficiency on various diseases clinically has necessitated the need to clarify the nature of the impact and interactions between quercetin on different types of targets; and the other concept probably would be a mechanism to determine how efficient and practical it would be to increase the production of quercetin using the available proteomic and genomic details, as the role of quercetin is limitless.

## Conflicts of Interest

Nidhi Rani, Lakshmi Palanisamy Thanga Velan, Saravanan Vijaykumar and Annamalai Arunachalam declare that they have no conflict of interest.

## Author Contribution

Nidhi Rani made substantial contributions to conception and design, acquisition of data, analysis and interpretation of data and participated in drafting the article. Lakshmi Palanisamy Thanga Velan takes the overall responsibility in supervising the designed work and involved in revising the article critically for the intellectual content. Saravanan Vijayakumar made substantial contributions to data collection and integration and participated in drafting and critical revision of the article. Annamalai Arunachalam contributed to the literary resources and enabled for the technical drafting of the article.

## Acknowledgement

The authors acknowledge the library facilities extended by both Pondicherry University and Karunya University.

## REFERENCES

- Liu XY, Wang Q, Xia SJ, Huang JH, Shen ZY, Xu H. Characteristics of lymphocyte nuclear factor-kappaB signal transduction kinase expression in aging process and regulatory effect of epimedium flavonoids. *Chin J Integr Med* 2011;17:704-709.
- Liu PX, Gao J, Chen YJ, Long W, Shen X, Tang WS. Anticancer activity of total flavonoids isolated from Xianhe Yanling Recipe. *Chin J Integr Med* 2011;17:459-463.
- Prabhakar PK, Doble M. Mechanism of action of natural products used in the treatment of diabetes mellitus. *Chin J Integr Med* 2011;17:563-574.
- Shi XL, Liu K, Wu LG. Interventional value of total flavonoids from *Rhizoma Drynariae* on Cathepsin K, a potential target of osteoporosis. *Chin J Integr Med* 2011;17:556-560.
- Harborne JB, Williams CA. Advances in flavonoid research since 1992. *Phytochemistry* 2000;55:481-504.
- Russo M, Spagnuolo C, Tedesco I, Bilotto S, Russo GL. The flavonoid quercetin in disease prevention and therapy: facts and fancies. *Biochem Pharmacol* 2012;83:6-15.
- Williams CA, Grayer RJ. Anthocyanins and other flavonoids. *Nat Prod Rep* 2004;21:539-573.
- Wiczowski W. PMK. Food flavonoids. *Pol. J Food Nutr Sci* 2004;13:101-114.
- Lu Y, Foo LY. Polyphenolics of *Salvia*—a review. *Phytochemistry* 2002;59:117-140.
- Berardini N, Fezer R, Conrad J, Beifuss U, Carle R, Schieber A. Screening of mango (*Mangifera indica* L.) cultivars for their contents of flavonol O- and xanthone

- C-glycosides, anthocyanins, and pectin. *J Agric Food Chem* 2005;53:1563-1570.
11. Ryan D, Robards K, Lavee S. Determination of phenolic compounds in olives by reversed-phase chromatography and mass spectrometry. *J Chromatography A* 1999;832:87-96.
  12. Materska M, Piacente S, Stochmal A, Pizza C, Oleszek W, Perucka I. Isolation and structure elucidation of flavonoid and phenolic acid glycosides from pericarp of hot pepper fruit *Capsicum annuum* L. *Phytochemistry* 2003;63:893-898.
  13. Kuti JO, Konuru HB. Antioxidant capacity and phenolic content in leaf extracts of tree spinach (*Cnidioscolus spp.*) *J Agric Food Chem* 2004;52:117-121.
  14. Rice-Evans C, Packer L., ed. *Flavonoids in health and disease*. New York: Marcel Dekker; 1997:137-161.
  15. Hollman PCH, Bijlsman MNCP, van Gameren Y, Cnossen EPJ, de Vries JHM, Katan MB. The sugar moiety is a major determinant of the absorption of dietary flavonoid glycosides in man. *Free Radical Res* 1999;31:569-573.
  16. Ross JA, Kasum CM. Dietary flavonoids: bioavailability, metabolic effects, and safety. *Annu Rev Nutr* 2002;22:19-34.
  17. Erlund I, Marniemi J, Hakala P, Alfthan G, Meririnne E, Aro A. Consumption of black currants, lingonberries and bilberries increases serum quercetin concentrations. *Eur J Clin Nutr* 2003;57:37-42.
  18. Slimestad RTK, Natel HS, Johannessen T, Giske NH. Flavonoids from black chokeberries, *Aronia melano-carpa*. *J Food Comp Anal* 2005;18:61-68.
  19. Kalinova J, Triska J, Vrchtova N. Distribution of vitamin E, squalene, epicatechin, and rutin in common buckwheat plants (*Fagopyrum esculentum Moench*). *J Agric Food Chem* 2006;54:5330-5335.
  20. Chang Q, Wong YS. Identification of flavonoids in Hakmeitau beans (*Vigna sinensis*) by high-performance liquid chromatography-electrospray mass spectrometry (LC-ESI/MS). *J Agric Food Chem* 2004;52:6694-6699.
  21. Harborne JB. The flavonoids: advances in research since 1986. *J Chem Educ* 1995;72-73.
  22. Flamini G, Antognoli E, Morelli I. Two flavonoids and other compounds from the aerial parts of *Centaurea bracteata* from Italy. *Phytochemistry* 2001;57:559-564.
  23. Murota K, Terao J. Antioxidative flavonoid quercetin: implication of its intestinal absorption and metabolism. *Arch Biochem Biophys* 2003;417:12-17.
  24. Fisher MB, Campanale K, Ackermann BL, Vanden BM, Wrighton SA. *In vitro* glucuronidation using human liver microsomes and the pore-forming peptide alamethicin. *Drug Metab Dispos* 2000;28:560-566.
  25. Boersma MG, van der Woude H, Bogaards J, Boeren S, Vervoort J, Cnubben NH, et al. Regioselectivity of phase II metabolism of luteolin and quercetin by UDP-glucuronosyl transferases. *Chem Res Toxicol* 2002;15:662-670.
  26. Oliveira EJ, Watson DG, Grant MH. Metabolism of quercetin and kaempferol by rat hepatocytes and the identification of flavonoid glycosides in human plasma. *Xenobiotica* 2002;32:279-287.
  27. Gee JM, DuPont MS, Rhodes MJ, Johnson IT. Quercetin glucosides interact with the intestinal glucose transport pathway. *Free Radic Biol Med* 1998;25:19-25.
  28. Gee JM, DuPont MS, Day AJ, Plumb GW, Williamson G, Johnson IT. Intestinal transport of quercetin glycosides in rats involves both deglycosylation and interaction with the hexose transport pathway. *J Nutr* 2000;130:2765-2771.
  29. Day AJ, Gee JM, DuPont MS, Johnson IT, Williamson G. Absorption of quercetin-3-glucoside and quercetin-4'-glucoside in the rat small intestine: the role of lactase phlorizin hydrolase and the sodium-dependent glucose transporter. *Biochem Pharmacol* 2003;65:1199-1206.
  30. Kwon O, Eck P, Chen S, Corpe CP, Lee JH, Kruhlak M, et al. Inhibition of the intestinal glucose transporter GLUT2 by flavonoids. *FASEB J* 2007;21:366-377.
  31. O'Leary KA, Day AJ, Needs PW, Mellon FA, O'Brien NM, Williamson G. Metabolism of quercetin-7- and quercetin-3-glucuronides by an *in vitro* hepatic model: the role of human beta-glucuronidase, sulfotransferase, catechol-O-methyltransferase and multi-resistant protein 2 (MRP2) in flavonoid metabolism. *Biochem Pharmacol* 2003;65:479-491.
  32. Sesink AL, Arts IC, de Boer VC, Breedveld P, Schellens JH, Hollman PC, et al. Breast cancer resistance protein (Bcrp1/Abcg2) limits net intestinal uptake of quercetin in rats by facilitating apical efflux of glucuronides. *Mol Pharmacol* 2005;67:1999-2006.
  33. De Santi C, Pietrabissa A, Mosca F, Rane A, Pacifici GM. Inhibition of phenol sulfotransferase (SULT1A1) by quercetin in human adult and foetal livers. *Xenobiotica* 2002;32:363-368.
  34. Blaut M, Schoefer L, Braune A. Transformation of flavonoids by intestinal microorganisms. *Int J Vitam Nutr Res* 2003;73:79-87.
  35. Raghavan K, Buolamwini JK, Fesen MR, Pommier Y, Kohn KW, Weinstein JN. Three-dimensional quantitative structure-activity relationship (QSAR) of HIV integrase inhibitors: a comparative molecular field analysis (CoMFA) study. *J Med Chem* 1995;38:890-897.
  36. Lu H, Chang DJ, Baratte B, Meijer L, Schulze-Gahmen U. Crystal structure of a human cyclin-dependent kinase 6 complex with a flavonol inhibitor, fisetin. *J Med Chem* 2005;48:737-743.
  37. Sharma SK, Parasuraman P, Kumar G, Surolia N, Surolia A. Green tea catechins potentiate triclosan binding to enoyl-ACP reductase from *Plasmodium falciparum* (PfENR). *J*

- Med Chem 2007;50:765-775.
38. Nahrstedt A, Butterweck V. Lessons learned from herbal medicinal products: the example of St. John's Wort (perpendicular). *J Nat Prod* 2010;73:1015-1021.
  39. Penissi AB, Rudolph MI, Piezzi RS. Role of mast cells in gastrointestinal mucosal defense. *Biocell* 2003;27:163-172.
  40. Fox CC, Wolf EJ, Kagey-Sobotka A, Lichtenstein LM. Comparison of human lung and intestinal mast cells. *J Allergy Clin Immunol* 1988;81:89-94.
  41. Wadsworth TL, Koop DR. Effects of the wine polyphenolics quercetin and resveratrol on pro-inflammatory cytokine expression in RAW 264.7 macrophages. *Biochem Pharmacol* 1999;57:941-949.
  42. Kempuraj D, Madhappan B, Christodoulou S, Boucher W, Cao J, Papadopoulou N, et al. Flavonols inhibit proinflammatory mediator release, intracellular calcium ion levels and protein kinase C theta phosphorylation in human mast cells. *Br J Pharmacol* 2005;145:934-944.
  43. Park HH, Lee S, Son HY, Park SB, Kim MS, Choi EJ, et al. Flavonoids inhibit histamine release and expression of proinflammatory cytokines in mast cells. *Arch Pharm Res* 2008;31:1303-1311.
  44. Chirumbolo S, Marzotto M, Conforti A, Vella A, Ortolani R, Bellavite P. Bimodal action of the flavonoid quercetin on basophil function: an investigation of the putative biochemical targets. *Clin Mol Aller* 2010;8:13.
  45. Lee EJ, Ji GE, Sung MK. Quercetin and kaempferol suppress immunoglobulin E-mediated allergic inflammation in RBL-2H3 and Caco-2 cells. *Inflamm Res* 2010;59:847-854.
  46. Rommel C, Camps M, Ji H. PI3K delta and PI3K gamma: partners in crime in inflammation in rheumatoid arthritis and beyond? *Nat Rev Immunol* 2007;7:191-201.
  47. Miura K, MacGlashan DW Jr. Expression of protein kinase C isozymes in human basophils: regulation by physiological and nonphysiological stimuli. *Blood* 1998;92:1206-1218.
  48. Brock C, Schaefer M, Reusch HP, Czupalla C, Michalke M, Spicher K, et al. Roles of G beta gamma in membrane recruitment and activation of p110 gamma/p101 phosphoinositide 3-kinase gamma. *J Cell Biol* 2003;160:89-99.
  49. Ferrero-Miliani L, Nielsen OH, Andersen PS, Girardin SE. Chronic inflammation: importance of NOD2 and NALP3 in interleukin-1beta generation. *Clin Exp Immunol* 2007;147:227-235.
  50. Dogne JM, Hanson J, Pratico D. Thromboxane, prostacyclin and isoprostanes: therapeutic targets in atherogenesis. *Trends Pharmacol Sci* 2005;26:639-644.
  51. Raetz CR, Whitfield C. Lipopolysaccharide endotoxins. *Annu Rev Biochem* 2002;71:635-700.
  52. Lee KM, Hwang MK, Lee DE, Lee KW, Lee HJ. Protective effect of quercetin against arsenite-induced COX-2 expression by targeting PI3K in rat liver epithelial cells. *J Agric Food Chem* 2010;58:5815-5820.
  53. Ortega MG, Saragusti AC, Cabrera JL, Chiabrando GA. Quercetin tetraacetyl derivative inhibits LPS-induced nitric oxide synthase (iNOS) expression in J774A.1 cells. *Arch Biochem Biophys* 2010;498:105-110.
  54. Miyake K. Innate recognition of lipopolysaccharide by Toll-like receptor 4-MD-2. *Trends Microbiol* 2004;12:186-192.
  55. Palsson-McDermott EM, O'Neill LA. Signal transduction by the lipopolysaccharide receptor, Toll-like receptor-4. *Immunology* 2004;113:153-162.
  56. Karin M, Ben-Neriah Y. Phosphorylation meets ubiquitination: the control of NF-[kappa]B activity. *Annu Rev Immunol* 2000;18:621-663.
  57. Magnani M, Crinelli R, Bianchi M, Antonelli A. The ubiquitin-dependent proteolytic system and other potential targets for the modulation of nuclear factor- $\kappa$  B (NF- $\kappa$  B). *Curr Drug Targets* 2000;1:387-399.
  58. Tian B, Brasier AR. Identification of a nuclear factor kappa B-dependent gene network. *Recent Prog Horm Res* 2003;58:95-130.
  59. Noda M, Takeda K, Sugimoto H, Hosoi T, Takechi K, Hara T, et al. Purification and characterization of human fibroblast derived differentiation inducing factor for human monoblastic leukemia cells identical to interleukin-6. *Anticancer Res* 1991;11:961-968.
  60. Kim Y, Jang DS, Park SH, Yun J, Min BK, Min KR, et al. Flavonol glycoside gallate and ferulate esters from *Persicaria lapathifolia* as inhibitors of superoxide production in human monocytes stimulated by unopsonized zymosan. *Planta Med* 2000;66:72-74.
  61. Hsu HY, Wen MH. Lipopolysaccharide-mediated reactive oxygen species and signal transduction in the regulation of interleukin-1 gene expression. *J Biol Chem* 2002;277:22131-22139.
  62. Kim BH, Lee IJ, Lee HY, Han SB, Hong JT, Ahn B, et al. Quercetin 3-O-beta-(2''-galloyl)-glucopyranoside inhibits endotoxin LPS-induced IL-6 expression and NF-kappa B activation in macrophages. *Cytokine* 2007;39:207-215.
  63. Kristiansen OP, Mandrup-Poulsen T. Interleukin-6 and diabetes: the good, the bad, or the indifferent? *Diabetes* 2005;54 (Suppl) 2:S114-124.
  64. Dubinski A, Zdrojewicz Z. The role of interleukin-6 in development and progression of atherosclerosis. *Pol Merkur Lekarski* 2007;22:291-294.
  65. Dowlati Y, Herrmann N, Swardfager W, Liu H, Sham L, Reim EK, et al. A meta-analysis of cytokines in major depression. *Biol Psychiatry* 2010;67:446-457.
  66. Swardfager W, Lanctot K, Rothenburg L, Wong A, Cappell J, Herrmann N. A meta-analysis of cytokines in Alzheimer's

- disease. *Biol Psychiatry* 2010;68:930-941.
67. Tackey E, Lipsky PE, Illei GG. Rationale for interleukin-6 blockade in systemic lupus erythematosus. *Lupus* 2004;13:339-343.
  68. Smith PC, Hobisch A, Lin DL, Culig Z, Keller ET. Interleukin-6 and prostate cancer progression. *Cytokine Growth Factor Rev* 2001;12:33-40.
  69. Nishimoto N. Interleukin-6 in rheumatoid arthritis. *Curr Opin Rheumatol* 2006;18:277-281.
  70. Hollman PC, Katan MB. Dietary flavonoids: intake, health effects and bioavailability. *Food Chem Toxicol* 1999;37:937-942.
  71. Vasquez-Martinez Y, Ohri RV, Kenyon V, Holman TR, Sepulveda-Boza S. Structure-activity relationship studies of flavonoids as potent inhibitors of human platelet 12-hLO, reticulocyte 15-hLO-1, and prostate epithelial 15-hLO-2. *Bioorg Med Chem* 2007;15:7408-7425.
  72. Chopra M, Fitzsimons PE, Strain JJ, Thurnham DI, Howard AN. Nonalcoholic red wine extract and quercetin inhibit LDL oxidation without affecting plasma antioxidant vitamin and carotenoid concentrations. *Clin Chem* 2000;46:1162-1170.
  73. Ansari MA, Abdul HM, Joshi G, Opii WO, Butterfield DA. Protective effect of quercetin in primary neurons against Abeta(1-42): relevance to Alzheimer's disease. *J Nutr Biochem* 2009;20:269-275.
  74. Ha SB, Smith AP, Howden R, Dietrich WM, Bugg S, O'Connell MJ, et al. Phytochelatase genes from *Arabidopsis* and the yeast *Schizosaccharomyces pombe*. *Plant Cell* 1999;11:1153-1164.
  75. Ong CS, Tran E, Nguyen TT, Ong CK, Lee SK, Lee JJ, et al. Quercetin-induced growth inhibition and cell death in nasopharyngeal carcinoma cells are associated with increase in Bad and hypophosphorylated retinoblastoma expressions. *Oncol Rep* 2004;11:727-733.
  76. Halliwell B, Gutteridge JM, Cross CE. Free radicals, antioxidants, and human disease: where are we now? *J Lab Clin Med* 1992;119:598-620.
  77. Cos P, Ying L, Calomme M, Hu JP, Cimanga K, van Poel B, et al. Structure-activity relationship and classification of flavonoids as inhibitors of xanthine oxidase and superoxide scavengers. *J Nat Prod* 1998;61:71-76.
  78. Ryu YB, Curtis-Long MJ, Lee JW, Kim JH, Kim JY, Kang KY, et al. Characteristic of neuraminidase inhibitory xanthenes from *Cudrania tricuspidata*. *Bioorg Med Chem* 2009;17:2744-2750.
  79. Hao Z, Duncan GS, Chang CC, Elia A, Fang M, Wakeham A, et al. Specific ablation of the apoptotic functions of cytochrome C reveals a differential requirement for cytochrome C and Apaf-1 in apoptosis. *Cell* 2005;121:579-591.
  80. Tan J, Wang B, Zhu L. Regulation of survivin and Bcl-2 in HepG2 cell apoptosis induced by quercetin. *Chem Biodivers* 2009;6:1101-1110.
  81. Locksley RM, Killeen N, Lenardo MJ. The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell* 2001;104:487-501.
  82. Koschny R, Ganten TM, Sykora J, Haas TL, Sprick MR, Kolb A, et al. TRAIL/bortezomib cotreatment is potentially hepatotoxic but induces cancer-specific apoptosis within a therapeutic window. *Hepatology* 2007;45:649-658.
  83. Siegelin MD, Reuss DE, Habel A, Rami A, von Deimling A. Quercetin promotes degradation of survivin and thereby enhances death-receptor-mediated apoptosis in glioma cells. *Neuro Oncol* 2009;11:122-131.
  84. Galluzzo P, Martini C, Bulzomi P, Leone S, Bolli A, Pallottini V, et al. Quercetin-induced apoptotic cascade in cancer cells: antioxidant versus estrogen receptor alpha-dependent mechanisms. *Mol Nutr Food Res* 2009;53:699-708.
  85. Subramanian M, Shaha C. Oestrogen modulates human macrophage apoptosis via differential signalling through oestrogen receptor-alpha and beta. *J Cell Mol Med* 2009;13:2317-2329.
  86. Koishi M, Hosokawa N, Sato M, Nakai A, Hirayoshi K, Hiraoka M, et al. Quercetin, an inhibitor of heat shock protein synthesis, inhibits the acquisition of thermotolerance in a human colon carcinoma cell line. *Jpn J Cancer Res* 1992;83:1216-1222.
  87. Elia G, Amici C, Rossi A, Santoro MG. Modulation of prostaglandin A1-induced thermotolerance by quercetin in human leukemic cells: role of heat shock protein 70. *Cancer Res* 1996;56:210-217.
  88. Hansen RK, Oesterreich S, Lemieux P, Sarge KD, Fuqua SA. Quercetin inhibits heat shock protein induction but not heat shock factor DNA-binding in human breast carcinoma cells. *Biochem Biophys Res Commun* 1997;239:851-856.
  89. Aalinkeel R, Bindukumar B, Reynolds JL, Sykes DE, Mahajan SD, Chadha KC, et al. The dietary bioflavonoid, quercetin, selectively induces apoptosis of prostate cancer cells by down-regulating the expression of heat shock protein 90. *Prostate* 2008;68:1773-1789.
  90. Hirai I, Okuno M, Katsuma R, Arita N, Tachibana M, Yamamoto Y. Characterisation of anti-*Staphylococcus aureus* activity of quercetin. *Int J Food Sci Technol* 2010;45:1250-1254.
  91. Lakshmi PTV, Radhika S, Annamalai A. Molecular docking analysis of phyto-ligands with multi drug resistant  $\beta$ -lactamases of *Staphylococcus aureus*. *Trends Bioinf* 2011;4:23-34.
  92. Tasdemir D, Lack G, Brun R, Ruedi P, Scapozza L, Perozzo R. Inhibition of plasmodium falciparum fatty acid biosynthesis: evaluation of FabG, FabZ, and FabI as drug targets for flavonoids. *J Med Chem* 2006;49:3345-3353.
  93. Afzelius L, Zamora I, Masimirembwa CM, Karlen A, Andersson TB, Mecucci S, et al. Conformer- and alignment-independent model for predicting structurally diverse competitive CYP2C9

- inhibitors. *J Med Chem* 2004;47:907-914.
94. Takemura H, Itoh T, Yamamoto K, Sakakibara H, Shimoi K. Selective inhibition of methoxyflavonoids on human CYP1B1 activity. *Bioorg Med Chem* 2010;18:6310-6315.
95. Androutsopoulos VP, Papakyriakou A, Vourloumis D, Spandidos DA. Comparative CYP1A1 and CYP1B1 substrate and inhibitor profile of dietary flavonoids. *Bioorg Med Chem* 2011;19:2842-2849.
96. Gao Y, Wang HY. Casein kinase 2 is activated and essential for Wnt/beta-catenin signaling. *J Biol Chem* 2006;281:18394-18400.
97. Sarno S, de Moliner E, Ruzzene M, Pagano MA, Battistutta R, Bain J, et al. Biochemical and three-dimensional-structural study of the specific inhibition of protein kinase CK2 by [5-oxo-5,6-dihydroindolo-(1,2-a)quinazolin-7-yl] acetic acid (IQA). *Biochem J* 2003;374:639-646.
98. Pauff JM, Hille R. Inhibition studies of bovine xanthine oxidase by luteolin, silibinin, quercetin, and curcumin. *J Nat Prod* 2009;72:725-731.
99. Wong IL, Chan KF, Tsang KH, Lam CY, Zhao Y, Chan TH, et al. Modulation of multi drug resistance protein 1 (MRP1/ABCC1)-mediated multi drug resistance by bivalent apigenin homodimers and their derivatives. *J Med Chem* 2009;52:5311-5322.
100. Ji XD, Melman N, Jacobson KA. Interactions of flavonoids and other phytochemicals with adenosine receptors. *J Med Chem* 1996;39:781-788.
101. Innocenti A, Beyza Ozturk Sarikaya S, Gulcin I, Supuran CT. Carbonic anhydrase inhibitors. Inhibition of mammalian isoforms I -XIV with a series of natural product polyphenols and phenolic acids. *Bioorg Med Chem* 2010;18:2159-2164.
102. Sarikaya SB, Gulcin I, Supuran CT. Carbonic anhydrase inhibitors: Inhibition of human erythrocyte isozymes I and II with a series of phenolic acids. *Chem Biol Drug Des* 2010;75:515-520.
103. Jung HA, Oh SH, Choi JS. Molecular docking studies of phlorotannins from *Eisenia bicyclis* with BACE1 inhibitory activity. *Bioorg Med Chem Lett* 2010;20:3211-3215.
104. Murthi KK, Dubay M, McClure C, Brizuela L, Boisclair MD, Worland PJ, et al. Structure-activity relationship studies of flavopiridol analogues. *Bioorg Med Chem Lett* 2000;10:1037-1041.
105. Zhang LM, Zhang YZ, Liu YQ, Gong ZH, Zhao YM, Li YF. CTN-986, a compound extracted from cottonseeds, increases cell proliferation in hippocampus *in vivo* and in cultured neural progenitor cells *in vitro*. *Eur J Pharmacol* 2009;607:110-113.
106. Walker EH, Pacold ME, Perisic O, Stephens L, Hawkins PT, Wymann MP, et al. Structural determinants of phosphoinositide 3-kinase inhibition by wortmannin, LY294002, quercetin, myricetin, and staurosporine. *Mol Cell* 2000;6:909-919.
107. McGovern SL, Shoichet BK. Kinase inhibitors: not just for kinases anymore. *J Med Chem* 2003;46:1478-1483.
108. Lo Piparo E, Scheib H, Frei N, Williamson G, Grigorov M, Chou CJ. Flavonoids for controlling starch digestion: structural requirements for inhibiting human alpha-amylase. *J Med Chem* 2008;51:3555-3561.
109. Holder S, Zemska M, Zhang C, Tabrizid M, Bremer R, Neidigh JW, et al. Characterization of a potent and selective small-molecule inhibitor of the PIM1 kinase. *Mol Cancer Ther* 2007;6:163-172.
110. DuPriest MT, Griffin BW, Kuzmich D, McNatt LG. Spiro[fluoreneisothiazolidin]one dioxides: new aldose reductase and L-hexonate dehydrogenase inhibitors. *J Med Chem* 1991;34:3229-3234.
111. Costantino L, Rastelli G, Gamberini MC, Vinson JA, Bose P, Iannone A, et al. 1-Benzopyran-4-one antioxidants as aldose reductase inhibitors. *J Med Chem* 1999;42:1881-1893.
112. Malleron JL, Roussel G, Gueremy G, Ponsinet G, Robin JL, Terlain B, et al. Penta- and hexadienoic acid derivatives: a novel series of 5-lipoxygenase inhibitors. *J Med Chem* 1990;33:2744-2749.
113. Deng S, Palu K, West BJ, Su CX, Zhou BN, Jensen JC. Lipoxygenase inhibitory constituents of the fruits of noni (*Morinda citrifolia*) collected in Tahiti. *J Nat Prod* 2007;70:859-862.
114. Bruneau P, Delvare C, Edwards MP, McMillan RM. Indazolinones, a new series of redox-active 5-lipoxygenase inhibitors with built-in selectivity/oral activity. *J Med Chem* 1991;34:1028-1036.
115. Cushman M, Nagarathnam D, Burg DL, Geahlen RL. Synthesis and protein-tyrosine kinase inhibitory activities of flavonoid analogues. *J Med Chem* 1991;34:798-806.
116. Huang H, Jia Q, Ma J, Qin G, Chen Y, Xi Y, et al. Discovering novel quercetin-3-O-amino acid-esters as a new class of Src tyrosine kinase inhibitors. *Eur J Med Chem* 2009;44:1982-1988.
117. Checa A, Ortiz AR, de Pascual-Teresa B, Gago F. Assessment of solvation effects on calculated binding affinity differences: trypsin inhibition by flavonoids as a model system for congeneric series. *J Med Chem* 1997;40:4136-4145.
118. McGovern SL, Caselli E, Grigorieff N, Shoichet BK. A common mechanism underlying promiscuous inhibitors from virtual and high-throughput screening. *J Med Chem* 2002;45:1712-1722.
119. Chen RM, Hu LH, An TY, Li J, Shen Q. Natural PTP1B inhibitors from *Broussonetia papyrifera*. *Bioorg Med Chem Lett* 2002;12:3387-3390.
120. Da Settimo F, Primofiore G, Da Settimo A, La Motta C, Simorini F, Novellino E, et al. Novel, highly potent aldose reductase inhibitors: cyano(2-oxo-2,3-dihydroindol-3-yl) acetic acid derivatives. *J Med Chem* 2003;46:1419-1428.

121. Costantino L, Rastelli G, Vescovini K, Cignarella G, Vianello P, Del Corso A, et al. Synthesis, activity, and molecular modeling of a new series of tricyclic pyridazinones as selective aldose reductase inhibitors. *J Med Chem* 1996;39:4396-4405.
122. Zapata-Torres G, Opazo F, Salgado C, Munoz JP, Krautwurst H, Mascayano C, et al. Effects of natural flavones and flavonols on the kinase activity of Cdk5. *J Nat Prod* 2004;67:416-420.
123. Nichols DE, Nichols CD. Serotonin receptors. *Chem Rev* 2008;108:1614-1641.
124. Gafner S, Dietz BM, McPhail KL, Scott IM, Glinski JA, Russell FE, et al. Alkaloids from *Eschscholzia californica* and their capacity to inhibit binding of [<sup>3</sup>H]8-Hydroxy-2-(di-N-propylamino)tetralin to 5-HT<sub>1A</sub> receptors *in vitro*. *J Nat Prod* 2006;69:432-435.
125. Takasawa R, Takahashi S, Saeki K, Sunaga S, Yoshimori A, Tanuma S. Structure-activity relationship of human GLO I inhibitory natural flavonoids and their growth inhibitory effects. *Bioorg Med Chem* 2008;16:3969-3975.
126. Schuster D, Nashev LG, Kirchmair J, Laggner C, Wolber G, Langer T, et al. Discovery of nonsteroidal 17β-hydroxysteroid dehydrogenase 1 inhibitors by pharmacophore-based screening of virtual compound libraries. *J Med Chem* 2008;51:4188-4199.
127. Freitas RF, Prokopczyk IM, Zottis A, Oliva G, Andricopulo AD, Trevisan MT, et al. Discovery of novel trypanosoma *cruxi* glyceraldehyde-3-phosphate dehydrogenase inhibitors. *Bioorg Med Chem* 2009;17:2476-2482.
128. Lee C, Lee JM, Lee NR, Kim DE, Jeong YJ, Chong Y. Investigation of the pharmacophore space of severe acute respiratory syndrome coronavirus (SARS-CoV) NTPase/helicase by dihydroxychromone derivatives. *Bioorg Med Chem Lett* 2009;19:4538-4541.
129. Chimenti F, Cottiglia F, Bonsignore L, Casu L, Casu M, Floris C, et al. Quercetin as the active principle of *Hypericum hircinum* exerts a selective inhibitory activity against MAO-A: extraction, biological analysis, and computational study. *J Nat Prod* 2006;69:945-949.
130. Chimenti F, Fioravanti R, Bolasco A, Chimenti P, Secci D, Rossi F, et al. A new series of flavones, thioflavones, and flavanones as selective monoamine oxidase-B inhibitors. *Bioorg Med Chem* 2010;18:1273-1279.
131. Ko S, Lee MK, Shin D, Park H. Structure-based virtual screening approach to the discovery of novel inhibitors of factor-inhibiting HIF-1: identification of new chelating groups for the active-site ferrous ion. *Bioorg Med Chem* 2009;17:7769-7774.
132. Muftuoglu Y, Mustata G. Pharmacophore modeling strategies for the development of novel nonsteroidal inhibitors of human aromatase (CYP19). *Bioorg Med Chem Lett* 2010;20:3050-3064.
133. Borbely G, Szabadkai I, Horvath Z, Marko P, Varga Z, Breza N, et al. Small-molecule inhibitors of NADPH oxidase 4. *J Med Chem* 2010;53:6758-6762.
134. Torres-Piedra M, Figueroa M, Hernandez-Abreu O, Ibarra-Barajas M, Navarrete-Vazquez G, Estrada-Soto S. Vasorelaxant effect of flavonoids through calmodulin inhibition: *Ex vivo*, *in vitro*, and *in silico* approaches. *Bioorg Med Chem* 2011;19:542-546.
135. Pick A, Muller H, Mayer R, Haenisch B, Pajeva IK, Weigt M, et al. Structure-activity relationships of flavonoids as inhibitors of breast cancer resistance protein (BCRP). *Bioorg Med Chem* 2011;19:2090-2102.
136. Goetter M, Schattel V, Koch P, Merfort I, Laufer S. Biological evaluation and structural determinants of p38α mitogen-activated-protein kinase and c-Jun-N-terminal kinase 3 inhibition by flavonoids. *Chembiochem* 2010;11:2579-2588.
137. Hrazdina G, Zobel AM, Hoch HC. Biochemical, immunological, and immunocytochemical evidence for the association of chalcone synthase with endoplasmic reticulum membranes. *Proc Natl Acad Sci U S A* 1987;84:8966-8970.
138. Koukol J, Conn EE. The metabolism of aromatic compounds in higher plants. IV. Purification and properties of the phenylalanine deaminase of *Hordeum vulgare*. *J Biol Chem* 1961;236:2692-2698.
139. Nakayama T. Enzymology of aurone biosynthesis. *J Biosci Bioeng* 2002;94:487-491.
140. Bednar RA, Hadcock JR. Purification and characterization of chalcone isomerase from soybeans. *J Biol Chem* 1988;263:9582-9588.
141. Turnbull JJ, Nakajima J, Welford RW, Yamazaki M, Saito K, Schofield CJ. Mechanistic studies on three 2-oxoglutarate-dependent oxygenases of flavonoid biosynthesis: anthocyanidin synthase, flavonol synthase, and flavanone 3β-hydroxylase. *J Biol Chem* 2004;279:1206-1216.
142. Owens DK, Alerding AB, Crosby KC, Bandara AB, Westwood JH, Winkel BS. Functional analysis of a predicted flavonol synthase gene family in *Arabidopsis*. *Plant Physiol* 2008;147:1046-1061.
143. Li Y, Baldauf S, Lim EK, Bowles DJ. Phylogenetic analysis of the UDP-glycosyltransferase multigene family of *Arabidopsis thaliana*. *J Biol Chem* 2001;276:4338-4343.
144. Lakshmi P. Medicinal plant informatics—an insight. *Pharm Commun* 2012;2:5-11.

(Received December 5, 2012)  
Edited by WANG Wei-xia